



Since January 2020 Elsevier has created a COVID-19 resource centre with free information in English and Mandarin on the novel coronavirus COVID-19. The COVID-19 resource centre is hosted on Elsevier Connect, the company's public news and information website.

Elsevier hereby grants permission to make all its COVID-19-related research that is available on the COVID-19 resource centre - including this research content - immediately available in PubMed Central and other publicly funded repositories, such as the WHO COVID database with rights for unrestricted research re-use and analyses in any form or by any means with acknowledgement of the original source. These permissions are granted for free by Elsevier for as long as the COVID-19 resource centre remains active.



Contents lists available at ScienceDirect

Bioorganic & Medicinal Chemistry Letters

journal homepage: www.elsevier.com/locate/bmcl

Exploring the purine core of 3'-C-ethynyladenosine (EAdo) in search of novel nucleoside therapeutics



Fabian Hulpia^a, Jan Balzarini^b, Dominique Schols^b, Graciela Andrei^b, Robert Snoeck^b, Serge Van Calenbergh^{a,*}

^aLaboratory for Medicinal Chemistry, Ghent University, Ottergemsesteenweg 460, B-9000 Ghent, Belgium

^bKU Leuven, Rega Institute for Medical Research, Minderbroedersstraat 10, B-3000 Leuven, Belgium

ARTICLE INFO

Article history:

Received 4 February 2016

Revised 1 March 2016

Accepted 2 March 2016

Available online 3 March 2016

Keywords:

Nucleosides

3'-C-Ethynyladenosine

Vorbrüggen coupling

ABSTRACT

A series of new nucleoside analogues based on a C-3 branched ethynyl sugar derivative as present in 3'-ethynylcytidine (ECyT) and -adenosine (EAdo), combined with modified purine bases was synthesized and evaluated against a broad array of viruses and tumour cell lines. The pronounced cytostatic activity of EAdo was confirmed. EAdo and its 2,6-diaminopurine analogue showed inhibitory activity against vaccinia virus (EC₅₀: 0.31 and 51 μM, respectively). Derivative **10** on the other hand was found active against varicella zoster virus (EC₅₀: 4.68 μM).

© 2016 Elsevier Ltd. All rights reserved.

Nucleoside analogues have been used as therapeutics for the treatment of cancer and viral infections for over 50 years. Despite this fact, new agents with improved efficacy and tolerability have still been discovered over the past decade.¹

Recently, a strategy relying on building nucleosides with a fixed sugar and varying heterocyclic bases has shown to reveal intriguing biological activity.^{2,3} Additionally, previous focussed profiling (e.g., only HIV, HCV or anticancer) might have overlooked starting points for other indications (e.g., cytotoxicity in antiviral assays or activity against other viruses). Furthermore, re-investigating certain sugar modifications has already shown to be a fruitful approach with some derivatives ultimately being advanced into clinical trials.⁴

With this in mind, research was initiated to explore the untapped potential of certain nucleoside scaffolds. We therefore became interested in a C-3 branched sugar motif, as is present in a former Phase-II candidate, 3'-C-ethynylcytidine (**2**, ECyT). After the discovery in the mid-90s by Matsuda et al.,⁵ SAR of its sugar⁶ as well as its heterocyclic⁷ part has been reported, but failed to produce analogues with improved activity. The adenosine (**1**, EAdo) congener of ECyT however, which also showed antitumour properties,⁵ only received scarce follow-up^{8,9} (see Fig. 1).

Based on these findings, the C-3 ethynyl ribofuranose unit was believed a good sugar starting point for the 'mix-and-match' strat-

egy mentioned above. In this study we concentrated on the poorly explored purine analogues.

In order to prioritize, we focused on substituents present in FDA-approved nucleosides as well as clinical phase analogues. This translated into modifications of EAdo at both the C-2 and C-6 position of the purine ring to give a heterocycle pattern as is, for example, found in the two FDA-approved nucleosides nelarabine **3** and clofarabine **4**.¹ Additionally, two modifications (cyclopentylamine, 3-chlorobenzylamine) were included as they are preferred in purinergic receptor ligands.¹⁰

The synthesis of the key ribofuranose building block **4** was accomplished using known literature procedures.^{7,11} Next, three different purines were subjected to 'classical' Vorbrüggen conditions to achieve glycosylation with **4**¹² (Scheme 1). Deprotection or nucleophilic aromatic substitution with either NH₃ in MeOH or NaOMe in MeOH yielded the corresponding target nucleosides **1**,⁵ **7**, **8**, **12** and **13**. Compounds **9** and **10** were obtained by nucleophilic aromatic substitution with the appropriate amine, immediately followed by deprotection. Initially, synthesis of **14** and **15** was also attempted from **11** and **6**, respectively. However, due to prolonged exposure and elevated temperature to force nucleophilic aromatic ring substitution, the desired product could not be isolated. Instead, the product formed was the enol ether resulting from a 5-*exo* dig cyclization of the 5'-OH onto the alkyne (**16** and **17**). Therefore, ring substitution was performed before glycosylation (Scheme 2), resulting in **20** and **21**. Final deprotection of these intermediates furnished **14** and **15**.

* Corresponding author. Tel.: +32 9 264 81 24; fax: +32 9 264 81 46.

E-mail address: Serge.VanCalenbergh@ugent.be (S. Van Calenbergh).

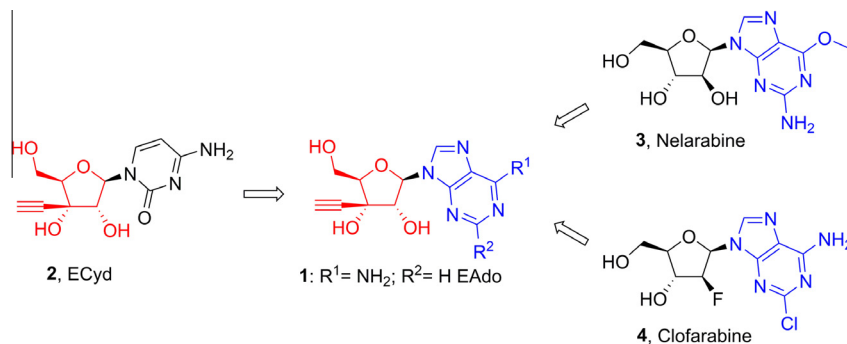
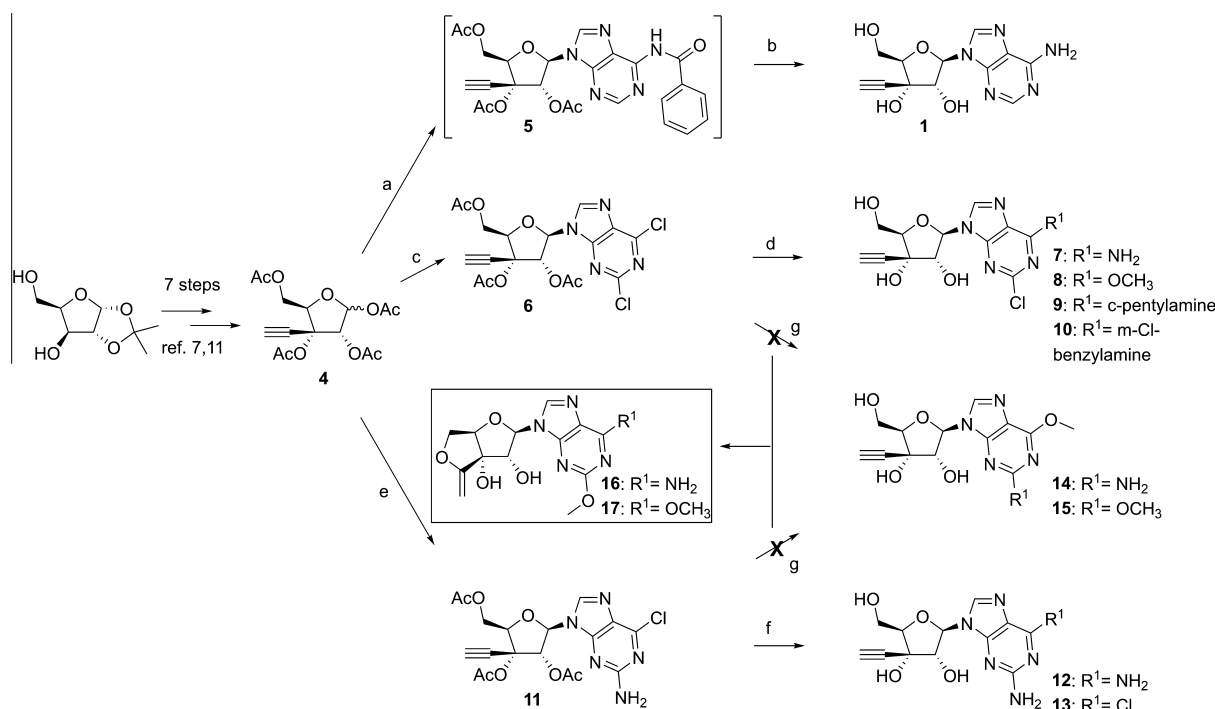
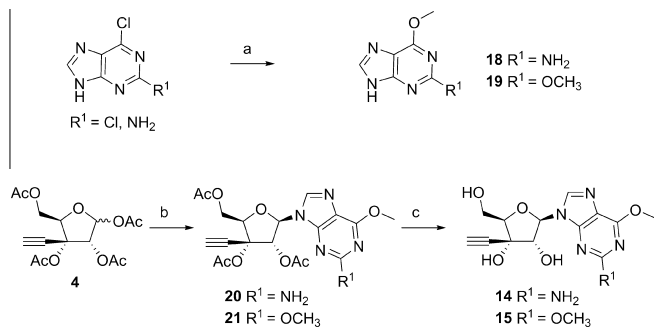


Figure 1.



Scheme 1. Reagents and conditions: (a) (1) *N*-benzoyladenine, HMDS, cat. (NH₄)₂SO₄, reflux, (2) TMSOTf, MeCN, reflux; (b) 7 N NH₃ in MeOH (40% over 2 steps (a + b)); (c) (1) 2,6-dichloropurine, HMDS, cat. (NH₄)₂SO₄, reflux, (2) TMSOTf, 1,2-dichloroethane, reflux (72%); (d) 7 N NH₃ in MeOH for **7** (31%), NaOMe in MeOH for **8** (72%), for **9**: (1) *c*-pentylamine, EtOH, reflux, (2) 7 N NH₃ in MeOH (85%); for **10**: (1) 3-chlorobenzylamine, EtOH, reflux, (2) 7 N NH₃ in MeOH (77%); (e) (1) 2-amino-6-chloropurine, HMDS, cat. (NH₄)₂SO₄, reflux, (2) TMSOTf, 1,2-dichloroethane, reflux (43%); (f) 7 N NH₃ in MeOH for **12** (25%) and NaOMe in MeOH for **13** (63%); (g) NaOMe in MeOH, rt to reflux, 38% for **16** and 29% for **17**.



Scheme 2. Reagents and conditions: (a) Na, MeOH, reflux, 91% for **18**; 83% for **19**; (b) (1) 2-amino-6-methoxypurine (**18**) or 2,6-dimethoxypurine (**19**), HMDS, cat. (NH₄)₂SO₄, reflux, (2) TMSOTf, 1,2-dichloroethane, reflux; 29% for **20**; 36% for **21**; (c) NaOMe in MeOH; 62% for **14**; 66% for **15**.

All final compounds were investigated for their potential activity against a broad array of viruses including herpes simplex virus-1 and -2 (HSV), cytomegalovirus (CMV), varicella zoster virus (VZV), vaccinia virus (VV), adenovirus-2, influenza A virus (H1N1, H3N2), influenza B virus, feline corona virus, feline Herpes virus, para-influenza virus, reovirus-1, sindbis virus, Coxsackie virus B4, Punta Toro virus, vesicular stomatitis virus, respiratory syncytial virus (RSV), HIV-1 and HIV-2. Additionally, inhibition of cell proliferation of murine leukemia (L1210), human CD₄⁺ T-lymphocyte (CEM) and human cervix carcinoma cells (HeLa) was also evaluated (Table 1).

EAdo **1** was confirmed to exhibit potent cytostatic activity, while all other derivatives were found to be poorly cytostatic (IC₅₀: 104 > 250 μM). Compound **10** was found to be moderately cytotoxic (22–50 μM) in the three cell lines tested, but at least 30 times less potent than EAdo. All nucleosides were found to be inactive in the antiviral assays up to a concentration of 100 μM,

Table 1
Inhibition of proliferation

	L1210 IC ₅₀ (μM)	CEM IC ₅₀ (μM)	HeLa IC ₅₀ (μM)
1	0.73 ± 0.14	0.61 ± 0.08	0.29 ± 0.11
7	167 ± 37	>250	>250
8	>250	210 ± 56	175 ± 106
9	170 ± 28	104 ± 33	150 ± 1
10	50 ± 14	22 ± 10	43 ± 20
12	193 ± 80	≥250	123 ± 7
13	>250	>250	>250
14	>250	>250	>250
15	>250	>250	>250

except for three. EAdo **1** showed potent anti-vaccinia virus activity at subtoxic concentrations [EC₅₀: 0.35 ± 0.05 μM; MCC (minimal cytotoxic concentration): 20 μM]. Diaminopurine derivative **13**, showed weak activity against vaccinia virus (EC₅₀: 51 ± 6 μM; MCC: >100 μM). Finally, *m*-chlorobenzylamino derivative **10** showed moderate activity against VZV (EC₅₀: 4.68 μM; MCC: 100 μM; CC₅₀: 34.46 μM), with no markedly different results obtained in either TK⁺ or TK⁻ strains.

Compounds **9** and **10** were also evaluated for their agonistic behaviour at the adenosine A₃-receptor. Both were found to bind only weakly (70 ± 9% and 48 ± 6% inhibition at 10 μM for **9** and **10**, respectively), which is in line with previous observations.¹³

Interestingly, compound **7** did not show any cytostatic activity, even though the C-2 chloro substituent should make it more resistant towards adenosine deaminase,¹⁴ the enzyme responsible for the breakdown of EAdo.⁹ Furthermore, lack of phosphorylation by cellular kinase(s) could also be a contributor to the observed results, and further investigation on a prodrug approach that allows intracellular release of the monophosphate form might be more promising.

In conclusion, a subset of purine-modified nucleosides based on C-3 branched chain sugar matched with different purines was synthesized and evaluated against a broad array of viruses and tumour cell lines. The potent cytostatic activity of EAdo was confirmed. This compound was found inhibitory to vaccinia virus at subtoxic concentrations. Two of the newly synthesized compounds were found active antivirally. While their activity is only moderate, they could serve as a starting point for further structural elaboration to improve antiviral activity. Furthermore, these results indicate the usefulness of the ‘mix-and match’ approach in finding novel biologically active nucleosides.

Acknowledgements

F.H. is indebted to the Flanders Research Fund (FWO) for a PhD-scholarship and D.S., G.A., R.S. and J.B. to the KU Leuven for financial support (GOA 15/19 TBA). The authors wish to thank Prof. K. Jacobson for performing the adenosine A₃-receptor assays on derivatives **9** and **10**.

Supplementary data

Supplementary data associated with this article can be found, in the online version, at <http://dx.doi.org/10.1016/j.bmcl.2016.03.005>.

References and notes

- Jordheim, L. P.; Durantel, D.; Zoulim, F.; Dumontet, C. *Nat. Rev. Drug Disc.* **2013**, *12*, 447.
- Zhou, L.; Zhang, H.; Tao, S.; Ehteshami, M.; Cho, J. H.; McBrayer, T. R.; Tharnish, P.; Whitaker, T.; Amblard, F.; Coats, S. J.; Schinazi, R. F. *ACS Med. Chem. Lett.* **2016**, *7*, 17.
- Yin, Z.; Chen, Y.-L.; Schul, W.; Wang, Q.-Y.; Gu, F.; Duraiswamy, J.; Kondreddi, R. R.; Niyomrattanakit, P.; Lakshminarayana, S. B.; Goh, A.; Xu, H. Y.; Liu, W.; Liu, B.; Lim, J. Y. H.; Ng, C. Y.; Qing, M.; Lim, C. C.; Yip, A.; Wang, G.; Chan, W. L.; Tan, H. P.; Lin, K.; Zhang, B.; Zou, G.; Bernard, K. A.; Garrett, C.; Beltz, K.; Dong, M.; Weaver, M.; He, H.; Pichota, A.; Dartois, V.; Keller, T. H.; Shi, P.-Y. *Proc. Natl. Acad. Sci. U.S.A.* **2009**, *106*, 20435.
- Smith, D. B.; Martin, J. A.; Klumpp, K.; Baker, S. J.; Blomgren, P. A.; Devos, R.; Granycome, C.; Hang, J.; Hobbs, C. J.; Jiang, W.-R.; Laxton, C.; Pogam, S. L.; Leveque, V.; Ma, H.; Maile, G.; Merrett, J. H.; Pichota, A.; Sarma, K.; Smith, M.; Swallow, S.; Symons, J.; Vesey, D.; Najera, L.; Cammack, N. *Bioorg. Med. Chem. Lett.* **2007**, *17*, 2570.
- Hattori, H.; Tanaka, M.; Fukushima, M.; Sasaki, T.; Matsuda, A. *J. Med. Chem.* **1996**, *39*, 5005.
- Hattori, H.; Nozawa, E.; Iino, T.; Yoshimura, Y.; Shuto, S.; Shimamoto, Y.; Nomura, M.; Fukushima, M.; Tanaka, M.; Sasaki, T.; Matsuda, A. *J. Med. Chem.* **1998**, *41*, 2892.
- Hrdlicka, P. J.; Jepsen, J. S.; Nielsen, C.; Wengel, J. *Bioorg. Med. Chem.* **2005**, *13*, 1249.
- Endo, Y.; Obata, T.; Nomura, M.; Fukushima, M.; Yamada, Y.; Matsuda, A.; Sasaki, T. *Nucleosides Nucleotides Nucleic Acids* **2007**, *26*, 691.
- Tritsch, D.; Jung, P. M. J.; Burger, A.; Biellmann, J.-F. *Bioorg. Med. Chem. Lett.* **2000**, *10*, 139.
- Jacobson, K. A.; Gao, Z.-G.; Tchilibon, S.; Duong, H. T.; Joshi, B. V.; Sonin, D.; Liang, B. T. *J. Med. Chem.* **2005**, *48*, 8103.
- Kim, J.; Weledji, Y. N.; Greenberg, M. M. *J. Org. Chem.* **2004**, *69*, 6100.
- Cosyn, L.; Gao, Z.-G.; Van Rompaey, P.; Lu, C.; Jacobson, K. A.; Van Calenberg, S. *Bioorg. Med. Chem.* **2006**, *14*, 1403.
- Cappellacci, L.; Franchetti, P.; Pasqualini, M.; Petrelli, R.; Vita, P.; Lavecchia, A.; Novellino, E.; Costa, B.; Martini, C.; Klotz, K.-N.; Grifantini, M. *J. Med. Chem.* **2005**, *48*, 1550.
- Cristalli, G.; Costanzi, S.; Lambertucci, C.; Lupidi, G.; Vittori, S.; Volpini, R.; Camaioni, E. *Med. Res. Rev.* **2001**, *21*, 105.