



Complete Genome Sequence of the *Arcobacter bivalviorum* Type Strain LMG 26154

William G. Miller,^a Emma Yee,^a James L. Bono^b

^aProduce Safety and Microbiology Research Unit, U.S. Department of Agriculture, Agricultural Research Service, Albany, California, USA

^bMeat Safety and Quality Research Unit, U.S. Department of Agriculture, Agricultural Research Service, Clay Center, Nebraska, USA

ABSTRACT *Arcobacters* are routinely recovered from marine environments, and multiple *Arcobacter* species have been isolated from shellfish. *Arcobacter bivalviorum* was recovered from mussels collected in the Ebro Delta in northeastern Spain. This report describes the complete whole-genome sequence of the *A. bivalviorum* type strain LMG 26154 (= F4^T = CECT 7835^T).

Members of the genus *Arcobacter* have been isolated from a wide range of hosts and environments (1–3). While many *Arcobacter* species are routinely isolated from terrestrial food animals, at least seven *Arcobacter* species have been recovered from shellfish (4–13). *Arcobacter bivalviorum* is a catalase-positive, urease-negative arcobacter that was isolated originally from mussels collected in the Ebro Delta in Catalonia, Spain (6). The *A. bivalviorum* type strain LMG 26154 (= F4^T = CECT 7835^T) was isolated in March 2007 from mussels of the Ebro Delta. In this study, we report the first closed genome sequence of the *A. bivalviorum* type strain LMG 26154.

Arcobacter bivalviorum was grown at 28°C aerobically for 48 h on anaerobe basal agar (Oxoid) amended with 5% horse blood, and genomic DNA was extracted as described (14) from a 5- μ l loop of cells. Shotgun and paired-end Roche 454 reads were generated as described (14) and assembled into 40 contigs using Newbler (version 2.6) with default parameters. A single chromosomal scaffold was produced from 21 unique contigs and closed into a circular, contiguous sequence by using the 19 remaining contigs that represent repeat regions, the Perl script `contig_extender3` (14), and directed PCR amplification followed by Sanger sequencing. PacBio sequencing was performed as described (14), generating a single circular chromosomal sequence which was inserted into a SeqMan version 8.0.2 (DNASTAR, Madison, WI) assembly comprised of 454 and Sanger reads, creating a composite 454/PacBio chromosomal sequence and assembly, and additionally verifying the order of the 454 contigs within the original assembly. Base calls within this composite 454/PacBio chromosomal sequence were verified using Illumina HiSeq reads (SeqWright, Houston, TX), as follows. The HiSeq reads were also assembled within Newbler as described above; the resulting trimmed contigs, with a per-base quality score of ≥ 40 , were assembled onto the SeqMan 454/PacBio assembly. Sequences between the Illumina contigs were queried for single nucleotide polymorphisms (SNPs) using the HiSeq reads and the “find variations/SNPs” module within Geneious version 11.0.2 (Biomatters Ltd., Auckland, New Zealand). The final coverage across the closed genome (i.e., a single circular chromosome with no gaps or ambiguous bases) was 1,327 \times . Chromosomal assembly was also validated using an optical restriction map (restriction enzyme XbaI; OpGen, Gaithersburg, MD).

A summary of the LMG 26154^T genome features is presented in Table 1. *A. bivalviorum* strain LMG 26154^T has a circular genome of 2,684,688 bp with an average GC content of 28.1%. Protein-, rRNA-, and tRNA-encoding genes were identified and

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TABLE 1 Sequencing metrics and genomic data for *A. bivalviorum* strain LMG 26154^T

Feature	Value(s) ^a
Sequencing metrics	
454 (shotgun) platform	
No. of reads	117,700
No. of bases	65,973,662
Average length (bases)	560.5
Coverage (×)	24.6
454 (paired end) platform	
No. of reads	89,211
No. of bases	28,498,539
Average length (bases)	319.5
Coverage (×)	10.6
Illumina HiSeq platform	
No. of reads	18,785,810
No. of bases	1,878,581,000
Average length (bases)	100
Coverage (×)	699.8
PacBio platform	
No. of reads	356,337
No. of bases	1,588,024,540
Average length (bases)	4,456.5 ^b
Coverage (×)	591.5
Genomic data	
Chromosome	
Size (bp)	2,684,688
G+C content (%)	28.11
No. of CDS ^c	2,584
Assigned function (% CDS)	961 (37.2)
General function annotation (% CDS)	1,018 (39.4)
Domain/family annotation only (% CDS)	180 (7.0)
Hypothetical (% CDS)	425 (16.4)
Pseudogenes	16
Genomic islands/CRISPRs	
No. of genetic islands	4
No. of CDS in genetic islands	49
No. of CRISPR-Cas loci	0
Gene content/pathways	
Signal transduction	
Che proteins	<i>cheABCDRVW(Y)</i> ₂
No. of methyl-accepting chemotaxis proteins	23
No. of response regulators	55
No. of histidine kinases	67, [1]
No. of response regulator/histidine kinase fusions	6
No. of diguanylate cyclases	30, [1]
No. of diguanylate phosphodiesterases (HD-GYP, EAL)	9, 5
No. of diguanylate cyclase/phosphodiesterases	12
No. of other	10
Motility	
Flagellin genes	<i>flaAB</i>
Restriction/modification	
No. of type I systems (<i>hsd</i>)	0
No. of type II systems	1
No. of type III systems	0
Transcription/translation	
No. of transcriptional regulatory proteins	64, [1]
Non-ECF ^d σ factor	σ^{70}
No. of ECF σ factors	2
No. of tRNAs	47
No. of ribosomal loci	4
Nitrogen fixation (<i>nif</i>)	No
Osmoprotection	<i>ectABC, proVWX</i>
Pyruvate → acetyl-CoA	
Pyruvate dehydrogenase (E1/E2/E3)	Yes
Pyruvate:ferredoxin oxidoreductase	<i>por</i>
Urease	No
Vitamin B ₁₂ biosynthesis	Yes

^aNumbers in brackets indicate pseudogenes or fragments.

^bMaximum length, 29,273 bp.

^cNumbers do not include pseudogenes. CDS, coding sequences.

^dECF, extracytoplasmic function.

annotated as described (15), with the exception that the composite proteome used here for comparison included proteins derived from all currently completed *Campylobacter* and *Arcobacter* genomes. The genome is predicted to encode 2,584 putative protein-coding genes and 16 pseudogenes. Additionally, the LMG 26154^T genome contains 4 rRNA operons and 47 tRNA-encoding genes. Four genomic islands were identified in the LMG 26154^T chromosome. Two of these genomic islands are approximately 10 kb in size, and two are approximately 15 kb in size. No plasmids were identified in strain LMG 26154^T.

Strain LMG 26154^T contains 25 of the 30 genes necessary for *de novo* biosynthesis of adenosylcobalamin (coenzyme B₁₂) via the oxygen-independent (anaerobic) pathway (16). Orthologs of *cbiJ*, *cobA*, *cobC*, *fre*, and *pduX* were not identified; however, these enzymatic functions could be provided by other proteins or by orthologs with low similarity to proteins currently in the NCBI nr database.

Data availability. The complete genome sequence of *A. bivalviorum* strain LMG 26154^T has been deposited in GenBank under the accession number [CP031217](https://doi.org/10.1128/CP031217). Illumina HiSeq, 454, and PacBio sequencing reads have been deposited in the NCBI Sequence Read Archive (SRA) under the accession number [SRP154993](https://doi.org/10.1128/SRP154993).

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REFERENCES

- Collado L, Figueras MJ. 2011. Taxonomy, epidemiology, and clinical relevance of the genus *Arcobacter*. *Clin Microbiol Rev* 24:174–192. <https://doi.org/10.1128/CMR.00034-10>.
- Forsythe SJ. 2006. *Arcobacter*, p. 181–221. In Motarjemi Y, Adams M (ed), *Emerging foodborne pathogens*. Woodhead Publishing Ltd, Cambridge, England.
- Wesley IV, Miller WG. 2010. *Arcobacter*: an opportunistic human foodborne pathogen? p. 185–212. In Scheld WM, Grayson ML, Hughes JM (ed), *Emerging Infections* 9. ASM Press, Washington, DC.
- Mizutani Y, Tanaka R. 2017. Genome sequence of *Arcobacter* sp. strain LA11, isolated from the abalone *Haliotis discus*. *Genome Announc* 5:e00032-17. <https://doi.org/10.1128/genomeA.00032-17>.
- Tanaka R, Cleenwerck I, Mizutani Y, Iehata S, Bossier P, Vandamme P. 2017. *Arcobacter haliotis* sp. nov., isolated from abalone species *Haliotis gigantea*. *Int J Syst Evol Microbiol* 67:3050–3056. <https://doi.org/10.1099/ijsem.0.002080>.
- Levicán A, Collado L, Aguilar C, Yustes C, Diéguez AL, Romalde JL, Figueras MJ. 2012. *Arcobacter bivalviorum* sp. nov. and *Arcobacter venerupis* sp. nov., new species isolated from shellfish. *Syst Appl Microbiol* 35:133–138. <https://doi.org/10.1016/j.syapm.2012.01.002>.
- Morejon IF, González A, Ferrús MA. 2017. Detection, identification, and antimicrobial susceptibility of *Arcobacter* spp. isolated from shellfish in Spain. *Foodborne Pathog Dis* 14:238–243. <https://doi.org/10.1089/fpd.2016.2202>.
- Diéguez AL, Balboa S, Magnesen T, Romalde JL. 2017. *Arcobacter lekithochrous* sp. nov., isolated from a molluscan hatchery. *Int J Syst Evol Microbiol* 67:1327–1332. <https://doi.org/10.1099/ijsem.0.001809>.
- Collado L, Cleenwerck I, Van Trappen S, De Vos P, Figueras MJ. 2009. *Arcobacter mytili* sp. nov., an indoxyl acetate-hydrolysis-negative bacterium isolated from mussels. *Int J Syst Evol Microbiol* 59:1391–1396. <https://doi.org/10.1099/ijms.0.003749-0>.
- Figueras MJ, Collado L, Levican A, Perez J, Solsona MJ, Yustes C. 2011. *Arcobacter molluscorum* sp. nov., a new species isolated from shellfish. *Syst Appl Microbiol* 34:105–109. <https://doi.org/10.1016/j.syapm.2010.10.001>.
- Figueras MJ, Levican A, Collado L, Inza MI, Yustes C. 2011. *Arcobacter ellisii* sp. nov., isolated from mussels. *Syst Appl Microbiol* 34:414–418. <https://doi.org/10.1016/j.syapm.2011.04.004>.
- Bonerba E, Mottola A, Parisi A, Di Pinto A, Serraino A, Bozzo G, Giacometti F, Ceci E, Tantillo G. 2015. Detection of *Arcobacter* spp. in *Mytilus galloprovincialis* samples collected from Apulia region. *Ital J Food Saf* 4:4583. <https://doi.org/10.4081/ijfs.2015.4583>.
- Levicán A, Rubio-Arcos S, Martínez-Murcia A, Collado L, Figueras MJ. 2015. *Arcobacter ebronensis* sp. nov. and *Arcobacter aquimarinus* sp. nov., two new species isolated from marine environment. *Syst Appl Microbiol* 38:30–35. <https://doi.org/10.1016/j.syapm.2014.10.011>.
- Miller WG, Yee E, Lopes BS, Chapman MH, Huynh S, Bono JL, Parker CT, Strachan NJ, Forbes KJ. 2017. Comparative genomic analysis identifies a *Campylobacter* clade deficient in selenium metabolism. *Genome Biol Evol* 9:1843–1858. <https://doi.org/10.1093/gbe/evx093>.
- Miller WG, Yee E, Chapman MH, Smith TP, Bono JL, Huynh S, Parker CT, Vandamme P, Luong K, Korlach J. 2014. Comparative genomics of the *Campylobacter lari* group. *Genome Biol Evol* 6:3252–3266. <https://doi.org/10.1093/gbe/evu249>.
- Fang H, Kang J, Zhang D. 2017. Microbial production of vitamin B₁₂: a review and future perspectives. *Microb Cell Fact* 16:15. <https://doi.org/10.1186/s12934-017-0631-y>.