## **Original Article**

# Effects of intermittent ladder-climbing exercise training on mitochondrial biogenesis and endoplasmic reticulum stress of the cardiac muscle in obese middle-aged rats

Kijin Kim\*, Nayoung Ahn, Suryun Jung, and Solee Park

Department of Physical Education, College of Physical Education, Keimyung University, Daegu 42601, Korea

#### **ARTICLE INFO**

Received May 26, 2017 Revised August 3, 2017 Accepted August 14, 2017

\***Correspondence** Kijin Kim E-mail: kjk744@kmu.ac.kr

#### **Key Words**

Aging Cardiac muscle ER stress High-fat diet-induced obesity Ladder climbing exercise Mitochondrial biogenesis ABSTRACT The aim of this study is to investigate the effects of intermittent ladderclimbing exercise training on mitochondrial biogenesis and ER stress of the cardiac muscle in high fat diet-induced obese middle-aged rats. We induced obesity over 6 weeks of period in 40 male Sprague-Dawley rats around 50 weeks old, and were randomly divided into four experimental groups: chow, HFD, exercise+HFD, and exercise+chow. The exercising groups underwent high-intensity intermittent training using a ladder-climbing and weight exercise 3 days/week for a total of 8 weeks. Highfat diet and concurrent exercise resulted in no significant reduction in body weight but caused a significant reduction in visceral fat weight (p < 0.05). Expression of PPARo increased in the exercise groups and was significantly increased in the highfat diet+exercise group (p<0.05). Among the ER stress-related proteins, the expression levels of p-PERK and CHOP, related to cardiac muscle damage, were significantly higher in the cardiac muscle of the high-fat diet group (p<0.05), and were significantly reduced by intermittent ladder-climbing exercise training (p<0.05). Specifically, this reduction was greater when the rats underwent exercise after switching back to the chow diet with a reduced caloric intake. Collectively, these results suggest that the combination of intermittent ladder-climbing exercise training and a reduced caloric intake can decrease the levels of ER stress-related proteins that contribute to cardiac muscle damage in obesity and aging. However, additional validation is required to understand the effects of these changes on mitochondrial biogenesis during exercise.

## INTRODUCTION

Aging results in the structural and functional degradation of the cardiac muscles accompanied by reduced contractility and muscle mass. Cell loss or reactive hypertrophy in aged heart results in functional degeneration, leading to eventual defects in heart function [1]. Changes in muscle cells during the aging process is mainly caused by metabolic changes from qualitative and quantitative changes in the mitochondria, fluctuations in the levels of oxidative stress factors in cells, changes in the Ca<sup>2+</sup> sig-

naling structure, and the transformation of type II muscle fibers to type I muscle fibers [2]. Since 95% of the energy required for contraction of the cardiac muscle are supplied by mitochondrial respiration, these metabolic changes have a significant impact on heart dysfunction and can lead to the onset of heart failure [3]. Thus, enhancing mitochondrial biogenesis of the heart is crucial for maintaining heart function and preventing cardiac diseases.

Along with aging, obesity is another key risk factor of cardiovascular diseases. Obese people have increased levels of plasma free fatty acids, with consequent activation of endoplasmic re-

This is an Open Access article distributed under the terms of the Creative Commons Attribution Non-Commercial License, which permits unrestricted non-commercial use, distribution, and reproduction in any medium, provided the original work is properly cited. Copyright © Korean J Physiol Pharmacol, pISSN 1226-4512, eISSN 2093-3827 Author contributions: S.P. performed animal experiment and western blotting analysis. S.J. interpreted data and wrote the manuscript. N.A. and K.K. designed study and revised article.

ticulum (ER) stress and a chronic inflammatory response with negative effects on the condition of the heart [4]. ER stress is strongly associated with various cardiovascular diseases, including heart failure, arteriosclerosis, and ischemic heart diseases [5-7]. A recent study showed that increased ER stress resulted in a 2.4fold reduction in mitochondrial biogenesis [8]. It is well known that the signaling pathway in ER stress mediated by three major ER stress sensors which are PKR-like ER kinase (PERK), inositol requiring enzyme 1 (IRE1) and activating transcription factor 6 (ATF6). PERK of them is a key ER stress sensor and molecular mediator of the ER-mitochondria contact sites, required to regulate inter-organellar cross-talk in ROS-induced cell death, a role which is not shared with other ER stress sensors, like IRE1 [9]. Therefore, PERK and pro-apoptotic C/EBP homologous protein (CHOP), a downstream of PERK, are key factors to understand an association between ER stress and mitochondria malfunction caused by obesity, but these are not clearly understood.

One of the most effective methods to stimulate mitochondrial biogenesis within the muscle tissue is through an exercise program. More specifically, high-intensity or long-term exercise training have been shown to increase the number and quality of mitochondria [10,11]. Previous studies have shown that endurance exercise training in elderly subjects increased the VO<sub>2</sub> peak [12-14], mitochondrial quantity [15], oxidase activity [12,14], muscular protein synthesis rate [16,17], mitochondrial protein gene transcripts [13], and the copy number of mitochondrial DNA [18]. Furthermore, physical exercise has shown to reduce the levels of oxides [19] and to protect muscle cells by increasing the activity of antioxidant enzymes [20-23] that is associated with peroxisome proliferator-activated receptors  $\beta/\delta$  (PPAR $\delta$ ) [24-26]. PPAR $\delta$  is an essential mediator to reduce ER stress in aortas of obese mice [27], and is required for exercise to attenuate ER stress [28], therefore, these findings led us to postulate that the PPAR $\delta$  induced by exercise reduces ER stress in cardiac muscle caused by obesityinduced ROS.

The aim of the present study was to determine the effects of PPAR $\delta$  on the mitochondrial enzymes and PERK/ pro-apoptotic C/EBP homologous protein (CHOP) pathway, a key ER stress pathway, in the cardiac muscle of high fat diet-induced obese middle-aged rats that were perform to high-intensity intermittent training using ladders and weights. The results of this study expect to provide key evidence to support the benefits of exercise training for maintaining cardiovascular health and preventing heart diseases through a mechanism of enhancing the obesity-incurred reduction of mitochondrial biogenesis.

## **METHODS**

## **Subjects**

After a 1-week adaptation period, 40 male, middle-aged (~50

weeks old) Sprague-Dawley rats were purchased and induced to become obese with 6 weeks of a high-fat diet (30%, 50%, and 20% calories from carbohydrate, fat, and protein, respectively). After obesity induction, the rats were randomly divided into four groups: the normal chow diet group (a high-protein diet consisting of 64.5%, 11.8%, and 23.5% calories from carbohydrate, fat, and protein, respectively; 3.2 kcal/g); the high-fat diet group (5.1 kcal/g); the chow+exercise group; and the high-fat diet+exercise group. The treatment regime continued for a total of 8 weeks. The exercising groups underwent high-intensity intermittent training using a ladder-climbing exercise 3 days/week. The chow diet groups were switched from the initial high-fat diet to the chow diet to reduce the total caloric intake and fat consumption.

The rats of all groups received concurrent supplements of vitamins (22 g/kg Teklad vitamins mix no. 40077), minerals (51 g/ kg Teklad mineral mix no. 170915), methionine (5 g/kg, Teklad Premier no. 10850), and choline chloride (1.3 g/kg) [29]. Drinking water was provided ad libitum in the form of secondary ion-exchange water. All rats were individually bred during the breeding period, and the day-night cycle was 12 h. Temperature and relative humidity were maintained around 24±1°C and 60%, respectively. The study design was approved by the Animal Research Ethics Committee of Keimyung University School of Medicine (KM-2012-30R).

### Ladder-climbing exercise

For adaptation, the rats were trained on the ladder-climbing exercise for the first week, using a 1-m ladder positioned at a 75° incline without adding a weight to the tail. After this adaptation period, weights of 30~50% body mass were added to the tails, and the weight or number of repetitions was steadily increased during the experimental period. For the first set, a weight of 70% body weight was added from the 2nd week until the end. An 80% weight was added for the 2nd and 3rd sets, a 90% weight was added for 6th to 8th sets. This protocol for rodent's resistance exercise was followed or modified from Kraemer et al.'s protocol [30]. There were 8 repetitions per set, with a 2 min resting period between sets. The frequency of the ladder-climbing exercise was 3 days per week [31,32].

#### **Tissue extraction and analysis**

After 8 weeks of the treatment, the rats were given a 48 h resting period to eliminate any last-bout exercise effect. The rats were then fasted for 12 h, and anesthetized using Zoletil 50 (10 mg/ kg body weight, Virbac Korea) and Rumpun 2% (0.04 ml/kg, Bayer Korea). First, the abdominal cavity was opened and blood was extracted from the abdominal arteries. The heart was then extracted after completely removing the blood via perfusion using phosphate-buffered saline. The weight was measured once

after removing the water contents, and then the heart was snapfrozen and stored at -80°C until protein analysis. The extracted heart was homogenized using ice-cold buffer [250 mM sucrose, 10 mM HEPES/1 mM ethylenediaminetetraacetic acid (EDTA, pH 7.4), 1 mM Pefabloc (Roche), 1 mM EDTA, 1 mM NaF, 1 µg/ ml aprotinin, 1 µg/ml leupeptin, 1 µg/ml pepstatin, 0.1 mM bpV (phen), 2 mg/ml glycerophosphate]. The homogenized sample was subjected to three freeze/thaw cycles and centrifuged (700 g, 10 min). The supernatant was collected, and the protein level was quantified through the Bradford assay [33]. The prepared sample was dissolved in Laemmli buffer, loaded onto a sodium dodecyl sulfate-polyacrylamide gel, electrophoresed, and then transferred to a nitrocellulose membrane. The membrane was blocked for 60 min using 5% non-fat dry milk and Tris-buffered saline+0.1% Tween 10 (TBST; pH 7.5), washed with TBST, and incubated with the following primary antibodies overnight at 4°C: phospho-protein kinase RNA-like endoplasmic reticulum kinase (PERK, Cell Signaling Technology, Beverly, MA, USA), PERK (Cell Signaling Technology, Beverly, MA, USA), peroxisome proliferator-activated receptor (PPAR) β/δ (Santa Cruz Biotechnology, Santa Cruz, CA, USA), C/EBP homologous protein (CHOP, Cell Signaling Technology, Beverly, MA, USA), 78-kDa glucose-regulated protein (GRP78, Santa Cruz Biotechnology, Santa Cruz, CA, USA), succinate dehydrogenase (Santa Cruz Biotechnology, Santa Cruz, CA, USA), cytochrome C (Santa Cruz Biotechnology, Santa Cruz, CA, USA), peroxisome proliferator-activated receptor-coactivator  $1\alpha$  (PGC- $1\alpha$ ), and  $\beta$ -actin (Calbiochem, Germany). After another wash with TBST, the samples were treated with the secondary antibody (anti-mouse or anti-rabbit, Santa Cruz Biotechnology, Santa Cruz, CA, USA) for 60 min. The bands were visualized using ECL (Genekhan Scientific, St. Louis, MO, USA), and the relative intensity of the bands was assessed using SigmaGel (Jandel Scientific Corp., Erkrath, Germany).

#### Statistical analysis

SPSS 10.0 was used to determine the statistical significance in the means and standard deviations of each variable among treatment groups with one-way analysis of variance. Post-hoc analyses were done using the Tukey method. Statistical significance was defined at p<0.05.

## RESULTS

#### Body weight and visceral fat weight

Feeding of the high-fat diet for 8 weeks resulted in a significant increase of both body weight and visceral fat weight for the middle-aged rats (p<0.05). Concurrent ladder climbing exercise with the high-fat diet did not result in a significant reduction of body weight, but did lead to a significant reduction of visceral fat weight (p<0.05). The groups that were switched to the chow diet for 8 weeks had significantly reduced body and visceral fat weights (p<0.05) compared to those of the high-fat diet group. The group on the chow diet with exercise showed a reduced body weight and visceral fat weight compared to the chow diet-only group, but the difference was not statistically significant (Fig. 1).

#### Mitochondrial biogenesis enzymes

The protein expression levels of cytochrome C and succinate dehydrogenase were measured to evaluate the effects of diet and/or exercise on mitochondrial biogenesis within the cardiac muscle. Eight weeks of ladder-climbing exercise significantly increased the mitochondrial enzyme levels (p<0.05). As shown in Fig. 2, the mitochondrial enzyme levels were particularly significantly higher in the chow diet+exercise group compared to those in the other three groups (p<0.05).

#### PPAR $\delta$ and PGC-1 $\alpha$ protein levels

To identify the potential mechanism by which the 8-week ladder-climbing exercise promoted mitochondrial biogenesis within the cardiac muscle of middle-aged rats, the protein levels



**Fig. 1. Body weight and visceral fat weight.** (A) Changes in body weight after 8-week intervention. <sup>a</sup>Significantly different from CD, CE (p<0.05). (B) Changes of visceral fat weight after 8-week intervention. <sup>+</sup>significantly different from CD, CE (p<0.05); \*Significantly different from CD, CE, FE (p<0.05). CD, normal chow diet; CE, chow diet+exercise; FD, high-fat diet only; FE, high-fat diet+exercise.



**Fig. 2. Expression levels of mitochondrial enzymes in the cardiac muscles.** <sup>+</sup>Significantly different from CD, FD (p<0.05); <sup>+</sup>Significantly different from CD, FD, FE (p<0.05). CD, normal chow diet; CE, chow diet+exercise; FD, high-fat diet only; FE, high-fat diet+exercise. SUD, succinate dehydrogenase (A); Cyto C (B), cytochrome C.

of PPAR $\delta$  and PGC-1 $\alpha$  were measured. The level of PPAR $\delta$  in the cardiac muscle was significantly increased in the high-fat diet+exercise group, but there were no significant changes in the other three groups. The level of PGC-1 $\alpha$  did not differ among any of the treatment groups (Fig. 3).

### **ER stress-related biomarkers**

To assess the effects of diet and/or exercise on ER stress within the cardiac muscle, the levels of phospho (p)-PERK, PERK, and CHOP proteins (markers of myocardial damage) and GRP78 protein (marker of myocardial protection) were measured. Maintenance on the high-fat diet resulted in a significant increase in p-PERK and CHOP protein levels (p<0.05), whereas the ladderclimbing exercise significantly reduced these levels down to values comparable to those of the chow diet group (p<0.05). More specifically, the CHOP protein level in the chow diet+exercise group was significantly lower than that of the chow diet-only group (p<0.05), representing the lowest level of CHOP protein overall. However, there was no significant difference in the GRP78 protein level among the groups (Fig. 4).



Fig. 3. Changes in PPAR $\beta$  (A) and PGC-1 $\alpha$  (B) protein expression in the cardiac muscle after 8-week diet and/or exercise intervention. \*Significantly different from CD, CE, FD (p<0.05). CD, normal chow diet; CE, chow diet+exercise; FD, high-fat diet only; FE, high-fat diet+exercise.

## DISCUSSION

In this study, the effects of intermittent ladder-climbing exercise training on mitochondrial biogenesis within the cardiac muscle of middle-aged rats were assessed by exploring the roles of a biogenesis-promoting mechanism (PGC-1 $\alpha$ ) and biogenesisinhibiting mechanism (ER stress). Recent studies have demonstrated that ER stress is a key factor contributing to the quantitative and functional degradation of the cardiac skeletal muscle during the aging process [34]. The ER is one of the organelles found in all eukaryotic cells, composed of a web-shape mixture of tubules, vesicles, and cisternae. The ER produces proteins that are distributed to various compartments of the cell. When an immature protein enters the ER that cannot be handled by the organelle owing to changes in the physiological or pathological environment or when the calcium within the ER is depleted, the ER cannot function properly, reaching a state of ER stress [35-37]. ER stress is strongly associated with various cardiovascular diseases such as heart failure, cardiotoxicity, arteriosclerosis, ischemia, and heart diseases [5], and many studies have been performed to investigate the potential of exercise training for reversing these changes in ER stress to prevent or treat these conditions. Murlasits et al. [38] placed 16-week-old rats on a 60 min running exercise routine at an intensity of VO<sub>2</sub>max >70% for 5 days, and found no



Fig. 4. Changes in (A) p-PERK, and (B) GRP78 (blue) and CHOP (red) protein expression levels in the cardiac muscle after 8 weeks of diet and/or exercise intervention. (A) <sup>+</sup>Significantly different from CD (p < 0.05); \*Significantly different from CD, CE, FE (p<0.05). (B) <sup>+</sup>Significantly different from CD, FE (p<0.05); \*Significantly different from CD, CE, FE (p<0.05). CD, normal chow diet; CE, chow diet+exercise; FD, high-fat diet only; FE, high-fat diet+exercise.

effect of this short-term endurance training on the expression of ER stress-related proteins in the cardiac muscle, such as GRP78, GRP94, or calreticulin. However, Gonzalez et al. [39] reported that endurance training for 3 months resulted in significantly increased expression of GRP78 within the soleus and extensor digitorum longus muscle. GRP78 and GRP94 are ER chaperones that protect myocardial cells against damage (i.e., cell death) induced by myocardial ischemia when activated [40,41]. However, the effect of exercise on ER stress is still unclear, requiring continuous and detailed studies to determine the underlying mechanisms.

High-intensity or long-term exercise training increases the number and quality of mitochondria by effectively stimulating mitochondrial biogenesis [10]. Although the reasons for this adaptation mechanism are still unclear, recent microarray analyses have suggested that this phenomenon is caused by the exercise-induced promotion of the transcription of mitochondrial proteins to consequently increase the expression of PGC-1 $\alpha$  [42-47]. However, in our study, in spite of no significant change in the level of myocardial PGC-1 $\alpha$  after 8 weeks of a ladder-climbing exercise, the PPAR $\delta$  level increased in the exercise groups, with a significant increase in the high-fat diet+exercise group compared to that in the other groups. PPARs are nuclear receptors that use free fatty acids as ligands for their activation [48], and show tissue-specific expression patterns. PPAR $\alpha$  and PPAR $\gamma$  are specifically

ring continuous ROS and ER mechanisms. catalase [25] j g increases the showed that rely stimulating 1 (GPx1), SC known that e as well as ant therefore, an ladder-climb

Nuclear respiratory factor-1 (NRF-1) is a key transcription factor for nuclear-encoded subunits of the four mitochondrial respiratory chain complexes and ATP synthase [53,54], and PPAR $\delta$  has been identified as a transcription factor for NRF-1 [54]. PPAR $\delta$ overexpression has been shown to increase SUO and Cyt C as well as NADH and ATP synthase prior to the increase in PGC-1 $\alpha$  expression [55]. Therefore, we suggest that intermittent ladderclimbing exercise training is supposed to increase mitochondrial biogenesis through the expression of PPAR $\delta$  without a significant change of PGC-1 $\alpha$  in aged myocardial tissue. The results of the present study showed that the higher levels of the myocardial damage-associated proteins p-PERK and CHOP in the cardiac muscle of high fat diet-induced obese middle-aged

expressed in the liver and fat tissue, respectively, and PPAR $\delta$  ex-

pression is extremely high in tissues with relatively high metabolic

activity, including the muscles, liver, and fat tissues [49]. Previous

studies [50-52] have demonstrated that PPAR $\delta$  could play a role

in increasing running endurance and the oxidative metabolic

activity of the skeletal muscle in rats. Wang et al. [52] established transgenic mice with overexpression of PPAR $\delta$  in the skeletal muscle, which had increased levels of mitochondrial enzymes without a corresponding increase in the PGC-1 $\alpha$  mRNA level.

in the cardiac muscle of high fat diet-induced obese middle-aged rats could be significantly reduced by 8 weeks of intermittent ladder-climbing exercise training. High fat diet increases the production of mitochondrial ROS which is clearly associated with insulin resistance [56-59]. Reactive oxygen species (ROS) has been identified as crucial regulators of ER function and unfolded protein response activation (UPR), ER stress occur concurrently with an increased ROS production [60]. ROS modulates the cross-talk between ER and mitochondria during apoptosis caused by ER stress [61,62]. Previous study has shown that PERK, a key ER stress sensor of the UPR, is an essential component of mitochondria-associated ER membrane (MAMs) [9]. MAMs were established a physical and functional connection between ER and mitochondria [63]. Thus, an increased PERK phosphorylation in cardiac muscle due to chronic high fat diet involved in an increase ROS and ER stress. Superoxide dismutase2 (SOD2) [24] and catalase [25] promoter has PPAR response element, Fan et al. [26] showed that PPARS activation increases Glutathione Peroxidase 1 (GPx1), SOD3 and catalase gene in skeletal muscle. It is well known that exercise training decreases ROS and increases PPAR\delta as well as antioxidants in cardiac muscle (REF). In present study, therefore, an increased PPAR $\delta$  in cardiac muscle by intermittent ladder-climbing exercise training of rat resulted in a decrease PERK/CHOP pathway that is associate with ROS [9].

This reduction was even greater in the group with lower caloric intake owing to switching from the high-fat diet to chow diet with concurrent exercise training. These results indicate that intermittent ladder-climbing exercise training combined with a reduced caloric intake can attenuate the levels of ER stress-related proteins that would induce myocardial damage. However, there was no influence of exercise or diet on GRP78 levels, which is a protein



Fig. 5. Effect of ladder-climbing exercise training on mitochondrial biogenesis and ER stress of the cardiac muscle. We put forward a hypothesis that the concurrent stimulation from exercise training is more effective through inhibition of PERK and CHOP expression or the activation of PGC-1 $\alpha$  and PPARb in protecting the cardiac muscle compared to a weight loss scheme that is entirely dependent on diet and reduced caloric intake. However, we could not suggest the significant relationship between PGC-1 $\alpha$  and PPAR $\beta$  and the activation GRP78 expression.

related to myocardial protection. Previous studies reported that acute exercise significantly increased the GRP78 level in the liver, but the effects on other tissues were not consistent [64]. Similarly, in our study, there was no change in GRP78 expression in the cardiac tissue, although changes are possible in other tissues.

The results of the present study suggest that changes in ER stress-related factors caused by intermittent high-intensity training in the context of aging do not play a substantial role in the activation of mitochondrial biogenesis, or that these changes are mutually independent of each other. However, considering previous findings suggesting that ER stress can lead to the suppression of mitochondrial biogenesis [65], it is crucial to clearly identify this potential association through additional analyses of reactive oxygen species induction and defense-related factors [66]. Moreover, the exercise routine used in this study, i.e., adding weights to the tail with ladder climbing, is considered an intermittent highintensity type of training, and it is possible that persistence in the stimulation from exercise was not sufficient to alter the PGC-1 $\alpha$ level in the cardiac muscle. Finally, the use of middle-aged rats rather than young, healthy rats should also be taken into consideration when interpreting these results. Therefore, along with age- and tissue-specific changes, associations between ER stress, PPAR $\delta$ , and PGC-1 $\alpha$  should be identified through future studies.

One particularly interesting finding of this study was that switching the high fat-induced obese rats to a chow diet for 8 weeks caused a significant decrease in body weight and visceral fat weight, but did not affect myocardial mitochondrial biogenesis. Civitarese et al. [67] analyzed the effect of diet (i.e., reduced caloric intake) or endurance training over 6 months on the mitochondrial biogenesis of the skeletal muscle in 36 obese human subjects, and claimed that reduced caloric intake improved mitochondrial function. However, conflicting results have emerged from animal studies evaluating the effects of reduced caloric intake on mitochondrial biogenesis of the skeletal muscle [68-71]. Furthermore, even among studies on humans, there are many reports showing no increase in the activation of oxidative enzymes in obese subjects after weight loss [72,73].

The results indicated that 8 weeks of a ladder-climbing exercise increased mitochondrial biogenesis within the cardiac muscle of middle-aged obese rats. Although there was no difference in the mitochondrial biogenesis-promoting mechanism (i.e., the PGC-1 $\alpha$  protein level) among the different groups, 8 weeks of the ladder-climbing exercise significantly reduced ER stress-related factors, which are in turn biogenesis-inhibiting factors (Fig. 5). Although our data are not sufficient to draw any conclusion on these effects at this point, at least for the cardiac muscle, the results nevertheless indicate that concurrent stimulation from exercise training is more effective in protecting the cardiac muscle compared to a weight loss scheme that is entirely dependent on diet and reduced caloric intake. However, in this study, we used obese, middle-aged rats rather than rats with cardiac defects. Therefore, future studies focusing on subjects or models with cardiac defects should help to provide a clearer explanation of the effects of exercise training and energy control diet on protection of the cardiac muscle.

## ACKNOWLEDGEMENTS

This research was supported by the Bisa Research Grant of Keimyung University in 2015.

# **CONFLICTS OF INTEREST**

The authors declare no conflicts of interest.

## REFERENCES

- 1. Higami Y, Shimokawa I. Apoptosis in the aging process. *Cell Tissue Res.* 2000;301:125-132.
- 2. Di Lisa F, Bernardi P. Mitochondrial function and myocardial aging. A critical analysis of the role of permeability transition. *Cardiovasc Res.* 2005;66:222-232.
- Rimbaud S, Garnier A, Ventura-Clapier R. Mitochondrial biogenesis in cardiac pathophysiology. *Pharmacol Rep.* 2009;61:131-138.
- 4. Palomer X, Capdevila-Busquets E, Botteri G, Salvadó L, Barroso E, Davidson MM, Michalik L, Wahli W, Vázquez-Carrera M. PPARβ/ δ attenuates palmitate-induced endoplasmic reticulum stress and induces autophagic markers in human cardiac cells. *Int J Cardiol.* 2014;174:110-118.

- 5. Colgan SM, Tang D, Werstuck GH, Austin RC. Endoplasmic reticulum stress causes the activation of sterol regulatory element binding protein-2. *Int J Biochem Cell Biol.* 2007;39:1843-1851.
- 6. Myoishi M, Hao H, Minamino T, Watanabe K, Nishihira K, Hatakeyama K, Asada Y, Okada K, Ishibashi-Ueda H, Gabbiani G, Bochaton-Piallat ML, Mochizuki N, Kitakaze M. Increased endoplasmic reticulum stress in atherosclerotic plaques associated with acute coronary syndrome. *Circulation*. 2007;116:1226-1233.
- 7. Wu J, Kaufman RJ. From acute ER stress to physiological roles of the Unfolded Protein Response. *Cell Death Differ*. 2006;13:374-384.
- Nisha VM, Anusree SS, Priyanka A, Raghu KG. Apigenin and quercetin ameliorate mitochondrial alterations by tunicamycininduced ER stress in 3T3-L1 adipocytes. *Appl Biochem Biotechnol.* 2014;174:1365-1375.
- 9. Verfaillie T, Rubio N, Garg AD, Bultynck G, Rizzuto R, Decuypere JP, Piette J, Linehan C, Gupta S, Samali A, Agostinis P. PERK is required at the ER-mitochondrial contact sites to convey apoptosis after ROS-based ER stress. *Cell Death Differ*. 2012;19:1880-1891.
- Holloszy JO. Regulation by exercise of skeletal muscle content of mitochondria and GLUT4. J Physiol Pharmacol. 2008;59 Suppl 7:5-18.
- Terada S, Yokozeki T, Kawanaka K, Ogawa K, Higuchi M, Ezaki O, Tabata I. Effects of high-intensity swimming training on GLUT-4 and glucose transport activity in rat skeletal muscle. *J Appl Physiol* (1985). 2001;90:2019-2024.
- Coggan AR, Spina RJ, King DS, Rogers MA, Brown M, Nemeth PM, Holloszy JO. Skeletal muscle adaptations to endurance training in 60- to 70-yr-old men and women. *J Appl Physiol (1985)*. 1992;72:1780-1786.
- Kohrt WM, Malley MT, Coggan AR, Spina RJ, Ogawa T, Ehsani AA, Bourey RE, Martin WH 3rd, Holloszy JO. Effects of gender, age, and fitness level on response of VO2max to training in 60-71 yr olds. J Appl Physiol (1985). 1991;71:2004-2011.
- 14. Short KR. Changes of deleted mtDNA after endurance exercise. *Eur J Appl Physiol*. 2003;90:223.
- 15. Jubrias SA, Esselman PC, Price LB, Cress ME, Conley KE. Large energetic adaptations of elderly muscle to resistance and endurance training. *J Appl Physiol (1985)*. 2001;90:1663-1670.
- Balagopal P, Schimke JC, Ades P, Adey D, Nair KS. Age effect on transcript levels and synthesis rate of muscle MHC and response to resistance exercise. *Am J Physiol Endocrinol Metab.* 2001;280:E203-208.
- Hasten DL, Pak-Loduca J, Obert KA, Yarasheski KE. Resistance exercise acutely increases MHC and mixed muscle protein synthesis rates in 78-84 and 23-32 yr olds. *Am J Physiol Endocrinol Metab.* 2000;278:E620-626.
- Menshikova EV, Ritov VB, Fairfull L, Ferrell RE, Kelley DE, Goodpaster BH. Effects of exercise on mitochondrial content and function in aging human skeletal muscle. *J Gerontol A Biol Sci Med Sci.* 2006;61:534-540.
- Kavazis AN, McClung JM, Hood DA, Powers SK. Exercise induces a cardiac mitochondrial phenotype that resists apoptotic stimuli. *Am J Physiol Heart Circ Physiol*. 2008;294:H928-935.
- Hamilton KL, Powers SK, Sugiura T, Kim S, Lennon S, Tumer N, Mehta JL. Short-term exercise training can improve myocardial tolerance to I/R without elevation in heat shock proteins. *Am J Physiol Heart Circ Physiol.* 2001;281:H1346-1352.
- 21. Morán M, Delgado J, González B, Manso R, Megías A. Responses of

rat myocardial antioxidant defences and heat shock protein HSP72 induced by 12 and 24-week treadmill training. *Acta Physiol Scand.* 2004;180:157-166.

- Radak Z, Taylor AW, Ohno H, Goto S. Adaptation to exerciseinduced oxidative stress: from muscle to brain. *Exerc Immunol Rev.* 2001;7:90-107.
- Starnes JW, Taylor RP. Exercise-induced cardioprotection: endogenous mechanisms. *Med Sci Sports Exerc.* 2007;39:1537-1543.
- 24. Ding G, Fu M, Qin Q, Lewis W, Kim HW, Fukai T, Bacanamwo M, Chen YE, Schneider MD, Mangelsdorf DJ, Evans RM, Yang Q. Cardiac peroxisome proliferator-activated receptor γ is essential in protecting cardiomyocytes from oxidative damage. *Cardiovasc Res.* 2007;76:269-279.
- Girnun GD, Domann FE, Moore SA, Robbins ME. Identification of a functional peroxisome proliferator-activated receptor response element in the rat catalase promoter. *Mol Endocrinol.* 2002;16:2793-2801.
- 26. Fan W, Waizenegger W, Lin CS, Sorrentino V, He MX, Wall CE, Li H, Liddle C, Yu RT, Atkins AR, Auwerx J, Downes M, Evans RM. PPARδ promotes running endurance by preserving glucose. *Cell Metab.* 2017;25:1186-1193.e4.
- 27. Cheang WS, Tian XY, Wong WT, Lau CW, Lee SS, Chen ZY, Yao X, Wang N, Huang Y. Metformin protects endothelial function in dietinduced obese mice by inhibition of endoplasmic reticulum stress through 5' adenosine monophosphate-activated protein kinaseperoxisome proliferator-activated receptor  $\delta$  pathway. *Arterioscler Thromb Vasc Biol.* 2014;34:830-836.
- 28. Cheang WS, Wong WT, Zhao L, Xu J, Wang L, Lau CW, Chen ZY, Ma RC, Xu A, Wang N, Tian XY, Huang Y. PPARδ is required for exercise to attenuate endoplasmic reticulum stress and endothelial dysfunction in diabetic mice. *Diabetes*. 2017;66:519-528.
- 29. Hancock CR, Han DH, Chen M, Terada S, Yasuda T, Wright DC, Holloszy JO. High-fat diets cause insulin resistance despite an increase in muscle mitochondria. *Proc Natl Acad Sci U S A*. 2008;105:7815-7820.
- 30. Kraemer WJ, Flanagan SD, Volek JS, Nindl BC, Vingren JL, Dunn-Lewis C, Comstock BA, Hooper DR, Szivak TK, Looney DP, Maresh CM, Hymer WC. Resistance exercise induces region-specific adaptations in anterior pituitary gland structure and function in rats. J Appl Physiol (1985). 2013;115:1641-1647.
- Lee S, Barton ER, Sweeney HL, Farrar RP. Viral expression of insulin-like growth factor-I enhances muscle hypertrophy in resistancetrained rats. *J Appl Physiol* (1985). 2004;96:1097-1104.
- 32. Jung S, Ahn N, Kim S, Byun J, Joo Y, Kim S, Jung Y, Park S, Hwang I, Kim K. The effect of ladder-climbing exercise on atrophy/hypertrophy-related myokine expression in middle-aged male Wistar rats. J Physiol Sci. 2015;65:515-521.
- Bradford MM. A rapid and sensitive method for the quantitation of microgram quantities of protein utilizing the principle of proteindye binding. *Anal Biochem.* 1976;72:248-254.
- 34. Ozcan U, Cao Q, Yilmaz E, Lee AH, Iwakoshi NN, Ozdelen E, Tuncman G, Görgün C, Glimcher LH, Hotamisligil GS. Endoplasmic reticulum stress links obesity, insulin action, and type 2 diabetes. *Science*. 2004;306:457-461.
- 35. Mori K. Tripartite management of unfolded proteins in the endoplasmic reticulum. *Cell*. 2000;101:451-454.
- 36. Oyadomari S, Koizumi A, Takeda K, Gotoh T, Akira S, Araki E,

Mori M. Targeted disruption of the Chop gene delays endoplasmic reticulum stress-mediated diabetes. *J Clin Invest.* 2002;109:525-532.

- Schröder M, Kaufman RJ. The mammalian unfolded protein response. Annu Rev Biochem. 2005;74:739-789.
- Murlasits Z, Lee Y, Powers SK. Short-term exercise does not increase ER stress protein expression in cardiac muscle. *Med Sci Sports Ex*erc. 2007;39:1522-1528.
- González B, Hernando R, Manso R. Stress proteins of 70 kDa in chronically exercised skeletal muscle. *Pflugers Arch*. 2000;440:42-49.
- 40. Vitadello M, Penzo D, Petronilli V, Michieli G, Gomirato S, Menabò R, Di Lisa F, Gorza L. Overexpression of the stress protein Grp94 reduces cardiomyocyte necrosis due to calcium overload and simulated ischemia. *FASEB J.* 2003;17:923-925.
- 41. Zhang J, Jiang Y, Jia Z, Li Q, Gong W, Wang L, Wei D, Yao J, Fang S, Xie K. Association of elevated GRP78 expression with increased lymph node metastasis and poor prognosis in patients with gastric cancer. *Clin Exp Metastasis.* 2006;23:401-410.
- Baar K, Wende AR, Jones TE, Marison M, Nolte LA, Chen M, Kelly DP, Holloszy JO. Adaptations of skeletal muscle to exercise: rapid increase in the transcriptional coactivator PGC-1. *FASEB J*. 2002;16:1879-1886.
- 43. Goto M, Terada S, Kato M, Katoh M, Yokozeki T, Tabata I, Shimokawa T. cDNA Cloning and mRNA analysis of PGC-1 in epitrochlearis muscle in swimming-exercised rats. *Biochem Biophys Res Commun.* 2000;274:350-354.
- 44. Higashida T, Kreipke CW, Rafols JA, Peng C, Schafer S, Schafer P, Ding JY, Dornbos D 3rd, Li X, Guthikonda M, Rossi NF, Ding Y. The role of hypoxia-inducible factor-1a, aquaporin-4, and matrix metalloproteinase-9 in blood-brain barrier disruption and brain edema after traumatic brain injury. *J Neurosurg.* 2011;114:92-101.
- 45. Terada S, Goto M, Kato M, Kawanaka K, Shimokawa T, Tabata I. Effects of low-intensity prolonged exercise on PGC-1 mRNA expression in rat epitrochlearis muscle. *Biochem Biophys Res Commun.* 2002;296:350-354.
- 46. Terada S, Tabata I. Effects of acute bouts of running and swimming exercise on PGC-1alpha protein expression in rat epitrochlearis and soleus muscle. *Am J Physiol Endocrinol Metab.* 2004;286:E208-216.
- 47. Wende AR, Huss JM, Schaeffer PJ, Giguère V, Kelly DP. PGC-1alpha coactivates PDK4 gene expression via the orphan nuclear receptor ERRalpha: a mechanism for transcriptional control of muscle glucose metabolism. *Mol Cell Biol.* 2005;25:10684-10694.
- Wang YX, Lee CH, Tiep S, Yu RT, Ham J, Kang H, Evans RM. Peroxisome-proliferator-activated receptor delta activates fat metabolism to prevent obesity. *Cell*. 2003;113:159-170.
- 49. Braissant O, Foufelle F, Scotto C, Dauça M, Wahli W. Differential expression of peroxisome proliferator-activated receptors (PPARs): tissue distribution of PPAR-alpha, -beta, and -gamma in the adult rat. *Endocrinology*. 1996;137:354-366.
- Luquet S, Lopez-Soriano J, Holst D, Fredenrich A, Melki J, Rassoulzadegan M, Grimaldi PA. Peroxisome proliferator-activated receptor delta controls muscle development and oxidative capability. *FASEB J*. 2003;17:2299-2301.
- 51. Tanaka T, Yamamoto J, Iwasaki S, Asaba H, Hamura H, Ikeda Y, Watanabe M, Magoori K, Ioka RX, Tachibana K, Watanabe Y, Uchiyama Y, Sumi K, Iguchi H, Ito S, Doi T, Hamakubo T, Naito M, Auwerx J, Yanagisawa M, Kodama T, Sakai J. Activation of peroxisome proliferator-activated receptor delta induces fatty acid beta-

oxidation in skeletal muscle and attenuates metabolic syndrome. *Proc Natl Acad Sci U S A*. 2003;100:15924-15929.

- 52. Wang YX, Zhang CL, Yu RT, Cho HK, Nelson MC, Bayuga-Ocampo CR, Ham J, Kang H, Evans RM. Regulation of muscle fiber type and running endurance by PPARdelta. *PLoS Biol.* 2004;2:e294.
- Kelly DP, Scarpulla RC. Transcriptional regulatory circuits controlling mitochondrial biogenesis and function. *Genes Dev.* 2004;18: 357-368.
- 54. Scarpulla RC. Nuclear activators and coactivators in mammalian mitochondrial biogenesis. *Biochim Biophys Acta*. 2002;1576:1-14.
- 55. Koh JH, Hancock CR, Terada S, Higashida K, Holloszy JO, Han DH. PPAR $\beta$  is essential for maintaining normal levels of PGC-1 $\alpha$  and mitochondria and for the increase in muscle mitochondria induced by exercise. *Cell Metab.* 2017;25:1176-1185.e5.
- 56. Anderson EJ, Lustig ME, Boyle KE, Woodlief TL, Kane DA, Lin CT, Price JW 3rd, Kang L, Rabinovitch PS, Szeto HH, Houmard JA, Cortright RN, Wasserman DH, Neufer PD. Mitochondrial H2O2 emission and cellular redox state link excess fat intake to insulin resistance in both rodents and humans. *J Clin Invest.* 2009;119:573-581.
- 57. Barbosa MR, Sampaio IH, Teodoro BG, Sousa TA, Zoppi CC, Queiroz AL, Passos MA, Alberici LC, Teixeira FR, Manfiolli AO, Batista TM, Cappelli AP, Reis RI, Frasson D, Kettelhut IC, Parreirase-Silva LT, Costa-Neto CM, Carneiro EM, Curi R, Silveira LR. Hydrogen peroxide production regulates the mitochondrial function in insulin resistant muscle cells: effect of catalase overexpression. *Biochim Biophys Acta*. 2013;1832:1591-1604.
- 58. Ding H, Heng B, He W, Shi L, Lai C, Xiao L, Ren H, Mo S, Su Z. Chronic reactive oxygen species exposure inhibits glucose uptake and causes insulin resistance in C2C12 myotubes. *Biochem Biophys Res Commun.* 2016;478:798-803.
- Henriksen EJ. Effects of H2O2 on insulin signaling the glucose transport system in mammalian skeletal muscle. *Methods Enzymol.* 2013;528:269-278.
- Malhotra JD, Kaufman RJ. Endoplasmic reticulum stress and oxidative stress: a vicious cycle or a double-edged sword? *Antioxid Redox Signal.* 2007;9:2277-2293.
- 61. Pinton P, Giorgi C, Pandolfi PP. The role of PML in the control of apoptotic cell fate: a new key player at ER-mitochondria sites. *Cell Death Differ*. 2011;18:1450-1456.
- 62. Sano R, Annunziata I, Patterson A, Moshiach S, Gomero E, Opferman J, Forte M, d'Azzo A. GM1-ganglioside accumulation at the mitochondria-associated ER membranes links ER stress to Ca<sup>2+</sup>-dependent mitochondrial apoptosis. *Mol Cell*. 2009;36:500-511.
- 63. Hayashi T, Rizzuto R, Hajnoczky G, Su TP. MAM: more than just a housekeeper. *Trends Cell Biol.* 2009;19:81-88.
- 64. Hunter RB, Mitchell-Felton H, Essig DA, Kandarian SC. Expression of endoplasmic reticulum stress proteins during skeletal muscle disuse atrophy. *Am J Physiol Cell Physiol*. 2001;281:C1285-1290.
- 65. Chen Y, Pandiri I, Joe Y, Kim HJ, Kim SK, Park J, Ryu J, Cho GJ, Park JW, Ryter SW, Chung HT. Synergistic effects of cilostazol and probucol on ER stress-induced hepatic steatosis via heme oxygenase-1-dependent activation of mitochondrial biogenesis. Oxid Med Cell Longev. 2016;2016:3949813.
- 66. St-Pierre J, Drori S, Uldry M, Silvaggi JM, Rhee J, Jäger S, Handschin C, Zheng K, Lin J, Yang W, Simon DK, Bachoo R, Spiegelman BM. Suppression of reactive oxygen species and neurodegeneration

by the PGC-1 transcriptional coactivators. Cell. 2006;127:397-408.

- Civitarese AE, Carling S, Heilbronn LK, Hulver MH, Ukropcova B, Deutsch WA, Smith SR, Ravussin E. Calorie restriction increases muscle mitochondrial biogenesis in healthy humans. *PLoS Med.* 2007;4:e76.
- Ballor DL. Effect of dietary restriction and/or exercise on 23-h metabolic rate and body composition in female rats. *J Appl Physiol* (1985). 1991;71:801-806.
- 69. Dulloo AG, Girardier L. 24 hour energy expenditure several months after weight loss in the underfed rat: evidence for a chronic increase in whole-body metabolic efficiency. *Int J Obes Relat Metab Disord*. 1993;17:115-123.
- 70. McCarter RJ, Palmer J. Energy metabolism and aging: a lifelong

study of Fischer 344 rats. Am J Physiol. 1992;263:E448-452.

- Nisoli E, Tonello C, Cardile A, Cozzi V, Bracale R, Tedesco L, Falcone S, Valerio A, Cantoni O, Clementi E, Moncada S, Carruba MO. Calorie restriction promotes mitochondrial biogenesis by inducing the expression of eNOS. *Science*. 2005;310:314-317.
- 72. Kempen GI, Miedema I, van den Bos GA, Ormel J. Relationship of domain-specific measures of health to perceived overall health among older subjects. *J Clin Epidemiol.* 1998;51:11-18.
- Simoneau JA, Veerkamp JH, Turcotte LP, Kelley DE. Markers of capacity to utilize fatty acids in human skeletal muscle: relation to insulin resistance and obesity and effects of weight loss. *FASEB J.* 1999;13:2051-2060.