

Article



Growth Performance and Hematological Changes in Growing Sika Deers Fed with Spent Mushroom Substrate of *Pleurotus ostreatus*

Chongshan Yuan, Min Wu, Xinyuan Chen, Changze Li, Aiwu Zhang * and Wenfa Lu *

College of Animal Science and Technology, Jilin Agricultural University, Changchun 130118, China; 18844146800@163.com (C.Y.); koo331500@163.com (M.W.); chenxy9797@163.com (X.C.); mbconfuse@163.com (C.L.)

* Correspondence: zhangaiwu@jlau.edu.cn (A.Z.); wenfa2004@163.com (W.L.); Tel.: +86-138-441-02196 (W.L.)

Simple Summary: With the rapid development of the mushroom industry, a large number of spent mushroom substrate (SMS) has also been produced. SMS can be easily digested by ruminants and is suitable for feeding animals, such as cows, sheep, as well as deer. The results of this study show that the dietary spent mushroom substrate of *Pleurotus ostreatus* (SMS-MP) has no obvious effect on the physiological condition of growing sika deer, at the same time it can reduce the cost of feeding and avoid environmental pollution caused by improper disposal of SMS-MP.

Abstract: The purpose of this experiment is to expand the feed of growing sika deer and to explore the effects on growing sika deer of the spent mushroom substrate of *Pleurotus ostreatus* (SMS-MP). Twelve immature female growing sika deer were randomly assigned to four groups. The ratios of SMS-MP to replace concentration supplements were 0%, 10%, 20%, and 30%, respectively, and the growth performance, feed intake and apparent digestibility, serum biochemical indexes, blood physiological indexes, serum immune globulin and plasma amino acid of growing sika deer were measured. The results of the current study confirmed the applicability of SMS-MP as a feed ingredient in growing sika deer diets. There was no significant change in growth performance and hematology of growing sika deer when the concentrate supplement was replaced with 10–20% SMS-MP. However, replacing 30% of concentrate supplements with SMS-MP in the growing sika deer diet resulted in significantly decreased Hb and HCT levels. It can be concluded that, as a waste resource, adding a small amount of SMS-MP has no significant effect on the growth of sika deer, and at the same time can reduce the consumption of concentrate supplements, thereby improving the economic benefits of sika deer breeding.

Keywords: average daily weight gain; blood index; digestibility; growing sika deer; spent mushroom substrate

1. Introduction

With the increase in global mushroom production [1], the production of waste spent mushroom substrate (SMS) as a by-product of mushroom cultivation has been growing rapidly [2]. In Asia, one of the major problems in mushroom production is the treatment and disposal of the spent mushroom substrate (SMS) which is a co-product of the mushroom industry [3–5]. Recently, there has been increasing public concern on the effects of SMS disposal on the environment. It would be more economical and beneficial if SMS could be recycled and reused. SMS could potentially be a source of planting industry, such as accelerating the maturation of compost [6], enhancing soil organic matter and nutrient contents [7], and increasing the morphological growth of tomato and pepper seedlings [8]. SMS is an organic by-product rich in protein, polysaccharides, which also contains several bioactive compounds resulting from fungal growth, such as vitamins and some trace



Citation: Yuan, C.; Wu, M.; Chen, X.; Li, C.; Zhang, A.; Lu, W. Growth Performance and Hematological Changes in Growing Sika Deers Fed with Spent Mushroom Substrate of *Pleurotus ostreatus. Animals* **2022**, *12*, 765. https://doi.org/10.3390/ ani12060765

Academic Editor: Tomás Landete-Castillejos

Received: 30 January 2022 Accepted: 16 March 2022 Published: 18 March 2022

Publisher's Note: MDPI stays neutral with regard to jurisdictional claims in published maps and institutional affiliations.



Copyright: © 2022 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (https:// creativecommons.org/licenses/by/ 4.0/). elements such as Fe, Ca, Zn and Mg, and therefore can potentially be a value-added product. One of the recycling methods is to use it as animal feed since the ingredients used as SMS are used as animal feeds as well. Many studies have already been performed in regards to the advantages and processing methods of feed based on SMS [9,10]. SMS has been widely reported to be used in feed processing [11] and animal feed presently [12–14]. SMS can be easily digested by ruminants and are suitable for feeding animals, such as cows [15] and sheep [16]. Furthermore, SMS is considered to be an animal feed with nutritional value [17]. One of the more popularly cultivated mushroom species worldwide belongs to the genus Pleurotus ostreatus, it can be cultivated on a variety of substrates such as rice straw, wheat straw, corncob, cottonseed hull as well as other substrates [18]. *Pleurotus ostreatus* increase the nutritional content of SMS during growth, at the same time, different components of SMS also affect the content of mushroom nutrients [19]. Most sika deer in China are fed in captivity. According to ancient Chinese medicine monographs, sika deer antler has long been used as a traditional tonic or alternative medicine [20]. At the same time, raising female sika deer is very important to expand the group size and increase the scale of antler production. To our knowledge, there are no published reports on using SMS as a diet for sika deer. This study is designed to evaluate the effectiveness of dietary spent mushroom substrate of *Pleurotus ostreatus* (SMS-MP) on the average daily weight gain, blood index, digestibility of female growing sika deer in order to reduce breeding costs while avoiding environmental pollution caused by improper disposal.

2. Materials and Methods

2.1. Animals, Diets, and Experimental Designs

Twelve healthy immature female growing sika deer with the similar initial body weight (P > 0.05) were randomly assigned to four groups (three deer / group), living in the artificial feedlot with no grass, tree canopy cover, etc. The four groups were named I, II, III and IV, respectively, in which a different experimental diet was given to each group. Three levels of corncob SMS-MP (dried in the open air) were adopted in the four groups as follows: group 1 (I): basal diet, without substitute of SMS-MP; group 2 (II): replace 10% of concentrate with SMS-MP; group 3 (III): replace 20% of concentrate with SMS-MP; group 4 (IV): replace 30% of concentrate with SMS-MP. Each group was fed concentrate supplement with the same weight and offered three times per day at 3:30, 10:30, 16:30. Silage and water was *ad libiturn*. The growth trial lasted for 31 days according to a randomized design, during which feed intake was recorded daily, and the feed efficiency and the average daily gain (ADG) were calculated. The composition and nutrition of concentrate supplement are presented in Table 1, and the chemical composition of feeds are presented in Table 2. The main component of SMS-MP was corncob. The chemical composition of dried SMS-MP is presented in Table 3.

| Ingredients | Percentage |
|---------------|------------|
| Corn | 78.55 |
| Soybean meal | 20.00 |
| NaCl | 1.20 |
| Stone powder | 0.15 |
| $Ca(HCO_3)_2$ | 0.10 |

 Table 1. Ingredients of concentrate supplement (%).

2.2. Sample Collection and Treatment

2.2.1. Samples of Silage, Concentrate Supplement and Feces

Silage, concentrate supplement and feces were weighed and recorded daily. Samples of silage, concentrate supplement and feces were dried in a forced-air oven at 65 °C for at least 48 h until sub-samples reached constant weight, and then ground in a Wiley mill to pass a 2 mm screen, prepare for preservation until laboratory analysis.

| Parameter | I | II | III | IV |
|-----------------------------|------------------|----------------|------------------|----------------|
| Gross Energy (GE, MJ/kg) | 15.87 ± 0.78 | 15.79 ± 0.83 | 15.39 ± 0.54 | 15.19 ± 0.35 |
| Organic matter (OM) | 96.04 ± 0.04 | 97.03 ± 0.12 | 95.75 ± 0.07 | 95.23 ± 0.16 |
| Ether extract (EE) | 16.57 ± 1.26 | 16.23 ± 0.60 | 15.93 ± 1.03 | 15.29 ± 0.47 |
| Crude protein (CP) | 9.16 ± 0.19 | 8.89 ± 0.08 | 7.70 ± 0.31 | 6.25 ± 0.12 |

Table 2. Chemical composition of dried concentrate supplements (%, DM).

Table 3. Chemical composition of dried SMS-MP (%, DM).

| Parameter | SMS-MP |
|------------------|------------------|
| GE (MJ/kg) | 14.79 ± 1.07 |
| OM | 93.23 ± 0.33 |
| EE | 9.13 ± 0.96 |
| СР | 3.01 ± 0.82 |
| Crude fiber (CF) | 31.70 ± 1.07 |
| Phosphorus (P) | 0.22 ± 0.01 |
| Calcium (Ca) | 1.46 ± 0.59 |

2.2.2. Blood Sampling

On the morning of the end of the trial, all of the growing sika deer were anesthetized with xylazine hydrochloride, which was administered using a blow gun-dart syringe at a dosage of 1 mL per deer. Then, 5 mL of blood was collected with vacuum tubes (Vacutainer; Becton Dickinson and Co. Rutherford, NJ, USA) containing sodium heparin, 2 mL of sample was taken for the detection of blood physiological indexes, and the other 3 mL samples were centrifuged at 3000 rpm to take the plasma to test the amino acids. Another 5 mL of whole blood was put into a vacuum tube, and the serum obtained from whole blood was used to detect serum biochemical indexes and immunoglobulins. Samples were stored at -20 °C for later analysis. After sample collection, the sika deer were injected with 1 mL Nikethamide immediately to wake up the growing sika deer.

2.3. Determination

2.3.1. Body Weight Gain Determination

Body weight gain was obtained by the difference in weight between the first day and the last day of the experiment.

2.3.2. Feed Intake and Nutrient Digestibility

Feed intake was recorded daily by weighing the silage and concentrate complement offered, and the refusals of the previous day. In the last 8 days of the experiment, samples of silage, concentrate supplement and feces were collected per day.

Dry matter (DM) content of concentrate supplement, silage and feces samples was determined in a forced-air oven at 105 °C to constant weight. N (CP = N × 6.25) determinations were carried out using the LECO FP-528N/Protein tester (LECO, Corporation). Ether extract (EE) and crude fiber (CF) were analyzed in duplicate as extraction disappearance in anhydrous diethyl ether using Official Analytical Chemists (AOAC, 1998) [21]. Gross energy concentration in feces and feedstuffs was determined using an adiabatic bomb calorimeter (Changji, Shanghai, China). Ash was determined after thermal decomposition in a muffle furnace at 525 °C for 12 h, and organic matter (OM) was determined by difference.

A nutrient digestibility trial was carried out in which acid insoluble ash was used as an internal indigestibility marker, and digestibility coefficients of digestion were calculated according to the following equation:

$$N = 100 - 100 \times \frac{D}{F} \times \frac{D_1}{F_1} \tag{1}$$

where *N* represents apparent digestibility of nutrients, *D* is the content of indicator in the diet, *F* is the content of indicator in the feces, F_1 is the content of nutrition in the feces; D_1 is the content of nutrition in the diet.

2.3.3. Analysis of Blood

Plasma-free amino acid was determined using high-performance liquid chromatography on an L3000 AA Analyzer (Shimadzu Ltd., Kyoto, Japan). Additional procedural details are described elsewhere [22].

The concentration of albumin (ALB), total protein (TP), globulin (GLO), calcium (Ca), glucose (GLU), urea nitrogen (BUN), amylase (AMY), cholesterol (CHOL), alanine aminotransferase (ALT), total bilirubin (TBIL), alkaline phosphatase (ALP), inosine (CRE), urea nitrogen (BUN), creatine kinase (CK) in serum were analyzed by BC-5300vet automatic blood cell analyzer.

The values of white blood cells (WBC), red blood cells (RBC), hemoglobin (Hb), hematocrit (HCT), MCV (Mean erythrocyte volume), mean corpuscular hemoglobin (MCH), mean corpuscular-hemoglobin concentration (MCHC), variation of RBC distribution width (RDW-CV) were determined using a hematological analyzer (Sysmex K-1000, TAO Medical Electronics Co., Kobe, Japan).

IgA, IgG and IgM of serum were determined using an enzyme-linked immunoassay kit (Shanghai Enzyme Biotechnology Co., Ltd., Shanghai, China), each index corresponding to a specific kit, according to the manufacturer's instructions.

2.4. Statistical Analysis

Statistical analysis was via least-squares analysis of variance (ANOVA), following the general linear models procedure of SPSS (SPSS 19.0 for Windows; SPSS Inc., Chicago, IL, United States). Each experiment was repeated at least three times. Data are presented as the mean \pm SD. The variances were first analyzed using a homogeneity test. If the data met the assumption of homoscedasticity, the significance of differences in the means was determined by one-way ANOVA and subsequent LSD test. *P* < 0.01 was considered as greatly significant; *P* < 0.05 was considered as significant.

3. Results

3.1. Effects on Growth

From Table 4, we can see that there is no difference in IW, FW and ADG (P > 0.05). It can also be seen that there are no differences in F/W and E/W among the four groups (P > 0.05).

Table 4. Effects of dietary supplementation with different inclusion of SMS-MP on growth performance of growing sika deer.

| Items | Ι | II | III | IV |
|---------------------------------------|------------------|------------------|------------------|------------------|
| initial weight (IW, kg) | 53.27 ± 2.65 | 52.02 ± 3.73 | 51.80 ± 3.59 | 52.35 ± 2.63 |
| final weight (FW, kg) | 57.03 ± 1.81 | 56.03 ± 4.21 | 54.35 ± 3.71 | 56.23 ± 2.08 |
| average daily weight gain (ADG, g) | 125.56 ± 57.38 | 133.89 ± 17.82 | 119.17 ± 60.10 | 129.44 ± 30.97 |
| weight gain to feed ratio (F/W, %) | 10.17 ± 4.08 | 7.91 ± 1.13 | 9.51 ± 4.45 | 9.16 ± 1.94 |
| net energy/weight gain (E/W, %) | 139.63 ± 39.85 | 124.89 ± 17.99 | 123.67 ± 28.99 | 139.18 ± 29.54 |

3.2. Effects on Feed Intake and Apparent Digestibility

There were no differences in nutrient intake (P > 0.05) during the whole experiment. It can also be noted that there were no differences in apparent digestibility of EE, CP and DM among the four groups. Furthermore, the digestibility of OM in group I was greater than that in group III (P < 0.05) as indicated in Table 5.

| Items | Ι | II | III | IV |
|---|---------------------------|--------------------------|------------------------------|---------------------------------|
| Dry matter intake $(g \cdot d^{-1})$ | 1126.95 ± 176.51 | 1065.67 ± 216.43 | 1114.36 ± 131.39 | 1125.29 ± 164.35 |
| Organic matter intake (g·d ⁻¹) | 1071.88 ± 166.25 | 1019.73 ± 203.77 | 1058.39 ± 123.76 | 1065.76 ± 154.80 |
| Digestibility of OM (%) | 70.23 ± 6.94 $^{\rm a}$ | $68.15\pm9.10~^{\rm ab}$ | $63.01 \pm 5.12^{\text{ b}}$ | 66.60 ± 10.60 ^{ab} |
| Digestibility of EE (%) | 79.23 ± 4.52 | 80.41 ± 5.23 | 77.37 ± 3.40 | 77.00 ± 7.60 |
| Digestibility of CP (%) | 68.46 ± 8.57 | 64.92 ± 14.96 | 69.56 ± 9.85 | 72.90 ± 10.79 |
| Digestibility of DM (%) | 68.85 ± 6.82 | 65.48 ± 9.06 | 61.56 ± 4.99 | 65.74 ± 10.01 |

Table 5. Effects of dietary supplementation with different inclusion of SMS-MP on feed intake and apparent digestibility of growing sika deer.

Note: ^{a, b} Values within a row with different superscripts differ significantly (P < 0.05).

3.3. Effects on Serum Biochemical Indices

Based on the serum biochemical indexes of the tested deer (Table 6), SMS-MP silage had no significant effects on the serum concentrations of AST, BUN, CRN, ALP, ALB, TP or ALT (P > 0.05). Serum concentrations of γ -GT in group I was greater than that in other groups (P < 0.05). GLUC in group III was significantly higher than group II and I (P < 0.05), and extremely significantly higher than that of group IV (P < 0.01).

Table 6. Effects of dietary supplementation with different inclusion of SMS-MP on serum biochemical indexes of growing sika deer.

| Items | I | II | III | IV |
|---------------|----------------------------|-------------------------------|------------------------------|------------------------------|
| AST (U/L) | 51.67 ± 7.64 | 54.67 ± 4.62 | 54.67 ± 1.53 | 49.67 ± 2.08 |
| BUN (mmol/L) | 4.72 ± 0.59 | 5.29 ± 1.45 | 4.70 ± 1.00 | 6.45 ± 1.27 |
| CRE (umol/L) | 137.00 ± 4.36 | 137.33 ± 10.02 | 136.00 ± 15.72 | 141.00 ± 7.81 |
| γ-GT (U/L) | $19.33 \pm 3.21 \ ^{ m b}$ | $25.67\pm4.04~^{\rm a}$ | $19.33 \pm 2.31 \ { m b}$ | $21.67\pm2.08~^{\mathrm{b}}$ |
| GLUC (mmol/L) | 6.99 ± 0.99 $^{ m ABb}$ | $7.28 \pm 1.31 ^{\text{ABb}}$ | $9.99\pm1.31~^{\mathrm{aA}}$ | $6.31\pm0.74~^{ m cB}$ |
| ALP (U/L) | 238.00 ± 44.19 | 354.67 ± 68.97 | 313.00 ± 149.92 | 228.33 ± 72.59 |
| ALB(g/L) | 24.33 ± 0.58 | 25.33 ± 0.58 | 25.00 ± 1.73 | 25.33 ± 0.58 |
| TP(g/L) | 54.60 ± 1.42 | 55.67 ± 0.55 | 54.53 ± 4.01 | 55.03 ± 1.07 |
| ALT (U/L) | 54.00 ± 22.72 | 60.00 ± 4.58 | 54.33 ± 6.66 | 57.33 ± 4.04 |

Note: ^{a, b} Values within a row with different superscripts differ significantly (P < 0.05). ^{A, B} Values within a row with different superscripts differ extremely significantly (P < 0.01). AST: aspartate aminotransferase; BUN: blood urea nitrogen; CRE: creatinine; γ -GT: gamma-glutamyl transpeptidase; GLUC: blood sugar; ALP: alkaline phosphatase; ALB: albumin; TP: total protein; ALT: alanine aminotransferase.

3.4. Effects on Blood Physiological Indexes

The parameters of serum physiological indexes are shown in Table 7. RBC and Hb in IV were significantly lower than in other groups (P < 0.05).

Table 7. Effects of dietary supplementation with different inclusion of SMS-MP on blood physiological indexes of growing sika deer.

| Items | Ι | II | III | IV |
|---------------------------|--------------------------------|--------------------------|---------------------------------|-----------------------------|
| WBC (10 ⁹ /L) | 2.97 ± 1.57 | 3.01 ± 1.01 | 3.28 ± 1.44 | 1.68 ± 0.84 |
| RBC (10 ¹² /L) | 6.79 ± 0.63 $^{\rm a}$ | 6.71 ± 0.50 $^{\rm a}$ | 7.92 ± 1.61 $^{\rm a}$ | $5.79\pm0.40^{\text{ b}}$ |
| Hb (g/L) | 92.67 ± 4.73 $^{\mathrm{a}}$ | $99.33\pm4.51~^{\rm a}$ | 103.33 ± 15.53 ^a | 81.00 ± 8.72 ^b |
| HCT (%) | 0.51 ± 0.23 | 0.63 ± 0.07 | 0.43 ± 0.14 | 0.46 ± 0.22 |
| MCV (fL) | 74.93 ± 32.01 | 93.20 ± 11.79 | 58.73 ± 33.75 | 79.17 ± 36.83 |
| MCH (pg) | 13.70 ± 0.95 | 14.60 ± 0.44 | 13.13 ± 0.84 | 14.00 ± 1.15 |
| MCHC (g/L) | 215.00 ± 112.41 | 158.00 ± 16.52 | 263.33 ± 103.37 | 210.00 ± 108.86 |
| RDW-CV (%) | 0.21 ± 0.01 | 0.22 ± 0.01 | 0.22 ± 0.01 | 0.20 ± 0.01 |

Note: ^{a, b} Values within a row with different superscripts differ significantly (P < 0.05). WBC: white blood cell; RBC: red blood cell; Hb: hemoglobin; HCT: hematocrit; MCV: mean red blood cell volume; MCH: mean corpuscular hemoglobin; MCHC: mean red blood cell hemoglobin concentration; RDW-CV: coefficient of variation of red blood cell distribution width.

There were no differences in IgA and IgG during the whole experiment. Higher content of IgM was observed with the substitution of SMS for 30% of the concentrate supplement (P < 0.01) as indicated in Table 8.

Table 8. Effects of dietary supplementation with different inclusion of SMS-MP on serum immune globulin of growing sika deer (mg/mL).

| Items | Ι | II | III | IV |
|-------|--------------------------|-------------------|------------------------|------------------------|
| IgA | 28.72 ± 10.53 | 41.59 ± 6.84 | 41.01 ± 6.38 | 28.75 ± 9.32 |
| IgG | 47.15 ± 16.84 | 20.68 ± 0.54 | 47.52 ± 20.88 | 44.50 ± 49.80 |
| IgM | 1.97 ± 0.10 $^{\rm B}$ | $2.96\pm0.16\ ^B$ | $2.84\pm0.02~^{\rm B}$ | $3.12\pm0.49~^{\rm A}$ |

Note: ^{A, B} Values within a row with different superscripts differ extremely significantly (P < 0.01). IgA: Immunoglobulin A; IgG: Immunoglobulin G; IgM: Immunoglobulin M.

3.6. Effects on Plasma Blood Amino Acid

Effects of SMS-MP on Blood amino acids are shown in Table 9. Parameters of blood amino acid were not affected by dietary SMS-MP level (P > 0.05).

Table 9. Effects of dietary supplementation with different inclusion of SMS-MP on plasma amino acid of growing sika deer (μ mol/mL).

| Items | Ι | II | III | IV |
|-------|---------------------|------------------|---------------------|------------------|
| Ala | 28.32 ± 17.17 | 21.11 ± 0.54 | 20.41 ± 7.29 | 22.48 ± 2.52 |
| Gly | 690.29 ± 228.19 | 637.32 ± 71.88 | 678.78 ± 156.98 | 669.74 ± 58.88 |
| Ile | 162.04 ± 66.29 | 109.80 ± 31.29 | 99.86 ± 37.55 | 132.25 ± 11.00 |
| Leu | 101.60 ± 53.48 | 71.69 ± 9.02 | 73.04 ± 38.69 | 86.14 ± 5.24 |
| Met | 75.60 ± 16.53 | 82.07 ± 17.13 | 67.91 ± 42.62 | 106.60 ± 4.90 |
| Phe | 63.63 ± 13.03 | 70.78 ± 1.99 | 63.60 ± 9.25 | 72.60 ± 2.01 |
| Pro | 1.79 ± 0.50 | 1.67 ± 0.08 | 1.84 ± 0.77 | 1.55 ± 0.32 |
| Ser | 2.46 ± 0.44 | 3.06 ± 0.13 | 2.98 ± 0.94 | 3.11 ± 0.07 |
| Thr | 6.37 ± 1.34 | 7.82 ± 0.05 | 8.39 ± 5.47 | 7.91 ± 0.10 |
| Tyr | 0.62 ± 0.25 | 0.90 ± 0.00 | 1.42 ± 1.67 | 0.90 ± 0.00 |
| Val | 117.96 ± 1.69 | 121.11 ± 1.78 | 132.14 ± 16.00 | 118.05 ± 1.97 |

Note: Ala: alanine; Gly: Glycine; Ile: Isoleucine; Leu: Leucine; Met: methionine; Phe: Phenylalanine; Pro: Proline; Ser: serine; Thr: threonine; Tyr: tyrosine; Val: valine.

4. Discussion

The present experiments were designed to explore the effect of adding SMS-MP to concentrate supplements on the growth performance of growing sika deer. Moreover, adding a small amount of SMS-MP to concentrate supplements had no effect on the health of growing sika deer.

Above all, there was no significant difference in the ADG of growing sika deer in each group, meanwhile, the ratio of F/W in the control group was higher than other groups, but was not significantly different. Thus, it can be concluded that SMS-MP is conducive to the growth of sika deer, improving the utilization rate of feed and reducing the cost of breading. The feed intake of sika deer can be affected by both physiological factors and external factors, including dietary habits, lifestyle behaviors, physiological factors, environmental factors and animal learning abilities [23]. The biodegradation of straw with *Pleurotus ostreatus* increased its nutritional value and digestibility in ruminant diets [24]. According to this study, compared with the control group, SMS-MP had no effect on feed intake of dry matter and organic matter, the apparent digestibility of EE, meanwhile, CP and DM had no effect on growing sika deer. The findings of the current study support that of previous research which determined that SMS-MP could be used as ruminant feed without any harmful effects on eating behavior in cattle [14]. In this study, we found that the CP in concentrate supplements would decrease with the addition of SMS-MP, and the addition of a small amount of SMS-MP did not affect growing sika deer, indicating that the

protein consumption of growing sika deer can be reduced, thereby avoiding environmental pollution caused by excessive consumption of N elements. However, in this study, the SMS-MP replacement of 20% reduces the digestibility of OM, which may be caused by the low organic content of SMS-MP compared with the concentrate supplement and further research is needed.

In ruminants, blood BUN can be influenced by the quantity of dietary protein, level of feed intake, and protein degradability in the rumen [25]. Blood glucose level is used as an indicator of ruminant energy status. In the current study, sika deer replacement of 30% with SMS-MP had lower glucose concentration, and this result may be attributed to lower energy consumption as compared to other groups. There was no significantly different level, however, higher BUN concentration was founded in group IV, it might be concluded that, higher nitrogen (N) due to replacement of 30% with SMS-MP. As an ectoenzyme, γ -GT plays a key role in GSH homeostasis by breaking down extracellular GSH and providing cysteine [26]. γ -GT from mammals have been found to play roles in cell defense against oxidative stress, Parkinson's disease, other neurodegenerative diseases, and cardiovascular disease [27,28]. Compared with other groups, a remarkable increase in the concentration of γ -GT was observed for replacement of 10% with SMS-MP in concentrate supplement. It showed that a small amount of SMS-MP in diets can improve the antioxidant capacity of growing sika deer. Jae et al [29] identified that Hb and HCT levels in elk-fed diets with SMS were significantly increased. As the results showed that the contents of Hb and HCT increased with SMS feeding, but were not significantly different. Regardless of that, replacing 30% of concentrate supplements with SMS-MP in the growing sika deer diet may result in decreased Hb and HCT levels significantly. It means that the addition of high content of SMS-MP may have an adverse effect on the growth of sika deer.

IgM is the first antibody isotype to appear during evolution, ontogeny and immune responses. IgM not only serves as the first line of host defense against infections but also plays an important role in immune regulation and immunological tolerance [30]. Natural IgM autoantibodies (IgM–NAA) are rapidly produced to inhibit pathogens and abrogate inflammation mediated by invading microorganisms and host neoantigens [31]. As reported by Rahman et al. [32], oxidative stress could be inhibited by IgM anti-MDA. Moreover, it is reported that supplementation of cordyceps militaris spent mushroom substrate at 2 g/kg of diet increases immunoglobulin secretion of growing pigs [33]. The findings of our current study imply that SMS-MP inclusion in the diet may have modulating effects on the immune system by activating monocytes, although no significant differences were seen between the SMS-MP groups and control.

Blood or plasma-free amino acid pools represent only a very small proportion of tissue-free amino pools. Plasma-free amino acid profiles have been used as a tool to study nutritional and metabolic aspects of protein status. Static measurements of plasma amino acid levels have limited value as sensitive indicators of protein status [34]. Determination of metabolite or nutrient levels in readily available body fluids (blood and urine) is a long-established practice in studying metabolic changes in intact animals [35]. Plasma alpha amino nitrogen and free amino acid (AA) levels have been determined in an effort to assess protein status more clearly. The sum total of all factors influencing total body AA flux, that is protein synthesis, protein degradation, tissue uptake and efflux (including the role of hormones or dietary factors on these processes) and influx from the small intestine and AA catabolism are reflected by the concentration of AA in the plasma [36]. In this study, there was no significant difference in plasma amino acid content between the groups (P > 0.05). It can be concluded that SMS-MP has no significant effect on the plasma amino acids in the sika deer.

Our previous unpublished study found that replacement of SMS-MP in adult male sika deer and sika doe concentrate supplements for 4 and 11 months, respectively, had no effect on their health. It can be indicated that SMS-MP can be safely replaced with the concentrate supplements of growing sika deer for a long time. The breeding scale of sika deer in China is very small. Due to the actual operation restrictions of the experiment, we can only select a small number of samples (three deer/group) for preliminary experiments. Our results preliminarily indicated that a small amount of SMS-MP can be replaced with the concentrate supplements of growing sika deer. Later, it needs to be extended to larger groups to observe the effect of SMS-MP on growing sika deer.

5. Conclusions

The results of the current study confirmed the applicability of SMS-MP as a feed ingredient in growing sika deer diets. A quantity of 10% and 20% with SMS-MP can be safely replaced with the concentrate supplement of growing sika deer. The economic benefits of growing sika deer breeding can be improved by making full use of SMS-MP to reduce the consumption of concentrate supplements.

Author Contributions: C.Y., M.W., X.C., C.L., W.L. and A.Z. designed the study and C.Y. wrote the paper. M.W. performed the experiments. X.C. did the statistical analysis. C.L. collected the samples and data. W.L. and A.Z. edited and reviewed the paper. All authors have read and agreed to the published version of the manuscript.

Funding: This research was funded by the Development project of Jilin Science and Technology Department (No. 20200404012YY); Liaoyuan City Science and Technology Bureau Industrial Development Project (No. 202100005).

Institutional Review Board Statement: The authors confirm that the ethical policies of the journal, as noted on the journal's author guidelines page, have been adhered to and the appropriate ethical review committee approval has been received. This study was conducted at Jilin Agricultural University in P.R. China and approved by the Experimental Animal Welfare and Ethics Committee of Jilin Agricultural University. All experimental protocols were also in accordance with the Code of Ethics of the EU Directive 2010/63/EU for the care and use of experimental animals.

Informed Consent Statement: Not applicable.

Data Availability Statement: The datasets generated and/or analysed during the current study are available from the corresponding author on reasonable request. And they will be provided during review.

Acknowledgments: The SMS-MP was purchased from Jilin Agricultural University.

Conflicts of Interest: The authors declare no conflict of interest.

References

- Kumla, J.; Suwannarach, N.; Sujarit, K.; Penkhrue, W.; Kakumyan, P.; Jatuwong, K.; Vadthanarat, S.; Lumyong, S. Cultivation of mushrooms and their lignocellulolytic enzyme production through the utilization of agro-industrial waste. *Molecules* 2020, 25, 2811. [CrossRef] [PubMed]
- Zhou, J.; Ge, W.; Zhang, X.; Wu, J.; Chen, Q.; Ma, D.; Chai, C. Effects of spent mushroom substrate on the dissipation of polycyclic aromatic hydrocarbons in agricultural soil. *Chemosphere* 2020, 259, 127462. [CrossRef] [PubMed]
- Royse, D.J.; Rhodes, T.W.; Ohga, S.; Sanchez, J.E. Yield, mushroom size and time to production of *Pleurotus cornucopiae* (oyster mushroom) grown on switch grass substrate spawned and supplemented at various rates. *Bioresour. Technol.* 2004, 91, 85–91. [CrossRef]
- Ko, H.G.; Park, S.H.; Kim, S.H.; Park, H.G.; Park, W.M. Detection and recovery of hydrolytic enzymes from spent compost of four mushroom species. *Folia Microbiol.* 2005, *50*, 103–106. [CrossRef] [PubMed]
- 5. Lau, K.; Tsang, Y.; Chiu, S. Use of spent mushroom compost to bioremediate PAH-contaminated samples. *Chemosphere* 2003, 52, 1539–1546. [CrossRef]
- Hu, T.; Wang, X.; Zhen, L.; Gu, J.; Zhang, K.; Wang, Q.; Ma, J.; Peng, H.; Lei, L.; Zhao, W. Effects of inoculating with lignocellulose-degrading consortium on cellulose-degrading genes and fungal community during co-composting of spent mushroom substrate with swine manure. *Bioresour. Technol.* 2019, 291, 121876. [CrossRef] [PubMed]
- Lou, Z.; Sun, Y.; Bian, S.; Baig, S.A.; Hu, B.; Xu, X. Nutrient conservation during spent mushroom compost application using spent mushroom substrate derived biochar. *Chemosphere* 2017, 169, 23–31. [CrossRef]
- 8. Meng, X.; Dai, J.; Zhang, Y.; Wang, X.; Zhu, W.; Yuan, X.; Yuan, H.; Cui, Z. Composted biogas residue and spent mushroom substrate as a growth medium for tomato and pepper seedlings. *J. Environ. Manag.* **2018**, *216*, 62–69. [CrossRef] [PubMed]
- Phan, C.-W.; Sabaratnam, V. Potential uses of spent mushroom substrate and its associated lignocellulosic enzymes. *Appl. Microbiol. Biotechnol.* 2012, 96, 863–873. [CrossRef] [PubMed]
- Zhu, H.; Sheng, K.; Yan, E.; Qiao, J.; Lv, F. Extraction, purification and antibacterial activities of a polysaccharide from spent mushroom substrate. *Int. J. Biol. Macromol.* 2012, *50*, 840–843. [CrossRef]

- 11. Kwak, W.; Kim, Y.; Seok, J.; Oh, Y.; Lee, S. Molasses and microbial inoculants improve fermentability and silage quality of cotton waste-based spent mushroom substrate. *Bioresour. Technol.* 2009, 100, 1471–1473. [CrossRef] [PubMed]
- Xu, C.; Cai, Y.; Zhang, J.; Matsuyama, H. Feeding value of total mixed ration silage with spent mushroom substrate. *Anim. Sci. J.* 2010, *81*, 194–198. [CrossRef] [PubMed]
- Kwak, W.S.; Jung, S.H.; Kim, Y.I. Broiler litter supplementation improves storage and feed-nutritional value of saw-dust-based spent mushroom substrate. *Bioresour. Technol.* 2008, 99, 2947–2955. [CrossRef] [PubMed]
- 14. Katya, K.; Yun, Y.H.; Park, G.; Lee, J.Y.; Yoo, G.; Bai, S.C. Evaluation of the Efficacy of Fermented By-product of Mushroom, Pleurotus ostreatus, as a Fish Meal Replacer in Juvenile Amur Catfish, Silurus asotus: Effects on Growth, Serological Characteristics and Immune Responses. *Asian-Australas. J. Anim. Sci.* 2014, *27*, 1478–1486. [CrossRef]
- Liu, Y.; Zhao, C.; Lin, D.; Lin, H.; Lin, Z. Effect of water extract from spent mushroom substrate after *Ganoderma balabacense* cultivation by using JUNCAO technique on production performance and hematology parameters of dairy cows. *Anim. Sci. J.* 2015, *86*, 855–862.
- Seok, J.S.; Kim, Y.I.; Lee, Y.H.; Choi, D.Y.; Kwak, W.S. Effect of feeding a by-product feed-based silage on nutrients intake, apparent digestibility, and nitrogen balance in sheep. J. Anim. Sci. Technol. 2016, 58, 25. [CrossRef]
- 17. Medina, E.; Paredes, C.; Murcia, M.D.P.; Bustamante, M.A.; Moral, R. Spent mushroom substrates as component of growing media for germination and growth of horticultural plants. *Bioresour. Technol.* **2009**, *100*, 4227–4232. [CrossRef]
- Mensah, D.L.N.; Addo, P.; Dzomeku, M.; Obodai, M. Bioprospecting of powdered pineapple rind as an organic supplement of composted sawdust for *Pleurotus ostreatus* mushroom cultivation. *Food Sci. Nutr.* 2017, *6*, 280–286. [CrossRef]
- 19. Bhattacharjya, D.K.; Paul, R.K.; Miah, M.N.; Ahmed, K.U. Comparative study on nutritional composition of oyster mushroom (*Pleurotus ostreatus* Fr.) cultivated on different sawdust substrates. *Biores. Commun.* **2015**, *1*, 93–98.
- Jia, B.-Y.; Ba, H.-X.; Wang, G.-W.; Yang, Y.; Cui, X.-Z.; Peng, Y.-H.; Zheng, J.-J.; Xing, X.-M.; Yang, F.-H. Transcriptome analysis of sika deer in China. *Mol. Genet. Genom.* 2016, 291, 1941–1953. [CrossRef]
- De Smedt, J. AOAC validation of qualitative and quantitative methods for microbiology in foods. *Int. J. Food Microbiol.* 1998, 45, 25–28. [CrossRef]
- Kim, S.W.; Morris, J.G.; Rogers, Q.R. Dietary soybean protein decreases plasma taurine in cats. J. Nutr. 1995, 125, 2831–2837. [CrossRef] [PubMed]
- 23. Laeger, T.; Görs, S.; Metges, C.; Kuhla, B. Effect of feed restriction on metabolites in cerebrospinal fluid and plasma of dairy cows. *J. Dairy Sci.* **2012**, *95*, 1198–1208. [CrossRef] [PubMed]
- Kholif, A.E.A.A.; Khattab, H.M.; El-Shewy, A.; Salem, A.Z.M.; El-Sayed, M.; Gado, H.; Mariezcurrena, M.D. Nutrient digestibility, ruminal fermentation activities, serum parameters and milk production and composition of lactating goats fed diets containing rice straw treated with *Pleurotus ostreatus*. *Asian-Australas*. J. Anim. Sci. 2014, 27, 357–364. [CrossRef] [PubMed]
- Karnezos, T.P.; Matches, A.G.; Preston, R.L.; Brown, C.P. Corn supplementation of lambs grazing alfalfa. J. Anim. Sci. 1994, 72, 783–789. [CrossRef] [PubMed]
- Dominici, S.; Valentini, M.; Maellaro, E.; Del, B.B.; Paolicchi, A.; Lorenzini, E.; Tongiani, R.; Comporti, M.; Pompella, A. Redox modulation of cell surface protein thiols in U937 lymphoma cells: The role of gamma-glutamyl transpeptidase-dependent H₂O₂ production and S-thiolation. *Free Radic. Biol. Med.* **1999**, *27*, 623–635. [CrossRef]
- Wada, K.; Hiratake, J.; Irie, M.; Okada, T.; Yamada, C.; Kumagai, H.; Suzuki, H.; Fukuyama, K. Crystal structures of Esche-richia coli gamma-glutamyltranspeptidase in complex with azaserine and acivicin: Novel mechanistic implication for inhibition by glutamine antagonists. J. Mol. Biol. 2008, 380, 361–372. [CrossRef] [PubMed]
- 28. Regula, J.; Krejpcio, Z.; Staniek, H. Bioavailability of iron from cereal products enriched with dried shittake mushrooms (*Lentinula edodes*) as determined by iron regeneration efficacy method in female rats. J. Med. Food **2010**, 13, 1189–1194. [CrossRef]
- Park, J.H.; Kim, S.W.; Do, Y.J.; Kim, H.; Ko, Y.G.; Yang, B.S.; Shin, D.; Cho, Y.M. Spent mushroom substrate influences elk (*Cervus Elaphus Canadensis*) hematological and serum biochemical parameters. *Asian-Australas. J. Anim. Sci.* 2012, 25, 320–324. [CrossRef]
- Liu, J.; Wang, Y.; Xiong, E.; Hong, R.; Lu, Q.; Ohno, H.; Wang, J.-Y. Role of the IgM Fc receptor in immunity and tolerance. *Front. Immunol.* 2019, 10, 529. [CrossRef]
- 31. Lobo, P.I. Role of Natural IgM autoantibodies (IgM-NAA) and IgM anti-leukocyte antibodies (IgM-ALA) in regulating inflammation. *Curr. Top. Microbiol. Immunol.* **2017**, *408*, 89–117. [CrossRef] [PubMed]
- Rahman, M.; Sing, S.; Golabkesh, Z.; Fiskesund, R.; Gustafsson, T.; Jogestrand, T.; Frostegard, A.G.; Hafstrom, I.; Liu, A.; Frostegard, J. IgM antibodies against malondialdehyde and phosphorylcholine are together strong protection markers for ath-erosclerosis in systemic lupus erythematosus: Regulation and underlying mechanisms. *Clin. Immunol.* 2016, 166–167, 27–37. [CrossRef] [PubMed]
- 33. Boontiam, W.; Wachirapakorn, C.; Wattanachai, S. Growth performance and hematological changes in growing pigs treated with Cordyceps militaris spent mushroom substrate. *Vet.-World* **2020**, *13*, 768. [CrossRef] [PubMed]
- Bergen, W.G. Free Amino Acids in Blood of Ruminants—Physiological and Nutritional Regulation. J. Anim. Sci. 1979, 49, 1577–1589. [CrossRef] [PubMed]

- 35. Bergen, W.G.; Henneman, H.A.; Magee, W.T. Effect of dietary protein level and protein source on plasma and tissue free amino acids in growing sheep. *J. Nutr.* **1973**, *103*, 575–585. [CrossRef] [PubMed]
- Rochell, S.J.; Helmbrecht, A.; Parsons, C.M.; Dilger, R.N. Influence of dietary amino acid reductions and Eimeria acervulina infection on growth performance and intestinal cytokine responses of broilers fed low crude protein diets. *Poult. Sci.* 2016, 95, 2602–2614. [CrossRef] [PubMed]