



# **The Bile Acid Membrane Receptor TGR5 in Cancer: Friend or Foe?**

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**Abstract:** The G-protein-coupled bile acid receptor, Gpbar1 or TGR5, is characterized as a membrane receptor specifically activated by bile acids. A series of evidence shows that TGR5 induces protein kinase B (AKT), nuclear factor kappa-B (NF-κB), extracellular regulated protein kinases (ERK1/2), signal transducer and activator of transcription 3 (STAT3), cyclic adenosine monophosphate (cAMP), Ras homolog family member A (RhoA), exchange protein activated by cAMP (Epac), and transient receptor potential ankyrin subtype 1 protein (TRPA1) signaling pathways, thereby regulating proliferation, inflammation, adhesion, migration, insulin release, muscle relaxation, and cancer development. TGR5 is widely distributed in the brain, lung, heart, liver, spleen, pancreas, kidney, stomach, jejunum, ileum, colon, brown adipose tissue (BAT), white adipose tissue (WAT), and skeletal muscle. Several recent studies have demonstrated that TGR5 exerts inconsistent effects in different cancer cells upon activating via TGR5 agonists, such as INT-777, ursodeoxycholic acid (UDCA), and taurolithocholic acid (TLCA). In this review, we discuss both the 'friend' and 'foe' features of TGR5 by summarizing its tumor-suppressing and oncogenic functions and mechanisms.

Keywords: TGR5; bile acids; cancer; oncogenic functions

### 1. Introduction

G-protein-coupled receptors (GPCRs) are a large family of receptors that are abundantly distributed in different tissues in mammals, with numerous significant effects, such as anti-inflammatory effects, immune regulation, regulation of energy metabolism, regulation of glucose metabolism, anti-aging effects, protection against radiation, anti-cancer effects, and neuroprotection [1–4]. GPCRs have the same seven transmembrane domains on the cell membrane [5]. Upon GPCRs being activated by ligands of drugs, lipids, and hormones, extracellular signals are transduced into intracellular signals by cascade amplification, thereby exerting important effects and regulating physiological and pathological processes in human beings and animals [6]. These key roles of GPCRs are related to cellular signaling pathways, such as protein kinase B (AKT), nuclear factor kappa-B (NF- $\kappa$ B), extracellular regulated protein kinases (ERK1/2), signal transducer and activator of transcription 3 (STAT3), and cyclic adenosine monophosphate (cAMP) [7,8].

The G-protein-coupled bile acid receptor (Gpbar1), also called the TGR5 receptor or M-BAR, belongs to the family of GPCRs. It is reported that TGR5 is widely expressed in different cells and tissues, for instance in the endocrine gland, adipocytes, muscles, immune organs, spinal cord, and the enteric nervous system [9]. In recent years, a large number of findings have revealed that TGR5 is not only the second bile acid receptor,



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**Copyright:** © 2022 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (https:// creativecommons.org/licenses/by/ 4.0/). but also a metabolic regulator taking part in energy homeostasis, bile acid homeostasis, and glucose metabolism via NF- $\kappa$ B, AKT, and ERK signaling pathways [10–12]. Upon the binding of ligands in TGR5 domains, activated TGR5 transduces the special extracellular signal into an intracellular signal to regulate downstream cascades, such as 6 $\alpha$ -ethyl-23(*S*)-methyl-cholic acid (6-EMCA, INT-777), chenodeoxycholic acid (CDCA), lithocholic acid (LCA), taurochenodeoxycholic acid (TCDCA), ursodeoxycholic acid (UDCA), and tauroursodeoxycholic acid (TUDCA) [13–18] (Figure 1).



Figure 1. The chemical structures of the bile acids.

We comprehensively analyzed TGR5's affinity energy with a number of bile acids using Autodock software (Figure 2). The results show that TGR5's affinity energy with INT-777, CDCA, LCA, TCDCA, UDCA, and TUDCA is -8.8, -7.5, -6.7, -8.2, -7.5, and -7.7 kcal/mol, respectively. Our data demonstrate why INT-777 can be widely used to study TGR5. INT-777, as a special TGR5 agonist, reduced the severity of AP in mice, which was manifested as decreased pancreatic tissue damage as well as a decrease in the expression of serum enzymes (amylase and lipase), interleukin-1 beta (IL-1β), interleukin-6 (IL-6), tumor necrosis factor- $\alpha$  (TNF- $\alpha$ ), and necrosis-related proteins (RIP3 and p-MLKL) caused by inhibiting the reactive oxygen species/nucleotide-binding oligomerization domain (NOD)-like receptor containing the pyrin domain (3ROS/NLRP3) inflammasome pathway [19]. In 3T3-L1 cells, CDCA remarkably inhibits ligand-stimulated peroxisome proliferator-activated receptor  $\gamma$  (PPAR $\gamma$ ) transcriptional activity to decrease adipocyte differentiation [20]. LCA alleviates high-glucose-induced cardiac hypertrophy via enhancing sarcoplasmic/endoplasmic reticulum Ca<sup>2+</sup> ATPase 2a (SERCA2a) and phosphorylated phospholamban (PLN) expression in H9C2 cells, thereby activating TGR5 [21]. The activation of TGR5 by TCDCA decreases TNF- $\alpha$ , IL-1 $\beta$ , IL-6, interleukin-8 (IL-8), and interleukin-12 (IL-12) expression through the cAMP-PKA-cAMP response element-binding protein (CREB) and Raf1-CREB signaling pathways in NR8383 cells [22]. UDCA can increase the transforming growth factor- $\beta$  (TGF- $\beta$ ) ubiquitination level at the site of K135 by means of Hsc70-interacting protein (CHIP) and phosphorylate TGF- $\beta$  at the T282 site via the TGR5-cAMP-PKA axis, commonly causing antitumor immunity [23]. TUDCA decreases DNA-dependent protein kinase (DNA-PK), tumor suppressor P53-binding protein-1 (53BP-1), and DNA ligase IV expression levels to impair DNA damage in embryos via activating the TGR5 receptor, thereby reducing endoplasmic reticulum (ER) stress [24]. However, a large array of evidence has shown that TGR5 is implicated in fatty metabolism, stress regulation, inflammation, and immunity. Hence, numerous results have revealed that the activation of TGR5 can prevent the development and migration of certain tumors; however, TGR5 is the opponent in other tumors.



Figure 2. Molecular docking of TGR5 to bile acids.

#### 2. TGR5 in Lung Cancer

Lung cancer is the leading cause of cancer deaths worldwide. In China, lung cancer is also one of the most common malignant tumors, accounting for 25% of the death toll of malignant tumors [25]. Lung cancer includes non-small cell, small cell, mesothelioma, thymoma, and neuroendocrine tumors [26]. Tests to diagnose lung cancer mainly include imaging tests, sputum cytology, and taking tissue samples (biopsy). Nowadays, a series of drugs and treatment methods can be used for the therapy as well as diagnosis of lung cancer, such as radiotherapy, chemotherapy, immunochemotherapy, molecular targeted therapy, Chinese medicine therapy, sotorasib (AMG-510), rybrevant (amivantamab-vmjw), capmatinib (tabrecta), lorlatinib (lorbrena), libtayo (cemiplimab), tepotinib (tepmetko), cisplatin, cyclophosphamide, and 5-FU [27-32]. However, many drugs directly or indirectly lead to severe adverse reactions, such as nausea, vomiting, and constipation [33]. Currently, Chinese traditional medicine is more widely used to regulate and treat physiological and pathological processes, such as dihydroartemisinin (DHA), which regulates immune cell heterogeneity by triggering a cascade reaction of cyclin-dependent kinases (CDK) and mitogen-activated protein kinase (MAPK) phosphorylation [34]; salvia miltiorrhiza and pueraria lobata, two eminent herbs in Xin-Ke-Shu (XKS), which ameliorate myocardial ischemia partially by modulating the accumulation of free fatty acids in rats [35]; and a monomeric polysaccharide from polygonatum sibiricum, which improved cognitive function in a model of Alzheimer's disease by reshaping the gut microbiota [36]. More and more research results have revealed that TCDCA, as one of the main active ingredients of bile acids, after combining and activating TGR5 in lung cancer cells of H1299, can increase cAMP content and elevate phosphorylation levels of protein kinase A (PKA) and CREB [22]. In addition, PKA can phosphorylate many downstream kinases, such as Raf, glycogen synthase kinase-3 (GSK3), and focal adhesion kinase (FAK), while the activation of the cAMP-PKA-CREB signaling pathway may promote lung cancer cell growth, migration, invasion, and metabolism [37]. Verma demonstrated that the expression of the dominant negative form of PKA (dnPKA) or treatment with the PKA-specific inhibitor, H89, greatly reduced the growth of small cell lung cancer (SCLC) tumors [38]. Hye-Sook Seo found that the expression levels of mRNA and CREB and phosphorylated CREB (pCREB) were significantly higher in most of the non-small cell lung cancer (NSCLC) cell lines than

in the normal human tracheobronchial epithelial (NHTBE) cells and adjacent normal lung tissue, respectively [39]. It is well known that NF- $\kappa$ B is a downstream protein of CREB, and the suppression of NF- $\kappa$ B activity can remarkably augment the development of tumor specimens in SCLC tumors. Additionally, a previous study by the present authors demonstrated that TGR5 activation strongly inhibited the Janus kinase-2 (JAK2)-STAT3 signaling pathway in vitro and in vivo. Jiang reported that the activation of TGR5 on membranes in NSCLC cell lines mediates the JAK2-STAT3 signaling pathway, exacerbating tumor cell development and migration [40]. Altogether, our summarized results strongly suggest that the cAMP-PKA-CREB and JAK2-STAT3 signaling pathways induced by TGR5 are promising therapeutic strategies and predict the effects of therapy for lung cancer (Figure 3).



Figure 3. TGR5 in lung cancer.

#### 3. TGR5 in Liver Cancer

Liver cancer is generally classified as primary or secondary. Primary cancer is commonly manifested in a malignant tumor beginning in the cells of the liver [41]. Researchers have generally divided primary cancer into three types: hepatocellular carcinoma (HCC) (hepatoma), cholangiocarcinoma (CCA), and angiosarcoma [42–44]. The latest statistics show that more than 900,000 people are diagnosed with HCC every year around the world, and HCC is a leading cause of cancer death worldwide, accounting for more than 800,000 deaths each year—thus ranking as the third most common cause of cancer-related death. HCC is more common in people who drink large amounts of alcohol and who have an accumulation of fat in the liver. Nowadays, blood tests, image tests (CT and MRI), and liver biopsies are usually used to diagnose HCC [45]. These methods can produce a range of adverse reactions in liver cancer patients. HCC treatments include surgery, liver transplant surgery, destroying cancer cells with heat or cold, delivering chemotherapy or radiation directly to cancer cells, radiation therapy, targeted drug therapy, immunotherapy, and clinical trials [46]. However, a great number of treatment and diagnosis methods of HCC definitely bring about several adverse reactions, such as an abdominal mass or lump, right-sided abdominal pain, right-shoulder blade pain, jaundice, itching, bloating, shortness of breath, unintentional weight loss and gain, loss of appetite, nausea and vomiting, fatigue and weakness, fever, and a general feeling of being unwell [47]. With a large number of people worldwide being infected by COVID-19, many traditional Chinese

medicines have been used against the coronavirus disease (COVID-19), such as Jinhua Qinggan granule, Lianhua Qingwen capsule, Xuebijing injection, a lung cleansing and detoxifying decoction, and Huashibaidu formula, etc. [47–51]. In recent years, bile acids, as one of the main active components of bile, have been used for experimental research into liver cancer via multiple pathways. Mounting research results have demonstrated that TGR5 is one of the most common therapy targets against HCC via the regulation of energy homeostasis and glucose metabolism. The  $\alpha$ 7-nicotinic acetylcholine receptor ( $\alpha$ 7-nAChR) is an oncogene and risk factor for HCC. In HCC, knocking-down  $\alpha$ 7-nAChR can reduce cell viability, inhibit cellular proliferation, attenuate migration and invasion, and diminish the sphere-formation ability of HCC, which is related to the phosphorylation of JAK2, STAT3, Ras homolog family member A (RhoA), Rho-associated protein kinase-1 (ROCK1), matrix metallopeptidase-2 (MMP2), and matrix metallopeptidase-9 (MMP9) in HCC-mediated by TGR5 [52]. However, the expression of matrix metalloproteinases (MMPs) is also higher in TGR5-/-mice, which may promote the development and migration of HCC [53]. In vivo, a lack of TGR5 in mice can promote diethylnitrosamine (DEN)-induced hepatocyte death, compensatory proliferation, and the gene expression of certain inflammatory cytokines, matrix metalloproteinasesacute, and liver carcinogenesis to a greater extent than wild-type (WT) mice; in vitro, TGR5 activation strongly inhibited the proliferation and migration of HCC via suppressing STAT3 signaling, and its DNA binding activity [54]. In summary, TGR5 receptor could be a new potential biomarker for the diagnosis and treatment of HCC in the future (Figure 4).



Figure 4. TGR5 in liver cancer.

### 4. TGR5 in Gastric Cancer

Although the occurrence of gastric cancer, also commonly called stomach cancer, has declined significantly over the past two decades, it is still among the most prevalent cancers worldwide [55,56]. According to recent research, the occurrence of gastric cancer relates to diet [57]. Helicobacter pylori infecting people's stomachs could become a special biomarker for the diagnosis and therapy of gastric cancer [58]. It can directly cause chronic gastric inflammation, which slowly progresses to atrophy, metaplasia, dysplasia, and gastric cancer when H. pylori enters into the stomach [59]. There are 1,000,000 people diagnosed with gastric cancer worldwide every year, who are mainly distributed in countries in East Asia, Eastern Europe, and Central and South America [45]. Gastric cancer patients normally experience bloating after eating, heartburn, a lack of appetite, nausea, and an upset stomach [60].

Tumor removal and chemotherapy are the foremost treatment methods for gastric cancer patients [61]. However, these methods pose a risk and garner great cost for gastric cancer patients and their families. TGR5 can be activated by traditional Chinese medicines, thereby mediating a great number of signal transduction pathways for the treatment of gastric cancer, such as 23(S)-mCDCA. TGR5 overexpression in the gastric cancer cell line SGC7901 activated by 23(S)-mCDCA greatly inhibited the gene expression of interferon-inducible protein 10 (IP10), TNF- $\alpha$ , and chemoattractant protein-1 (MCP1) induced by NF- $\kappa$ B, as well as LPS. Sirtuin-1 (SIRT1), a class-III protein deacetylase, regulates cell death and metabolism via multiple physiological effects, such as DNA damage, anti-inflammation, and cellular oxidative stress [62,63]. The activation of SIRT1 not only inhibited the mRNA expression of STAT3 and c-Myc, but also suppressed the phosphorylation of NF-κB p65 [64–66]. Thus, SIRT1 has a repressive function on gastric cancer via inhibiting the activation of STAT3 and NF- $\kappa$ B. However, TGR5 activation strongly antagonizes the STAT3 signal pathway through suppressing the phosphorylation of STAT3 and its transcription activity induced by LPS. The activation of TGR5 on the gastric cancer cell line SGC7901 by 23(S)-mCDCA significantly inhibited the downstream gene expression of STAT3, as well as MMP2, complement component 3 (C3), c-Myc, interleukin 6 receptor (IL-6R), epidermal growth factor receptor (EGFR), endothelial PAS domain protein 1 (EPAS), suppressor of cytokine signaling 3 (SOCS3), MMP7, and MMP14 [67]. In summary, TGR5 activation can inhibit the proliferation and migration of gastric cancer cells via the suppression of STAT3 and NF-κB signal pathways, and thus TGR5 could be used to diagnose and treat gastric cancer in the future (Figure 5).



Figure 5. TGR5 in gastric cancer.

# 5. TGR5 in Colorectal Cancer

Colorectal cancer (CRC), sometimes called colon cancer, results in 900,000 deaths every year and has been considered the fourth leading cause of mortality related to cancer diseases worldwide [45]. In 2020 in the USA, approximately 147,950 individuals were diagnosed with CRC and 53,200 died from the disease, including 17,930 cases and 3640 deaths in individuals aged younger than 50 years [68]. CRC development in patients can lead to the occurrence of ulcerative colitis and Crohn's disease with age [69]. Mounting evidence shows that the risk factors for CRC include a lack of regular physical activity, a diet low in fruit and vegetables, a low-fiber and high-fat diet, being overweight or obese, alcohol consumption, and tobacco use, etc. [70]. Today, surgery, radiation therapy, and chemotherapy are the key components of CRC therapy [71]. CRC therapy can cause a range of side effects, such as nausea, vomiting, loss of appetite, diarrhea, abdominal pain, etc. [68]. Bile acids are one of the main components of bile, exerting anti-inflammation and immune functions via the cAMP-PKA-CREB and Raf1-CREB signaling pathways mediated by TGR5 [22]. Continuous primary inflammation might trigger the production and development of CRC. In HCT116 cells, SW480 cells, and DSS-induced CRC mice, UDCA, as one of the main active components of bile, suppresses the malignant progression of CRC via TGR5 mediating the cAMP-PKA-RhoA signal pathway to antagonize Yes-associated protein (YAP) [72]. Moreover, LCA remarkably activates TGR5 to repress the production of pro-inflammation cytokines in the colon, decreasing the development and migration of CRC [73]. Taken together, TGR5 combined and activated by bile acids is considered as a novel treatment target for CRC (Figure 6).



Figure 6. TGR5 in colorectal cancer.

#### 6. TGR5 in Other Cancers

The membrane bile acid receptor TGR5 is a multiple-target receptor, and it can mediate several cell signal pathways to regulate itching, inflammation, proliferation, migration,

insulin release, monocyte adhesion, and muscle relaxation, such as NF-KB, ERK, AKT, STAT3, transient receptor potential ankyrin 1 (TRPA1), exchange protein activated by cAMP (Epac), and RhoA [4,74–76]. Therefore, TGR5 is not only a metabolism regulator, but also has multiple functions in other cancers, such as endometrial cancer, breast cancer, and pancreatic cancer. First, CDCA at low concentrations significantly inhibited Ishikawa cell growth by inducing a remarkable increase in cyclin D1 protein and mRNA expression via TGR5 mediating the ERK-CREB signal pathway, suggesting that CDCA activated the TGR5-dependent CREB signaling pathway to promote human endometrial cancer cell proliferation [77]. Second, in MCF-7 and MDA-MB-231 breast cancer cells, LCA exerted anti-proliferative and pro-apoptotic effects through TGR5 mediating the cAMP-PKA-CREP signal pathway, thereby negatively regulating the gene and protein expressions of P53 and Bcl-2 to suppress the migration and development of breast cancer—suggesting that TGR5 is a novel target for interventions in breast cancer [78]. In addition, LCA can increase oxidative stress in breast cancer via TGR5. In the breast cancer cell line MCF-7, LCA decreased nuclear factor-2 (NRF2) expression and increased Kelch-like ECH associating protein 1 (KEAP1) expression via the activation of TGR5 and constitutive androstane receptor (CAR); in breast cancer patients, the overexpression of inducible nitric oxide synthases (iNOS), neuronal nitric oxide synthases (nNOS), chimeric antigen receptor (CAR), KEAP1, NADPH oxidase 4 (NOX4), and TGR5 or the downregulation of nuclear factor erythroid 2-related factor 2 (NRF2) were correlated with better survival, except for triple negative cases; therefore, TGR5 activation by LCA significantly suppressed the proliferation of breast cancer cells via oxidative stress [79]. Moreover, in pancreatic cancer, TGR5 had a significantly higher expression in the cancerous tissues than the adjacent normal tissues (81.6% vs. 36.8%), and Cox proportional hazards regression analysis confirmed that TGR5 expression was an independent predictor of the overall survival of patients with pancreatic cancer (p = 0.019), suggesting that TGR5 might serve as an important therapeutic target for pancreatic cancer [80] (Figure 7).



Figure 7. TGR5 in other cancers.

# 7. Conclusions

Since its identification in 2002, TGR5 has been found to be ubiquitously expressed in humans and animals, and is known to activate various intracellular signaling pathways upon interaction with bile acids. It has been continuously reported that the activation of TGR5 by bile acids mediated the cAMP-PKA-CREB, JAK2-STAT3, cAMP-PKA-RhoA, and cAMP-PKA-CREP signaling pathways, affecting the proliferation and migration of lung cancer, liver cancer, gastric cancer, colorectal cancer, endometrial cancer, breast cancer, and pancreatic cancer through regulating some special gene and protein expression, such as Raf, GSK3, FAK, Yap, cMyc, IL-6R, EGFR, EPAS, SOCS3, MMP7, and MMP14. In this review, we found that TGR5-dependent signaling pathways can promote the development and migration of lung cancer, endometrial cancer, and pancreatic cancer; meanwhile, it can inhibit the proliferation and migration of liver cancer, gastric cancer, and breast cancer. Hence, TGR5 has double-regulatory functions in the development process of cancer.

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### References

- 1. Sun, L.; Ye, R.D. Role of G protein-coupled receptors in inflammation. Acta Pharmacol. Sin. 2012, 3, 342–350. [CrossRef] [PubMed]
- Zhou, H.; Zhou, S.; Shi, Y.; Wang, Q.; Wei, S.; Wang, P.; Cheng, F.; Auwerx, J.; Schoonjans, K.; Lu, L. TGR5/Cathepsin E signaling regulates macrophage innate immune activation in liver ischemia and reperfusion injury. *Am. J. Transplant.* 2021, *4*, 1453–1464. [CrossRef] [PubMed]
- 3. Chen, X.S.; Lou, G.Y.; Meng, Z.P.; Huang, W.D. TGR5: A Novel Target for Weight Maintenance and Glucose Metabolism. *J. Diabetes Res.* 2011, 2011, 853501. [CrossRef] [PubMed]
- 4. Guo, C.; Chen, W.D.; Wang, Y.D. TGR5, Not Only a Metabolic Regulator. Front. Physiol. 2016, 7, 646. [CrossRef] [PubMed]
- Baidya, M.; Chaturvedi, M.; Dwivedi-Agnihotri, H. Allosteric modulation of GPCR-induced β-arrestin trafficking and signaling by a synthetic intrabody. *Nat. Commun.* 2022, 13, 4634. [CrossRef] [PubMed]
- Liu, N.; Wang, Y.; Li, T.; Feng, X. G-Protein Coupled Receptors (GPCRs): Signaling Pathways, Characterization, and Functions in Insect Physiology and Toxicology. *Int. J. Mol. Sci.* 2021, 10, 5260. [CrossRef]
- National Center for Biotechnology Information. PubChem Pathway Summary for Pathway R-HSA-388396, GPCR Downstream Signalling, Source: Reactome. 2022. Available online: https://pubchem.ncbi.nlm.nih.gov/pathway/Reactome:R-HSA-388396 (accessed on 12 August 2022).
- Yu, H.; Lee, H.; Herrmann, A. Revisiting STAT3 signalling in cancer: New and unexpected biological functions. *Nat. Rev. Cancer* 2014, 14, 736–746. [CrossRef]
- Duboc, H.; Taché, Y.; Hofmann, A.F. The bile acid TGR5 membrane receptor: From basic research to clinical application. *Dig. Liver Dis.* 2014, 4, 302–312. [CrossRef]
- 10. Yang, H.; Luo, F.; Wei, Y.; Jiao, Y.; Qian, J.; Chen, S.; Gong, Y.; Tang, L. TGR5 protects against cholestatic liver disease via suppressing the NF-κB pathway and activating the Nrf2/HO-1 pathway. *Ann. Transl. Med.* **2021**, *14*, 1158. [CrossRef]
- 11. Shotaro, M.; Takashi, S.; Yuki, Y.; Makoto, S.; Ryuichiro, S. Maslinic acid activates mTORC1 and human TGR5 and induces skeletal muscle hypertrophy. *Biosci. Biotech. Bioch.* 2021, *11*, 2311–2321.
- 12. Velazquez-Villegas, L.; Perino, A.; Lemos, V. TGR5 signalling promotes mitochondrial fission and beige remodelling of white adipose tissue. *Nat. Commun.* **2018**, *9*, 245. [CrossRef] [PubMed]
- Zuo, G.; Zhang, T.; Huang, L.; Araujo, C.; Peng, J.; Travis, Z.; Okada, T.; Ocak, U.; Zhang, G.; Tang, J.; et al. Activation of TGR5 with INT-777 attenuates oxidative stress and neuronal apoptosis via cAMP/PKCε/ALDH2 pathway after subarachnoid hemorrhage in rats. *Free Radic. Biol. Med.* 2019, 143, 441–453. [CrossRef] [PubMed]

- Zhou, L.; Wang, X.D.; Mao, W.; Li, P.F. Effects of taurochenodeoxycholic acid on Ca<sup>2+</sup>/CaM signalling mediated by the tgr5 signalling pathway. *Die Pharmazie*. 2016, *7*, 390–393.
- Xu, J.; Li, H.; Xia, Z. Propofol ameliorates hyperglycemia-induced cardiac hypertrophy and dysfunction via heme oxygenase-1/STAT3 signaling pathway in rats. *Crit Care Med.* 2014, *8*, e583-94. [CrossRef] [PubMed]
- Qi, Y.; Shi, L.; Duan, G.; Ma, Y.; Li, P. Taurochenodeoxycholic acid increases cAMP content via specially interacting with bile acid receptor TGR5. *Molecules* 2021, 23, 7066. [CrossRef] [PubMed]
- 17. Iguchi, Y.; Nishimaki-Mogami, T.; Yamaguchi, M.; Teraoka, F.; Kaneko, T.; Une, M. Effects of chemical modification of ursodeoxycholic acid on TGR5 activation. *Biol. Pharm. Bull.* **2011**, *34*, 1–7. [CrossRef]
- Naomi, D.; Karina, G.; Luke, C. Tauroursodeoxycholic acid/TGR5 signaling promotes survival and early development of glucose-stressed porcine embryos. *Biol Reprod.* 2021, 105, 76–86.
- Li, B.; Yang, N.; Li, C.; Li, C.; Gao, K.; Xie, X.; Dong, X.; Yang, J.; Yang, Q.; Tong, Z.; et al. INT-777, a bile acid receptor agonist, extenuates pancreatic acinar cells necrosis in a mouse model of acute pancreatitis. *Biochem. Biophys. Res. Commun.* 2018, 1, 38–44. [CrossRef]
- Chen, X.; Yan, L.; Guo, Z.; Chen, Y.; Li, M.; Huang, C.; Chen, Z.; Meng, X. Chenodeoxycholic acid attenuates high-fat dietinduced obesity and hyperglycemia via the G protein-coupled bile acid receptor 1 and proliferator-activated receptor γ pathway. *Exp. Ther. Med.* 2017, *6*, 5305–5312. [CrossRef]
- Cheng, K.C.; Chang, W.T.; Kuo, F.Y. TGR5 activation ameliorates hyperglycemia-induced cardiac hypertrophy in H9c2 cells. Sci. Rep. 2019, 9, 3633. [CrossRef]
- Qi, Y.C.; Duan, G.Z.; Mao, W.; Liu, Q.; Zhang, Y.L.; Li, P.F. Taurochenodeoxycholic acid mediates cAMP-PKA-CREB signaling pathway. *Chin. J. Nat. Med.* 2020, 12, 898–906. [CrossRef]
- Shen, Y.; Lu, C.; Song, Z.; Qiao, C.; Wang, J.; Chen, J.; Zhang, C.; Zeng, X.; Ma, Z.; Chen, T.; et al. Ursodeoxycholic acid reduces antitumor immunosuppression by inducing CHIP-mediated TGF-β degradation. *Nat. Commun.* 2022, 1, 3419. [CrossRef] [PubMed]
- 24. Dicks, N.; Gutierrez, K.; Currin, L. Tauroursodeoxycholic acid acts via TGR5 receptor to facilitate DNA damage repair and improve early porcine embryo development. *Mol. Reprod. Dev.* **2019**, *87*, 161–173. [CrossRef] [PubMed]
- 25. Wu, F.; Wang, L.; Zhou, C. Lung cancer in China: Current and prospect. Curr. Opin. Oncol. 2021, 1, 40–46. [CrossRef]
- Thomas, A.; Hassan, R. Immunotherapies for non-small-cell lung cancer and mesothelioma. *Lancet Oncol.* 2012, 7, e301–e310. [CrossRef]
- Hirsch, F.R.; Scagliotti, G.V.; Mulshine, J.L.; Kwon, R.; Curran, W.J.; Wu, Y.L.; Paz-Ares, L. Lung cancer: Current therapies and new targeted treatments. *Lancet* 2017, 10066, 299–311. [CrossRef]
- Lynch, T.J.; Kass, F.; Kalish, L.A.; Elias, A.D.; Strauss, G.; Shulman, L.N.; Sugarbaker, D.J.; Skarin, A.; Frei, E. Cisplatin, 5-fluorouracil, and etoposide for advanced non-small cell lung cancer. *Cancer* 1993, *10*, 2953–2957. [CrossRef]
- 29. Wu, Z.X.; Li, J.; Dong, S.; Lin, L.; Zou, C.; Chen, Z.S. Tepotinib hydrochloride for the treatment of non-small cell lung cancer. *Drug Today* **2021**, *4*, 265–275. [CrossRef]
- Zheng, X.; Luo, J.; Liu, W.; Ashby, C.R.; Chen, Z.S.; Lin, L. Sotorasib: A treatment for non-small cell lung cancer with the KRAS G12C mutation. *Drug Today* 2022, 4, 175–185. [CrossRef]
- 31. Simone, C.B.; Burri, S.H.; Heinzerling, J.H. Novel radiotherapy approaches for lung cancer: Combining radiation therapy with targeted and immunotherapies. *Transl. Lung Cancer Res.* **2015**, *5*, 545–552.
- 32. Vansteenkiste, J.F.; Van De Kerkhove, C.; Wauters, E.; Van Mol, P. Capmatinib for the treatment of non-small cell lung cancer. *Expert Rev. Anticancer Ther.* 2019, *8*, 659–671. [CrossRef] [PubMed]
- 33. Morias, S.; Loredana, G.; Michala, S.; Eileen, G.; Andrew, P.; Justin, S.; Thanh, G.; Eva, B. Treatment-related adverse effects in lung cancer patients after stereotactic ablative radiation therapy. *J. Oncol.* **2018**, *2018*, 6483626. [CrossRef] [PubMed]
- 34. Li, Q.; Yuan, Q.; Jiang, N. Dihydroartemisinin regulates immune cell heterogeneity by triggering a cascade reaction of CDK and MAPK phosphorylation. *Signal Transduct. Target. Ther.* **2022**, *7*, 222. [CrossRef] [PubMed]
- Sun, L.; Jia, H.; Yu, M.; Yang, Y.; Li, J.; Tian, D.; Zhang, H.; Zou, Z. Salvia miltiorrhiza and Pueraria lobata, two eminent herbs in Xin-Ke-Shu, ameliorate myocardial ischemia partially by modulating the accumulation of free fatty acids in rats. *Phytomedicine* 2021, 89, 153620. [CrossRef]
- Luo, S.; Zhang, X.; Huang, S.; Feng, X.; Zhang, X.; Xiang, D. A monomeric polysaccharide from Polygonatum sibiricum improves cognitive functions in a model of Alzheimer's disease by reshaping the gut microbiota. *Int. J. Biol. Macromol.* 2022, 213, 404–415. [CrossRef]
- Zhang, H.; Kong, Q.; Wang, J.; Jiang, Y.; Hua, H. Complex roles of cAMP-PKA-CREB signaling in cancer. *Exp. Hematol. Oncol.* 2020, 1, 32. [CrossRef]
- Xia, Y.; Zhan, C.; Feng, M.; Leblanc, M.; Ke, E.; Yeddula, N.; Verma, I.M. Targeting CREB pathway suppresses small cell Lung cancer. *Mol. Cancer Res.* 2018, 5, 825–832. [CrossRef]
- Seo, H.S.; Liu, D.D.; Bekele, B.N.; Kim, M.K.; Pisters, K.; Lippman, S.M.; Wistuba, I.I.; Koo, J.S. Cyclic AMP response elementbinding protein overexpression: A feature associated with negative prognosis in never smokers with non-small cell lung cancer. *Cancer Res.* 2008, 15, 6065–6073. [CrossRef]
- 40. Liu, X.; Chen, B.; You, W.; Xue, S.; Qin, H.; Jiang, H. The membrane bile acid receptor TGR5 drives cell growth and migration via activation of the JAK2/STAT3 signaling pathway in non-small cell lung cancer. *Cancer Lett.* **2018**, *412*, 194–207. [CrossRef]

- 41. Li, X.; Ramadori, P.; Pfister, D.; Seehawer, M.; Zender, L.; Heikenwalder, M. The immunological and metabolic landscape in primary and metastatic liver cancer. *Nat. Rev. Cancer* **2021**, *9*, 541–557. [CrossRef]
- 42. Llovet, J.M.; Kelley, R.K.; Villanueva, A.; Singal, A.G.; Pikarsky, E.; Roayaie, S.; Lencioni, R.; Koike, K.; Zucman-Rossi, J.; Finn, R.S. Hepatocellular carcinoma. *Nat. Rev. Dis. Primers* **2021**, *1*, 6. [CrossRef] [PubMed]
- 43. Brindley, P.J.; Bachini, M.; Ilyas, S.I.; Khan, S.A.; Loukas, A.; Sirica, A.E.; Teh, B.T.; Wongkham, S.; Gores, G.J. Cholangiocarcinoma. *Nat. Rev. Dis. Primers* **2021**, *1*, 65. [CrossRef] [PubMed]
- 44. Young, R.J.; Brown, N.J.; Reed, M.W.; Hughes, D.; Woll, P.J. Angiosarcoma. Lancet Oncol. 2010, 10, 983–991. [CrossRef]
- 45. World Health Organization. Cancer Today. 2020. Available online: https://gco.iarc.fr/today/fact-sheets-cancers (accessed on 1 December 2020).
- American Society of Clinical Oncology. Liver Cancer: Statistics. 2022. Available online: https://www.cancer.net/cancer-types/ liver-cancer/statistics (accessed on 1 February 2022).
- Lisa Fayed. Symptoms of Liver Cancer. 2021. Available online: https://www.verywellhealth.com/liver-cancer-symptoms-514170 (accessed on 25 June 2021).
- Chen, H.; Song, Y.P.; Gao, K.; Zhao, L.T.; Ma, L. Efficacy and safety of Jinhua Qinggan granules for coronavirus disease 2019 (COVID-19): A protocol of a systematic review and meta-analysis. *Medicine* 2020, 24, e20612. [CrossRef] [PubMed]
- 49. Fan, S.J.; Liao, J.K.; Wei, L.; Wang, B.Y.; Kai, L.; Tan, D.X. Treatment efficacy of Lianhua Qingwen capsules for eraly-stage COVID-19. *Am. J. Transl. Res.* 2022, *2*, 1332–1338.
- 50. Guo, H.; Zheng, J.; Huang, G.; Xiang, Y.; Lang, C.; Li, B.; Huang, D.; Sun, Q.; Luo, Y.; Zhang, Y.; et al. Xuebijing injection in the treatment of COVID-19: A retrospective case-control study. *Ann. Palliat. Med.* **2020**, *5*, 3235–3248. [CrossRef]
- Cao, P.; Wu, S.L.; Wu, T.T.; Deng, Y.H.; Zhang, Q.L.; Wang, K.P.; Zhang, Y. The important role of polysaccharides from a traditional Chinese medicine-Lung Cleansing and Detoxifying Decoction against the COVID-19 pandemic. *Carbohyd. Polym.* 2020, 240, 116346. [CrossRef]
- 52. Cai, Y.; Zeng, M.; Chen, Y.Z. The pharmacological mechanism of Huashi Baidu Formula for the treatment of COVID-19 by combined network pharmacology and molecular docking. *Ann. Palliat. Med.* **2021**, *4*, 3864–3895. [CrossRef]
- 53. Li, C.L.; Lin, Y.K.; Chen, H.A.; Huang, C.Y.; Huang, M.T.; Chang, Y.J. Smoking as an independent risk factor for hepatocellular carcinoma due to the *α*7-nachr modulating the JAK2/STAT3 signaling axis. *J. Clin. Med.* **2019**, *9*, 1391. [CrossRef]
- 54. Chen, W.D.; Yu, D.; Forman, B.M.; Huang, W.; Wang, Y.D. Deficiency of G-protein-coupled bile acid receptor Gpbar1 (TGR5) enhances chemically induced liver carcinogenesis. *Hepatology* **2013**, *2*, 656–666. [CrossRef]
- 55. Ma, S.C.; Zhao, Y.; Zhang, T.; Ling, X.L.; Zhao, D. Association between the ERCC1 rs11615 polymorphism and clinical outcomes of oxaliplatin-based chemotherapies in gastrointestinal cancer: A meta-analysis. *OncoTargets Ther.* **2015**, *8*, 641–648.
- 56. Jemal, A.; Bray, F.; Center, M.M.; Ferlay, J.; Ward, E.; Forman, D. Global cancer statistics. *CA Cancer J. Clin.* **2011**, *61*, 69–90. [CrossRef]
- 57. Richa, S.N.; Sageena, G. Dietary factors associated with gastric cancer-a review. Transl. Med. Commun. 2022, 7, 7. [CrossRef]
- 58. Cai, H.; Ye, F.; Michel, A.; Murphy, G.; Sasazuki, S.; Taylor, P.R.; Qiao, Y.L.; Park, S.K.; Yoo, K.Y.; Jee, S.H.; et al. Helicobacter pylori blood biomarker for gastric cancer risk in East Asia. *Int. J. Epidemiol.* **2016**, *3*, 774–781. [CrossRef] [PubMed]
- 59. Fox, J.G.; Wang, T.C. Inflammation, atrophy, and gastric cancer. J. Clin. Investig. 2007, 1, 60–69. [CrossRef]
- 60. American Society of Clinical Oncology. Stomach Cancer: Symptoms and Signs. 2022. Available online: https://www.cancer.net/ cancer-types/stomach-cancer/symptoms-and-signs (accessed on 12 August 2022).
- 61. Meyer, H.J.; Wilke, H. Treatment strategies in gastric cancer. Dtsch. Arztebl. Int. 2011, 41, 698–705. [CrossRef]
- Guo, C.; Qi, H.; Yu, Y.; Zhang, Q.; Su, J.; Yu, D.; Huang, W.; Chen, W.D.; Wang, Y.D. The G-protein-coupled bile acid receptor Gpbar1 (TGR5) inhibits gastric inflammation through antagonizing NF-κB signaling pathway. *Front. Pharmacol.* 2015, *6*, 287. [CrossRef]
- Lu, J.; Zhang, L.; Chen, X.; Lu, Q.; Yang, Y.; Liu, J.; Ma, X. SIRT1 counteracted the activation of STAT3 and NF-κB to repress the gastric cancer growth. *Int. J. Clin. Exp. Med.* 2014, 12, 5050–5058.
- 64. Bosch-Presegue, L.; Vaquero, A. The dual role of sirtuins in cancer. Gene Cancer 2011, 2, 648–662. [CrossRef]
- 65. Yuan, H.; Su, L.; Chen, W.Y. The emerging and diverse roles of sirtuins in cancer: A clinical perspective. *Onco Targets Ther.* **2013**, *6*, 1399–1416.
- Kume, S.; Haneda, M.; Kanasaki, K.; Sugimoto, T.; Araki, S.; Isshiki, K.; Isono, M.; Uzu, T.; Guarente, L.; Kashiwagi, A.; et al. SIRT1 inhibits transforming growth factor beta-induced apoptosis in glomerular mesangial cells via Smad7 deacetylation. *J. Biol. Chem.* 2007, 282, 151–158. [CrossRef] [PubMed]
- Guo, C.; Su, J.; Li, Z.; Xiao, R.; Wen, J.; Li, Y.; Zhang, M.; Zhang, X.; Yu, D.; Huang, W.; et al. The G-protein-coupled bile acid receptor Gpbar1 (TGR5) suppresses gastric cancer cell proliferation and migration through antagonizing STAT3 signaling pathway. *Oncotarget* 2015, *33*, 34402–34413. [CrossRef] [PubMed]
- 68. American Society of Clinical Oncology. Colorectal Cancer. 2022. Available online: https://www.cancer.net/cancer-types/ colorectal-cancer/statistics (accessed on 1 May 2022).
- 69. Rogler, G. Chronic ulcerative colitis and colorectal cancer. Cancer Lett. 2014, 2, 235–241. [CrossRef] [PubMed]
- Xi, Y.; Xu, P. Global colorectal cancer burden in 2020 and projections to 2040. *Transl. Oncol.* 2021, *10*, 101174. [CrossRef] [PubMed]
  McQuade, R.M.; Stojanovska, V.; Bornstein, J.C.; Nurgali, K. Colorectal cancer chemotherapy: The evolution of treatment and new approaches. *Curr. Med. Chem.* 2017, *15*, 1537–1557. [CrossRef]

- 72. Zhang, H.; Xu, H.; Zhang, C. Ursodeoxycholic acid suppresses the malignant progression of colorectal cancer through TGR5-YAP axis. *Cell Death Discov.* 2021, 7, 207. [CrossRef]
- Ward, J.B.J.; Lajczak, N.K.; Kelly, O.B.; O'Dwyer, A.M.; Giddam, A.K.; Ni Gabhann, J.; Franco, P.; Tambuwala, M.M.; Jeferies, C.A.; Keely, S.; et al. Ursodeoxycholic acid and lithocholic acid exert anti-infammatory actions in the colon. *Am. J. Physiol. Gastrointest. Liver Physiol.* 2017, 312, G550–G558. [CrossRef]
- Pols, T.W.; Noriega, L.G.; Nomura, M.; Auwerx, J.; Schoonjans, K. The bile acid membrane receptor TGR5: A valuable metabolic target. Dig. Dis. 2011, 1, 37–44. [CrossRef]
- 75. Pols, T.W.; Noriega, L.G.; Nomura, M.; Auwerx, J.; Schoonjans, K. The bile acid membrane receptor TGR5 as an emerging target in metabolism and inflammation. *J. Hepatol.* **2011**, *6*, 1263–1272. [CrossRef]
- 76. Thomas, C.; Gioiello, A.; Noriega, L.; Strehle, A.; Oury, J.; Rizzo, G.; Macchiarulo, A.; Yamamoto, H.; Mataki, C.; Pruzanski, M.; et al. TGR5-mediated bile acid sensing controls glucose homeostasis. *Cell Metab.* **2009**, *3*, 167–177. [CrossRef]
- Casaburi, I.; Avena, P.; Lanzino, M.; Sisci, D.; Giordano, F.; Maris, P.; Catalano, S.; Morelli, C.; Andò, S. Chenodeoxycholic acid through a TGR5-dependent CREB signaling activation enhances cyclin D1 expression and promotes human endometrial cancer cell proliferation. *Cell Cycle* 2012, *14*, 2699–2710. [CrossRef] [PubMed]
- Luu, T.H.; Bard, J.M.; Carbonnelle, D.; Chaillou, C.; Huvelin, J.M.; Bobin-Dubigeon, C.; Nazih, H. Lithocholic bile acid inhibits lipogenesis and induces apoptosis in breast cancer cells. *Cell. Oncol.* 2018, 1, 13–24. [CrossRef] [PubMed]
- 79. Kovács, P.; Csonka, T.; Kovács, T.; Sári, Z.; Ujlaki, G.; Sipos, A.; Karányi, Z.; Szeőcs, D.; Hegedűs, C.; Uray, K.; et al. Lithocholic Acid, a Metabolite of the Microbiome, Increases Oxidative Stress in Breast Cancer. *Cancers* **2019**, *9*, 1255. [CrossRef] [PubMed]
- 80. Zhao, R.Y.; He, S.J.; Ma, J.J.; Hu, H.; Gong, Y.P.; Wang, Y.L.; Hu, B.J.; Xie, J.Z.; Tu, W.Z.; Huang, Q.; et al. High expression of TGR5 predicts a poor prognosis in patients with pancreatic cancer. *Int. J. Clin. Exp. Pathol.* **2018**, *7*, 3567–3574.