Review Endocrinology and hormone therapy in breast cancer **New insight into estrogen receptor-** α **function and its implication for endocrine therapy resistance in breast cancer**

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Abstract

Estrogen and its receptor (ER) are critical for development and progression of breast cancer. This pathway is targeted by endocrine therapies that either block ER functions or deplete ER's estrogen ligand. While endocrine therapies are very effective, de novo and acquired resistance are still common. Laboratory and clinical data now indicate that bidirectional molecular crosstalk between nuclear or membrane ER and growth factor receptor pathways such as HER2/neu is involved in endocrine resistance. Preclinical data suggest that blockade of selected growth factor receptor signaling can overcome this type of resistance, and this strategy is already being tested in clinical trials

Introduction

Estrogen receptors (ERs) play a key role in normal breast development and in the development and progression of breast cancer. Endocrine therapy for this disease, which had its start more than 100 years ago when patients were first treated with ovarian ablation [1], blocks the function of ERs and thereby represents the first molecularly targeted therapy for cancer. Long-term studies of patients treated with adjuvant endocrine therapy suggest that this therapy is curative in many patients. Now, many types of endocrine therapy are available that inhibit the function of ERs in different ways. In addition to ovarian ablation, selective ER modulators such as tamoxifen bind ERs and partially block their function [2]. Selective ER downregulators such as fulvestrant completely block the function of ERs and downregulate the receptor proteins [3]. Finally, aromatase inhibitors in postmenopausal women reduce estrogen to very low levels - a strategy that may be the most effective endocrine therapy used today [4,5].

However, endocrine therapies do not always work in patients, despite the presence of ERs in their tumors [6]. Some tumors

are initially inhibited by endocrine therapy only to develop acquired resistance with prolonged treatment. Other tumors are *de novo* resistant to endocrine therapy. Clinical experience indicates that tumors resistant to one form of therapy may still be sensitive to other hormonal treatments [7], suggesting that resistance does not always indicate global estrogen independence; resistance can be treatment specific. Insights into the mechanisms of endocrine therapy resistance have come from provocative studies on the biology of ERs and the various signaling pathways in the cell with which they communicate. Potential strategies to overcome this resistance have been successful in preclinical models and are now being tested in patients.

Structure and function of estrogen receptors

ERs are members of a large family of nuclear transcriptional regulators that are activated by steroid hormones such as estrogen [8]. The ERs exist as two isoforms, α and β , that are encoded by two different genes [9]. These receptor proteins have a similar structure but evidence indicates that they have very different functions. ER proteins consist of a DNA binding region and at least two transcriptional activation functions (AFs) known as AF-1 and AF-2 [10]. A ligand binding domain that interacts with estradiol is also located in the AF-2 region [10,11]. The DNA binding domain of ERs allows the receptors to bind to specific sequences in the promoters of estrogen-regulated genes. AF-1 and AF-2 are important domains for regulating transcription of these genes; AF-1 is constitutively active but is enhanced by growth factor signaling [12], whereas AF-2 is ligand-depend-active with estrogen but not with tamoxifen [13].

Although ER- α and ER- β are both expressed in normal mammary gland, it appears that ER- α , but not ER- β , is critical

AF = activation function; AIB = amplified in breast cancer; AP = activating protein; EGF = epidermal growth factor; ER = estrogen receptor; ERK = extracellular signal-regulated kinase; IGF = insulin-like growth factor; MAPK = mitogen-activated protein kinase; USF = upstream stimulatory factor.

for normal gland development [14]. A dramatic increase in ER- α content is observed in premalignant hyperproliferative breast lesions and in many breast cancers compared with normal mammary glands [15]. These observations are consistent with the crucial role of the ER- α isoform in breast cancer etiology and progression, and with the role played by estrogens as tumor promoters. Growing evidence suggests that ER- β may antagonize the function of ER- α , and high levels of ER- β are associated with a more favorable response to tamoxifen treatment [16]. However, the overall role of ER- β in regulating breast cancer is yet to be defined, and therefore further discussion in this review focuses mostly on ER- α .

Classical ER- α function

Classically, ER- α regulates the expression of specific genes in the nucleus. This function is sometimes referred to as nuclear initiated steroid signaling [17]. Estrogen binding to ER- α results in phosphorylation of the receptor protein, dissociates chaperone proteins such as heat shock protein 90, and induces a change in protein conformation [11]. This 'activated' ER- α then dimerizes with another receptor protein and the dimer binds to specific DNA sequences sometimes called estrogen response elements in the promoter region of estrogen responsive genes (Fig. 1a) [10]. This complex then recruits a variety of coregulatory proteins such as amplified in breast cancer 1 (AIB1 or SRC3), which function as coactivators to enhance transcription of genes [18]. However, estrogen bound ER- α is also capable of downregulating expression of many genes [19]. Downregulation of gene expression by estrogen may be explained by the recruitment of corepressor proteins to the ER complex at certain gene promoter sequences.

The coregulatory proteins complexed with ER- α on the promoters of target genes are very important for the function of ER-a [20,21]. Coactivators recruit other proteins including acetyltransferases, which alter chromatin structure to facilitate transcription [20-22]. AIB1 is a very important ER- α coactivator protein and could even function as an oncogene when overexpressed in preclinical models [23]. Reducing the level of AIB1 impedes ER- α function and can inhibit the growth of human breast tumor xenografts in mice [24]. AIB1 is also important in breast cancer in patients. It is overexpressed in 65% of breast cancers and is occasionally gene-amplified, suggesting that it plays an important role in the development and progression of breast cancer [25,26]. Preclinical studies and clinical studies in patients suggest that high levels of AIB1 may contribute to tamoxifen resistance by enhancing the estrogen agonist activity of the drug [27-29]. This may be particularly true in tumors that also express high levels of HER2. Kinases activated by HER2 phosphorylate both ER and AIB1, resulting in an increase in ER- α activity [29]. In this situation, tamoxifen-bound ER- α may recruit coactivators such as AIB1 rather than corepressors, resulting in tamoxifen-stimulated growth as a mechanism for resistance to the drug.



Nuclear ER- α functions. (a) Enhanced transcription of genes with an estrogen response element (ERE) in the promoter. (b) Enhanced transcription of genes by modifying activity of transcription complexes at other promoter sites (activator protein [AP]-1). AIB, amplified in breast cancer; At, acetyltransferase; E, estradiol; ER, estrogen receptor- α .

ER- α has also been shown to modulate gene expression in other ways. Through protein-protein interactions ER can bind to transcriptional complexes on other regulatory DNA sequences such as activating protein (AP)-1, Sp1, and upstream stimulatory factor (USF) sites, as well as other poorly defined promoter regions that are not typical estrogen response elements (Fig. 1b) [30-33]. In this circumstance ER- α is not the primary transcription factor binding DNA but it is tethered to other transcription factor proteins such as c-jun or c-fos. It then functions as a coactivator protein by stabilizing DNA binding of the transcription complex or by recruiting other coactivator proteins to these complexes [34,35]. Several genes important for tumor cell proliferation and survival may be regulated by estrogen in this way, and data suggest that modulation of gene expression at AP-1 sites may be the most critical element in estrogen-mediated tumor growth [30,36].

Through these direct transcriptional mechanisms, estrogen regulates the expression of hundreds of genes, many of which are important for cell proliferation, antiapoptosis, invasion and metastasis, and angiogenesis. c-Myc, vascular endothelial growth factor, bcl-2, insulin-like growth factor (IGF)-R1, insulin receptor substrate-1, transforming growth factor- α , cyclin-D₁, and IGF-2 are all regulated by estrogen [34,37-41]. Downregulation of genes related to apoptosis or cell cycle inhibition may explain the curative potential of adjuvant endocrine therapy. Altered gene expression may also contribute to certain forms of endocrine therapy resistance. As mentioned above, the ER- α coactivator AIB1

can bind to tamoxifen-bound ER- α complexes under some circumstances, potentially reducing the estrogen antagonist activity of the drug [29]. Increased AP-1 transcriptional activity has been identified in preclinical models and in patients with tamoxifen resistance [42,43]. More recent data suggest that ER- α crosstalk with growth factor receptor signaling pathways can modulate ER- α transcriptional activity and may also contribute to resistance to tamoxifen or other endocrine therapies [37,38].

Modification of transcriptional function of estrogen receptor- α by growth factor receptor signaling

Numerous growth factors and signaling molecules including IGF-1, epidermal growth factor (EGF), heregulin, transforming growth factor- α , dopamine, cyclic AMP, and membrane permeable phosphatase inhibitors can all potentiate the nuclear transcriptional effects of ER- α [40,44-46]. This functional activation of ER-a results from direct phosphorylation of the ER- α protein at multiple sites, mainly at the AF-1 domain of the receptor [47], or from phosphorylation of ER- α coactivators and corepressors [48]. This potentiation of ER- α transcriptional activity due to phosphorylation of the ligand independent AF-1 domain has been termed ligand independent activation of ER- α , because it can occur in the absence of estrogen. Multiple kinases involved in growth factor, stress, or cytokine induced signaling can phosphorylate ER- α or its coregulatory proteins. These include extracellular signal regulated kinase (ERK)1/2 and p38 mitogen-activated protein kinases (MAPKs), cyclin A/CDK-2, CDK-7, c-Src, protein kinase A, pp90rsk1, Akt, IkB kinase, and Jun amino terminal kinase [12,49-57]. Phosphorylation of ER- α on these various sites affects protein dimerization, DNA binding, ligand binding, nuclear localization, interactions with coactivators and corepressors, and transcriptional activation of genes [53,54,58-60]. Phosphorylation of ER- α coregulatory proteins augments their activity on ER-a dependent transcription, alters nuclear localization, affects their binding to ER- α , and enhances their ability to recruit other transcriptional coregulators [56,57,61]. Phosphorylation of ER- α corepressors such as N-CoR also changes their affinity for ER- α binding and modifies nuclear localization [62-64]. Thus, estrogen regulated gene expression is a coordinated effort that depends on the receptor itself, its coregulatory molecules, specific gene promoter sequences, the presence or absence of other transcription factors, and input from a variety of intracellular signaling pathways.

Membrane initiated (nongenomic) steroid signaling

ER- α functions not only as a transcription factor in the nucleus; new data suggest that it can activate intracellular signaling pathways outside the nucleus in the cytoplasm or the plasma membrane (Fig. 2) [65]. It has been observed for decades that estrogens can have very rapid effects on cells, too rapid to invoke transcriptional mechanisms [66]. ERs have been localized outside the nucleus by biochemical analyses and by direct visualization using immuno-





Membrane or nongenomic functions of ER- α . Estrogen (E) or tamoxifen binds membrane or cytoplasmic estrogen receptor (ER)- α to trigger complexes with growth factor signaling molecules (such as insulin-like growth factor [IGF]-R1, insulin receptor substrate [IRS]-1, or phosphatidylinositol-3-OH kinase [PI3K]), which activate mitogenactivated protein kinase (MAPK) or AKT. ER- α also acting as a G-protein-coupled receptor activates Src, which activates matrix metalloproteinase (MMP)-2, which cleaves heparin-binding epidermal growth factor (Hb-EGF) from the membrane to bind and activate EGF receptor (EGFR). EGFR also activates MAPK and AKT, which in turn can phosphorylate and activate nuclear ER- α and amplified in breast cancer (AIB)1, or directly contribute to tumor growth and survival (dotted line).

cytochemistry or more sophisticated microscopy [67,68]. This nongenomic mode of action of ERs, as well as of other steroid receptors, is also known as membrane-initiated steroid signaling [17]. Many studies have now shown that non-nuclear ER- α can exist in complexes with several signaling molecules. Estrogen treatment of cells induces the interaction between ER- α and IGF-1 receptor, which activates signaling via ERK1/2 MAPK [69]. Estrogen also stimulates association between ER- α and the p85 subunit of phosphatidylinositol-3-OH kinase, resulting in its activation [70]. ER- α interacts with other IGF signaling intermediates including insulin receptor substrate 1 [71], which then translocates to the nucleus where it may exist in transcription complexes. ER-a can also interact with Shc, which links the receptor to other signaling intermediates in the membrane [72]. Thus, ER can complex with many elements in the IGF-1R signaling cascade, thereby activating this pathway in response to estrogen.

ER- α has also been shown to act as a G-protein-coupled receptor at the membrane, where it binds to caveolin-1 (Fig. 2) [73-75]. In this way, ER- α can activate c-Src, in turn leading to activation of metalloproteinases that then cleave heparin binding EGF from the cell. This growth factor, acting in an autocrine manner, binds EGF receptors on the cell membrane, thereby activating downstream kinases such as ERK1/2 MAPK and phosphatidylinositol-3-OH kinase. The observation that tamoxifen, like estrogen, can activate

membrane ER suggests a plausible mechanism for tamoxifen resistance in tumors in which this pathway is active. Pure ER antagonists such as fulvestrant do not activate membrane ER [69,75]. This type of ER- α activity may not be prominent in most breast cancers that lack strong receptor tyrosine kinase activity, but this activity can be potentiated in tumors that overexpress tyrosine kinase receptors [29,76] or in those that expresshigh levels of other proteins that bind ER- α and sequester it in the cytoplasm or membrane. Such proteins as modulator of nongenomic activity of ER (MNAR) [77-79] and members of the metastasis associated gene family (MTA) [80,81] may be important in membrane initiated ER- α signaling by trapping it outside the nucleus.

Thus, ER-α may stimulate breast cancer growth in at least two ways. It increases the expression of growth regulatory genes via its transcriptional effects in the nucleus and it activates the growth factor signaling cascade via its nongenomic or membrane activity outside the nucleus. In some tumors, particularly those with amplification of HER2 or overexpression of other growth factor receptor tyrosine kinases, a vicious cycle of cell proliferative and cell survival stimuli is initiated by the binding of estrogen to its receptor. ER- α by a variety of mechanisms activates the growth factor signaling cascade, while at the same time the growth factor receptor kinase cascade phosphorylates and activates ER- α and its coregulating proteins. Membrane or nonnuclear ER-α signaling has been described in experimental model systems and needs to be confirmed in clinical breast cancer. Tumors with active ER- α /growth factor receptor crosstalk, especially those that are largely dependent on the membrane/non-nuclear ER- α signaling, could be expected to be very dependent on estrogen for growth and might then be highly responsive to treatment with estrogen deprivation therapy, such as aromatase inhibition. These tumors should also be responsive to selective ER downregulators, such as fulvestrant, which function as complete antagonists on both nuclear and non-nuclear ER activities. Tamoxifen, on the other hand, because it activates membrane ER- α signaling might not be as effective, and in fact it might even stimulate growth of such tumors [29]. As described below, clinical data do support these paradigms. An understanding of these ER activities at the molecular level may yield new strategies to prevent or overcome resistance to tamoxifen and other forms of treatment.

Blocking estrogen receptor/growth factor receptor crosstalk to overcome treatment resistance

The molecular functions of ER- α and its crosstalk with growth factor receptors are exploitable for treatment purposes. Preclinical experimental models first suggested that tumors overexpressing HER2, in addition to ER- α , might be tamoxifen resistant [82]. Since then, the bulk of clinical evidence indicates that patients with HER2-overexpressing ER- α -positive breast cancers derive less benefit from tamoxifen than do those with HER2-negative breast cancers [29,83-85]. A recent clinical report [86] also suggested that activation of ER- α function by growth factor signaling is important for tamoxifen resistance. Tamoxifen-treated patients whose tumors expressed high levels of both AIB1 and HER2 had extremely poor disease-free survival compared with patients whose tumors overexpressed neither or only one of the two proteins. This clinical observation makes biologic sense because kinases activated downstream from HER2 phosphorylate and functionally activate AIB1 and ER- α , thereby reducing the antagonist activity of the tamoxifenbound receptor.

Prospective trials in patients with metastatic disease also suggest that tamoxifen resistance is associated with high levels of expression of receptor tyrosine kinases such as HER2 and EGF receptor [87]. Furthermore, because resistance to selective ER modulators such as tamoxifen is mediated, at least in part, via activation of the nongenomic actions of ER, other hormonal treatments that can block this ER activity should be effective second-line therapies for patients whose tumors acquired resistance to tamoxifen. Importantly, aromatase inhibitors and the ER downregulator fulvestrant, which can deactivate both the genomic and the nongenomic effects of ER, are indeed effective therapies in tamoxifen resistant patients [7]. Finally, three studies in which patients were given neoadjuvant endocrine therapy as initial treatment further support these hypotheses [88-90]. Two of these trials randomized patients to tamoxifen or an aromatase inhibitor. Both found that tumor response to tamoxifen was about half of that observed with the aromatase inhibitor in tumors overexpressing HER2. The response rates were more similar for tumors that were negative for HER2. In fact, the response rates to the aromatase inhibitor in these two trials, as well as in a third nonrandomized trial, were at least as high or higher in tumors overexpressing HER2 than in HER2 negative tumors. These neoadjuvant trials do not provide data on the duration of response to the aromatase inhibitors in HER2 positive disease. Preclinical studies suggest that growth factor receptor signaling can also contribute to acquired resistance to estrogen deprivation therapies [91]. Nevertheless, the substantial initial tumor response in the HER2 positive subset of patients suggests that these tumors are indeed very dependent on estrogen for growth. Analysis of completed large adjuvant trials with patients classified by ER- α , progesterone receptor, and HER2 status are needed to confirm the superior efficacy of aromatase inhibitors compared with tamoxifen in such patients.

If the growth factor receptor tyrosine kinase signaling pathway is involved in resistance to hormonal therapy, then an obvious question is whether blockade of these pathways can delay or overcome treatment resistance (Fig. 3). Compeling data from *in vitro* and *in vivo* experimental models suggest that this strategy is worth testing in patients. In a xenograft model of an ER- α positive, HER2 overexpressing tumor, tamoxifen-stimulated growth mediated by growth





Blockage of growth factor receptor signaling pathways to overcome tamoxifen resistance. Tamoxifen (T) bound estrogen receptor (ER)- α in the membrane can activate epidermal growth factor receptor (EGFR), setting up a signaling cascade that leads to tamoxifen resistance. Blocking EGFR activation would block activation of mitogen-activated protein kinase (MAPK), AKT, ER- α and amplified in breast cancer (AIB)1, and would restore tamoxifen's antagonist activity on tumor growth.

factor receptor activation by nongenomic ER- α signaling is the mechanism for *de novo* resistance [29]. Simultaneous treatment of mice with the EGF receptor tyrosine kinase inhibitor gefitinib blocked activation of HER2 signaling by tamoxifen, reduced phosphorylation of ER- α and AlB1, and completely restored tamoxifen's antagonist activity on tumor growth. Similar data have been reported in other experimental models [85]. In a recent study [92], lapatinib – a dual EGF receptor/HER2 tyrosine kinase inhibitor – effectively inhibited the growth of non-HER2 overexpressing tumors that had developed acquired resistance to tamoxifen. Gefitinib and lapatinib are now in clinical trials to test the concept that simultaneous treatment with growth factor inhibitors combined with ER-targeted therapy is more effective than either treatment by itself.

Conclusion

Although ER- α was discovered nearly 40 years ago, scientists continue to unravel its complex role in cellular function. It is clear that ER- α has multiple activities and that it often does not act alone but rather in concert with other signaling pathways that can modify its output. Some of these pathways lead to specific changes in receptor activity that contribute to resistance to hormonal therapies. Because there are many signaling molecules that can modulate the function of ERs, an accurate molecular profile of each patient's tumor will be required in the future to design the optimal therapeutic approach by identifying the major pathways that are working together to drive the growth of a particular tumor. The fact that targeting ER- α by itself offers long-term disease control in many patients that is tantamount

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to cure indicates that the mechanisms regulating breast cancer growth may not be as complicated as some may have thought, and that accurate identification of the pathway or pathways that regulate tumor cell survival and proliferation and then blocking only those pathways may prove to be an extraordinarily successful strategy.

Competing interests

The author(s) declare that they have no competing interests.

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