

Reduction in Operating Room Airborne Particle Burden and Time-Dependent Contamination of Sterile Instrument Trays With the Use of a Novel Air Filtration System

Review began 05/24/2022

Review ended 07/11/2022

Published 07/14/2022

© Copyright 2022

Hijji et al. This is an open access article distributed under the terms of the Creative Commons Attribution License CC-BY 4.0., which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

Fady Y. Hijji¹, Andrew D. Schneider¹, Jeffrey T. Reeves¹, Michael L. Wilson¹, Logan Nye¹, Joseph G. Lyons¹, Michael J. Prayson¹, Louis J. Rubino¹

1. Orthopaedic Surgery, Boonshoft School of Medicine, Dayton, USA

Corresponding author: Joseph G. Lyons, jglyons245@gmail.com

Abstract

Introduction

Postoperative infections represent a substantial burden to patients and healthcare systems. To improve patient care and reduce healthcare expenditures, interventions to reduce surgical infections must be employed. The crystalline C-band ultraviolet (UV-C) air filtration technology (Aerobiotix Inc., Miamisburg, OH, USA) has been designed to reduce airborne bioburden through high-quality filtration and germicidal irradiation. The purpose of this study was to assess the ability of a novel UV-C air filtration device to reduce airborne particle counts and contamination of surgical instrument trays in an operating room (OR) setting.

Materials and methods

Thirty sterile instrument trays were opened in a positive-air-flow OR. The trays were randomly assigned to one of two groups (UV-C or control, n=15 per group). In the UV-C group, the UV-C filtration device was used and in the control, it was not. All trays were opened with the use of a sterile technique and left exposed in the OR for four hours. Air was sampled by a particle counter to measure the numbers of 5µm and 10µm particles. Culture specimens were obtained from the trays to assess for bacterial contamination. Outcome data were collected at 30-minute intervals for the duration of the four-hour study period.

Results

Use of the UV-C device resulted in statistically significant reductions in the numbers of 5µm (average of 64.9% reduction when compared with the control, p<0.001) and 10µm (average of 65.7% reduction when compared with the control, p<0.001)-sized particles detectable in the OR. There was no significant difference in the overall rates of contamination (33.3% in the control group vs. 26.7% in the UV-C group, p=1.0) or the time to contamination (mean survival of 114 minutes in the control group vs. 105 minutes in the UV-C group, p=0.72) of surgical instrument trays with the use of the UV-C device.

Conclusions

The results demonstrate that the UV-C filtration device can successfully reduce airborne bioburden in standard ORs, suggesting that it may have the potential to reduce the risk for wound and hardware infections. Further clinical trials are necessary to better determine the effect of this air filtration system on postoperative infection rates.

Categories: Orthopedics, Quality Improvement, Healthcare Technology

Keywords: orthopedic infections, airborne infection control, microbial contamination, air-contamination, operating room safety, peri-prosthetic joint infection, prosthetic infection, post surgical infection, airborne contamination, ultraviolet filtration

Introduction

Postoperative infection is one of the most challenging complications following orthopedic surgery today. Implant-associated and prosthetic infections represent a substantial burden to patients and healthcare systems [1,2]. The health care costs of prosthetic joint infections alone are enormous, in the billions of dollars annually in the United States alone [2,3]. Implant-associated infections are also associated with substantial morbidity and mortality, with five-year mortality rates after the diagnosis reported at greater than 25% [4,5]. In order to improve patient outcomes and reduce healthcare expenditures, interventions to reduce surgical infections must be employed.

Airborne contamination has been attributed as a potential cause of surgical site infections for decades [6-9]. Aerosolized particles including dust, skin scales, and respiratory particulates arising from patients and

How to cite this article

Hijji F Y, Schneider A D, Reeves J T, et al. (July 14, 2022) Reduction in Operating Room Airborne Particle Burden and Time-Dependent Contamination of Sterile Instrument Trays With the Use of a Novel Air Filtration System. Cureus 14(7): e26864. DOI 10.7759/cureus.26864

surgical staff frequently carry a variety of organisms including *Staphylococcus aureus* and Coagulase-negative staphylococci [7]. These particles eventually settle and can contaminate surgical instruments and/or surgical wounds. Increased airborne particle counts in the operating room (OR) have previously been associated with implant contamination and surgical site infections [1,9-11]. In an effort to minimize intraoperative airborne contamination, hospitals have employed a variety of technologies including high-efficiency particulate arrestance (HEPA) filtration, positive air pressurization, and surgical helmet systems [1]. Guidelines have also been developed to improve OR air quality, however, many of these guidelines do not provide specific criteria to adequately eliminate microbial aerosols or minimum particle count standards [1,12]. Additionally, necessary OR activity such as personnel traffic and surgical gowning have been demonstrated to increase viable airborne particulates that are not well controlled by current OR air systems [13-16]. As a result, poor OR air quality has remained a healthcare issue and a contributing factor in surgical infections.

In order to mitigate airborne contamination and surgical infections, multiple innovative technologies are emerging. One such technology is the crystalline C-band UV (UV-C) air filtration (Aerobiotix Inc., Miamisburg, OH, USA), designed to reduce contamination through high-quality filtration and germicidal irradiation [17]. In this context, the purpose of the present study was to evaluate the ability of a UV-C air filtration device to reduce airborne particles and bacteria in an OR setting. Specifically, this study sought to assess the effects of a specific UV-C filtration device on particle counts and rates of contamination of surgical instrument trays.

Materials And Methods

Group assignment

Thirty sterile surgical instrument trays were randomly assigned to either a control or test group (n=15 per group). All trays underwent data collection in a single, unoccupied positive-air-flow OR during normal daytime hours. Data collection was split across two separate weekend days with an equal number of control and test group trays for each day. On each day, the control surgical tray contamination rate was assessed first in the OR under standard conditions. Test surgical tray contamination rate was then assessed after the control group in the OR with the Illuvia™ HUAIRS (HEPA-UV Air Recirculation System) UV-C air filtration device (Aerobiotix, Inc., Miamisburg, OH, USA) running. The Illuvia™ UV-C filtration device was turned on 30 minutes before test group data collection per the manufacturer's recommendations.

Operating room set-up

Surgical instrument trays were opened using standard sterile technique and placed on top of two sterilely draped back tables. The tables were positioned in the OR in relation to the centrally-placed patient table to simulate a standard total hip arthroplasty. The Illuvia™ UV-C filtration device was positioned diagonally from the air return grates following manufacturer specifications. The position of all equipment in the OR remained the same between groups.

Particle count

A Particles Plus 8305 Handheld Particle Counter (Particles Plus, Inc., Stoughton, MA, USA) was placed atop the OR circulator desk in front of the air return grates. The particle counter was used to obtain the counts of 5µm and 10µm-sized particles within the room; these sizes were selected as they reflect the size range of microbial-carrying particles [18]. Particle counts were obtained immediately after opening surgical trays and every 30 minutes thereafter until an end time point of four hours for each group. Readings were obtained in triplicate at each time point.

Surgical instrument contamination rate

Trays in each group were assessed for contamination using aerobic culture swabs and tubes, as previously described [19]. Each tray was subdivided into four quadrants which were thoroughly swabbed upon opening and every thirty minutes thereafter until an end time point of four hours. All trays remained uncovered during the entire testing period for each group. Cultures were obtained by a single individual who remained sterile in standard OR attire for the entirety of each testing period. To simulate normal OR traffic and airflow disruption, a single individual would pass through the OR from the non-sterile hallway into the sub-sterile core every 10 minutes.

All culture samples were handled by the testing facility's internal lab. Cultures were kept for a minimum of 72 hours. All species grown from cultures were identified without antibiotic sensitivity data. Trays were considered contaminated upon confirmation of a single positive culture from any quadrant within the tray. The time to contamination and the proportion of trays that remained sterile were recorded for all groups.

Statistical analysis

Statistical analyses were performed using Statistical Package For The Social Sciences (SPSS) software version 22 (IBM Corp., Chicago, IL, USA). Continuous variables were compared using a two-tailed Student's T-test.

Categorical variables were analyzed via Fischer's exact test. The time to contamination was compared among the groups with the use of a Kaplan-Meier survivorship curve and the log-rank test. Statistical significance was accepted at $p < 0.05$.

Results

Particle count

Use of the Illuvia™ UV-C filtration unit resulted in significant reductions in the numbers of both the 5µm and 10µm-sized particles when compared with the control. The numbers of 5µm particles were reduced by an average of 64.9% when compared with the control ($p < 0.001$) and the numbers of 10µm particles were reduced by an average of 65.7% ($p < 0.001$) across all time points (Table 1).

Particle Size	Control (SD)	UV-C (SD)	P-value
5 µm	3020 (3136)	1059 (1129)	<0.001
10 µm	1373 (1428)	471 (643)	<0.001

TABLE 1: Mean airborne particle counts

SD: Standard deviation, UV-C: Ultraviolet C

Surgical instrument contamination rate

None of the trays (0/15) yielded positive cultures upon opening at time point zero. The overall contamination rate for the control group was 33.3% (5/15) compared to 26.7% (4/15) for the Illuvia™ UV-C test group ($p = 1.0$). The control group exhibited contamination of one tray at 30 minutes (6.7% total), two trays at 90 minutes (20% total), one tray at 150 minutes (26.7% total), and one tray at 210 minutes (33.3% total). The Illuvia™ UV-C group exhibited contamination of two trays at 60 minutes (13.3% total), one tray at 90 minutes (20% total), and one tray at 210 minutes (26.7% total). The estimated mean survival times for the control and Illuvia™ UV-C groups were 114 and 105 minutes, respectively (Figure 1). When analyzed by the log rank test this was not significantly different ($p = 0.72$).

Overall Comparisons

	Chi-Square	df	Sig.
Log Rank (Mantel-Cox)	.127	1	.722

Test of equality of survival distributions for the different levels of Group.

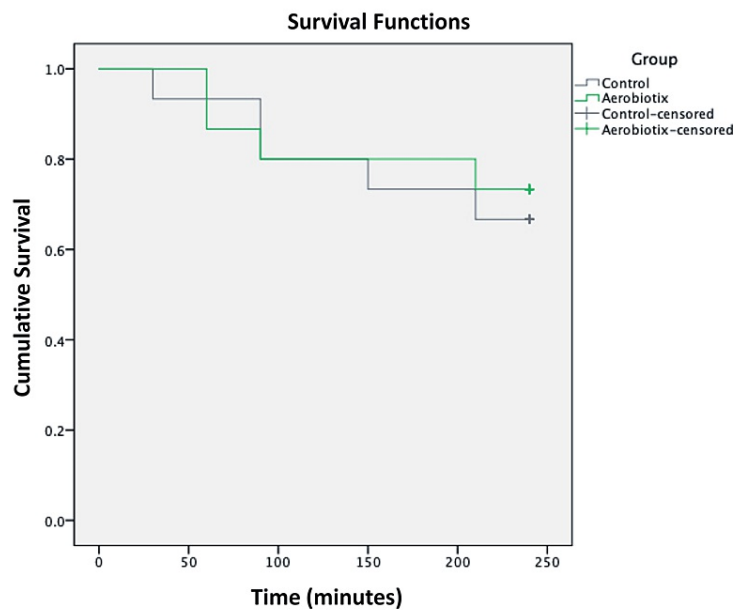


FIGURE 1: Kaplan-Meier estimates of survival (i.e., absence of bacterial contamination) with log rank (Mantel-Cox) test for surgical instrument trays with the UV-C unit (Aerobiotix) and without (Control).

Results of the log rank test are displayed in the above graph.

Only one tray (3.3%) in the control group yielded more than one positive culture at different time points. The nine total trays (including both the control and Illuvia™ UV-C cohorts) that had positive cultures grew: 33% (3/9) *Micrococcus* species, 33% (3/9) *Bacillus* species, 22% (2/9) *Paenibacillus* species, 11% (1/9) *Streptomyces* species, and 11% (1/9) *Staphylococcus epidermidis* (Table 2).

Tray	Time (minutes)	Organism
Control 1	150	<i>Micrococcus luteus</i>
Control 2	30	<i>Bacillus</i> species
Control 6	90	<i>Micrococcus luteus</i>
Control 6	240	<i>Paenibacillus</i> species
Control 12	210	<i>Paenibacillus urinalis</i>
Control 15	90	<i>Staphylococcus epidermidis</i>
UV-C 7	90	<i>Micrococcus</i> species
UV-C 9	60	<i>Bacillus</i> species
UV-C 10	60	<i>Streptomyces</i> species
UV-C 13	210	<i>Bacillus</i> species

TABLE 2: Organisms grown from contaminated (i.e., bacterial culture positive) surgical instrument trays

UV-C: Ultraviolet C

Discussion

The present study sought to determine the effect of a UV-C air filtration device on airborne particulate burden and rates of surgical tray contamination in a simulated OR setting. The results demonstrated that the Illuvia™ UV-C system significantly reduced the number of viable aerosolized particles in the OR, and there was no significant difference in the rates of contamination or “survivability” of opened surgical instrument trays over four hours.

These results suggest that the Illuvia™ UV-C filtration device is successful in reducing air contamination in standard ORs. Devices utilizing HEPA filters have previously been attributed to substantial reductions in air contamination, with some reviews reporting a 99.97% reduction in airborne particles larger than 0.5µm [1,10,17,20,21]. Specifically, in a pilot study assessing the efficacy of the Illuvia™ UV-C filtration unit on particle counts in unoccupied ORs with simulated OR traffic, Curtis et al. demonstrated significant reductions in both viable and total particle counts in ORs containing the device [17]. Similarly, Parvizi et al. reported a 53.4% reduction in OR air contamination following the application of this device [1]. Interventions to reduce air particulate burden in the OR are becoming increasingly important due to the potential effects of air contamination on surgical outcomes [9,10]. However, despite recent engineering standards and practice requirements, the air quality in standard ORs frequently does not reach recommended levels, although there is no universally agreed-upon standard [1,22]. Furthermore, factors such as OR traffic and surgical gowning have been attributed to air contamination that is not adequately controlled by current technologies [13,15,16]. As such, the employment of supplemental systems such as a UV-C air filtration device may be necessary to adequately reduce airborne particulate burdens and improve air quality in modern ORs.

The reduction in air particulates with the Illuvia™ UV-C air filtration system may be partly attributed to the method of air flow as well as the use of UV-C light. Laminar air flow systems have been historically employed in ORs as a method of reducing air contamination and infection rates [6,7,21]. However, recent studies have brought into question the utility of standard laminar air flow filtration devices, noting a potentially increased risk of surgical infections with this type of air delivery [21,23,24]. This increased risk has been attributed to air turbulence that occurs in fringe areas that are not directly underneath the laminar air flow canopies [7,21]. This air can potentially contaminate the tables and instruments that are often located in these areas of the OR. As such, the World Health Organization (WHO) has advised against the use of laminar air flow as a means for reducing surgical infection risk [1]. The Illuvia™ UV-C system employs nonturbulent, unidirectional air flow with a neutral pressure system to filter air in the fringe areas to reduce air entrainment while still avoiding mixing of sterile and contaminated air. In addition, this air filtration system utilizes UV-C germicidal irradiation which has been demonstrated to reduce airborne bacterial counts and inactivate bacterial spores [25,26]. The combination of these mechanisms may have contributed to the substantial decrease in air particulate for the particle sizes known to frequently carry microorganisms.

The reduction of aerosolized particulates in the OR with the Illuvia™ UV-C filtration system may result in

reduced surgical site contamination and subsequent risk for postoperative infections. In a study assessing the rate of prosthetic joint infection (PJI) following total knee arthroplasty, Cook et al. demonstrated a substantially decreased rate of PJI in patients who underwent surgery in an OR with an Illuvia™ UV-C system [27]. Darouiche et al. performed a randomized controlled trial assessing airborne contamination and infection rates following total hip arthroplasty, vascular bypass graft implantation, and instrumented spinal procedures performed with and without an air filtration device [10]. The use of the filtration device was associated with a substantial reduction in both airborne colony forming units (CFUs) and postoperative infections, with no reported infections in the air filtration group. Additionally, high density of airborne CFUs was identified as a significant risk factor for infection. In recent years, elevated airborne particulate counts have become more frequently attributed to instrument contamination as well as surgical site and prosthetic infections [9,24,25,27-30]. While the present study was unable to detect a difference in tray contamination between the Illuvia™ UV-C and the control group, this may be attributable to limitations in the number of trays and data collection in an ideal OR setting. However, the substantial reduction in viable airborne particles with the Illuvia™ UV-C unit in the present study may indicate a potential for reducing surgical site contamination and subsequent infections *in vivo*. As such, further clinical trials employing the Illuvia™ UV-C system are necessary to better illustrate the relationship between the device and its effect on surgical infection rates.

This study has several limitations. First, the present study utilized an OR with a positive pressure ventilation system. As such, these results may not be generalizable to all ORs. Second, the cultures and particle counts were obtained in a clean, unoccupied OR that had not been utilized since the previous night. As such, the particle counts and tray contamination rates may be lower than that found in an occupied OR that contains multiple operative cases throughout the day. Third, while there were a large number of cultures obtained, the total number of overall trays and contaminated trays in the present study was low. As such, the power of the study is limited and may have contributed to the inability to detect a difference in contamination rates between the groups. Fourth, contaminated trays as identified by positive cultures cannot be distinguished from potentially false positive results that occur from contamination during culturing in the laboratory. However, the frequency of contaminated trays in the present study is similar to that of a previous study using similar methods [19], suggesting that the contamination rates observed here are likely reliable.

Conclusions

Postoperative infections are a significant cause of morbidity, mortality, and cost in the current healthcare system. Airborne contamination of surgical wounds and instruments in the OR has been determined to be a significant risk factor for surgical site infections. In order to improve OR air quality, new technologies and policies must be implemented. The present study demonstrated the efficacy of a UV-C air filtration device in reducing microbe-associated airborne particle counts in a simulated OR setting. These results suggest that this device may be effective in mitigating intraoperative airborne contamination, potentially reducing the risk for surgical wound and implant-associated infections. As such, this technology could potentially serve as an adjunctive risk-reduction strategy for patients undergoing orthopedic and other device-related surgical procedures at high risk of environmental contamination. Further clinical trials are necessary to better determine the effect of this air filtration system on postoperative infection rates.

Additional Information

Disclosures

Human subjects: All authors have confirmed that this study did not involve human participants or tissue.

Animal subjects: All authors have confirmed that this study did not involve animal subjects or tissue.

Conflicts of interest: In compliance with the ICMJE uniform disclosure form, all authors declare the following: **Payment/services info:** All authors have declared that no financial support was received from any organization for the submitted work. **Financial relationships:** All authors have declared that they have no financial relationships at present or within the previous three years with any organizations that might have an interest in the submitted work. **Other relationships:** Aerobiotix Inc. (Miamisburg, Ohio, USA)

provided the study materials for this investigation (equipment and supplies), including the UV-C filtration device which is the subject of the investigation, as well as the supplies used for data collection (air particle counter, cultures swabs, and required laboratory testing). No payments were made to any of the authors or to the authors' institution.

References

1. Parvizi J, Barnes S, Shohat N, Edmiston CE: Environment of care: Is it time to reassess microbial contamination of the operating room air as a risk factor for surgical site infection in total joint arthroplasty?. *Am J Infect Control*. 2017, 45:1267-1272. [10.1016/j.ajic.2017.06.027](https://doi.org/10.1016/j.ajic.2017.06.027)
2. Kurtz SM, Lau E, Watson H, Schmier JK, Parvizi J: Economic burden of periprosthetic joint infection in the United States. *J Arthroplasty*. 2012, 27:61-65. [10.1016/j.arth.2012.02.022](https://doi.org/10.1016/j.arth.2012.02.022)
3. Premkumar A, Kolin DA, Farley KX, Wilson JM, McLawhorn AS, Cross MB, Sculco PK: Projected economic burden of periprosthetic joint infection of the hip and knee in the United States. *J Arthroplasty*. 2021, 36:1484-1489. [10.1016/j.arth.2020.12.005](https://doi.org/10.1016/j.arth.2020.12.005)
4. Lange J, Troelsen A, Søballe K: Chronic periprosthetic hip joint infection. A retrospective, observational study on the treatment strategy and prognosis in 130 non-Selected patients. *PLoS One*. 2016, 11:e0163457.

- [10.1371/journal.pone.0163457](https://doi.org/10.1371/journal.pone.0163457)
5. Kurtz SM, Lau EC, Son M-S, Chang ET, Zimmerli W, Parvizi J: Are we winning or losing the battle with periprosthetic joint infection: trends in periprosthetic joint infection and mortality risk for the Medicare population. *J Arthroplasty*. 2018, 33:3238-3245. [10.1016/j.arth.2018.05.042](https://doi.org/10.1016/j.arth.2018.05.042)
 6. Dharan S, Pittet D: Environmental controls in operating theatres. *J Hosp Infect*. 2002, 51:79-84. [10.1053/jhin.2002.1217](https://doi.org/10.1053/jhin.2002.1217)
 7. Chauveaux D: Preventing surgical-site infections: measures other than antibiotics. *Orthop Traumatol Surg Res*. 2015, 101:77-83. [10.1016/j.otsr.2014.07.028](https://doi.org/10.1016/j.otsr.2014.07.028)
 8. Charnley J, Eftekhari N: Postoperative infection in total prosthetic replacement arthroplasty of the hip-joint with special reference to the bacterial content of the air of the operating room. *Br J Surg*. 2005, 56:641-649. [10.1002/bjs.1800560902](https://doi.org/10.1002/bjs.1800560902)
 9. Knobben BAS, Engelsma Y, Neut D, van der Mei HC, Busscher HJ, van Horn JR: Intraoperative contamination influences wound discharge and periprosthetic infection. *Clin Orthop Relat Res*. 2006, 452:236-241. [10.1097/01.blo.0000229339.11351.ea](https://doi.org/10.1097/01.blo.0000229339.11351.ea)
 10. Darouiche RO, Green DM, Harrington MA, Ehni BL, Kougias P, Bechara CF, O'Connor DP: Association of airborne microorganisms in the operating room with implant infections: a randomized controlled trial. *Infect Control Hosp Epidemiol*. 2017, 38:3-10. [10.1017/ice.2016.240](https://doi.org/10.1017/ice.2016.240)
 11. Byrne AM, Morris S, McCarthy T, Quinlan W, O'Byrne JM: Outcome following deep wound contamination in cemented arthroplasty. *Int Orthop*. 2007, 31:27-31. [10.1007/s00264-006-0121-z](https://doi.org/10.1007/s00264-006-0121-z)
 12. Sehulster L, Chinn RYW, CDC, HICPAC: Guidelines for environmental infection control in health-care facilities. Recommendations of CDC and the Healthcare Infection Control Practices Advisory Committee (HICPAC). *MMWR Recomm Rep*. 2003, 52:1-42.
 13. Mears SC, Blanding R, Belkoff SM: Door opening affects operating room pressure during joint arthroplasty. *Orthopedics*. 2015, 38:991-994. [10.3928/01477447-20151020-07](https://doi.org/10.3928/01477447-20151020-07)
 14. Andersson AE, Bergh I, Karlsson J, Eriksson BI, Nilsson K: Traffic flow in the operating room: an explorative and descriptive study on air quality during orthopedic trauma implant surgery. *Am J Infect Control*. 2012, 40:750-755. [10.1016/j.ajic.2011.09.015](https://doi.org/10.1016/j.ajic.2011.09.015)
 15. Noguchi C, Koseki H, Horiuchi H, et al.: Factors contributing to airborne particle dispersal in the operating room. *BMC Surg*. 2017, 17:78. [10.1186/s12893-017-0275-1](https://doi.org/10.1186/s12893-017-0275-1)
 16. Fraser JF, Young SW, Valentine KA, Probst NE, Spangehl MJ: The gown-glove interface is a source of contamination: a comparative study. *Clin Orthop Relat Res*. 2015, 475:2291-2297. [10.1007/s11999-014-4094-8](https://doi.org/10.1007/s11999-014-4094-8)
 17. Curtis GL, Faour M, Jawad M, Klika AK, Barsoum WK, Higuera CA: Reduction of particles in the operating room using ultraviolet air disinfection and recirculation units. *J Arthroplasty*. 2018, 33:196-200. [10.1016/j.arth.2017.11.052](https://doi.org/10.1016/j.arth.2017.11.052)
 18. Hansen D, Krabs C, Benner D, Brauksiepe A, Popp W: Laminar air flow provides high air quality in the operating field even during real operating conditions, but personal protection seems to be necessary in operations with tissue combustion. *Int J Hyg Environ Health*. 2005, 208:455-460. [10.1016/j.ijheh.2005.08.008](https://doi.org/10.1016/j.ijheh.2005.08.008)
 19. Dalstrom DJ, Venkatarayappa I, Manternach AL, Palcic MS, Heyse BA, Prayson MJ: Time-dependent contamination of opened sterile operating-room trays. *J Bone Joint Surg Am*. 2008, 90:1022-1025. [10.2106/JBJS.G.00689](https://doi.org/10.2106/JBJS.G.00689)
 20. Soots G, Leclerc H, Pol A, Savage C, Fieve R: Air-borne contamination hazard in open heart surgery. Efficiency of HEPA air filtration and laminar flow. *J Cardiovasc Surg*. 1982, 23:155-162.
 21. McHugh SM, Hill ADK, Humphreys H: Laminar airflow and the prevention of surgical site infection. More harm than good?. *Surg*. 2015, 13:52-58. [10.1016/j.surge.2014.10.003](https://doi.org/10.1016/j.surge.2014.10.003)
 22. Sossai D, Dagnino G, Sanguineti F, Franchini F: Mobile laminar air flow screen for additional operating room ventilation: reduction of intraoperative bacterial contamination during total knee arthroplasty. *J Orthop Traumatol*. 2011, 12:207-211. [10.1007/s10195-011-0168-5](https://doi.org/10.1007/s10195-011-0168-5)
 23. Merollini KMD, Zheng H, Graves N: Most relevant strategies for preventing surgical site infection after total hip arthroplasty: guideline recommendations and expert opinion. *Am J Infect Control*. 2013, 41:221-226. [10.1016/j.ajic.2012.03.027](https://doi.org/10.1016/j.ajic.2012.03.027)
 24. Pasquarella C, Pitzurra O, Herren T, Poletti L, Savino A: Lack of influence of body exhaust gowns on aerobic bacterial surface counts in a mixed-ventilation operating theatre. A study of 62 hip arthroplasties. *J Hosp Infect*. 2003, 54:2-9. [10.1016/S0195-6701\(03\)00077-X](https://doi.org/10.1016/S0195-6701(03)00077-X)
 25. Lidwell OM: Ultraviolet radiation and the control of airborne contamination in the operating room. *J Hosp Infect*. 1994, 28:245-248. [10.1016/0195-6701\(94\)90088-4](https://doi.org/10.1016/0195-6701(94)90088-4)
 26. Evans RP: Current concepts for clean air and total joint arthroplasty: laminar airflow and ultraviolet radiation: a systematic review. *Clin Orthop Relat Res*. 2011, 469:945-953. [10.1007/s11999-010-1688-7](https://doi.org/10.1007/s11999-010-1688-7)
 27. Cook TM, Piatt CJ, Barnes S, Edmiston CE: The Impact of supplemental intraoperative air decontamination on the outcome of total joint arthroplasty: a pilot analysis. *J Arthroplasty*. 2019, 34:549-553. [10.1016/j.arth.2018.11.041](https://doi.org/10.1016/j.arth.2018.11.041)
 28. Lidwell OM, Lowbury EJJ, Whyte W, Blowers R, Stanley SJ, Lowe D: Airborne contamination of wounds in joint replacement operations: the relationship to sepsis rates. *J Hosp Infect*. 1983, 4:111-131. [10.1016/0195-6701\(83\)90041-5](https://doi.org/10.1016/0195-6701(83)90041-5)
 29. Gosden PE, MacGowan AP, Bannister GC: Importance of air quality and related factors in the prevention of infection in orthopaedic implant surgery. *J Hosp Infect*. 1998, 39:173-180. [10.1016/S0195-6701\(98\)90255-9](https://doi.org/10.1016/S0195-6701(98)90255-9)
 30. Edmiston CE, Seabrook GR, Cambria RA, et al.: Molecular epidemiology of microbial contamination in the operating room environment: is there a risk for infection?. *Surgery*. 2005, 138:573-582. [10.1016/j.surg.2005.06.045](https://doi.org/10.1016/j.surg.2005.06.045)