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Bats and their virome: an important source of emerging viruses capable of infecting humans

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Bats are being increasingly recognized as an important reservoir of zoonotic viruses of different families, including SARS coronavirus, Nipah virus, Hendra virus and Ebola virus. Several recent studies hypothesized that bats, an ancient group of flying mammals, are the major reservoir of several important RNA virus families from which other mammalian viruses of livestock and humans were derived. Although this hypothesis needs further investigation, the premise that bats carry a large number of viruses is commonly accepted. The question of whether bats have unique biological features making them ideal reservoir hosts has been the subject of several recent reviews. In this review, we will focus on the public health implications of bat derived zoonotic viral disease outbreaks, examine the drivers and risk factors of past disease outbreaks and outline research directions for better control of future disease events.

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Introduction

Approximately 75% of emerging infectious diseases are zoonoses [1,2]. The rate of emergence of zoonotic viruses appears to be increasing and/or our ability to detect new viruses is improving. Viruses are well adapted to their reservoir hosts and therefore exhibit stability within their host's cellular and ecological environments and display little or no clinical disease in these species. However, when a virus jumps the species barrier and spills over into humans, the effects can be devastating. Only when these viruses from wildlife spillover to humans or their domesticated animals and cause mortalities, does this become a significant concern for public health. High mortality rates often characterize these spillover events as do high economic losses. This is particularly true for the bat borne viruses that have emerged in the last 20 years such as Severe Acute Respiratory Syndrome coronavirus (SARS-CoV), Nipah virus (NiV) Hendra virus (HeV) and Ebola virus (EBOV). International travel has contributed to the transmission of many infectious diseases and the bat borne zoonotic viruses are no exception with the most notable being the transmission of SARS-CoV from the key port in Hong Kong to Canada, the Americas, Europe, Asia and Australasia resulting in 8422 cases with 916 deaths (10.9% case fatality) [3].

Bats, order Chiroptera, comprise greater than 20% of living mammalian species with more than 1100 species across 17 families [4]. They are among the most ancient of mammals, their extensive speciation occurred before development of most modern mammals, and they are the only mammals capable of powered flight. Bats in general are also very long-lived and more widely dispersed globally than other mammals; they play vital roles as pollinators of hundreds of species of plants and trees and in the control of arthropod populations. More recently, resurgence in interest in bat biology and ecology has been sparked by their recognition as the known reservoir hosts of some of the most deadly viral zoonoses [5], to which they appear resistant to any pathogenic effects. Whether bats are unique or special as a reservoir of viruses has been the subject of several recent publications [6,7] and will not be covered in this review. This review will focus on the ecology, pathogen-host interface and public health considerations of selected bat zoonotic viruses.

Key examples of bat zoonotic viruses Hendra virus (HeV)

HeV first emerged in 1994 in an outbreak of respiratory disease that infected 20 horses and two humans, resulting in the death of all horses and one of the humans [8,9]. There have been a total of 39 spillover events identified resulting in the infection of 78 horses, one dog and seven humans. In humans the case fatality rate from HeV infection is 57%. The four species of flying fox, *Pteropus poliocephalus*, *P. alecto*, *P. scapulatus* and *P. conspicillatus* have been found to be seropositive for HeV antibodies [10] and all have detectable virus in their urine [11[•],12]. All human cases of HeV infection have been associated with veterinarians and the continual outbreaks have resulted in many veterinarians leaving the equine field due to fears of HeV infection and legal liability [13].

Nipah virus (NiV)

There have been two clusters of Nipah virus outbreaks detected since the end of the 1990s, one in Malaysia/

Singapore and the other in Bangladesh/India. NiV first emerged in Malaysia, causing an outbreak of neurological and respiratory disease affecting pigs between September 1998 and June 1999. Transmission from pigs to humans resulted in 283 human cases and 109 deaths (39% case fatality) in Malaysia and 11 cases and one death in Singaporean abattoir workers [14,15]. The outbreak was controlled by the slaughtering of over 1.1 million pigs [16]. The reservoir hosts of NiV have been identified as *Pteropus vampyrus* and *Pteropus hypomenalus* in Malaysia [17,18,19[•]] and *P. giganteus* in Bangladesh and India [20,21].

Since 2001, outbreaks of NiV in Bangladesh and India have occurred almost on an annual basis with a total of 302 cases and 210 human deaths (69.5% case fatality) [22,23]. In Bangladesh and India, NiV shedding in *P. giganteus* bats appears to result in direct transmission to humans via consumption of raw date palm sap contaminated with bat excreta (saliva, urine, faeces) without transmission through an intermediate host [24^{••},25]. Person to person transmission has also been observed [26,27].

Severe Acute Respiratory Syndrome coronavirus (SARS-CoV)

In 2003, a global outbreak of SARS started in southern China and Hong Kong leading to 8422 cases and 916 human deaths (10.9% case fatality) worldwide [28]. For the transmission of SARS-CoV to humans, the virus required rapid adaption through the intermediary host which was thought to be the palm civet (*Paguma* sp.) where adaption allowed for transmission of the sufficiently fit virus to humans [29^{••}]. The cave dwelling fruit bat *Rousettus leschenaultia* was identified as the reservoir host of SARS-like coronaviruses [29^{••}]. The SARS-CoV outbreak was estimated to have cost \$US54 billion globally [30].

Ebola virus (EBOV)

The transmission of EBOV to humans has usually occurred through the capture and slaughtering of animals, commonly non-human primates, for 'bush meat' [31]. The fruit bats *Hypsignathus monstrosus*, *Epomops franqueti* and *Myonycteris torquata* are possible reservoirs for EBOV in Africa based on serological surveys [32^{••}]. The initial outbreaks of EBOV in 1976 in Sudan and Zaire were exacerbated by the reuse of contaminated needles [33,34]. Person to person transmission occurs through contact with bodily fluids (blood, semen, organs, urine, faeces and secretions) [35] including contact with cadavers [36].

Initially identified in imported macaques in the USA, Ebola Reston has recently emerged in pigs in the Philippines and poses a concern for Public Health and agriculture. Of particular significance was the co-infection of the pigs with porcine reproductive and respiratory syndrome virus and porcine circovirus type 2. Serological studies identified six people that had seroconverted to Ebola Reston virus, all of which has contact with sick pigs [37]. Importantly, experimental infection with Ebola Reston virus found that the disease was asymptomatic in pigs despite shedding, indicating a risk for farm and abattoir workers [38]. Antibodies to Ebola Reston virus were detected in *Rousettus amplexicaudatus* bats implicating this bat as a potential reservoir host [39].

In addition to the 'high impact' bat zoonotic viruses discussed above, there are other bat viruses which have caused zoonotic infections. See Table 1 for a summary list of major bat zoonotic viruses recently emerged.

Understanding the ecology of disease emergence

There are many potential drivers that can contribute to a spillover of zoonotic bat-borne disease (Table 1 and Figure 1). These factors can be extrinsic and/or intrinsic. Extrinsic factors such as environmental and anthropogenic stressors can have an effect on the ecology of disease in bats. Environmental stressors such as climatic events (typhoons/cyclones and droughts) that destroy habitat and food resources have been hypothesized to have an impact on the health of bats [40-43,44[•]]. In addition, human activities are selecting for some species of bats that are synanthrophic, and so are benefiting from living close to humans, thereby increasing their numbers and the risk of transmission of disease to humans [45]. Human activities are artificially increasing animal densities by changing the land use and this is increasing the contact between humans, domesticated animals and bats [46]. In the cases of Hendra virus, Nipah virus and Menangle virus, domesticated animals are the amplifying hosts of these zoonotic viruses [15,47,48]. Habitat change such as deforestation force changes in roosting sites and can lead to alterations in the population density and the migratory patterns of bats. These anthropogenic activities may be impacting our ecosystem in such a way that the equilibrium is disturbed and spillover of zoonotic viruses readily occur [49-51].

In the case of the NiV outbreak in Malaysia, the drivers of the outbreak were primarily agricultural intensification and more specifically the co-location of pig farms and fruit orchards. It is believed that the fruiting trees overhanging the intensive pig farms attracted flying foxes, leading to NiV spillover into pigs [52]. It has been hypothesized that the Nipah virus outbreak in pigs was contributed by the El Nino Southern Oscillation induced drought and subsequent forest fires in Indonesia that produced a smoke haze that led to the migration of flying foxes into Malaysia and the subsequent spillover of NiV into pigs [52]. However, this hypothesis was refuted by a later study which suggested that there were multiple introductions of NiV into the piggery. It was thought that repeated introductions of NiV allowed for viral persistence to be established





Examples of drivers responsible for zoonotic virus spillover from bats.

within the pig population which in turn resulted in transmission of NiV to humans [53]. NiV underwent a host shift from fruit bats into pigs that allowed for high levels of virus to be shed and persist within a new host. Management practices of relocating pigs to grower farms further spread the virus. Trade allowed for further NiV transmission with pigs being transported to Singapore for processing increasing the spread of the outbreak [14].

The drivers for transmission of NiV in Bangladesh differed to those in Malaysia. In Bangladesh, cultural practices of consumption and trade of date palm sap have allowed for the transmission of NiV from bats to humans [24,25]. Traditional social practices of family members caring for the sick in the absence of barrier nursing have resulted in person-to-person transmission further contributing to outbreaks of disease [26,27].

Similarly, cultural traditions of eating wild animal meat or 'bush meat' in Africa and Asia have led to outbreaks of EBOV and SARS-CoV. In the case of EBOV, the transmission has been further exacerbated by the lack of barrier nursing and the use of traditional burial practices where mourners make contact with the deceased [36]. Desire for wild animal meat including bats, and trade in live wet markets have allowed susceptible animals to come into contact with bats enabling the subsequent transmission of SARS-CoV into humans [54,55]. Once established, person-to-person transmission occurred and the virus was widely spread by international travellers [56]. Importantly, international collaboration allowed for the aetiology of the outbreak to be rapidly determined. This in turn allowed for the development of diagnostic assays for the detection of the virus and greatly assisted in the control of the outbreak [3]. Recently, a novel coronavirus isolated from a Saudi make patient suffering pneumonia and renal failure was determined to be most similar to bat coronaviruses through an international collaboration [57,58[•]]. Following the posting on Promed of this case [57], another coronavirus with 99.5% similarity to the virus was identified as the causative agent of a Saudi man from Qatar (and hospitalized in London) suffering from a severe respiratory illness [59]. This allowed the development of real-time molecular assays for the rapid detection of these viruses [60]. More recently, another Saudi man has been diagnosed with a similar coronavirus infection [61].

Overall, ecology-based management of drivers that can lead to spillover are likely to be more effective than movement or culling, as they have the potential to reduce disease susceptibility in the reservoir host and opportunities for transmission [42,44].

Understanding the virus-bat interface

Our understanding of virus-host interactions is in its infancy. The virus-bat interface is impacted by the extrinsic factors mentioned above, as well as viral and

Table 1

Virus	Drivers	Risk factors/modes of transmission	References
SARS-CoV	Economic growth Desire for game meat Live wild animal trading in wet markets International travel	Slaughtering Social/cultural practices Farming of wild animals Laboratory acquired infection	[29**,54,55,93]
Ebola virus	Desire for game meat (Bush meat) Live wild animal trading Burial practices	Slaughtering/hunters Social/cultural Practices Poor health care practices	[31]
Marburg virus	Infected monkeys used for research Mining Tourism	Laboratory acquired infection Caves (eco-tourism) Mining	[84,94–96]
Hendra virus	Population growth/urbanization/ human encroachment/ synanthrophy Climate change Starvation Reproductive stress	Inadequate PPE for veterinarians Intermit contact with horses (cuts, abrasions, respiratory secretions) For example performing necropies	[13,46,62]
Nipah virus (Bangladesh)	Date palm juice Cultural tradition	Drinking date palm juice Caring for infected patients	[26,27,90,97,98]
Nipah virus (Malaysia)	Agricultural intensification (dual land use) Encroachment into forested areas Movement of pigs to grower piggeries within Malaysia Food processing in Singapore Trade Habitat destruction Stress	Piggery workers (aerosols, husbandry practices) Abbatoir workers (slaughtering)	[53,99,100]
Reoviruses (Melaka virus and related viruses)	Urbanization Tourism	Close proximity to bats Eco-tourism	[101,102]
Menangle virus	Agricultural practices (transportation of pigs within farm) Movement of pigs to grower piggeries Close proximity to flying fox colony	Piggery workers (husbandry practices — birthing and necropies without PPE)	[47]
Lyssaviruses for example Rabies, Australian bat lyssavirus (ABLV), European bat lyssaviruses (EBLV) 1 and 2, Lagos bat virus, Duvenhage virus	Urbanization Deforestation Synanthrophy	Bat carers Cohabitation with bats in houses	[42,44 *]

host factors (intrinsic factors). There are multiple intrinsic factors of the reservoir host such as age, body condition, reproductive status, sex and social status that are impacted by stress. For instance, stresses due to starvation and the breeding season for males may play an important role in the epidemiology of disease as stress caused by these factors may dampen the immune response making individuals more susceptible to infection [62]. In the case of EBOV, it is thought that starvation leads to fruit bats and primates coming into close proximity during their quest for food, facilitating spillover [63]. Transmission of viruses could also occur following changes in the hierarchy of the colony that lead to fighting for dominance, and during courtship and mating via grooming and biting [7]. Seasonal periods when juveniles have waning maternal antibody and are therefore susceptible to virus infection have been associated with increased transmission of Marburg virus to humans [64^{••}] and have been thought to have a role in increasing the incidence of transmission within bat colonies for HeV as well as Marburg virus [46,62,64]. Bats live on average 3.5 times longer than a mammal of similar size [65], hence longevity in bats promotes persistence of viruses in the reservoir host, while the ability to fly allows long-distance dispersal of the infectious agent. Aerosols, direct contact and arthropods could also serve as vectors for transmission within the colony or to other species.

Little is known about the diversity of viruses, the amount of virus present, the mechanisms of shedding,

the incidence of supershedders or the contact rates between infectious and susceptible individuals. From metagenomic analyses, bats harbour a range of viruses and there is a possibility of multiple viral infections in bats spilling over [66–68]. Notably, all of the zoonotic viruses of bat origin so far identified are RNA viruses. Many of these highly pathogenic viruses display a broad cell tropism, being able to infect a wide range of cells and hosts (HeV, NiV, EBOV, SARS-CoV) [69–72]. Viruses such as HeV, lyssaviruses and NiV show high genome conservation within their bat hosts, suggesting that they are under strong selective constraints [11[•],73–76].

Public health considerations: prevention and control strategies

Understanding the ecology of bat-borne viral pathogens and identifying the triggers of an outbreak will assist in the control or reduction of emerging zoonotic disease outbreaks. By understanding the mechanisms of emergence, outbreak management plans can be developed and risk mitigation processes can be implemented. Once identified, risk reduction strategies can be implemented through education of the general public, doctors, veterinarians and policy makers [77]. Measures such as the wearing of appropriate protective equipment (PPE) when caring for patients or animals and restriction on the sale and consumption of game meat would reduce the risks of transmission of bat-borne viruses.

Following outbreaks of disease, public health measures implemented have included enhanced surveillance and increased infection control, whether it is in hospitals in the case of SARS-CoV, EBOV and NiV, or during veterinary procedures in the case of HeV. Quarantine and contact tracing to limit the spread of viruses have also implemented in outbreaks [16,78,79^{••},80–84]. A communication strategy is implemented to inform the public aims to reduce further spread by avoidance of risky activities or alteration of activities. For example, following the recognition of SARS, response teams were formed and communication to the public was instigated including global alerts from the World Health Organization [85].

Implementation of prevention and control measures can be carried out at many different levels. At the farming level, changing agricultural practices by the creation of buffers between fruiting trees and domesticated animals would significantly reduce the transmission for HeV and NiV, which is believed to have already played an important role in preventing further NiV outbreaks in Malaysia [86,87]. The introduction of biocontainment measures within piggeries, including surveillance of pigs being transferred between farms and sent to abattoirs, is another effective approach which can be applied at the farming and trade level [87–89]. Since some of these zoonotic agents involve an intermediate host of livestock importance, it is highly important that veterinarians wear appropriate PPE when performing procedures on animals to minimize the risk of transmission of zoonotic diseases.

In addition to the above general strategies, some diseasespecific prevention and control measures can also be applied. In Bangladesh, the installation of barriers on the date palms that prevent the bats from accessing the collection vessels is a simple strategy currently being investigated to control transmission of NiV. This strategy also improves the quality of sap and therefore results in a higher price [90,91,92^{••}]. For prevention of HeV infection in humans, a One Health approach is being adopted. This involved the development of a recombinant proteinbased vaccination program for horses in high-risk areas. The vaccination aims to achieve two purposes: prevent horses from HeV infection and, more importantly, block the horse-to-human transmission. With the recent release of a vaccine against HeV in early November 2012, it will be interesting to see whether this One Health strategy is effective in interrupting the zoonotic transmission cycle of HeV.

Strategies for the development of surveillance for new and emerging diseases and the management of bats need to be developed. With increased interaction between humans and their domesticated animals and bats, increasing rates of infections will continue to occur. In the past, there has been passive surveillance on dead animals, and it is time to form an international consortium for active surveillance of different bat populations to detect potential zoonotic agents as well as unknown viruses of low pathogenicity that could combine with other viruses to be pathogenic [42]. The use of new technologies such as high throughput sequencing and multiplex serological tests should be an integral part of this effort to increase our ability for pre-emergence monitoring of potential zoonotic pathogens.

Future directions

With human activity increasingly overlapping the habitats of bats, there is no doubt that zoonotic viruses will continue to emerge from these species. In order to help predict and prevent the emergence of viruses, we need a greater understanding of the infection dynamics within their hosts and to understand the impact of human changes to the environment on the potential for virus spillover. A fully integrated One Health approach with international scientists, ecologists, veterinarians, health professionals, social scientists and politicians working together is required to minimize the impact of bat borne zoonotic diseases. It is critical that we are able to coexist with bats as these unique creatures are vital to our ecosystem.

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References and recommended reading

Papers of particular interest, published within the period of review, have been highlighted as:

- of special interest
- •• of outstanding interest
- 1. Taylor LH, Latham SM, Woolhouse ME: Risk factors for human disease emergence. *Philos Trans R Soc Lond B Biol Sci* 2001, 356:983-989.
- Jones KE, Patel NG, Levy MA, Storeygard A, Balk D, Gittleman JL, Daszak P: Global trends in emerging infectious diseases. *Nature* 2008, 451:990-993.
- 3. World Health Organization: **SARS: lessons from a new disease**. *The World Health Report 2003.* 2004:71-82.
- 4. Nowak R: *Walker's Bats of the World*. The Johns Hopkins University Press; 1994.
- Calisher CH, Childs JE, Field HE, Holmes KV, Schountz T: Bats: important reservoir hosts of emerging viruses. *Clin Microbiol Rev* 2006, 19:531-545.
- Wang LF, Walker PJ, Poon LL: Mass extinctions, biodiversity and mitochondrial function: are bats "special" as reservoirs for emerging viruses? *Curr Opin Virol* 2011, 1:649-657.
- Wong S, Lau S, Woo P, Yuen KY: Bats as a continuing source of emerging infections in humans. *Rev Med Virol* 2007, 17:67-91.
- Murray K, Rogers RJ, Selvey LA, Selleck P, Hyatt AD, Gould AR, Gleeson LJ, Hooper PT, Westbury H: A novel morbillivirus pneumonia of horses and its transmission to humans. *Emerg Infect Dis* 1995, 1:31-33.
- Selvey LA, Wells RM, McCormack JG, Ansford AJ, Murray K, Rogers RJ, Lavercombe PS, Selleck P, Sheridan JW: Infection of humans and horses by a newly described morbillivirus. *Med J Aust* 1995, 162:642-645.
- Young PL, Halpin K, Selleck PW, Field H, Gravel JL, Kelly MA, Mackenzie JS: Serologic evidence for the presence in *Pteropus* bats of a paramyxovirus related to equine morbillivirus. *Emerg Infect Dis* 1996, 2:239-240.
- Smith I, Broos A, de Jong C, Zeddeman A, Smith C, Smith G, Moore F, Barr J, Crameri G, Marsh G et al.: Identifying Hendra

virus diversity in pteropid bats. *PLoS One* 2011, **6**:e25275. First report of the isolation of HeV from the urine of naturally infected flying foxes, and reported that the genome is highly conserved between bat, horses and human isolates

- 12. Field H, de Jong C, Melville D, Smith C, Smith I, Broos A, Kung YH, McLaughlin A, Zeddeman A: **Hendra virus infection dynamics in Australian fruit bats**. *PLoS One* 2011, **6**:e28678.
- Mendez DH, Judd J, Speare R: Unexpected result of Hendra virus outbreaks for veterinarians, Queensland, Australia. Emerg Infect Dis 2012, 18:83-85.
- Paton NI, Leo YS, Zaki SR, Auchus AP, Lee KE, Ling AE, Chew SK, Ang B, Rollin PE, Umapathi T *et al.*: Outbreak of Nipah-virus infection among abattoir workers in Singapore. *Lancet* 1999, 354:1253-1256.
- Chua KB, Bellini WJ, Rota PA, Harcourt BH, Tamin A, Lam SK, Ksiazek TG, Rollin PE, Zaki SR, Shieh W *et al.*: Nipah virus: a recently emergent deadly paramyxovirus. *Science* 2000, 288:1432-1435.
- Field H, Young P, Yob JM, Mills J, Hall L, Mackenzie J: The natural history of Hendra and Nipah viruses. *Microbes Infect* 2001, 3:307-314.
- Yob JM, Field H, Rashdi AM, Morrissy C, van der Heide B, Rota P, bin Adzhar A, White J, Daniels P, Jamaluddin A et al.: Nipah virus infection in bats (order Chiroptera) in peninsular Malaysia. Emerg Infect Dis 2001, 7:439-441.
- Chua KB, Koh CL, Hooi PS, Wee KF, Khong JH, Chua BH, Chan YP, Lim ME, Lam SK: Isolation of Nipah virus from Malaysian Island flying-foxes. *Microbes Infect* 2002, 4:145-151.

 19. Chua KB: A novel approach for collecting samples from fruit
 bats for isolation of infectious agents. *Microbes Infect* 2003, 5:487-490.

Description of the successful isolation of NiV, other paramyxoviruses and bacteria using a method that did not involve capture of bats for sampling

- Epstein JH, Prakash V, Smith CS, Daszak P, McLaughlin AB, Meehan G, Field HE, Cunningham AA: Henipavirus infection in fruit bats (*Pteropus giganteus*), India. Emerg Infect Dis 2008, 14:1309-1311.
- Yadav PD, Raut CG, Shete AM, Mishra AC, Towner JS, Nichol ST, Mourya DT, Jonathan S: Short report: detection of Nipah virus RNA in fruit bat (*Pteropus giganteus*) from India. Am J Trop Med Hyg 2012, 87:576-578.
- 22. World Health Organization Regional Office for South East Asia: *Nipah virus infection* 2009.
- 23. World Health Organization Regional Office for South East Asia: Nipah virus outbreaks in the WHO South-East Asia Region 2012.
- 24. Luby SP, Rahman M, Hossain MJ, Blum LS, Husain MM, Gurley E,
- •• Khan R, Ahmed BN, Rahman S, Nahar N et al.: Foodborne transmission of Nipah virus, Bangladesh. Emerg Infect Dis 2006, **12**:14-16.

Recognized Date palm sap as a vehicle for transmission of NiV which is important for the control of transmission of the virus

- Rahman MMA, Hossain MJ, Sultana S, Homaira N, Khan SU, Gurley ES, Rollin PE, Lo MK, Comer JA, Lowe L et al.: Date palm sap linked to Nipah virus outbreak in Bangladesh, 2008. Vector Borne Zoonotic Dis 2012, 12:65-72.
- Gurley ES, Montgomery JM, Hossain MJ, Bell M, Azad AK, Islam MR, Molla MA, Carroll DS, Ksiazek TG, Rota PA *et al.*: Person-to-person transmission of Nipah virus in a Bangladeshi community. *Emerg Infect Dis* 2007, **13**:1031-1037.
- Blum LS, Khan R, Nahar N, Breiman RF: In-depth assessment of an outbreak of Nipah encephalitis with person-to-person transmission in Bangladesh: implications for prevention and control strategies. Am J Trop Med Hyg 2009, 80:96-102.
- World Health Organization: Summary table of SARS cases by country, 1 November 2002-7 August 2003 2003.
- 29. Li W, Shi Z, Yu M, Ren W, Smith C, Epstein JH, Wang H, Crameri G,
 Hu Z, Zhang H *et al.*: Bats are natural reservoirs of SARS-like

coronaviruses. *Science* 2005, **310**:676-679. Identification of bats as the reservoir host of SARS-like coronaviruses

- Lee J-W, McKibbon W: Estimating the global cost of SARS. In Learning from SARS: Preparing for the Next Disease Outbreak – Workshop Summary. Edited by Knobler S, Mahmoud A, Lemon S, Mad 109A, Sivitz L, Oberholtzer K. National Academies Press; 2004:
- Leroy EM, Epelboin A, Mondonge V, Pourrut X, Gonzalez J-P, Muyembe-Tamfum J-J, Formenty P: Human Ebola outbreak resulting from direct exposure to fruit bats in Luebo, Democratic Republic of Congo, 2007. Vector Borne Zoonotic Dis 2009, 9:723-728.
- 32. Leroy EM, Kumulungui B, Pourrut X, Rouquet P, Hassanin A,
 Yaba P, Délicat A, Paweska JT, Gonzalez J-P, Swanepoel R: Fruit

bats as reservoirs of Ebola virus. Nature 2005, 438:575-576. Detection of antibodies against EBOV in bats confirming them as a

reservoir host

- World Health Organization: Ebola haemorrhagic fever in Zaire, 1976. Bull World Health Organ 1978, 56:271-293.
- 34. World Health Organization: Ebola haemorrhagic fever in Sudan, 1976. Bull World Health Organ 1978, 56:247-270.
- 35. Bausch DG, Towner JS, Dowell SF, Kaducu F, Lukwiya M, Sanchez A, Nichol ST, Ksiazek TG, Rollin PE: Assessment of the risk of Ebola virus transmission from bodily fluids and fomites. *J Infect Dis* 2007, **196(Suppl)**:S142-S147.
- Dowell SF, Mukunu R, Ksiazek TG, Khan AS, Rollin PE, Peters CJ: Transmission of Ebola hemorrhagic fever: a study of risk factors in family members, Kikwit, Democratic Republic of the Congo, 1995. J Infect Dis 1995, 179:S87-S91.
- 37. World Health Organization: *Ebola Reston in pigs and humans in the Philippines* 2009.

- Marsh GA, Haining J, Robinson R, Foord A, Yamada M, Barr JA, Payne J, White J, Yu M, Bingham J et al.: Ebola Reston virus infection of pigs: clinical significance and transmission potential. J Infect Dis 2011, 204:S804-S809.
- Taniguchi S, Watanabe S, Masangkay JS, Omatsu T, Ikegami T, Alviola P, Ueda N, Iha K, Fujii H, Ishii Y et al.: Reston Ebolavirus antibodies in bats, the Philippines. Emerg Infect Dis 2011, 17:1559-1560.
- 40. Field HE: **Bats and emerging zoonoses: henipaviruses and SARS**. *Zoonoses Public Health* 2009, **56**:278-284.
- 41. Wood JLN, Leach M, Waldman L, Macgregor H, Fooks AR, Jones K, Restif O, Dechmann D, Hayman DTS, Baker KS et al.: A framework for the study of zoonotic disease emergence and its drivers: spillover of bat pathogens as a case study. *Philos Trans R Soc Lond B Biol Sci* 2012, 367:2881-2892.
- 42. Mathews F: **Zoonoses in wildlife integrating ecology into management**. *Advances in Parasitology*. Elsevier Ltd.; 2009:. 185–209.
- 43. Daszak P, Cunningham AA, Hyatt AD: Anthropogenic environmental change and the emergence of infectious diseases in wildlife. *Acta Trop* 2001, **78**:103-116.
- 44. Streicker DG, Recuenco S, Valderrama W, Gomez Benavides J,
- Vargas I, Pacheco V, Condori Condori RE, Montgomery J, Rupprecht CE, Rohani P et al.: Ecological and anthropogenic drivers of rabies exposure in vampire bats: implications for transmission and control. Proc R Soc Biol Sci 2012, 279:3384-3392.

A good example of how culling is not the answer to controlling bat borne viruses

- 45. McFarlane R, Sleigh A, McMichael T: **Synanthropy of wild** mammals as a determinant of emerging infectious diseases in the Asian-Australasian region. *EcoHealth* 2012, **9**:24-35.
- Plowright RK, Foley P, Field HE, Dobson AP, Foley JE, Eby P, Daszak P: Urban habituation, ecological connectivity and epidemic dampening: the emergence of Hendra virus from flying foxes (*Pteropus spp.*). *Proc Biol Sci* 2011, 278:3703-3712.
- Philbey A, Kirkland PD, Ross AD, Davis RJ, Gleeson A, Love RJ, Daniels PW, Gould A, Hyatt A: An apparently new virus (family Paramyxoviridae) infectious for pigs, humans, and fruit bats. *Emerg Infect Dis* 1998, 4:269-271.
- Murray K, Selleck P, Hooper P, Hyatt A, Gould AR, Gleeson LJ, Westbury H, Hiley L, Selvey LA, Rodwell B *et al.*: A morbillivirus that caused fatal disease in horses and humans. *Science* 1995, 268:94-97.
- 49. Wolfe ND, Daszak P, Kilpatrick AM, Burke DS: **Deforestation, and** prediction of zoonotic disease emergence. *Emerg Infect Dis* 2005, **11**:1822-1827.
- 50. Morse SS: Factors and determinants of disease emergence. *Rev Sci Tech* 2004, **23**:443-451.
- Field HE, Mackenzie JS, Daszak P: Henipaviruses: emerging paramyxoviruses associated with fruit bats. Curr Top Microbiol Immunol 2007, 315:133-159.
- 52. Chua KB, Chua BH, Wang CW: Anthropogenic deforestation, El Nino and the emergence of Nipah virus in Malaysia. *Malays J Pathol* 2002, **24**:15-21.
- Pulliam JRC, Epstein JH, Dushoff J, Rahman SA, Bunning M, Jamaluddin AA, Hyatt AD, Field HE, Dobson AP, Daszak P: Agricultural intensification, priming for persistence and the emergence of Nipah virus: a lethal bat-borne zoonosis. JR Soc Interface 2012, 9:89-101.
- Webster RG: Rapid review. Wet markets a continuing source of severe acute respiratory syndrome and influenza? *Lancet* 2004, 363:234-236.
- 55. Guan Y, Zheng BJ, He YQ, Liu XL, Zhuang ZX, Cheung CL, Luo SW, Li PH, Zhang LJ, Guan YJ *et al.*: **Isolation and characterization of viruses related to the SARS coronavirus from animals in southern China**. *Science* 2003, **302**:276-278.
- 56. Lam WK, Zhong NS, Tan WC: Overview on SARS in Asia and the world. *Respirology* 2003, 8(Suppl):S2-S5.

- 57. Promed, Novel coronavirus Saudi Arabia: human isolate. Archive Number: 20120920.1302733 2012.
- Zaki AM, van Boheemen S, Bestebroer TM, Osterhaus ADME,
 Fouchier RaM: Isolation of a novel coronavirus from a man with pneumonia in Saudi Arabia. N Engl J Med 2012 http://dx.doi.org/ 10.1056/NEJMoa1211721.

A recent emergence of a coronavirus that has caused mortalities that phylogenetically is closely related to bat coronaviruses. Another example of international collaboration to rapidly identify a virus

- Bermingham A, Chand M, Brown C, Aarons E, Tong C, Langrish C, Hoschler K, Brown K, Galiano M, Myers R *et al.*: Severe respiratory illness caused by a novel coronavirus, in a patient transferred to the United Kingdom from the Middle East, September 2012. *Euro Surveill* 2012, 17:1-5.
- Corman V, Eckerle I, Bleicker T, Zaki A, Landt O, Eschbach-Bludau M, van Boheemen S, Gopal R, Ballhause M, Bestebroer T et al.: Detection of a novel human coronavirus by real-time reverse-transcription polymerase chain reaction. Euro Surveill 2012, 17:1-6.
- Promed: Novel coronavirus Saudi Arabia (15): new case. Archive Number: 20121104.1391285 2012.
- Plowright RK, Field HE, Smith C, Divljan A, Palmer C, Tabor G, Daszak P, Foley JE: Reproduction and nutritional stress are risk factors for Hendra virus infection in little red flying foxes (*Pteropus scapulatus*). Proc Biol Sci 2008, 275:861-869.
- Gonzalez JP, Pourrut X, Leroy E: Ebolavirus and other filoviruses. Curr Top Microbiol Immunol 2007, 315:363-387.
- 64. Amman BR, Carroll SA, Reed ZD, Sealy TK, Balinandi S,
 Swanepoel R, Kemp A, Erickson BR, Comer JA, Campbell S et al.: Seasonal pulses of marburg virus circulation in juvenile *Rousettus aegyptiacus* bats coincide with periods of increased risk of human infection. *PLoS Pathog* 2012, 8:e1002877.

Extensive study of Marburg virus in bats and the linking of virus shedding to waning antibody levels in juveniles and therefore increased risk to humans during this time

- 65. Wilkinson GS, South JM: Life history, ecology and longevity in bats. *Aging Cell* 2002, 1:124-131.
- Li L, Victoria JG, Wang C, Jones M, Fellers GM, Kunz TH, Delwart E: Bat guano virome: predominance of dietary viruses from insects and plants plus novel mammalian viruses. *J Virol* 2010, 84:6955-6965.
- Ge X, Li Y, Yang X, Zhang H, Zhou P, Zhang Y, Shi Z: Metagenomic analysis of viruses from bat fecal samples reveals many novel viruses in insectivorous bats in China. J Virol 2012, 86:4620-4630.
- Donaldson EF, Haskew AN, Gates JE, Huynh J, Moore CJ, Frieman MB: Metagenomic analysis of the viromes of three North American bat species: viral diversity among different bat species that share a common habitat. *J Virol* 2010, 84:13004-13018.
- 69. Kaye M: SARS-associated coronavirus replication in cell lines. Emerg Infect Dis 2006, **12**:128-133.
- Aljofan M, Saubern S, Meyer AG, Marsh G, Meers J, Mungall BA: Characteristics of Nipah virus and Hendra virus replication in different cell lines and their suitability for antiviral screening. *Virus Res* 2009, **142**:92-99.
- 71. Westbury HA, Hooper PT, Selleck PW, Murray PK: Equine morbillivirus pneumonia: susceptibility of laboratory animals to the virus. *Aust Vet J* 1995, **72**:278-279.
- Van der Groen G, Webb P, Johnson K, Lange J, Linsday H, Eliott L: Growth of Lassa and Ebola viruses in different cell lines. *Ebola virus Haemorrhagic Fever*. Amsterdam: Elsevier/North-Holland Biomedical Press; 1978:. 255–260.
- Guyatt KJ, Twin J, Davis P, Holmes EC, Smith GA, Smith IL, Mackenzie JS, Young PL: A molecular epidemiological study of Australian bat lyssavirus. J Gen Virol 2003, 84:485-496.
- 74. Arankalle VA, Bandyopadhyay BT, Ramdasi AY, Jadi R, Patil DR, Rahman M, Majumdar M, Banerjee PS, Hati AK, Goswami RP

et al.: Genomic characterization of Nipah virus, West Bengal, India. Emerg Infect Dis 2011, **17**:907-909.

- Marsh GA, Todd S, Foord A, Hansson E, Davies K, Wright L, Morrissy C, Halpin K, Middleton D, Field HE *et al.*: Genome sequence conservation of Hendra virus isolates during spillover to horses, Australia. *Emerg Infect Dis* 2010, 16:1767-1769.
- Streicker DG, Turmelle AS, Vonhof MJ, Kuzmin IV, McCracken GF, Rupprecht CE: Host phylogeny constrains cross-species emergence and establishment of rabies virus in bats. *Science* 2010, 329:676-679.
- Brown C: Emerging zoonoses and pathogens of public health significance – an overview. Rev Sci Tech 2004, 23:435-442.
- Svoboda T, Henry B, Shulman L, Kennedy E, Rea E, Ng W, Wallington T, Yaffe B, Gournis E, Vicencio E et al.: Public health measures to control the spread of the severe acute respiratory syndrome during the outbreak in Toronto. N Engl J Med 2004, 350:2352-2361.
- 79. Knobler S, Mahmoud A, Lemon S, Mack A, Sivitz L, Oberholtzer K:
 Learning from SARS: Preparing for the Next Disease Outbreak-

Workshop Summary. National Academies Press; 2004. A good summary of the SARS outbreak and all the factors involved in the spread of the virus and its control

- Francesconi P, Yoti Z, Declich S, Onek PA, Fabiani M, Olango J, Andraghetti R, Rollin PE, Opira C, Greco D *et al.*: Ebola hemorrhagic fever transmission and risk factors of contacts, Uganda. *Emerg Infect Dis* 2003, 9:1430-1437.
- 81. Tsang T, Lam TH: SARS: public health measures in Hong Kong. *Respirology* 2003, 8(Suppl):S46-S48.
- Mahalingam S, Herrero LJ, Playford EG, Spann K, Herring B, Rolph MS, Middleton D, McCall B, Field H, Wang L-F: Hendra virus: an emerging paramyxovirus in Australia. *Lancet Infect Dis* 2012, 3099:1-9.
- 83. World Health Organization: Severe acute respiratory syndrome (SARS): status of the outbreak and lessons for the immediate future 2003.
- Timen A, Koopmans M, Vossen A, van Doornum G, Gunther S, van den Berkmortel F, Verduin K, Dittrich S, Emmerich P, Osterhaus A *et al.*: Response to imported case of Marburg hemorrhagic fever, the Netherlands. *Emerg Infect Dis* 2009, 15:1171-1175.
- 85. World Health Organization: *The World Health Report 2003. Shaping the future* 2003.
- Field H, Mackenzie J, Daszak P: Novel viral encephalitides associated with bats (Chiroptera) – host management strategies. Arch Virol Suppl 2004, S18:113-121.
- 87. Daszak P, Plowright RK, Epstein JH, Pulliam JR, Abdul Rahman S, Field HE, Jamaluddin A, Sharifah SH, Smith CS, Olival KJ et al.: The emergence of Nipah and Hendra virus: pathogen dynamics across wildlife livestock human continuum. In Disease Ecology: Community Structure and Pathogen Dynamics. Edited by Collinge S, Ray C. Oxford, United Kingdom: Oxford University Press; 2006:186-201.
- Daniels P, Ksiazek TG, Eaton BT: Laboratory diagnosis of Nipah and Hendra virus infections. *Microbes Infect* 2001, 3:289-295.

- Chua KB: Epidemiology, surveillance and control of Nipah virus infections in Malaysia. Malays J Pathol 2010, 32:69-73.
- Nahar N, Sultana R, Gurley ES, Hossain MJ, Luby SP: Date palm sap collection: exploring opportunities to prevent Nipah transmission. *EcoHealth* 2010, 7:196-203.
- 91. Khan SU, Gurley ES, Hossain MJ, Nahar N, Sharker MAY, Luby SP: A randomized controlled trial of interventions to impede date palm sap contamination by bats to prevent Nipah virus transmission in Bangladesh. *PLoS One* 2012, **7**:e42689.
- 92. Nahar N, Mondal UK, Sultana R, Hossain MJ, Khan MS, Gurley ES,
 Oliveras E, Luby SP: Piloting the use of indigenous methods to prevent Nipah virus infection by interrupting bats' access to date palm sap in Bangladesh. *Health Promot Int* 2012 http:// dx.doi.org/10.1093/heapro/das020.

Describes the implementation of practical solution to prevent the transmission of NiV in Bangladesh from contaminated date palm sap

- Lim PL, Kurup A, Gopalakrishna G, Chan KP, Wong CW, Ng LC, Se-Thoe SY, Oon L, Bai X, Stanton LW *et al.*: Laboratoryacquired severe acute respiratory syndrome. N Engl J Med 2004, 350:1740-1745.
- Adjemian J, Farnon EC, Tschioko F, Wamala JF, Byaruhanga E, Bwire GS, Kansiime E, Kagirita A, Ahimbisibwe S, Katunguka F et al.: Outbreak of Marburg hemorrhagic fever among miners in Kamwenge and Ibanda Districts, Uganda, 2007. J Infect Dis 2011, 204(Suppl):S796-S799.
- Bausch DG, Borchert M, Grein T, Roth C, Swanepoel R, Libande ML, Talarmin A, Bertherat E, Muyembe-Tamfum J-J, Tugume B *et al.*: Risk factors for Marburg hemorrhagic fever, Democratic Republic of the Congo. *Emerg Infect Dis* 2003, 9:1531-1537.
- Beer B, Kurth R, Bukreyev A: Characteristics of Filoviridae: Marburg and Ebola viruses. Naturwissenschaften 1999, 86:8-17.
- Luby SP, Gurley ES, Hossain MJ: Transmission of human infection with Nipah virus. *Clin Infect Dis* 2009, 49:1743-1748.
- Homaira N, Rahman M, Hossain MJ, Epstein JH, Sultana R, Khan MSU, Podder G, Nahar K, Ahmed B, Gurley ES et al.: Nipah virus outbreak with person-to-person transmission in a district of Bangladesh, 2007. Epidemiol Infect 2010, 138:1630-1636.
- Chua KB, Lam SK, Goh KJ, Hooi PS, Ksiazek TG, Kamarulzaman A, Olson J, Tan CT: The presence of Nipah virus in respiratory secretions and urine of patients during an outbreak of Nipah virus encephalitis in Malaysia. *J Infect* 2001, 42:40-43.
- 100. Chua KB: Nipah virus outbreak in Malaysia. J Clin Virol 2003, 26:265-275.
- 101. Chua KB, Crameri G, Hyatt A, Yu M, Tompang MR, Rosli J, McEachern J, Crameri S, Kumarasamy V, Eaton BT *et al.*: A previously unknown reovirus of bat origin is associated with an acute respiratory disease in humans. *Proc Natl Acad Sci U S* A 2007, **104**:11424-11429.
- 102. Chua KB, Voon K, Yu M, Keniscope C, Abdul Rasid K, Wang L-F: Investigation of a potential zoonotic transmission of orthoreovirus associated with acute influenza-like illness in an adult patient. *PLoS One* 2011, **6**:e25434.