


Understanding T cell phenotype for the design of effective chimeric antigen receptor T cell therapies

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ABSTRACT

Rapid advances in immunotherapy have identified adoptive cell transfer as one of the most promising approaches for the treatment of cancers. Large numbers of cancer reactive T lymphocytes can be generated *ex vivo* from patient blood by genetic modification to express chimeric antigen receptors (CAR) specific for tumor-associated antigens. CAR T cells can respond strongly against cancer cells, and adoptive transferred CAR T cells can induce dramatic responses against certain types of cancers. The ability of T cells to respond against disease depends on their ability to localize to sites, persist and exert functions, often in an immunosuppressive microenvironment, and these abilities are reflected in their phenotypes. There is currently intense interest in generating CAR T cells possessing the ideal phenotypes to confer optimal antitumor activity. In this article, we review T cell phenotypes for trafficking, persistence and function, and discuss how culture conditions and genetic makeups can be manipulated to achieve the ideal phenotypes for antitumor activities.

INTRODUCTION

Chimeric antigen receptor (CAR) T cell adoptive transfer has shown great promise and is currently approved by multiple countries in treating some hematological malignancies.¹ In this therapy, T cells from patients with cancer are genetically modified to express CARs specific for cancer antigens. CARs are synthetic receptors that usually comprise an extracellular domain specific for a tumor antigen, a cytoplasmic domain containing activation and costimulatory molecules (such as CD3 ϵ , CD28, and 4-1BB), and a hinge region that links the two domains. However, applying CAR T cell therapies to solid tumors has been hampered by additional challenges including limited infiltration and persistence of CAR T cells in the tumors, and a hostile immunosuppressive tumor microenvironment (TME).^{2–4}

Gene expression profiles and cell phenotypes determine if T cells can successfully traffic, persist and function in the tumor. Here, we discuss the progressive change in

phenotypes of T cells during the course of a natural T cell response as well as investigate studies that have attempted to emulate these phenotypes in CAR T cells, providing a rationale for the optimum CAR T cell phenotypes to achieve enhanced efficacy. We propose three central characteristics to be considered when designing cellular therapies: the ability to self-renew and persist, the ability to traffic to and penetrate disease sites, and the ability to function while resisting exhaustion induced by an immunosuppressive TME. These three characteristics can be revealed as phenotypes that specify markers for T cell differentiation, trafficking and exhaustion respectively. Herein, we discuss the relevance of these phenotypes in providing a rationale for the optimum design of CAR T cell therapy for cancers with a focus on solid tumors.

SELF-RENEWAL AND PERSISTENCE: T CELL DIFFERENTIATION PHENOTYPES

Naïve, memory and effector T cells

A resting naïve T cell (T_N) that circulates and traffics through the lymphoid tissues remains uncommitted to its effector and memory fates until receiving signals from antigen-presenting cells (APCs) alongside signaling downstream of their cytokine receptors. In response to the specific cytokine stimulation, the activated T cells will differentiate into a variety of phenotypes that together enable an effective immune response and establishment of immunological memory.^{5,6}

The conventional model of T cell differentiation proposes that T_N cells can differentiate into effector T cells (T_{EFF}), specialized killer cells that produce cytotoxic molecules and effector cytokines. Most of the T_{EFF} cells die during an immune response, but some of them survive and differentiate into memory T cells that are capable of mounting a memory response to a subsequent encounter of their

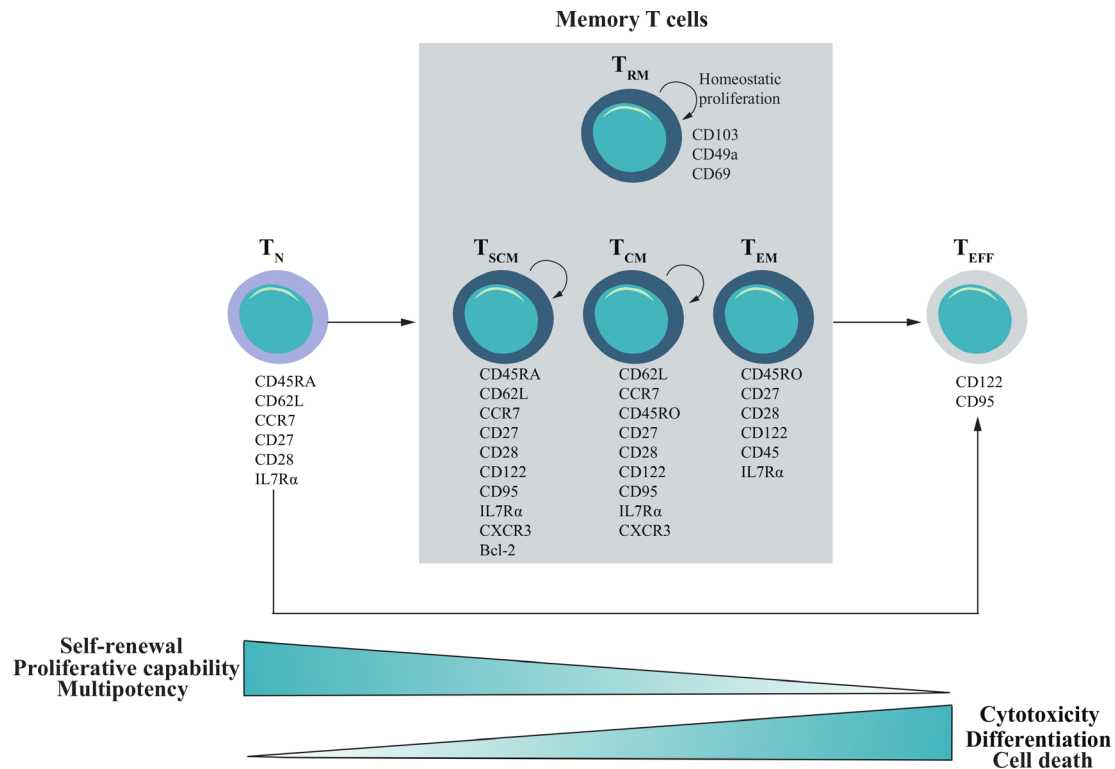


Figure 1 Phenotype definition and characteristics of T cell differentiation states. A widely accepted model of T cell differentiation in adoptive T cell transfer field dictates that T cells progress along a linear trajectory of development, with each differentiation phase displaying a unique phenotype with altered functionality and properties. This model proposes while some primed naïve T cell (T_N) cells will differentiate into effector T cells (T_{EFF}), some primed T_N cells will differentiate into memory T cells, including stem memory T cells (T_{SCM}), a subtype categorized by their ability to self-renew, while also possessing multipotent differentiation capacity, central memory T cells (T_{CM} , long-lived), effector memory T cells (T_{EM}), and tissue residential memory T cells (T_{RM}). These memory T cells can both self-renew, and give rise to a pool of T_{EFF} . Bcl 2, B-cell lymphoma 2; IL7R α , interleukin receptors.

TCR-specific antigens. Although this conventional model has been widely accepted and predicts that T_{EFF} cells can give rise to memory T cells, it has also been debated that T_{EFF} cells are terminally differentiated and do not give rise to memory cells. Instead, it was proposed that a proportion of memory cells are differentiated from the activated T_N cells that never experience a full-strength effector state.⁷ In fact, a number of T cell differentiation models have been proposed.⁷⁻⁹ In this review, we have adopted the nomenclatures that are widely used in the field of adoptive T cell transfer (ACT) in **figure 1**.

The T_{EFF} phenotype was originally considered ideal for ACT therapies due to their potent killing capacity. However, T_{EFF} cells have a poor ability to expand and persist in vivo. Transcriptome analysis has identified that a less differentiated T cell signature is associated with superior anti-cancer activity. Fraietta *et al*¹⁰ studied 41 chronic lymphocytic leukemia patients treated with CD19-CAR, and found that a good response was associated with CD27⁺CD45RO⁻CD8⁺ cell population at the time of leukapheresis. This phenotype is consistent with a resting antigen-experienced cell with long-lived memory characteristics.¹¹ This study links clinical responses to T cell differentiation status. Consequently, the CAR T cell

field is moving towards designing protocols to retain less differentiated T cells.

Initially, efforts were focused on generating T cell products with T_{CM} (CD62L⁺CCR7⁺) and T_{EM} (CD62L⁻CCR7⁻) cell phenotypes. Naturally occurring T_{CM} cells express the lymph node homing molecules CD62L and CCR7 and are considered to have enhanced replicative capacity but limited effector functions. T_{EM} cells are more cytolytic and express chemokine receptors and adhesion molecules necessary for trafficking to peripheral tissues, rather than lymph nodes. T_{EM} cells will terminally differentiate after a limited number of divisions, although some can convert to T_{CM} .¹² Klebanoff *et al*¹³ demonstrated that T cells generated in vitro with a CD62L⁺CCR7⁺ phenotype (T_{CM} -like) have a lower expression of genes associated with effector functions than the CD62L⁻CCR7⁻ cells (T_{EM} -like), predicting a less differentiated status. Importantly, the T_{CM} -like cells had a preference to traffic to secondary lymphoid tissues, which resulted in enhanced clearance of tumors in the mice.¹³ These findings exemplified the benefit of a low differentiation T cell status on ACT efficacy. Clinical protocols have been established for generating CAR T cell products with enriched memory-like T cells. For example, an early study transduced the enriched

CD8⁺CD45RA⁺CD62L⁺ T_{CM} cells from human peripheral blood mononuclear cells using lentiviral vectors encoding a CD19-CAR.¹⁴ These T_{CM}-CAR T cells retained their T_{CM}-like phenotype (CD62L⁺, CD28⁺) after expansion and demonstrated CD19-CAR mediated antitumor effect in vitro. Importantly, when adoptively transferred to NSG mice, these T_{CM}-like CAR T cells were capable of human interleukin (IL)-15-dependent homeostatic engraftment. Subsequently, the safety and feasibility using these T_{CM}-like CAR T cells were demonstrated in a phase I trials.¹⁵ Throughout this review, we refer to in vitro generated T cells with memory phenotypes as T memory-like cells.

Stem-like memory T cells

Recently, the stem-like memory T cells (T_{SCM})-like cells are considered a preferred phenotype for CAR T cell products because of their self-renewal capacity, ability to generate other T cell subsets and their increased ability to engraft.^{16,17} T_{SCM} cells are a T memory cell subtype categorized by their ability to self-renew while possessing multipotent differentiation capacity. T_{SCM} cells can give rise to long-lived T_{CM}, T_{EM} and a pool of short-lived T_{EFF}. T_{SCM} cells have stem-like properties and are phenotypically similar to T_N cells. Both subsets express CD45RA, CD62L, CCR7, CD27 and CD28, but diverge in their expression of CD122 and CD95, markers which demarcate antigen-experienced memory subsets (figure 1).¹⁶

The investigation of T_{SCM} cells has been difficult, due to the paucity of the T_{SCM} cells in the body, with only 2%–3% of circulating T cells in humans having the T_{SCM} cell phenotype. T_{SCM} cells were first described in mice, where a T cell subset with the naïve phenotype of CD44^{lo}CD62L^{hi} also expressed memory markers such as CD122, B-cell lymphoma 2 (Bcl-2), as well as stem cell antigen-1 (Sca-1).¹⁸ Sca-1 does not have a human ortholog, but Gattinoni *et al.*¹⁶ described the generation of human T_{SCM}-like cells by inducing Wnt signaling in T cells in vitro. Inhibition of this pathway with TWS119, a Wnt pathway activator, promoted the expression of TCF1 and LEF1, which have been determined to promote T cell persistence and self-renewal capacity.¹⁹ The resulting T_{SCM}-like cells retained a CD45RO⁺CD62L⁺ naïve-like phenotype but expressed high levels of memory markers including CD95 and CD122. In addition, these T_{SCM}-like cells expressed all the core phenotype markers including CD122, Bcl-2 and CXCR3,²⁰ similar to the murine T_{SCM} cells described previously.¹⁸

Direct evidence that a T_{SCM}-like phenotype is desirable for CAR T cell products comes from a clinical trial where CD19-CAR T cells were adoptively transferred to patients with B-cell malignancies. In this study, the CD19-CAR T cell in vivo expansion correlated with the frequency of a T_{SCM}-like phenotype within the infused product.²¹ In addition to the superior proliferative capacity,¹⁶ the tropism of T_{SCM}-like cells may also contribute to their enhanced engraftment. It was demonstrated that naturally occurring T_{SCM} cells in non-human primates have tropism for lymphoid organs, where they receive survival signals.²²

Although further studies are needed to confirm the tropism of the T_{SCM}-like cells in ACT settings, their phenotype including the expression of CD62L and CCR7, indicates that T_{SCM}-like cells may have a preference to traffic to lymphoid tissues. The resistance to apoptosis may also contribute to better engraftment. High expression of anti-apoptotic markers, such as BCL-2, is a common feature of T_{SCM}-like cells.¹⁶ In fact, adoptively transferred T_{SCM}-like cells were observed to persist while maintaining precursor potential in the patients of severe combined immunodeficiency disease for up to 12 years.²³

To date, a number of strategies have been developed to generate T_{SCM}-like CAR T cells for adoptive transfer. Early investigation demonstrated that Wnt signaling promotes the generation of T_{SCM}-like cells in vitro.²⁰ Subsequently, a clinical-grade platform using TWS119 was established to produce CD19-CAR T_{SCM}.²⁴ These CD19-CAR T_{SCM} cells exhibited enhanced metabolic fitness compared with CAR T cells cultured using a protocol without Wnt signaling induction. Controversially, a number of studies have demonstrated that β-catenin accumulation in T cells, an effect induced by TWS119, leads to reduced T cell expansion and impaired T cell effector function, which could potentially damage T cell-mediated immunity.^{25,26}

Subsequently, IL-7 and IL-15 were successfully used to generate T_{SCM}-like cells from T_N cells and demonstrated more efficiency in expanding and sustaining T_{SCM}-like cells than TWS119.¹⁷ IL-7 was shown to be critical for the T_{SCM}-like cells to preserve a CD45RA⁺CD62L⁺ phenotype, while IL-15 or IL-2 supported cell expansion, with IL-15 providing superior T_{SCM} phenotype preservation compared with IL-2 (17). In a separate study, it was demonstrated that the frequency of the CD45RA⁺CCR7⁺ T_{SCM} cell-like phenotype generated using γ-cytokines in the CAR T cell product correlated with expansion and persistence of CD19-CAR T cells in patients.²¹ Currently, the efficacy of IL-7 and IL-15 generated CD19 CAR T cells is being evaluated in a clinical trial (NCT02652910).

Interestingly, CAR T cells expanded in IL-15 alone also had a T_{SCM}-like phenotype. These T_{SCM}-like cells had reduced exhaustion markers, higher antiapoptotic properties and increased proliferative capacity compared with the cells cultured with IL-2. In addition, these T_{SCM}-like cells had decreased mammalian target of rapamycin complex 1 (mTORC1) activity, and improved mitochondrial fitness. The addition of the mTORC1 inhibitor rapamycin to the CAR T cell culture with IL-2 in the absence of IL-15 exhibited a similar T_{SCM} phenotype, suggesting that decreasing mTORC1 activity could preserve the T_{SCM}-like phenotype.²⁷ Other γ-cytokines, such as IL-21, have also been tested in culture to generate 'fit' T_{SCM}-like cells. Clinical procedures using CD8⁺CD62L⁺CD45RA⁺ enriched naïve cells cultured in the presence of IL-7, IL-21 and TWS119 demonstrated that TWS119 has a synergistic effect with IL-21 to enhance T_{SCM} generation, through stabilization of β-catenin and induction of maximal expression of TCF1 and LEF1. These T_{SCM}-like CAR T cells demonstrated enhanced metabolic fitness

compared with the current standard CAR T cells, and mediated robust, long-lasting antitumor activity in NSG mice.²⁴ A clinical trial (NCT01087294) has commenced to test these T_{SCM}-CAR T cell products.

Tissue resident T memory cells

After the resolution of an immune response, T cells that entered into an inflamed tissue can be retained in the tissue where they become resident T memory cells (T_{RM}) cells. The function of T_{RM} cells is to provide regional surveillance for reinfection, therefore T_{RM} cells do not recirculate in the blood like other T cells. Phenotypically, human and murine T_{RM} cells express similar markers which are essential for their tissue retention. These include the constitutive expression of CD69 and CD49a, which bind to extracellular matrix components (figure 1). T_{RM} cells also lack the expression of CD62L and CCR7 that normally enable entry into lymphoid tissue.

T_{RM} cells have been identified in almost all organs, and their expression profiles vary in different tissues, enabling their tissue-specific retention.²⁸ In the case of T_{RM} cells in epithelial tissues, integrin CD103 binds to E-cadherin and promotes T_{RM} adhesion to epithelial cells. In human tumors of epithelial origin, including the tumors of breast, lung, ovary and bladder, patients with the greatest portion of CD103⁺CD8⁺ T_{RM}-phenotype cells in their tumors have the best survival.²⁸ This phenomenon could be explained by an enhanced cytotoxic effect of the T_{RM} cells than the CD103⁻ T cells.²⁹

In addition to their direct cytotoxic activity, T_{RM} cells can produce large amounts of cytokines that can activate and attract other anti-tumor immune cells. The presence of T_{RM} is associated with a higher density of T cell infiltration in lung and breast tumors.^{30 31} T_{RM} cells have also been demonstrated to promote tumor antigen spreading by activating cross-presenting dendritic cells (DCs) to prime and expand new cytotoxic T cells with different tumor-antigen specificities. This resulted in broader protection against cancers, especially from cancer cells lacking T_{RM} targeted antigens.³² Recently, T_{RM} cells were proposed to be a mediator in checkpoint blockade therapies. Analysis of several tumor types have shown T_{RM} cells expressing high levels of programmed cell death protein 1 (PD1), exhibiting rapid expansion,³³ cytotoxic effects³⁴ and expression of cytotoxicity genes following anti-PD1 treatment.³⁵

Due to their quick and robust immune responses at the tumor site, promoting ACT cell differentiation into T_{RM} cells in vivo may be important for the long-term control of the disease. Recruitment of activated T cells into tissues depends on the cytokine/chemokine receptors and adhesion molecules expressed by T cells. Therefore, ACT products must express or obtain these factors in order to effectively traffic to tumors and establish residency. Additionally, strategies for reprogramming the TME for optimal T_{RM} generation is desired. To achieve this, a study used intratumoral delivery of the β -glucan curdlan, whose receptor dectin-1 is expressed by DCs and macrophages.

The β -glucan curdlan reprogrammed tumor infiltrating DCs and induced CD8⁺ T cells to differentiate into T_{RM} cells which resulted in inhibited cancer progression in a humanized murine breast cancer model.³⁶ Localized radiotherapy that has been widely used to reprogram TME³⁷ could also be considered to enhance T_{RM} generation. A recent study demonstrated that post irradiation, tumor infiltrating T cells were reprogrammed to express a similar transcriptome to that of T_{RM} cells, which are also radioresistant. These reprogrammed T cells retained their motility, ability to produce interferon (IFN)- γ and mediated antitumor responses.³⁸ Despite the potential benefit of an ACT product with a T_{RM}-like phenotype, little has been reported on generating such cells in vitro.

MIGRATION AND PENETRATION INTO TUMORS: T CELL TRAFFICKING MOLECULES

T cell infiltration from the blood into the tumor is a multi-step process. Activated T cells gain expression of tissue homing molecules, which include adhesion molecules and chemokine receptors.³⁹ The first step in T cell infiltration requires the slowing of T cell movement by the interaction of selectin ligands on T cells to the selectin molecules on endothelial cells. T cells then 'roll' slowly along the blood vessel surface, which provides an opportunity to encounter chemokines produced by the tissue. Chemokine receptors on T cells bind to the chemokines, thereby initiating the activation of T cell-expressed integrins, such as lymphocyte function-associated antigen-1 (LFA-1) and very late antigen-4 (VLA-4). The binding of the activated integrins to their cognate ligands on the vessel surface mediates firm arrest of T cells. This is followed by transmigration through endothelial cell junctions of the blood vessels into tissues along a gradient of chemokines. Inefficient trafficking and infiltration of T cells into the tumor have been identified as a major hurdle for using CAR T cells in treating solid tumors.³⁹ We show in figure 2 that three families of molecules are essential for T cell trafficking and can be exploited in the design of CAR T cells for improved trafficking.

Selectins and selectin ligands

Selectins are a family of calcium-dependent adhesion molecules and two members have been identified on endothelium. P-selectin (CD62P) is expressed by activated blood vessels and mediates weak interaction and rolling of T cells along the vascular endothelium. Decreased vascular P-selectin expression has been associated with cancer progression in melanoma⁴⁰ and colorectal cancer.⁴¹ It has been demonstrated that in malignant primary tumors and in metastatic lesions of melanoma or colorectal cancers, P-selectin expression was nearly absent and hardly any leukocytes infiltration was detected.^{40 41} P-selectin cannot be induced by inflammatory mediators (tumor necrosis factor (TNF)- α , IL-1 β , and lipopolysaccharide (LPS)) in humans.⁴²

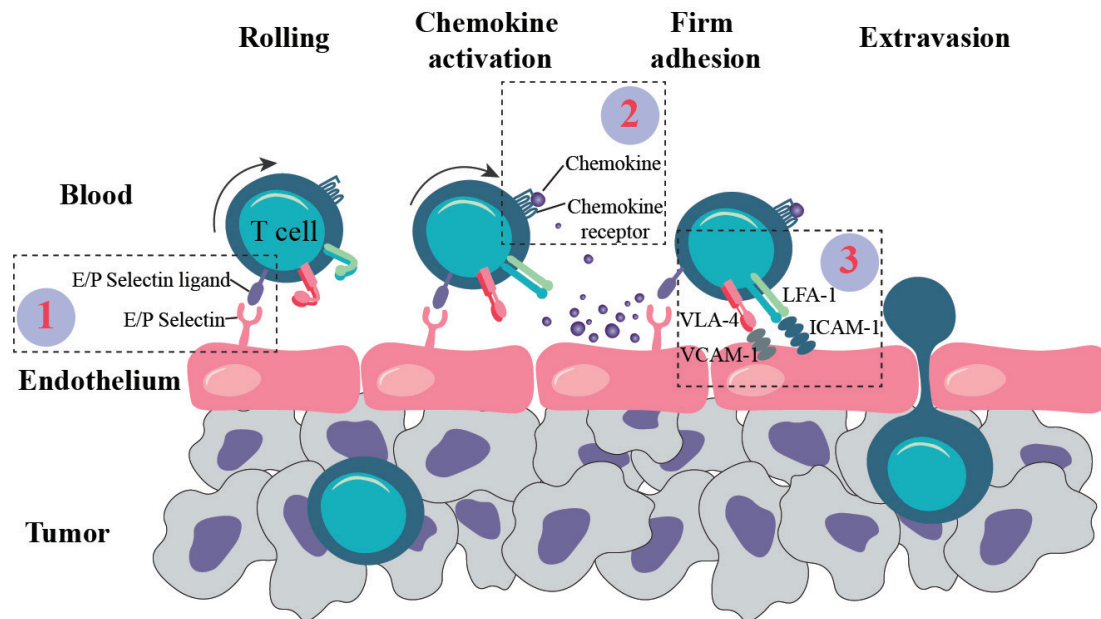


Figure 2 T cell homing events into tumor tissue. T cell infiltration into the tumor from the blood is a multistep process involving a variety of molecules such as (1) selectins and their ligands, (2) chemokines and their receptors, and (3) integrins and their ligands. Strategies manipulating these three family of molecules essential for T cell trafficking for enhanced CAR T cell infiltration into tumors are discussed in the main text. The first step in T cell infiltration requires the initiation of T cell rolling, or slowing of T cell movement, in the blood stream. This is facilitated by the interaction of selectin molecules on the blood vessel with their target ligands on the T cell. Next, chemokine receptors on the T cell bind to their cognate chemokines, produced by tissues or endothelium, thereby initiating the activation of integrins on the T cell. Integrins, when bound to their cognate ligands on the vessel surface, mediate firm arrest of T cell movement. Finally, the T cell is able to migrate through the cell junctions of the blood vessels into tissues, along a gradient of chemokines. CAR, chimeric antigen receptors; LFA-1, lymphocyte function-associated antigen-1; VLA-4, very late antigen-4.

E-selectin (CD62E) is not constitutively expressed by all endothelial cells but can be induced by TNF- α , IL-1 β , and LPS. E-selectin binding allows intimate contacts between the inflamed endothelium and T cells, slowing T cell rolling to much lower velocities than that of P-selectin, favoring subsequent T cell arrest.⁴³

A major ligand for both P- and E-selectins on T cells is P-selectin glycoprotein ligand-1 (PSGL-1), whose binding to selectins requires appropriate glycosylation. Glycosyltransferases add carbohydrate moieties to PSGL-1 that form the sialylated fucosylated O-linked glycan, sialyl Lewis x (sLe^x). Although PSGL-1 is expressed by most T cells, T_N lack the proper terminal glycosylation therefore cannot bind selectins.⁴⁴ Other selectin ligands on T cells have also been identified, including TIM1,⁴⁵ CD44 and CD43.⁴⁶ Blocking T cell binding to P/E selectin with a monoclonal antibody exhibited poor T cell infiltration into tumors.⁴⁷

Strategies enhancing expression of selectins and selectin ligands could be exploited for increasing engineered T cell trafficking to tumors. Because E-selectin expression can be induced by de novo synthesis on endothelial cells in response to inflammatory stimuli,⁴⁸ methods which temporarily enhance inflammation specifically in the TME could be used prior to adoptive transfer. In fact, intratumor injection of toll-like receptor (TLR) agonists enhanced adoptively transferred T cell infiltration into tumors and lead to significant tumor regression in murine

cancer models.⁴⁹ Other strategies such as using oncolytic virus coding cytokines, and local radiotherapy that enhances local E-selectin expression can also be explored. Oncolytic virus encoding TNF- α and IL-2 has been shown to possess potent antitumor activity in immunocompetent Syrian hamsters.⁵⁰ A clinical trial is currently in progress to study the use of this oncolytic virus in treating advanced melanoma (NCT04217473). Irradiation was found to induce the expression of E-selectin and ICAM on human endothelial cells. The induced E-selectin expression and increased adhesion of leukocytes was observed on the irradiated endothelial cells.⁵¹ Localized irradiation of tumors combined with ACT has demonstrated promise in murine tumor models, with increased T cell infiltration to the tumors that lead to tumor regression.⁵²

Glycosyltransferases could be exploited to enhance the expression of selectin-binding glycoforms of PSGL-1 and CD44 on CAR T cells. In humans, there are four glycosyltransferase isoenzymes specialized to create sLe^x: FT3, 5, 6 and 7.⁵³ Mondal *et al*⁵⁴ showed that CAR T cells expanded in standard culture conditions of CD3/CD28/IL2 had decreased expression of the sLe^x moiety and reduced ability to bind E-selectin compared with the T cells without stimulation. The activated CAR T cells uniformly expressed high levels of Type 2 sialyllactosamines, the precursor of sLe^x, that can be converted to sLe^x via cell surface fucosylation. After being surface fucosylated by the addition of purified FT6 enzyme, all the CAR T cells

displayed elevated levels of sLe^X. As a result, CAR T cells had strikingly higher rolling and adhesion on E-selectin⁺ cells and 10-fold enhanced infiltration into bone marrow, where E-selectin is constitutively expressed.⁵⁴

It will be interesting to investigate if other glycosyltransferases can also be used to enhance sLe^X expression on CAR T cells. Cytokines such as IL-4, IL-12 and G-CSF have been reported to increase the expression levels of glycosyltransferases in T cells⁵⁵ and CAR T cells secreting IL-12 have been reported to eradicate cancers.⁵⁶ Various mechanisms have been linked to the enhanced efficacy, and it remains a question whether sLe^X expression was increased on the IL-12-CAR T cells.

Chemokines and chemokine receptors

While a number of chemokines have been reported to be important for the migration of T cells into tumors, CXCR3 is one of the most important receptors in this process. CXCR3 has been found to be expressed on tumor infiltrating lymphocytes (TILs) isolated from human melanoma, colorectal and breast cancers, and a number of murine tumor models have shown the importance of CXCR3 in T cell trafficking into tumors.³⁹ In brief, CXCR3-dependent T cell recruitment into tumors has been proposed to operate on a positive feedback loop. CXCL9 and CXCL10, ligands for CXCR3, are produced by immune cells, endothelial cells and cancer cells in response to IFN- γ .⁵⁷ T cells that have been recruited into inflamed tissue can locally produce IFN- γ , increasing the expression of CXCR3-ligand chemokines which can recruit additional CXCR3⁺ T cells—strengthening the CXCR3-T cell infiltration loop.⁵⁸ CAR T cell therapies could benefit from treatments enhancing CXCR3 ligands in the TME. For example, reagents such as 5-aza-2'-deoxycytidine that enhance the expression of CXCL9 and CXCL10 in the TME⁵⁹ could be used to enhance CAR T cell infiltration to the TME. Similarly, reagents enhancing CXCR3 expression on CAR T cells should also be considered. Newick *et al*⁶⁰ has shown that genetic inhibition of protein kinase A activity in CAR T cells increased their CXCR3 expression and improved CAR T cell trafficking to CXCL10 resulting in enhanced tumor control in tumor-bearing mice.⁶⁰

There is often a mismatch of the chemokine receptors expressed by tumor reactive T cells and the chemokines in the tumor. Ensuring the homing phenotypes of the CAR T cells match the chemokines in the tumor can greatly enhance CAR T cell infiltration into the tumor. One approach is to transduce chemokine receptors into CAR T cells. This approach has been validated in solid tumor xenograft models. CAR T cells that were transduced to express CCR2b had improved infiltration into tumors in models of CCL2⁺ neuroblastoma and malignant pleural mesothelioma, resulting in increased treatment efficacy.^{61, 62} Enforcing expression of IL-8 receptors CXCR1/2 on CAR T cells, to target IL-8 secreting tumors, increased CAR T cell tumor infiltration and enhanced tumor control in a murine model of glioblastoma.⁶³

Recently, CAR T cells have been developed to produce CCL19 to recruit additional T cells and APCs to tumor tissues, and IL-7 to enhance the proliferation of survival of T cells. It has been demonstrated that enhanced interaction of APCs with CAR T cells *in vivo* is beneficial for the anti-tumor effect of CAR T cells.^{64–67} Tumors from IL-7/CCL19-CAR T cell treated mice showed significantly enhanced infiltration of T cells and APCs, and colocalisation of CAR T cells and APCs were significantly increased in the tumors. The treatment induced eradication of all the established tumors in murine models of mastocytoma and Lewis lung carcinoma. CAR T cells in long-term surviving mice developed a CD44⁺CD62L⁺ central memory phenotype.⁶⁸ Thus, this study elegantly combined the strategies enhancing CAR T cell memory and trafficking and generated a superior antitumor effect.

Integrins and integrin ligands

Rolling of T cells along endothelium is transient unless firm adhesion mediated by integrins occurs. Integrins are cell-surface heterodimers which consist of both an alpha (α) and a beta (β) chain. Multiple α and β integrin subunits are expressed on T cells and the α and β subunits couple to form integrins that bind their ligands on endothelium. The binding of integrins to their ligands transmits signals to T cells and endothelium bidirectionally.⁶⁹

In resting T cells, the integrin LFA-1 is in a default low affinity 'bent' conformation. On encountering chemokines on endothelium, LFA-1 is converted into an extended form with an intermediate affinity (figure 2). The subsequent conformational changes promote LFA-1 transition into an active high affinity and fully extended form, supporting T cell arrest through binding to integrin ligands on the endothelium.⁶⁹ Another important integrin is VLA-4. Similar to LFA-1, after the activation and conformational change, VLA-4 binds to its ligand vascular cell adhesion molecule-1 (VCAM-1) on blood vessels.⁶⁹

Although CAR T cells have been designed to target integrin expressing cancer cells,⁷⁰ to our knowledge, strategies altering CAR T cell expression of integrins for improved T cell infiltration are underexplored. Alternatively, treatments that induce the integrin receptor expression on tumor vessel cells can be used in combination with CAR T cells to improve T cell infiltration. It was reported that the administration of recombinant adenovirus encoding IL-12 upregulated VCAM-1 expression on tumor endothelium and mediated the recruitment of adoptively transferred T cells.⁷¹

RESISTING LOSS OF T CELL FUNCTION: MARKERS OF T CELL EXHAUSTION

In order to avoid causing damage to normal tissues, T cells downregulate their functions and develop a phenotype termed as exhaustion. An early study in chronic lymphocytic choriomeningitis virus (LCMV) infected mice found that some viral-specific T cells persisted for a long-time,

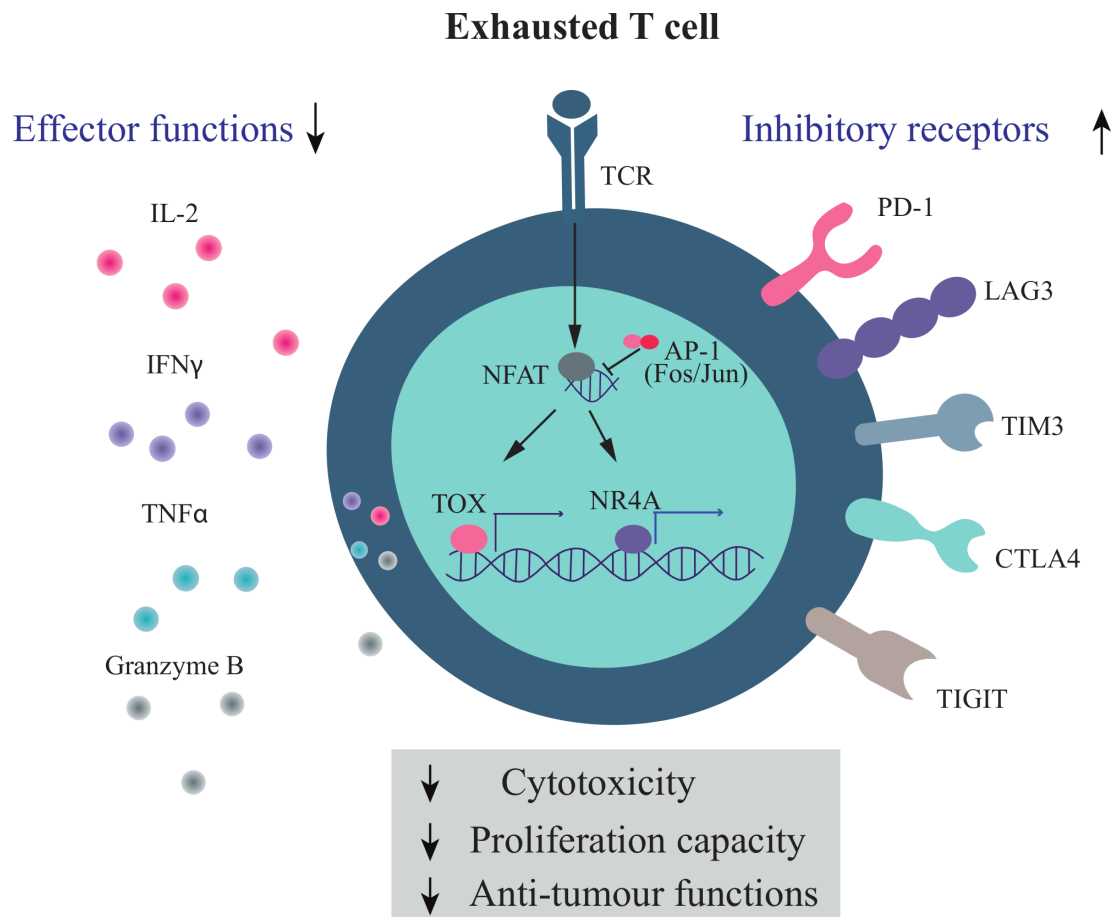


Figure 3 Markers of T cell exhaustion. T cell dysfunction occurs when there is chronic exposure to antigen, such as in the context of cancer. This loss of function is referred to as T cell exhaustion. T cell exhaustion includes the loss of IL-2 production, decreased proliferative capacity, and reduced cytotoxic abilities, including impairment in granzyme B, IFN- γ and TNF- α production. Along with the loss of functionality, T cells increase the expression of inhibitory receptors such as PD1, LAG3, Tim3, CTLA4, and TIGIT. Some of the key regulators of T cell exhaustion include the transcription factors TOX and NR4A (NR4A1, NR4A2, and NR4A3). IFN- γ , interferon- γ ; IL-2, interleukin 6; TNF- α , tumor necrosis factor- α .

proliferated *in vivo*, but were unable to generate an effective antiviral response. These T cells were engaged in continuous but ineffective actions and were likened to ‘Sisyphus’, the Greek king, who was tasked to perform the fruitless job of rolling a boulder up a hill only to see it roll down again.⁷²

Exhausted T cells (T_{ex} cells) represent a distinct population with a unique phenotypic, epigenetic and transcriptional profiles. The ontogeny of T_{ex} cells is still poorly understood, but the exhausted state can be induced in T cells at any differentiation stage and these T_{ex} cells continue to produce exhausted progeny. Thus, exhaustion is a parallel program to T cell differentiation.⁷³ The development of T_{ex} cells is progressive—beginning with the loss of IL-2 production, lower proliferative capacity, and reduced cytotoxic abilities. In further stages of dysfunction, T cells have an impairment in their ability to produce cytokines such as IFN- γ and TNF- α .⁷⁴ Correlating with the progressive accumulation of functional impairments is the expression of well-defined immune checkpoint markers, which include inhibitory receptors such as PD1, LAG3, TIM3, CTLA4, and TIGIT (figure 3).

Functionally impaired T cells have been widely documented in solid tumors such as renal cell carcinoma, non-small cell lung cancer and melanoma.^{75–79} While difficult to dissect the development of T_{ex} cells in humans, Schitinger *et al*⁸⁰ used a murine cancer model and demonstrated that the unresponsive T_{ex} cells could develop at the prephase and early phase of the tumor growth, due to persistent antigen exposure. Although T_{ex} cells in tumors are similar to those generated in chronic viral infection, they are not identical. For example, tumor T_{ex} cells lose Eomesodermin (EOMES) expression while LMCV T_{ex} cells express high levels of EOMES.⁸⁰ Factors such as the immunosuppressive TME and the lack of co-stimulation in tumor settings may contribute to the difference.

The identification of a PD1⁺TCF1⁺CD8⁺ T cell subset presents a link between T cell memory and exhaustion. These cells were identified in cancers of mice and humans and have stem cell-like properties while displaying exhausted features. PD1⁺TCF1⁺CD8⁺ T cells can expand to produce terminally differentiated PD1⁺TCF1⁺ T cells while maintaining a pool of progenitor PD1⁺TCF1⁺ T cells. Tumor control elicited by checkpoint



blockade treatments is dependent on the existence of intratumoral PD1⁺TCF1⁺CD8⁺ T cells and argues that checkpoint blockade expands and differentiates a stem cell-like progenitor cell population, rather than reverse a terminal T cell exhaustion program.⁸¹ Interestingly, a separate study demonstrated that a memory-precursor-like PD1⁺TCF1⁺CD8⁺ TIL subset expanded on checkpoint blockade and provided a wave of T_{EFF} cells to sustain an effective anti-tumor response. The checkpoint blockade impacted the PD1⁺ population indirectly due to the expression of checkpoint receptors on other immune cells in the TME.⁸² However, evidence has shown that PD1⁺ T cells have limited potential to give rise to durable memory T cells, and after the cessation of the PD-L1 blockade treatment, the reinvigorated T_{ex} cells became exhausted again. Although there was a transcriptional rewiring, T_{ex} cells were in a stable epigenetic state that cannot be reverted following therapy.⁸³

The effectiveness of CAR T cell treatment is dependent on the degree of T cell exhaustion. It was demonstrated that T cell exhaustion in cell culture limited the antitumor efficacy of CAR T cells. A GD2-CAR T cell containing CD3ζ and CD28 signaling domains showed high expression of surface inhibitory receptors such as PD1, TIM-3 and LAG3, exhaustion-associated transcription factors T-bet and Blimp-1, produced cytokines poorly in response to antigen compared with CD19.28ζ CAR T cells. The GD2-CAR T cells have basal levels of ζ-phosphorylation (tonic signaling) whereas CD19 CAR T cells do not. Ablating the tonic signaling (by exchanging the CAR costimulatory domain from CD28 to 4-1BB) reversed the exhaustion phenotype. Interestingly, varying degrees of exhaustion, including decreased tumor clearance in mouse models, reduced cytokine production, and increased expression of multiple inhibitory receptors, were identified in other CAR products, and 4-1BB signaling ameliorated exhaustion.⁸⁴

The nuclear factor TOX is a crucial regulator for the development of T_{ex} cells. Ectopic expression of TOX in T_{EFF} cells induced exhaustion and deletion of TOX in T_{ex} cells abrogated exhaustion.⁸⁵ In CD8⁺ T cells, inhibiting the interaction of NFAT1 with AP-1 increased the exhaustion transcriptional program in the T cells and decreased their ability to control infection and tumor growth in murine models.⁸⁶ In addition, NR4A and TOX also maintain the exhaustion program^{73 87} (figure 3).

A number of strategies have been used in CAR T cells to reduce exhaustion, including knocking out exhaustion-associated genes, such as NR4A and Cbl-b. The NR4A or Cbl-b knockout CAR T cells in the tumors had surface markers similar to T_{EFF} and increased cytokine production compared with wild type, which resulted in enhanced antitumor immunity.^{87 88} Rescuing AP-1 signaling could also reverse the exhausted state. Overexpression of JUN rescued CAR T cell exhaustion and greatly increased the anti-tumor efficacy of CAR T cells in multiple animal models.⁸⁹ CAR T cells deficient in TOX and TOX2 showed reduced expression of inhibitory receptors and were

more effective against human CD19 tumors,⁹⁰ demonstrating knocking out TOX and its related molecules can be a strategy to reduce an exhaustion phenotype.

Further studies are needed to optimize manufacturing conditions for CAR T cells to receive appropriate TCR stimulation, cytokines and other supplements. Additionally, other avenues to decrease CAR T cell exhaustion and increase CAR T cell fitness can be explored, such as genetic modifications of CAR T cells by knocking out/in exhaustion related molecules, as well as pairing CAR T cell therapy with other therapies which can reduce CAR T cell exhaustion.

CONCLUDING REMARKS

Key challenges facing effective CAR T cell therapies for solid cancers include inefficient infiltration into tumors, limited in vivo persistence and a hostile TME. Understanding intrinsic T cell factors and properties which occur during a natural immune response can help inform the design of a CAR T cell phenotype which can overcome these challenges.

Understanding T cell differentiation and the corresponding functional properties has led to new thinking around the optimal T cell differentiation state for ACT. Understanding the homing properties of T cells has allowed T cells engineered with receptors to increase tumor infiltration. Understanding the nuclear factors involved in T cell memory formation and exhaustion will allow for the generation of new therapeutics targeting these factors. Current research points in the direction of developing an ACT product with an early memory phenotype absent of exhaustion markers that would be an ideal phenotype for an effective treatment product.

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