

Empirical support for an involvement of the mesostriatal dopamine system in human fear extinction

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Exposure therapy for anxiety disorders relies on the principle of confronting a patient with the triggers of his fears, allowing him to make the unexpected safety experience that his fears are unfounded and resulting in the extinction of fear responses. In the laboratory, fear extinction is modeled by repeatedly presenting a fear-conditioned stimulus (CS) in the absence of the aversive unconditioned stimulus (UCS) to which it had previously been associated. Classical associative learning theory considers extinction to be driven by an aversive prediction error signal that expresses the expectation violation when not receiving an expected UCS and establishes a prediction of CS non-occurrence. Insufficiencies of this account in explaining various extinction-related phenomena could be resolved by assuming that extinction is an opponent appetitive-like learning process that would be mediated by the mesostriatal dopamine (DA) system. In accordance with this idea, we find that a functional polymorphism in the DA transporter gene, *DAT1*, which is predominantly expressed in the striatum, significantly affects extinction learning rates. Carriers of the 9-repeat (9R) allele, thought to confer enhanced phasic DA release, had higher learning rates. Further, functional magnetic resonance imaging revealed stronger hemodynamic appetitive prediction error signals in the ventral striatum in 9R carriers. Our results provide a first hint that extinction learning might indeed be conceptualized as an appetitive-like learning process and suggest DA as a new candidate neurotransmitter for human fear extinction. They open up perspectives for neurobiological therapy augmentation.

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Introduction

Classical associative learning theory explains fear conditioning by an aversive prediction error signal δ_{av} generated when an initially non-predictive conditioned stimulus (CS) is unexpectedly followed by an unconditioned stimulus (UCS). This establishes a UCS prediction, or aversive value V_{av} , for the CS that grows over successive pairings.^{1,2} If at some point the UCS is unexpectedly omitted, this generates a negative (oppositely signed) aversive prediction error that will reduce V_{av} . The latter mechanism is thought to underlie the extinction of conditioned fear responses by repeated unpaired CS presentations.¹ It is the theoretical basis of exposure therapy where a patient is repeatedly confronted with the trigger of his fears (the CS, for example, in an agoraphobic, an open space) and makes the experience that the predicted outcome (the UCS) is absent or less disastrous than expected (for example, he does not collapse).³

A frequent finding is that conditioned fear responses can return after successful extinction, indicating that the CS–UCS association (V_{av}) is not simply unlearned or erased during extinction but rather complemented by a competing inhibitory CS–noUCS association that may, or may not, dominate the CS–UCS association at future CS presentations.^{4–6} Moreover, there is compelling evidence for a partial segregation in

the neural systems subserving conditioning and extinction.^{7–12} The above simple account of extinction, as being solely mediated by the same learning system that also mediates conditioning, cannot accommodate these observations.

Alternatively, the omission of an expected aversive UCS could be conceptualized as an appetitive-like or reward prediction error δ_{app} and the consequential reduction of the UCS prediction V_{av} during extinction as generation of a reward-like safety prediction V_{app} . From this perspective, part of a solution for the above problem could be that extinction is driven by an opponent appetitive learning system. Reward learning has been strongly linked with the mesostriatal dopamine (DA) system.^{13–15} There is evidence that δ_{app} is signaled by a phasic increase in the firing of DAergic neurons that originate in the ventral tegmental area and substantia nigra and project to the ventral striatum (VS).¹⁵ It has therefore been hypothesized that VS DA release is involved in putative δ_{app} signaling during fear extinction as well.¹⁶ One rodent study that showed that DA signaling via D1 receptors is necessary for extinction¹⁷ further supports the potential link between fear extinction and the reward system. One goal of this study was therefore to test whether the VS encodes appetitive-like prediction error signals during extinction in humans.

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Extracellular DA levels in the striatum are prominently regulated by neuronal DA reuptake via the DA transporter (DAT).¹⁸ The human transporter gene *DAT1* features a frequent and functional variable number of tandem repeat (VNTR) polymorphism in a region that encodes the 3' untranslated region.¹⁹ The 40-bp VNTR element is mainly repeated either 9 or 10 times, with the 9-repeat (9R) form most likely reducing DAT expression^{20–24} (but see Dyck *et al.*²⁵) and thus presumably enhancing extrasynaptic striatal DA levels, in particular, during phasic DA release.¹⁸ Hence, if the mesostriatal DA system is involved in extinction in the fashion outlined above, one would expect the *DAT1* 9R allele to be associated with relatively enhanced extinction learning as well as with enhanced neural δ_{app} signaling in the VS. To examine this hypothesis, we conducted the experiment in a sample of normal healthy volunteers that were preselected on the basis of their *DAT1* genotype. In particular, we compared 9R carriers (genotypes 9/9 and 9/10) with non-9R carriers (genotypes 10/10). The grouping was chosen because of the relative scarcity of the 9R allele and to be in keeping with previous DA binding neuroimaging studies.^{20,21}

We also examined effects of interindividual variation in COMT (catechol-o-methyltransferase) function. In contrast to DAT, COMT is most strongly expressed in the prefrontal cortex (PFC)²⁶ where it degrades released DA, thereby regulating extracellular PFC DA levels.²⁷ The human *COMT* gene contains a functional single-nucleotide polymorphism that codes for a Val to Met change at position 158,²⁸ the Met variant of the protein being less active²⁹ and associated with higher prefrontal baseline synaptic DA.²⁷ Prefrontal DA appears to have a role in extinction in rats,³⁰ and a recent human study had suggested impaired extinction in *COMT* Met/Met carriers.³¹ Hence, including *COMT* genotype in the design allowed us to also explore potential contributions of extrastriatal DA to human fear extinction. As the Val and Met alleles are codominant,²⁷ participants were preselected in a way to obtain three similarly sized groups of Val/Val, Val/Met and Met/Met carriers. This resulted in a 2 by 3 (*DAT1* by *COMT*) factorial between-subject design.

Participants and methods

Participants. A total of 69 healthy male right-handed Caucasian participants with *DAT1* and *COMT* genotypes 9R-Val/Val ($n=13$), 9R-Val/Met ($n=10$), 9R-Met/Met ($n=10$), non-9R-Val/Val ($n=14$), non-9R-Val/Met ($n=12$) and non-9R-Met/Met ($n=10$) were examined. As participants were drawn from a bigger population to achieve a stratified and matched population, calculations of Hardy–Weinberg equilibrium (HWE) were only appropriate for the basic population ($n=450$). Genotype distributions were as follows *DAT1* 9/9: $n=28$; 9/10: $n=148$; 10/10: $n=252$ (HWE $\chi^2=0.97$), *COMT* Val/Val: $n=80$; Val/Met: $n=237$; Met/Met: $n=118$ (HWE $\chi^2=4.18$), and thus above the HWE threshold of $P=0.01$. Details on participant selection, sample characteristics and genotyping can be found in the Supplementary Methods and Supplementary Table 1. A different analysis of an overlapping sample has been reported before.³²

Experiment. Participants performed a simple uninstructed fear conditioning, extinction and reacquisition task, which has been described in detail elsewhere.³² Briefly, participants were first habituated to the CSs, the task and the scanner noise by presenting each CS four times before the actual experiment. In the subsequent acquisition phase (Acq), participants saw 18 pseudorandomized 5-s presentations of each of two geometric symbols (a circle, a triangle), one of which (CS+) was paired in 80% of cases with a painful electric stimulus (UCS) applied to the back of the right hand. The other symbol served as a control stimulus (CS–) for non-associative effects and was never paired with the UCS. Assignment of symbols as CS+ or CS– was counterbalanced across participants. In the extinction phase (Ext), both stimuli were again presented 18 times each, but in the absence of the UCS. The subsequent reacquisition phase (RAcq) was identical to the acquisition phase. The intertrial interval was jittered between 9–14 s, with an average of 11.5 s. We intermittently asked participants to give explicit ratings of their CS-evoked stress/fear/tension (at baseline, that is, after the habituation phase, and every 12 trials (six CS+ and six CS– trials) thereafter, resulting in three CS+ and three CS– ratings per phase). Throughout the experiment, the participants had to perform a speeded decision task on the geometric symbols (see Supplementary Methods). UCS intensity was individually adjusted before the experiment to achieve maximum tolerable pain.

Data acquisition and preprocessing. Acquisition and preprocessing of skin conductance and functional magnetic resonance imaging (fMRI) data followed standard procedures (see Supplementary Methods).

Data analysis

Fear ratings. All ratings were normalized by subtracting the baseline ratings given at the onset of the experiment (after habituation) such that positive ratings reflected an increase in fear relative to baseline and negative ratings a relative decrease in fear. In four participants, ratings were not acquired or lost because of technical problems. The remaining sample size for analysis of ratings was $n=65$.

Rescorla–Wagner model. The Rescorla–Wagner (RW) model is a simple and established associative learning model that formalizes the laws of learning outlined in the introduction. If learning is about enabling an organism to predict relevant future events from present stimuli, then classical conditioning should result in the organism being able to predict a UCS from the presentation of the CS. That is, the CS should activate a UCS expectation (or CS–UCS association or UCS prediction) that reflects the probability and magnitude of the UCS. This ‘associative strength’ or affective value of the CS is expressed in the V term of the RW equation below and will increase over the course of conditioning. It is thought to determine conditioned responding. Every violation of this expectation (for example, because a UCS occurs unexpectedly, as initially in the beginning of conditioning when the UCS prediction is still 0, or because an expected UCS does not occur, as in initial extinction) must result in an adjustment of the

expectation, that is, learning. Therefore, the update of V in the RW model is directly proportional to a prediction error term δ that reflects the difference between actual and predicted reinforcement (that is, the expectation violation).¹ This class of models is known to be relevant for learning about punishments and rewards, and has been successfully used to predict learning-related neural activation.^{33–39} Specifically, the RW model updates V at trial $t+1$ according to

$$V_{t+1} = V_t + \alpha(R - V_t),$$

with α being the constant learning rate (0–1), R being a fixed value assigned to the reinforcement/UCS and $(R - V_t)$ corresponding to the prediction error δ that is generated at the time of reinforcement.

We used this rule to model how participants change their aversive CS+ and CS– values V_{av} depending on their associated reinforcement pattern. A flow chart showing the separate analysis steps is provided in the Supplementary Methods. We first range corrected each participants' CS+ and CS– fear rating data (see Supplementary Figure 1a for sample average) according to

$$x_{i,corr,CS+orCS-} = (x_{i,CS+orCS-} - \min) / (\max - \min),$$

with $x_{i,CS+orCS-}$ ($i = 1, \dots, 10$) being the successive (CS+ or CS–) fear ratings, \min being the sample-wide minimum of all ratings (–58) and \max being the sample-wide maximum of all ratings (100). This resulted in individual rating time courses in which ratings ranged from 0 to 1 but retained interindividual differences in how participants used the rating scale. The (CS+ and CS–) baseline ratings $x_{1,CS+orCS-}$ (after habituation and before conditioning), which were 0 by definition in each participant (see above), became 0.36 ($= x_{1,corr,CS+orCS-}$).

After complete learning, the aversiveness R of the UCS is reflected in the aversiveness of the CS+, that is, in the last CS+ fear rating after acquisition ($x_{4,corr,CS+}$). With a partial reinforcement schedule of 80%, a participant's R in paired CS+ trials can thus be approximated as $x_{4,corr,CS+} / 0.8$.⁴⁰ R in CS– trials (0% reinforcement) was set at each participant's $x_{4,corr,CS-}$ rating. The same value of R was used for unpaired CS+ trials. See the Supplementary Methods for a more detailed explanation. As mentioned above, UCS (pain) intensity in this experiment was individually calibrated to each participant's subjectively tolerable maximum to eliminate interindividual differences in UCS processing. Concordantly, the calculated individual R -values for paired CS+ trials were not affected by genotype (*DAT1*: $F(1,64) = 1.24$, $P = 0.298$; *COMT*: $F(2,64) = 0.74$, $P = 0.438$; and interaction: $F(2,64) = 0.06$, $P = 0.978$). We nevertheless used individual R -values to factor out any potential interindividual differences in learning that might in fact merely result from differences in UCS processing.

$V_{av,CS+}$ and $V_{av,CS-}$ were modeled separately and set at 0.36 ($= x_{1,corr,CS+orCS-}$; see above) before learning. On the basis of the idea of dissociable neural systems for fear acquisition and extinction (and possibly reacquisition as well), we used three free parameters α_{Acq} , α_{Ext} and α_{RAcq} (one for each of the three experimental phases), which were adjusted to minimize the distance between the change in $V_{av,CS+}$ and

$V_{av,CS-}$ and the change in fear ratings $x_{corr,CS+}$ and $x_{corr,CS-}$ using a least-square approach. We did not use CS+ > CS– difference scores, and $V_{av,CS+}$ and $V_{av,CS-}$ were estimated at the same time within the same model. $V_{av,CS+}$ and $V_{av,CS-}$ time courses averaged across the entire sample are shown in Figure 1a. Average least sums of squares were similar between genotypes (*DAT1*: $F(1,64) = 0.55$, $P = 0.461$; *COMT*: $F(2,64) = 0.13$, $P = 0.878$; and interaction: $F(2,64) = 0.49$, $P = 0.613$). Model fits were substantially worse when assuming one identical learning rate across all three phases of the experiment (data not shown).

Note that the original RW formula assumes different learning rates for reinforced and non-reinforced CS trials, but this differentiation is not critical for most of the model's predictions¹ and has not been made in neuroimaging studies.^{33–39} Allowing different learning rates for reinforced and non-reinforced CS trials yielded worse fits (data not shown).

Imaging data. Analysis of imaging data was restricted to those 65 participants from which fear ratings were also available. In the comparison of *DAT1* 9R with non-9R carriers, group sizes were $n = 32$ and $n = 33$, respectively. To prepare the analysis, we used the sample-averaged learning rates $\alpha_{Acq} = 0.16$, $\alpha_{Ext} = 0.21$ and $\alpha_{RAcq} = 0.19$ to derive individual trial-by-trial V_{av} and δ_{av} estimates from the above modeling of the rating data, separately for acquisition, extinction and reacquisition. Averaging of learning rates was necessary to reduce noise in the data that resulted from a limited number of data points for fitting (10 ratings), and thus to obtain robust estimates. An exemplary individual δ_{av} time course is shown in Figure 1b. We emphasize that our estimated average learning rates are comparable to those used in previous neuroimaging studies.³⁴

This information was fed into the imaging data analysis that used a standard approach for fMRI, involving a general linear model (multiple regression) at the single-subject level and a random-effects analysis at the group level within the SPM5 software (www.fil.ion.ucl.ac.uk/spm).⁴¹ For each participant, regressors were defined that modeled the time course of the experimental events. Onsets of CSs, irrespective of whether they were a CS+ or a CS–, were modeled as categorical 'events', that is, one series of delta functions. This regressor was parametrically modulated in a trial-by-trial fashion by the individual's sequence of V_{av} estimates. Another categorical event regressor modeled CS (both CS+ and CS–) offsets and was parametrically modulated by the individual's sequence of δ_{av} estimates. This was done for acquisition, extinction and reacquisition separately. CS+ and CS– trials were not differentiated in this analysis, as the concept of predictions and prediction errors is a purely quantitative one that does not make qualitative distinctions between types of stimuli. We therefore assumed identical neural substrates for predictions and prediction errors, whether associated with a CS+ or a CS–. Additional categorical regressors modeled UCSs (events), key presses (events), and the occurrence of fear ratings (14-s box car). Each regressor was convolved with a canonical hemodynamic response function. Using these regressors in a general linear model of brain activation at each voxel yields parameter estimates of the contribution of

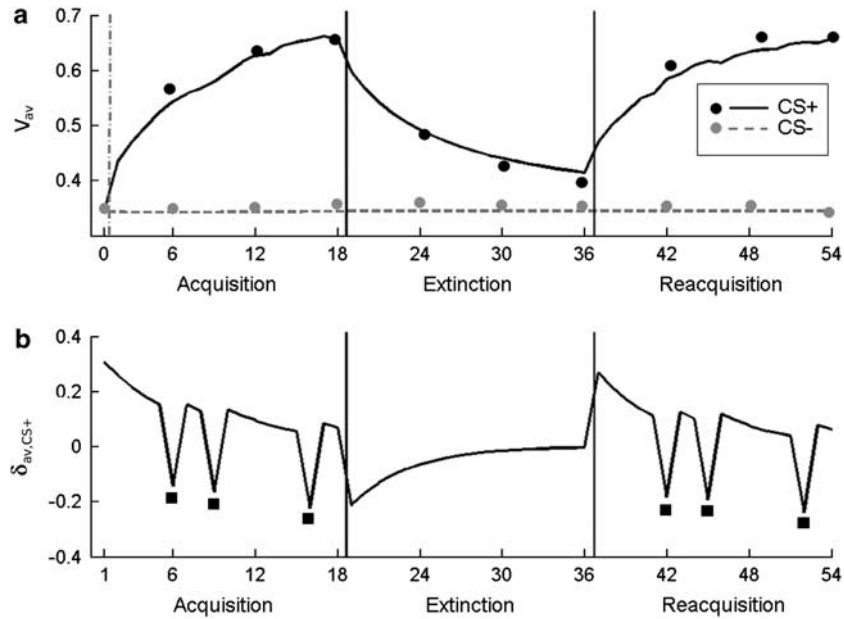


Figure 1 Formal modeling of fear ratings. (a) Lines show the sample average of the modeled trial-by-trial estimates of aversive conditioned stimulus values ($V_{av,CS+}$ and $V_{av,CS-}$). Dots show sample-average range-corrected fear ratings (made after every sixth CS+ and CS- trial). 0: baseline rating after habituation. (b) An example of a resulting individual time course of trial-by-trial aversive prediction error (δ_{av}) estimates associated with the CS+. Black squares mark unpaired CS+ trials during acquisition and reacquisition. CS+ trials during extinction were all unpaired. Prediction errors associated with the CS- were always 0 and are not shown for simplicity. x axis: CS+ or CS- trials.

each regressor to the fMRI signal measured in each voxel. The convolved regressors of interest ($V_{av,Acq}$, $\delta_{av,Acq}$, $V_{av,Ext}$, $\delta_{av,Ext}$, $V_{av,RAcq}$ and $\delta_{av,RAcq}$) were sufficiently decorrelated from each other and from the UCS regressor to allow for robust estimation (Pearson's R s for the correlations between $V_{av,Acq}$ and $\delta_{av,Acq}$: 0.01; $V_{av,Ext}$ and $\delta_{av,Ext}$: -0.21; $V_{av,RAcq}$ and $\delta_{av,RAcq}$: 0.01; $V_{av,Acq}$ and UCS: -0.12; $\delta_{av,Acq}$ and UCS: -0.43; $V_{av,Ext}$ and UCS: 0; $\delta_{av,Ext}$ and UCS: 0; $V_{av,RAcq}$ and UCS: -0.12; and $\delta_{av,RAcq}$ and UCS: -0.41). Note further that the use of a comparatively high reinforcement ratio of 80% during acquisition assured a high initial amount of prediction error signaling during extinction in combination with a relatively steep approach toward zero signal (see Figure 1b). This characteristic time course was sufficiently different in shape from the constant categorical CS offset regressor, of which it was a parametric modulator, for it to be able to explain additional variance. At the same time, choosing a partial reinforcement ratio also seemed preferable to a 100% schedule, because the latter would have generated a very steep approach toward zero, which would leave little room for modulation by individual-difference factors.

For the voxel-wise random-effects group analyses, the subject-specific parameter estimate images from the parametric δ_{av} and V_{av} regressors were spatially smoothed (FWHM 10 mm, resulting in total smoothing with an 11-mm kernel) to account for interindividual anatomical and functional variance, and to fulfil the requirements for later correction for multiple comparisons following Gaussian random field theory (see below). *DAT1* and *COMT* genotype effects were assessed using SPM's 'full factorial' model, which allows for correcting for possible non-sphericity of the error term (here unequal between-group variance). Separate models were

calculated for each effect of interest (for example, $V_{av,Acq}$). A design matrix included six regressors, one for each possible genotype combination. The significance of linear combinations of the regressors (for example, 1 1 1 -1 -1 -1 when asking which voxels show larger effects for *DAT1* 9R than for non-9R carriers in a given parameter estimate image) was assessed using one-tailed *t*-tests.

Correction for multiple comparisons following Gaussian random field theory (family-wise error method) at $P < 0.05$ was limited to the VS regions of interest (ROIs). Left- and right-sided ROIs were conservatively defined as spheres of 6-mm radius around coordinates.

The values around coordinates -27, 3, -9 and 27, -9, -9, respectively, which were taken from the first fMRI study that investigated neural reward prediction error coding using a formal associative learning model.³⁴ Where no anatomical hypothesis existed (exploratory analyses across the entire scan volume), an uncorrected threshold of $P < 0.001$ was used.

Results and discussion

Behavioral data. In the RW model of associative learning (see Participants and methods), the prediction error δ_{av} is weighted by a constant α , the learning rate, that determines how much a deviation from prediction at trial t is taken into account when formulating the prediction for the next trial $t+1$. A high learning rate signifies rapid prediction adjustment and thus quick learning. If extinction relies (in part) on a different learning system than conditioning, it may well show a different dynamic. We thus assessed learning

separately for the three phases of the experiment, allowing separate learning rates α_{Acq} , α_{Ext} and α_{RAcq} . These were treated as free parameters, which we optimized so that the individual V_{av} time courses fit changes in individual skin conductance responses (SCRs) or fear ratings across the entire experiment (see Supplementary Figure 1 for sample-average SCR and rating time courses).

An attempt to model SCRs failed because of excessive noise in the data. Modeling of fear ratings (Figure 1), followed by three separate two-way analyses of variance (one per phase, each with between-subject factors *DAT1* and *COMT*, and learning rate as the dependent variable) revealed significantly higher learning rates in *DAT1* 9R carriers compared with non-9R carriers during extinction (α_{Ext} : *DAT1* main effect $F(1,65) = 4.57$, $P = 0.037$) but not during acquisition (α_{Acq} : $F(1,65) = 0.13$, $P = 0.725$) or reacquisition (α_{RAcq} : $F(1,65) = 3.27$, $P = 0.075$; Figure 2). This suggests *DAT1* 9R carriers have a more sensitive extinction learning system and is consistent with the idea that striatal DA might positively contribute to extinction learning.

Learning rates were unaffected by *COMT* genotype (all $P > 0.267$), but there was a significant *DAT1* \times *COMT* interaction effect on learning rates in the reacquisition phase (α_{RAcq} : $F(2,65) = 4.48$, $P = 0.015$). A *post hoc* *t*-test showed that *DAT1* 9R carriers had significantly higher learning rates than non-9R carriers only in the *COMT* Val/Met group (9R: 0.42 ± 0.36 (mean \pm s.d.) versus non-9R: 0.09 ± 0.13 ; $t(22) = 2.55$, $P = 0.025$, two tailed; Supplementary Figure 2a). This incidental finding will be discussed further below. A standard, non-computational analysis of SCR and rating responses for genotype effects yielded no significant results.

Taken together, behavioral analysis suggested a significant contribution to extinction of *DAT1* in the predicted direction but found no evidence for an involvement of *COMT*. We note that the cited *COMT* study by Lonsdorf et al.³¹ also reported a negative result for SCR and that their *COMT* effect on startle potentiation might as well be explained by a modulation of fear memory consolidation rather than extinction learning itself. The current data from human subjects therefore do not lend

strong support to the idea³⁰ that prefrontal DA function is important for extinction.

Imaging data: entire sample. In the entire sample, exploratory analysis of extinction data for δ_{av} signals (the aversive prediction error characterized by phasic relative decreases in activation when a UCS is unexpectedly omitted, compare with Figure 1b) yielded no significant results. The putative appetitive-like prediction error δ_{app} is the mathematical inverse of δ_{av} and characterized by relative phasic increases at CS omission. Activity conforming to δ_{app} was found in, among others, left, and less so right, anterior insula, bilateral anterior lateral PFC, right ventrolateral PFC/lateral orbitofrontal cortex and right VS (ventral putamen and/or nucleus accumbens; $x, y, z = 14, 8, -6$; z -score = 3.22; $P < 0.001$ uncorrected; see Figure 3a; Supplementary Table 2), areas previously associated with δ_{app} coding in reward studies⁴² and with phasic activations to UCS omission during fear conditioning.⁴³ The VS activation was, however, not located within our conservatively defined (left or right) VS ROIs (see participants and methods for definition). As for δ_{av} , there were no suprathreshold V_{av} signals (the aversive CS value that decreases across extinction, compare with Figure 1a). By contrast, the putative reward-like safety value of the CS, V_{app} , which is the mathematical inverse of V_{av} and accordingly builds up across extinction, was found to be encoded in left orbitofrontal cortex/ventrolateral PFC, dorsomedial and lateral PFC, temporal cortex, left caudate, cerebellum and others (Supplementary Table 2). These observations might suggest that extinction is indeed primarily driven by reward-like safety learning. Results for the acquisition and reacquisition phases are reported in Supplementary Table 2.

Imaging data: genetic analysis. In the genetic analysis, we focused on the comparison of *DAT1* 9R with non-9R groups, as *COMT* genotype had not affected extinction learning rates in the behavioral analysis. If *DAT1* 9R carriers weight prediction errors during extinction more strongly (have higher learning rates), then they can be expected to show larger neural δ_{av} and/or δ_{app} signals during this phase. As predicted, 9R carriers had significantly stronger signal increases to UCS omission than non-9R carriers, corresponding to stronger δ_{app} coding, in our left VS ROI ($x, y, z = -32, 8, -6$; z -score = 2.99; $P = 0.03$ corrected). The activation was located in the putamen and extended into the anterior insula (Figure 3b). This result supports our conclusion from the behavioral analysis that *DAT1* 9R carriers have a more sensitive extinction learning system and is evidence for an involvement of the mesostriatal DA system in extinction. Further group differences, all going in the same direction, were observed in left anterior cingulate sulcus and other areas (exploratory analysis at $P < 0.001$ uncorrected; Supplementary Table 3). Group comparisons of CS value encoding (V_{av} or V_{app}) surprisingly showed stronger V_{app} signals in the non-9R compared with the 9R group in two frontal areas ($P < 0.001$ uncorrected; Supplementary Table 3). We stress the exploratory nature of the latter comparisons and the corresponding likelihood of false-positive results.

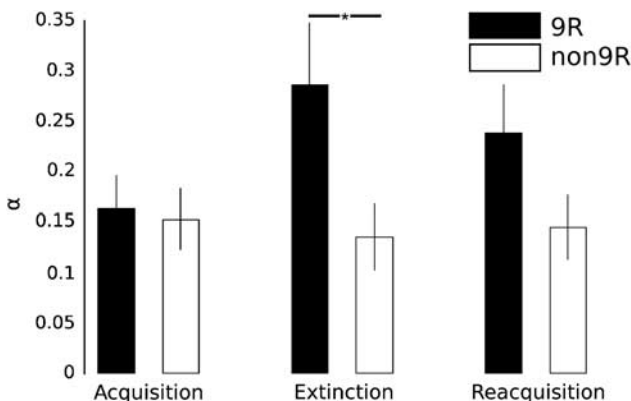


Figure 2 *DAT1* genotype affects learning rates during extinction. Formal modeling of fear rating data showed that 9-repeat (9R) carriers have significantly higher learning rates during extinction than non-9R carriers. Error bars: s.e.m. * $P < 0.05$ (*F* test).

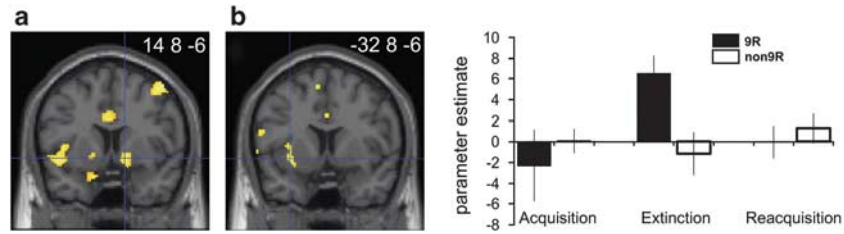


Figure 3 Ventral striatal (VS) prediction error signaling during extinction. (a) Appetitive prediction error (δ_{app}) signal in right VS in the entire sample. (b) Stronger δ_{app} signal in *DAT1* 9-repeat (9R) compared with non-9R carriers in left VS. Activations superimposed on a canonical structural image. Display threshold: $P < 0.01$ uncorrected. Bar graphs show average δ_{app} parameter estimates in extinction, as well as, for comparison, in acquisition and reacquisition in the peak voxel indicated by the hair cross. Error bars: s.e.m.

The unexpected behavioral finding of higher reacquisition learning rates in 9R compared with non-9R carriers specifically in the *COMT* Val/Met group (see above) was reflected in relatively higher δ_{app} signals in the right VS ROI in 9R-Val/Met participants ($x, y, z = 22, -8, -8$; z -score = 2.62; $P = 0.043$ corrected; Supplementary Figure 2b). The trial-by-trial variance that is captured by the parametric δ_{app} regressor during reacquisition mainly stems from the δ_{app} increases to the three unexpected UCS omissions (compare with the inverse of the curve shown in Figure 1b, reacquisition part). A ventral striatal *DAT1* effect on this type of signal suggests that the safety or appetitive-like mechanisms, which we propose are activated during extinction, carry over to the reacquisition situation. The observation that the *DAT1* effect is limited to *COMT* Val/Met carriers might speculatively be attributed to a situationally dependent influence of prefrontal DA on striatal DA function.^{44,45} In this context, it is worth noting that epistatic *DAT1* by *COMT* interactions in VS reward signaling have been observed before.⁴⁶ The exact nature of the effect must remain open until further investigation.

Supplementary Table 4 reports a genetic analysis of the VS ROIs from all experimental phases and contrasts for *DAT1*, *COMT* and *DAT1* by *COMT* effects.

Conclusion. To summarize, our combined behavioral and imaging data hint toward signaling of UCS omission during extinction by phasic DA release in the VS, in analogy to the role of the mesostriatal DA system in reward learning.¹⁵ They support a conceptualization of extinction as a reward-like safety learning process.¹⁶ More globally, such a conceptualization can be integrated within a perspective of opponent aversive and appetitive systems.^{47,48}

Several limitations of the current study should be noted. First, there are still controversies with respect to the actual impact of *DAT1* genotype on *in vivo* DAT function and striatal DA clearance (see introduction), and the prevailing hypothesis of stronger phasic DA signals in 9R carriers still has to be substantiated. Second, our approach is correlative, that is, we did not experimentally manipulate striatal DA signaling. Pharmacological manipulations in rodents have shown generally higher levels of conditioned freezing during extinction with DA antagonists given systemically^{16,49,50} or directly in the amygdala⁵¹ or nucleus accumbens.¹⁶ A systemic DA agonist reduced conditioned freezing during extinction.⁵⁰ Although these results could be taken to support a facilitatory role for DA in extinction learning, it should be noted that DAergic

manipulations also affect locomotion and baseline freezing, and it is therefore difficult to rule out that the enhanced conditioned freezing observed under DA antagonists might be explained by their motor side effects.^{16,49–51} Further, these studies have analyzed average freezing across the entire extinction session, a measure that might also be confounded by potential general fear-potentiating drug effects. It might therefore be advantageous to instead focus on rates of decay of freezing as a primary outcome measure for extinction in animals. In humans, where conditioned responding is normally not read out from motor behavior, pharmacological experiments might suffer from other confounds such as potential drug effects on arousal.³⁶ Such experiments will therefore have to carefully control for non-specific effects but might nevertheless be a valuable source of evidence. Third, showing a contribution of the mesostriatal DA system does not necessarily prove that extinction is appetitive, as the DA system is not exclusively involved in reward learning. Here, a direct demonstration of the appetitive nature of extinguished CSs would be beneficial. Another potentially interesting approach would be a direct formal comparison of extinction with a reward-learning task within the same sample. Fourth, our data do not exclude that non-appetitive, that is, aversive learning mechanisms also contribute to extinction. Fifth, although conditioning and extinction are generally considered relevant paradigms for the study of pathological anxiety and its therapy,³ it must be stressed that they cannot provide explanations for every aspect of anxiety and we can, in particular, not exclude that other mechanisms have a role in therapeutic fear relief. Sixth, our sample included mainly university students and exclusively comprised males. The latter selection criterion was introduced following reports of considerable gender and cycle effects on extinction⁵² and was intended to reduce variance, thus allowing us to limit sample size. Reproduction in other samples is therefore required. Seventh, where we reported results from exploratory whole-brain analyses, these are not corrected for multiple comparisons, as emphasized earlier. Again, reproduction will be paramount.

It is worth noting that we did not find evidence for a role for DA in fear acquisition, in line with one genetic study examining *COMT* genotype effects on conditioning.³¹ By contrast, a recent pharmacological fMRI study³⁶ had reported enhanced aversive prediction errors δ_{av} in the caudate nucleus and the substantia nigra/ventral tegmental area during conditioning in participants under amphetamine compared with participants

under haloperidol. As amphetamine participants also reported to feel less tired, drowsy and slowed, these results might also reflect a general attentional effect. It should, however, also be noted that the absence of *DAT1* or *COMT* effects on our and other peoples' measures of conditioning does not exclude a contribution of DA to fear acquisition. Further research will be necessary to settle this question.

To conclude, our findings highlight DA as a candidate neurotransmitter for fear extinction. This opens up interesting perspectives for neurobiological therapy augmentation, for instance, via adjunctive treatment with DAergic drugs. Experimental studies using the NMDA receptor agonist D-cycloserine to enhance the effects of exposure therapy have demonstrated the potential for such a strategy⁵³ (reviewed in Grillon⁵⁴). Pharmacological augmentation might be particularly useful in patients resistant to standard forms of behavior therapy. We would, however, caution against testing drugs in patients for which possible potentiating effects on fear expression or conditioning have not been carefully ruled out in previous non-clinical studies. Another promising lead for future studies would be to examine interactions with the endogenous opioid system, which, animal studies suggest, is another potential substrate of error signaling during fear extinction^{55,56} and therefore another interesting candidate neurotransmitter system for translational research.

Conflict of interest

The authors declare no conflict of interest.

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- Rescorla RA, Wagner AR. A theory of Pavlovian conditioning: variations in the effectiveness of reinforcement and nonreinforcement. In: Rescorla RA, Wagner AR, Black AH, Prokasy WF (eds). *Classical Conditioning II: Current Research and Theory*. Appleton-Century-Crofts: New York, NY, 1972, pp 64–99.
- Sutton RS, Barto AG. *Reinforcement Learning: an Introduction (Adaptive Computation and Machine Learning)*. MIT Press: Cambridge, Massachusetts, 1998.
- Rothbaum BO, Davis M. Applying learning principles to the treatment of post-trauma reactions. *Ann N Y Acad Sci* 2003; **1008**: 112–121.
- Pavlov IP. *Conditioned reflexes: an investigation of the physiological activity of the cerebral cortex*. Oxford University Press: Oxford, 1927.
- Myers KM, Davis M. Behavioral and neural analysis of extinction. *Neuron* 2002; **36**: 567–584.
- Bouton ME. Context and behavioral processes in extinction. *Learn Mem* 2004; **11**: 485–494.
- Marsicano G, Wotjak CT, Azad SC, Bisogno T, Rammes G, Cascio MG et al. The endogenous cannabinoid system controls extinction of aversive memories. *Nature* 2002; **418**: 530–534.
- Lin CH, Yeh SH, Lu HY, Gean PW. The similarities and diversities of signal pathways leading to consolidation of conditioning and consolidation of extinction of fear memory. *J Neurosci* 2003; **23**: 8310–8317.
- Gottfried JA, Dolan RJ. Human orbitofrontal cortex mediates extinction learning while accessing conditioned representations of value. *Nat Neurosci* 2004; **7**: 1144–1152.
- Heldt SA, Stanek L, Chhatwal JP, Ressler KJ. Hippocampus-specific deletion of BDNF in adult mice impairs spatial memory and extinction of aversive memories. *Mol Psychiatry* 2007; **12**: 656–670.
- Herry C, Ciocchi S, Senn V, Demmou L, Muller C, Luthi A. Switching on and off fear by distinct neuronal circuits. *Nature* 2008; **454**: 600–606.
- Jungling K, Seidenbecher T, Sosulina L, Lesting J, Sangha S, Clark SD et al. Neuropeptide S-mediated control of fear expression and extinction: role of intercalated GABAergic neurons in the amygdala. *Neuron* 2008; **59**: 298–310.
- Montague PR, Hyman SE, Cohen JD. Computational roles for dopamine in behavioural control. *Nature* 2004; **431**: 760–767.
- Pessiglione M, Seymour B, Flandin G, Dolan RJ, Frith CD. Dopamine-dependent prediction errors underpin reward-seeking behaviour in humans. *Nature* 2006; **442**: 1042–1045.
- Schultz W. Behavioral theories and the neurophysiology of reward. *Ann Rev Psychol* 2006; **57**: 87–115.
- Holtzman-Assif O, Laurent V, Westbrook RF. Blockade of dopamine activity in the nucleus accumbens impairs learning extinction of conditioned fear. *Learn Mem* 2010; **17**: 71–75.
- El Ghundi M, O'Dowd BF, George SR. Prolonged fear responses in mice lacking dopamine D1 receptor. *Brain Res* 2001; **892**: 86–93.
- Cragg SJ, Rice ME. DANCing past the DAT at a DA synapse. *Trends Neurosci* 2004; **27**: 270–277.
- Vandenberg DJ, Persico AM, Hawkins AL, Griffin CA, Li X, Jabs EW et al. Human dopamine transporter gene (*DAT1*) maps to chromosome 5p15.3 and displays a VNTR. *Genomics* 1992; **14**: 1104–1106.
- Heinz A, Goldman D, Jones DW, Palmour R, Hommer D, Gorey JG et al. Genotype influences *in vivo* dopamine transporter availability in human striatum. *Neuropsychopharmacology* 2000; **22**: 133–139.
- Jacobsen LK, Staley JK, Zoghbi SS, Seibyl JP, Kosten TR, Innis RB et al. Prediction of dopamine transporter binding availability by genotype: a preliminary report. *Am J Psychiatry* 2000; **157**: 1700–1703.
- Fuke S, Suo S, Takahashi N, Koike H, Sasagawa N, Ishiura S. The VNTR polymorphism of the human dopamine transporter (*DAT1*) gene affects gene expression. *Pharmacogenomics J* 2001; **1**: 152–156.
- Mill J, Asherson P, Browes C, D'Souza U, Craig I. Expression of the dopamine transporter gene is regulated by the 3' UTR VNTR: Evidence from brain and lymphocytes using quantitative RT-PCR. *Am J Med Genet* 2002; **114**: 975–979.
- VanNess SH, Owens MJ, Kilts CD. The variable number of tandem repeats element in *DAT1* regulates *in vitro* dopamine transporter density. *BMC Genet* 2005; **6**: 55.
- van Dyck CH, Malison RT, Jacobsen LK, Seibyl JP, Staley JK, Laruelle M et al. Increased dopamine transporter availability associated with the 9-repeat allele of the SLC6A3 gene. *J Nucl Med* 2005; **46**: 745–751.
- Matsumoto M, Weickert CS, Akil M, Lipska BK, Hyde TM, Herman MM et al. Catechol O-methyltransferase mRNA expression in human and rat brain: evidence for a role in cortical neuronal function. *Neurosci* 2003; **116**: 127–137.
- Tunbridge EM, Harrison PJ, Weinberger DR. Catechol-o-methyltransferase, cognition, and psychosis: Val158Met and beyond. *Biol Psychiatry* 2006; **60**: 141–151.
- Lachman HM, Papolos DF, Saito T, Yu YM, Szumlanski CL, Weinshilboum RM. Human catechol-O-methyltransferase pharmacogenetics: description of a functional polymorphism and its potential application to neuropsychiatric disorders. *Pharmacogenetics* 1996; **6**: 243–250.
- Chen J, Lipska BK, Halim N, Ma QD, Matsumoto M, Melhem S et al. Functional analysis of genetic variation in catechol-O-methyltransferase (*COMT*): effects on mRNA, protein, and enzyme activity in postmortem human brain. *Am J Hum Genet* 2004; **75**: 807–821.
- Morrow BA, Elsworth JD, Rasmusson AM, Roth RH. The role of mesoprefrontal dopamine neurons in the acquisition and expression of conditioned fear in the rat. *Neurosci* 1999; **92**: 553–564.
- Lonsdorf TB, Weike AI, Nikamo P, Schalling M, Hamm AO, Ohman A. Genetic gating of human fear learning and extinction: possible implications for gene-environment interaction in anxiety disorder. *Psychol Sci* 2009; **20**: 198–206.
- Raczka KA, Gartmann N, Mechias ML, Reif A, Buchel C, Deckert J et al. A neuropeptide S receptor variant associated with overinterpretation of fear reactions: a potential neurogenetic basis for catastrophizing. *Mol Psychiatry* 2010; **15**: 1045, 1067–1074.
- McClure SM, Berns GS, Montague PR. Temporal prediction errors in a passive learning task activate human striatum. *Neuron* 2003; **38**: 339–346.
- O'Doherty JP, Dayan P, Friston K, Critchley H, Dolan RJ. Temporal difference models and reward-related learning in the human brain. *Neuron* 2003; **38**: 329–337.
- Seymour B, O'Doherty JP, Dayan P, Koltzenburg M, Jones AK, Dolan RJ et al. Temporal difference models describe higher-order learning in humans. *Nature* 2004; **429**: 664–667.
- Menon M, Jensen J, Vitcu I, Graff-Guerrero A, Crawley A, Smith MA et al. Temporal difference modeling of the blood-oxygen level dependent response during aversive conditioning in humans: effects of dopaminergic modulation. *Biol Psychiatry* 2007; **62**: 765–772.
- Seymour B, Daw N, Dayan P, Singer T, Dolan RJ. Differential encoding of losses and gains in the human striatum. *J Neurosci* 2007; **27**: 4826–4831.
- Petrovic P, Kalisch R, Pessiglione M, Singer T, Dolan RJ. Learning affective values for faces is expressed in amygdala and fusiform gyrus. *Soc Cogn Affect Neurosci* 2008; **3**: 109–118.
- Schiller D, Levy I, Niv Y, LeDoux JE, Phelps EA. From fear to safety and back: reversal of fear in the human brain. *J Neurosci* 2008; **28**: 11517–11525.
- Dayan P. Classical conditioning and reinforcement learning. In: Dayan P, Abbott LF (eds). *Theoretical Neuroscience*. MIT Press: Cambridge, MA, 2001, pp 331.

41. Friston KJ, Ashburner JT, Kiebel SJ, Nichols TE, Penny WD. *Statistical Parametric Mapping: the Analysis of Functional Brain Images*. Academic Press: San Diego, CA, 2007.
42. O'Doherty JP. Reward representations and reward-related learning in the human brain: insights from neuroimaging. *Curr Opin Neurobiol* 2004; **14**: 769–776.
43. Spoomaker VI, Andrade KC, Schroter MS, Sturm A, Goya-Maldonado R, Samann PG et al. The neural correlates of negative prediction error signaling in human fear conditioning. *Neuroimage* 2011; **54**: 2250–2256.
44. Bilder RM, Volavka J, Lachman HM, Grace AA. The catechol-O-methyltransferase polymorphism: relations to the tonic-phasic dopamine hypothesis and neuropsychiatric phenotypes. *Neuropsychopharmacology* 2004; **29**: 1943–1961.
45. Meyer-Lindenberg A, Weinberger DR. Intermediate phenotypes and genetic mechanisms of psychiatric disorders. *Nat Rev Neurosci* 2006; **7**: 818–827.
46. Yacubian J, Sommer T, Schroeder K, Glascher J, Kalisch R, Leuenberger B et al. Gene-gene interaction associated with neural reward sensitivity. *Proc Natl Acad Sci USA* 2007; **104**: 8125–8130.
47. Solomon RL, Corbit JD. An opponent-process theory of motivation. I. Temporal dynamics of affect. *Psychol Rev* 1974; **81**: 119–145.
48. Dickinson A, Pearce JM. Inhibitory interactions between appetitive and aversive stimuli. *Psychol Bull* 1977; **84**: 690–711.
49. Mueller D, Bravo-Rivera C, Quirk GJ. Infralimbic D2 receptors are necessary for fear extinction and extinction-related tone responses. *Biol Psychiatry* 2010; **68**: 1055–1060.
50. Ponnusamy R, Nissim HA, Barad M. Systemic blockade of D2-like dopamine receptors facilitates extinction of conditioned fear in mice. *Learn Mem* 2005; **12**: 399–406.
51. Hikind N, Maroun M. Microinfusion of the D1 receptor antagonist, SCH23390 into the IL but not the BLA impairs consolidation of extinction of auditory fear conditioning. *Neurobiol Learn Mem* 2008; **90**: 217–222.
52. Milad MR, Goldstein JM, Orr SP, Wedig MM, Klibanski A, Pitman RK et al. Fear conditioning and extinction: influence of sex and menstrual cycle in healthy humans. *Behav Neurosci* 2006; **120**: 1196–1203.
53. Ressler KJ, Rothbaum BO, Tannenbaum L, Anderson P, Graap K, Zimand E et al. Cognitive enhancers as adjuncts to psychotherapy: use of D-cycloserine in phobic individuals to facilitate extinction of fear. *Arch Gen Psychiatry* 2004; **61**: 1136–1144.
54. Grillon C. D-cycloserine facilitation of fear extinction and exposure-based therapy might rely on lower-level, automatic mechanisms. *Biol Psychiatry* 2009; **66**: 636–641.
55. McNally GP, Westbrook RF. Opioid receptors regulate the extinction of Pavlovian fear conditioning. *Behav Neurosci* 2003; **117**: 1292–1301.
56. McNally GP, Pigg M, Weidemann G. Opioid receptors in the midbrain periaqueductal gray regulate extinction of pavlovian fear conditioning. *J Neurosci* 2004; **24**: 6912–6919.



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