

The therapeutic effects of bone marrow mesenchymal stem cells after optic nerve damage in the adult rat

HaiBo Tan^{1,*}

Xin Kang^{2,*}

ShiHeng Lu¹

Lin Liu¹

¹Department of Ophthalmology, Renji Hospital, The School of Medicine, Shanghai JiaoTong University, Shanghai, People's Republic of China;

²Department of Clinical Pharmacology, Changhai Hospital Affiliated Second Military Medical University, Shanghai, People's Republic of China

*These authors contributed equally to this work

Abstract: Optic nerve trauma is a common occurrence that results in irreversible blindness. Currently, no effective strategies are known to prevent optic nerve degeneration. We assessed the therapeutic effects of bone marrow mesenchymal stem cells (BMSCs) after optic nerve crush in the adult rat. Our results showed that BMSCs significantly promoted the regeneration of injured axons compared with phosphate buffered saline alone. Therefore, BMSC transplantation may be effective for the treatment of central nervous system disorders.

Keywords: bone marrow mesenchymal stem cell, axonal regeneration, rat optic nerve crush, nervous system disorders

Introduction

Optic nerve damage is an important cause of irreversible vision loss, which results from retinal ganglion cell (RGC) death and subsequent axonal degeneration.^{1,2} It is well known that in the central nervous system, the failure of axonal regeneration is due not only to the limited intrinsic growth capacity of adult neurons but also to the growth inhibitory environment.³ Therefore, therapies should focus on the protection of injured neurons and on the promotion of axonal regeneration.^{4–8} Cell transplantation therapy has been proven effective in this regard.⁹

Bone marrow mesenchymal stem cells (BMSCs) have both amplification and transplantation abilities.^{10,11} Furthermore, BMSCs have been shown to exert a neuroprotective effect on RGCs.¹² However, little is known about the effects of BMSCs on axonal regeneration of the injured optic nerve. Here, we explored the axonal regeneration-promoting effect of BMSCs after optic nerve crush (ONC).

Materials and methods

ONC and intravitreal injection

This experiment was approved by the Ethics Committee of Animal Research of Renji Hospital, The School of Medicine, Shanghai JiaoTong University. Eighty adult male Sprague Dawley rats (weight: 200–250 g) were provided by the Shanghai Institute of Traumatology and Orthopedics and were housed in a temperature- and humidity-controlled room with a 12:12 hour light:dark cycle.

Surgical procedures were performed as described in our previous study.¹³ The right optic nerve was crushed 1 mm behind the posterior eye pole for 9 seconds for all experimental groups. But the optic nerve was not crushed in the sham-surgery group. BMSCs were purchased from Shanghai Shengong Biotechnology (Shanghai Shengong Biotechnology, Shanghai, People's Republic of China). A total of 3 μ L of

Correspondence: Lin Liu
Department of Ophthalmology, Renji Hospital, The School of Medicine, Shanghai JiaoTong University, Shanghai 200025, People's Republic of China
Tel +86 136 2176 0754
Email thbbobo@126.com

phosphate buffered saline (PBS) or BMSCs (low number of transplantation cells: 3×10^4 ; high number of transplantation cells: 1×10^5) were intravitreally injected immediately, and 3, 6, and 9 days after ONC. Rats in the sham surgery group did not receive an intravitreal injection. Rats with lens injury were excluded. These rats were divided into four groups: the sham surgery group, PBS group, low BMSC group, and high BMSC group. Twenty rats in each experimental group were included in the final evaluation.

Processing of optic nerve samples

The procedures were performed as described in our previous study.¹³ The rats were sacrificed on day 15 after ONC and were transcardially perfused with 4% paraformaldehyde. The optic nerves were dissected and immediately fixed in 4% PFA in paraformaldehyde for 1 hour. Then, the tissue was embedded, frozen, and cut. Longitudinal sections (16 μm thickness) of the optic nerve were obtained and were placed on gelatin-coated glass slides for subsequent use.

Axon count

With respect to the axon count, the optic nerve sections were dehumidified at 37°C for 1 hour, and nonspecific binding was blocked by the application of 10% normal goat serum. To stain the regenerating fibers, an anti-GAP43 antibody (mouse anti-rat GAP-43 monoclonal antibody, 1:100, sc-17790; Santa Cruz Biotechnology Inc., Dallas, TX, USA) was applied and the sections were incubated at 4°C overnight. After two washes with PBS, the secondary

antibody (Cy-3-labeled anti-mouse fluorescein isothiocyanate antibody, 1:100; Thermo Fisher Scientific, Waltham, MA, USA) was applied and the sections were incubated for 45 minutes at room temperature. GAP-43-positive axons were quantified to assess the regeneration of the axons of RGCs as previously described.¹³

Western blot analysis

Western blot analysis was used to determine the expression levels of GAP-43 after ONC. The methods of Western blot analysis were described in our previous study.¹³

Statistical analyses

The data analyses were performed using SPSS version 19.0 software (IBM Corporation, Armonk, NY, USA), and the data are expressed as the mean \pm standard deviation. The differences between the groups were determined by one-way analysis of variance or by independent samples Student's *t*-test. A $P < 0.05$ was considered statistically significant.

Results

BMSC promotes the axonal regeneration of rat RGCs following ONC

Our results indicated very few axons beyond the crush site in the PBS-treated group. However, a large number of GAP-43-positive axons passed through the crush site in the BMSC-treated group (low or high transplantation group) on day 15 after ONC (Figure 1). Rats in the sham surgery group showed no significant staining (Figure 1). The data

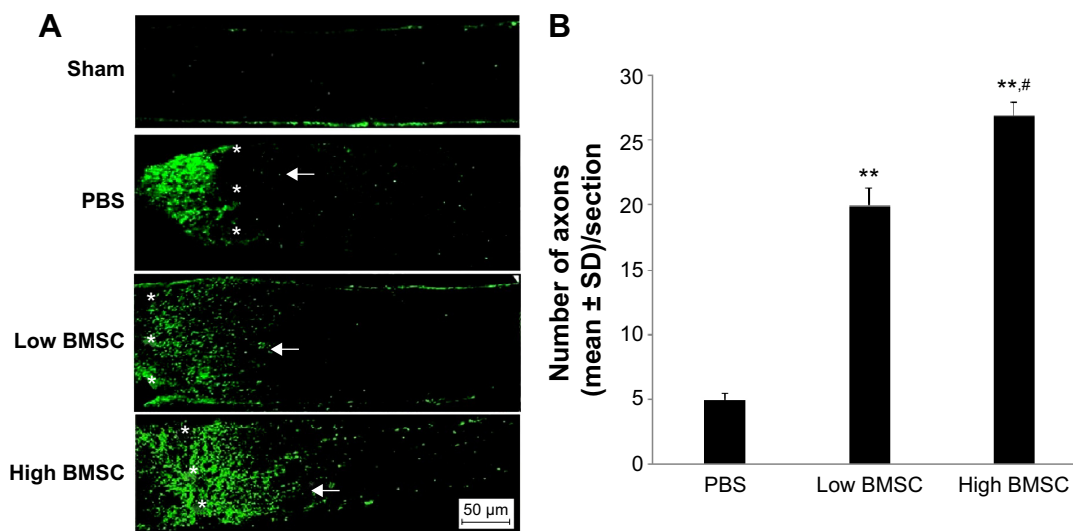


Figure 1 Representative photomicrographs of regenerating axons in all groups after ONC.

Notes: The numbers of axons/section are presented as the mean \pm SD ($n=10$). **(A)** asterisk, the crush site; arrow heads, GAP-43 immunofluorescence; scale bar, 50 μm .

(B) ** $P < 0.001$ compared with the PBS group; # $P < 0.05$ compared with the low BMSC group.

Abbreviations: ONC, optic nerve crush; SD, standard deviation; BMSC, bone marrow mesenchymal stem cell; PBS, phosphate buffered saline.

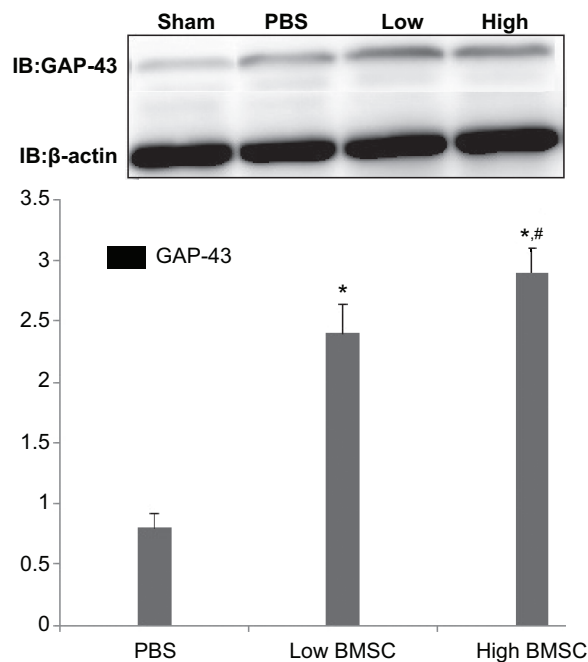


Figure 2 Western blot analysis of GAP-43 expression.

Notes: * $P < 0.001$, compared with the PBS group; # $P < 0.05$, compared with the low BMSC group.

Abbreviations: BMSC, bone marrow mesenchymal stem cell; IB, immunoblotting; PBS, phosphate buffered saline.

analysis suggested that more axons were present in the two BMSC groups (low and high) than in the PBS group ($P < 0.001$). Moreover, the high BMSC transplantation group was associated with higher numbers of regenerative axons in comparison with the low BMSC transplantation group.

BMSCs increase GAP-43 expression

According to Western blot analysis, GAP-43 expression was weak in the sham surgery group, but GAP-43 expression was differentially increased in the PBS and BMSC (low and high) groups ($P < 0.001$; Figure 2). Moreover, GAP-43 expression in the low and high BMSC transplantation groups was higher than that in the PBS group (Figure 2). In addition, the group with a high number of transplanted cells demonstrated increased axonal regeneration compared with the group with a low number of transplanted cells ($P > 0.05$; Figure 2).

Discussion

The present study explored the effect of transplanted BMSCs on axonal-regeneration after the ONC. GAP-43 expression in rats in the PBS and BMSC groups was significantly increased compared with rats in the sham surgery group, but the GAP-43 protein levels were higher in both BMSC groups than in the PBS group. This result suggested that BMSCs promoted axonal regeneration of the injured optic nerve.

The mechanism of how BMSC therapy might be effective for axonal regeneration after ONC has not yet been elucidated. The function of transplanted BMSCs may involve the regulation of immune system-related inflammation in the injured tissues. In experimental autoimmune encephalomyelitis, Semon et al demonstrated a significant reduction in inflammatory infiltrates by BMSCs.¹⁴ Furthermore, BMSC-conditioned medium significantly decreased cell proliferation and the secretion of pro-inflammatory factors through activated microglia.¹⁵ Li et al also indicated that BMSCs enhanced axonal remodeling in a model of experimental stroke.¹⁶ On the contrary, BMSCs promoted axonal regeneration via the increase in the production of trophic factors.¹⁷ In addition, no significant adverse effects were observed in the two BMSC groups.

In summary, our study demonstrates that BMSCs may be a potential therapeutic option for disease of the central nervous system.

Disclosure

The authors report no conflicts of interest in this work.

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