

Disseminated *Mycobacterium simiae* and *Mycobacterium avium* infection causing an immune reconstitution inflammatory syndrome in a female patient with HIV infection

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SUMMARY

This case study discusses the management of a disseminated *Mycobacterium simiae* and *Mycobacterium avium* infection causing an immune reconstitution inflammatory syndrome in a 52-year-old woman with HIV infection. Disseminated *M. avium* infections have extensively been described in HIV patients; however, reports of infections with *M. simiae* are rare. Treatment of *M. simiae* infections is challenging due to its high rates of natural drug resistances, and thus far, no standard treatment regimen exists.

BACKGROUND

Mycobacterium avium has been described frequently as a cause of pulmonary infection in patients with chronic pulmonary disease (eg, bronchiectasis and cystic fibrosis) and disseminated infection in patients with HIV/AIDS. Treatment recommendation for M. avium consists of a threedrug regimen including macrolide, rifamycin and ethambutol.² Mycobacterium simiae is a rare, slowly growing non-tuberculous mycobacterium (NTM), which was first discovered in monkeys in 1965 by Karassova et al (simia is the Latin word for monkey). 1-3 M. simiae has been reported in several regions of the world, mainly the USA and in the Middle East (Lebanon, Iran, Reunion Island, Israel and Texas).⁴⁻⁸ In the late 1990s, a nosocomial pseudo-outbreak in a hospital in Texas was documented; the reservoir was a contaminated water supply. In most published cases, M. simiae is reported to be an opportunistic pathogen-causing infections in immunocompromised patients (especially AIDS) or in patients with pre-existing pulmonary diseases. 4 10 However, there are cases of infections reported in immunocompetent patients: Cruz et al¹¹ describe a case of a healthy child with M. simiae lymphadenitis. M. simiae skin infection is reported in an immunocompetent adult due to an iatrogenic infection. 12 Balkis et al 13 describe a case of disseminated M. simiae infection in an immunocompetent 83-year-old man without pre-existing pulmonary disease with immune senescence being a possible risk factor. Treatment of M. simiae infections is challenging because of multiple intrinsic drug resistances, especially against tuberculostatic drugs. 14 15 Treatment regimen for M. simiae

infections containing a macrolide (azithromycin or clarithromycin), a fluorochinolone (eg, moxifloxacin) and a third antibiotic class (eg, clofazimine, streptomycin or an amionglycoside such as amikacin) have been used, but standardised treatment recommendations to this point are non-existent.

CASE PRESENTATION

A 52-year-old woman of African descent was newly diagnosed with AIDS due to a HIV type 1 infection in December 2019. At that time, the CD4 count was 13 cells/µL, and the viral load was about 240 000 copies/mL. In January 2020, an antiretroviral therapy (ART) with Descovy (emtricitabin +tenofoviralafenamid) and Tivicay (dolutegravir) was initiated. Bactrim (cotrimoxazol) was prescribed for pneumocystis prophylaxis. As part of the initial evaluation, the patient was tested for other infectious diseases: serological testing for hepatitis A indicated a past infection, hepatitis B and hepatitis C serological panels were negative, Cytomegalovirus (CMV) and Ebstein-Barr virus (EBV) PCR were also negative.

Two weeks after the beginning of ART, the patient presented with new onset of fever (39°C), fatigue and weight loss. Personal history and family history were unremarkable. No recent travel history. She reported regular medication intake. Physical examination did not show any abnormal findings apart from the elevated axillary temperature. At this time, leucocyte count was 1.74 G/L, haemoglobin was 67 g/L and the platelet count was 217 G/L. C reactive protein was 109 mg/L. Body mass index at presentation 17.2 kg/m². Chest radiograph and chest CT did not show any pulmonary infiltrates. Unspecific mediastinal lymphadenopathy was seen on chest CT. Urine analysis did not show signs of urinary tract infection. After another 2 weeks, the patient developed bilateral, painful, inguinal lymphadenopathy. A gynaecological infection or a gynaecological tumour as a cause of the lymphadenopathy were excluded after normal smear results and a normal pelvis MRI. Despite an empiric treatment with cefepime and doxycycline for 2 weeks, the patient was still febrile without clinical improvement.

Repeated blood cultures were performed without signs of bacterial growth. No other cause of the fever could be identified. Multiple examinations were initiated but no viral, bacterial or fungal infection

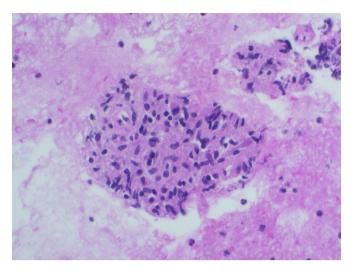


Figure 1 Cell block preparation from the fine needle aspiration (FNA) of an inguinal lymph node: granulomatous inflammation with typical aggregation of epithelioid histiocytes (H&E stain; 400×).

was identified: CMV PCR and EBV PCR were still negative, EBV and toxoplasmosis serologies indicated a past infection, *Crypto-coccus neoformans* antigen was negative and sputum PCR for atypical bacterial pneumonia was negative. Because of neutropaenia, a bone marrow biopsy was performed, which showed reactive changes compatible with HIV infection.

After additional 2 weeks, *M. simiae* grew in citrate blood cultures, bone marrow, lymph node, sputum and stool cultures. An inguinal lymph node biopsy confirmed granulomatous inflammation and copious acid-fast bacilli (figure 1). In one blood culture and in the lymph node biopsy *M. avium* was isolated in addition to *M. simiae*.

TREATMENT

Having diagnosed a disseminated mycobacterial infection with *M. simiae* and *M. avium*, we started a treatment regimen with azithromycin (oral 500 mg once daily), moxifloxacin (oral 400 mg once daily) and amikacin (intravenous 15 mg per kg ones daily). After 4 weeks, based on the drug susceptibility testing results (table 1), we switched from intravenous amikacin to clofazimine (oral 50 mg twice daily). Because of persistent fever, we repeatedly performed additional diagnostic rounds searching for other infections, but to no avail. Therefore, we started treatment with prednisone 20 mg for a presumptive immune reconstitution syndrome. The clinical condition of the patient improved rapidly, and we were able to gradually taper the prednisone dosage over weeks. The inguinal lymphadenopathy improved fast as well.

OUTCOME AND FOLLOW-UP

Blood culture showed mycobacterial growth up to 1 month after treatment start and turned negative after 7 weeks. Serial ECG monitoring was performed without prolongation of the correctet QT interval (QTc) . The patient reported significant improvement in general condition.

DISCUSSION

Here, we present a case with unmasking immune reconstitution inflammatory syndrome (IRIS) caused by a disseminated mycobacterial infection with *M. simiae* and *M. avium* in a 52-year-old woman with HIV infection 4 weeks after starting ART. Given

Table 1 Antibiotic susceptibility patterns of *M. simiae* and *M. avium*

| | M. simiae | | M. avium | |
|-----------------------------------|------------|----------------|------------|----------------|
| Drug | MIC (mg/L) | Interpretation | MIC (mg/L) | Interpretation |
| Clarithromycin | 4 | S | 4 | S |
| Ethambutol | >16 | na | 16 | na |
| Rifampicin | >8 | R | 1 | S |
| Rifabutin | >8 | R | 0.25 | S |
| Amikacin | 16 | S | 8 | S |
| Moxifloxacin | 0.5 | S | 1 | S |
| Clofazimine | ≤1 | na | ≤1 | na |
| Linezolid | >64 | R | 16 | I |
| Doxycyline | >16 | R | >16 | R |
| Trimethoprim/ sulfamethoxazole | 2/32 | S | 2/38 | S |
| Isoniazid | 8 | na | >8 | na |

Minimal inhibitory concentrations (MIC) were determined by broth microdilution as recommended by CLSI using Sensititre SLOWMYCOI plates (ThermoFisher, Waltham, MA). ClofazimineMIC was measured by quantitiative MGIT testing. ²⁶

Interpretation according to species-independent CLSI breakpoints for slowly growing $\overline{\text{NTM}}^{\,25}$

I, intermediate; *M. avium, Mycobacterium avium*; MIC, minimal inhibitory concentrations; *M. simiae, Mycobacterium simiae*; na, no breakpoints available; R, resistant; S, susceptible.

the temporal association of the beginning of the symptoms and introduction of the ART, and the extent of disease, it is suggestive that the patient suffered from disseminated *M. simiae* and *M. avium* infection already at the time of HIV diagnosis. Therefore, we hypothesise that the development of symptoms was mostly due to beginning IRIS after successful suppression of the HI viral load and immune reconstitution. Mycobacterial cultures are not routinely performed at the time of diagnosis, and therefore, we cannot prove whether the mycobacterial infection was ongoing at the time of HIV diagnosis or not.

The treatment for presumptive IRIS encompasses first treatment against the pathogen to reduce pathogen load and second slowing the fulminant immune response, ideally without hampering the 'normal' recovery of the immune system. ^{16–18}

The pathogen-directed therapy against M. simiae is complicated since optimal treatment regimens have not been studied. The standard treatment regimen for M. avium infections, consisting of a macrolide, ethambutol and a rifamycine, is not recommended for M. simiae infections due to high rates of resistances. Overall, M. avium seems to be more susceptible than M. simiae to antibiotics. This is consistent with the results from our patient, shown in table 1. For M. simiae infections, a combination of three drugs including a macrolide seems to be favourable. 19 20 Based on the in vitro investigations of van Ingen et al, moxifloxacin, a macrolide and a third antibiotic class (eg, clofazimine and streptomycin, an aminoglycoside such as amikacin) should be included in treatment regimens for a M. simiae infection.^{21 22} Azithromycin and clarithromycin are assumed to be equally effective. 20 23 Although moxifloxacin, clarithromycin and amikacin or clofazimine is not the standard treatment regimen for M. avium infections, M. avium mostly is also susceptible to

Learning points

- ► *Mycobacterium simiae* can cause severe disseminated disease in AIDS patients.
- Immune reconstitution due to antiretroviral treatment is a diagnostic and therapeutic challenge.

these drugs, as shown in our case (table 1). For lung infections and disseminated NTM infections treatment duration is recommended 12 months after sputum culture conversion.²⁰

The IRIS component of the infection was treated with prednisone after excluding other, active infections multiple times. ²⁴ The treatment with prednisone led to dramatic clinical improvement, cessation of fever and malaise and, subsequently, normalised appetite and weight gain.

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