



In Vitro Activity of Omadacycline and Five Comparators against Contemporary Ribotypes of *Clostridioides difficile* in Stockholm, Sweden

Angela Camporeale,^a Chaitanya Tellapragada,^a Jelena Kornijenko,^a Carl Erik Nord,^a Christian G. Giske^{a,b}

^aDepartment of Laboratory Medicine, Division of Clinical Microbiology, Karolinska Institutet, Stockholm, Sweden

^bDepartment of Clinical Microbiology, Karolinska University Hospital, Stockholm, Sweden

ABSTRACT *Clostridioides difficile* infection represents a growing clinical challenge. The new compound omadacycline is a potential treatment alternative, as many antibiotics have limited activity or are rarely used due to costs and side effects. The activity of omadacycline and five comparators was assessed with agar dilution on a 2015-to-2018 collection of 65 *C. difficile* isolates from Sweden. Omadacycline demonstrated *in vitro* activity against the contemporary ribotypes of *C. difficile*, and further clinical investigation is needed.

IMPORTANCE Evaluating the activity of novel antimicrobials like omadacycline is of great interest, as a reliable and efficient antimicrobial treatment for *Clostridioides difficile* infections is in demand.

KEYWORDS agar dilution method, *Clostridioides difficile*, omadacycline

Clostridioides difficile is an important nosocomial pathogen worldwide, causing *C. difficile* infection (CDI). Historically, metronidazole, given orally, has been the mainstay therapeutic agent for the treatment of CDI. A low efficacy of metronidazole among patients with a severe form of the disease and a high recurrence rate among patients despite receiving adequate treatment has prompted the need for treatment alternatives. Even though recurrence rates remain similar, comparative studies proved that vancomycin was superior, and therefore, it is now preferred as an initial treatment (1, 2). Additionally, fidaxomicin has been approved for CDI treatment and tigecycline was evaluated for treatment of patients with CDI. However, fidaxomicin is costly and does not seem to have a window for amortization due to its narrow use, whereas tigecycline is known for its side effects; thus, both are restricted in their utility (3, 4). Omadacycline is an aminomethylcycline antibiotic belonging to the tetracycline class that has been synthesized by chemical modifications of minocycline in order to overcome the most common class-related drug resistance mechanisms (efflux and ribosomal protection) found in bacterial pathogens (5). Several studies have previously reported that omadacycline has potent *in vitro* activity against a range of aerobic bacteria and a few anaerobic bacterial species (6–8). After FDA approval for the treatment of patients with acute bacterial skin and skin structure infections and community-acquired bacterial pneumonia in 2018, the clinical efficacy and safety of omadacycline were recently reported (9). Furthermore, omadacycline was reported as a potential therapeutic agent for CDI due to its ability to not induce toxin production and proliferation of *C. difficile* in an *in vitro* gut model (10). Herein, we report the *in vitro* activity of omadacycline and five comparators against a contemporary collection of *C. difficile* isolates from Stockholm, Sweden. (Preliminary results were submitted as an abstract for ECCMID 2020.)

A collection of clinical *C. difficile* isolates obtained from patients diagnosed with CDI in the Stockholm County region between 2015 and 2018 was used in the present study.

Citation Camporeale A, Tellapragada C, Kornijenko J, Nord CE, Giske CG. 2021. *In vitro* activity of omadacycline and five comparators against contemporary ribotypes of *Clostridioides difficile* in Stockholm, Sweden. *Microbiol Spectr* 9:e01440-21. <https://doi.org/10.1128/Spectrum.01440-21>.

Editor Bonnie Chase Prokesh, University of Texas Southwestern Medical Center

Copyright © 2021 Camporeale et al. This is an open-access article distributed under the terms of the [Creative Commons Attribution 4.0 International license](https://creativecommons.org/licenses/by/4.0/).

Address correspondence to Christian G. Giske, christian.giske@ki.se.

Received 31 August 2021

Accepted 3 September 2021

Published 6 October 2021

Data regarding the contemporary and prevalent ribotypes (RTs) in Sweden during 2015 to 2018 were extracted from the Swedish Public Health Agency database. *C. difficile* isolates belonging to nine different ribotypes were obtained from the Clinical Microbiology Laboratory, Karolinska University Hospital, Huddinge, Stockholm, Sweden, and one isolate belonging to ribotype 027 was obtained from the Department of Clinical Microbiology, Jönköping Hospital, Jönköping, Sweden. A reference strain of *C. difficile*, ATCC 43594 (ribotype 005), was used for quality control. All isolates were freshly subcultured before testing and confirmed as *C. difficile* using matrix-assisted laser desorption ionization–time of flight mass spectrometry (MALDI-TOF MS) (Bruker Diagnostics). The antibiotics tested alongside omadacycline were metronidazole, vancomycin, fidaxomicin, tigecycline, and moxifloxacin; these were incorporated within blood agar at various concentrations (Table 1) to perform the agar dilution method. Briefly, bacterial suspensions at 0.5 McFarland were grown on the aforementioned plates at 37°C for 48 h under anaerobic conditions. The MICs for omadacycline and comparator agents were evaluated according to the breakpoints and/or the epidemiological cut-offs (ECOFFs) recommended by EUCAST (https://www.eucast.org/fileadmin/src/media/PDFs/EUCAST_files/Breakpoint_tables/v_11.0_Breakpoint_Tables.pdf, accessed 4 January 2021). The values for all the study isolates were determined in triplicate, and the geometric mean MICs, MIC₅₀s, and MIC₉₀s are reported.

Sixty-five clinical *C. difficile* isolates were tested. The ribotypes included were 014 ($n = 20$ isolates), 002 ($n = 11$), 023 ($n = 8$), 001 ($n = 7$), 005 ($n = 5$), 078 ($n = 5$), 020 ($n = 4$), 078/126 ($n = 3$), 027 ($n = 1$), and 012 ($n = 1$). The distribution of the study isolates based on their MICs for the antibiotics tested is depicted in (Table 1). The geometric mean MIC, MIC₅₀, and MIC₉₀ for omadacycline were 0.677 mg/liter, 0.5 mg/liter, and 1 mg/liter respectively. The MIC₅₀ and MIC₉₀ of omadacycline reported from the present study are comparable with the results (MIC₅₀, 0.25 mg/liter, and MIC₉₀, 0.5 mg/liter) from a previous study that used the agar dilution method for MIC determination among 21 clinical isolates of *C. difficile* (8). In contrast, our results are higher than the MIC₅₀ (0.031 μ g/ml) and MIC₉₀ (0.031 μ g/ml) reported in a more recent study from Texas, USA (11). The ribotypes tested and, more importantly, the method (broth microdilution) employed for MIC determination in the study reported by Begum et al. were different from the ribotypes and method in our study. Moreover, broth microdilution has been proven to show discrepancies for anaerobic bacteria, especially *C. difficile*; hence, the results from the two studies should be compared with caution (12).

Among the five comparators tested in the present study, four antimicrobials have established ECOFFs and none of them have clinical breakpoints reported by EUCAST. Given this context, we did not use the standard susceptible-intermediate-resistant (S-I-R) definitions for characterizing the study isolates against the individual antibiotics tested. The geometric mean MIC of omadacycline (0.67 mg/liter) was comparable with the geometric mean MIC of metronidazole (0.55 mg/liter) and higher than the values for fidaxomicin (0.17 mg/liter) and tigecycline (0.07 mg/liter) among our strain collection. Despite the higher geometric mean MICs for omadacycline in comparison with those of fidaxomicin and tigecycline, the new agent can be a potential candidate for treatment of CDI due to its favorable pharmacokinetic and pharmacodynamic properties and its limited impact on the normal gut microbiota (10, 13, 14). Substantial variations in the geometric mean MICs were not observed among different ribotypes, with values ranging within 2 or 3 absolute 2-fold concentrations for all the antibiotics tested (Table 2). However, the geometric mean MICs for omadacycline were slightly higher among strains of ribotypes 078/126, 001, and 005 than among the other ribotypes tested in the present study. Furthermore, one isolate each belonging to ribotypes 078/126 and 014 had omadacycline MICs of 8 and 16 mg/liter, respectively. We foresee the need for further genetic characterization of these isolates to study the underlying mechanisms for higher MICs of omadacycline in these two isolates. One limitation of the present study is that we could determine the MIC of omadacycline against only one isolate of *C. difficile* belonging to the well-established virulent ribotype 027, which is more prevalent elsewhere. Infections

TABLE 1 Susceptibility overview of 65 *C. difficile* isolates for 9 antibiotics

Antimicrobials	Breakpoint (mg/liter) ^a	No. (%) of isolates with MIC (mg/liter) of:															GM	
		≤0.008	0.015	0.03	0.06	0.12	0.25	0.5	1	2	4	8	16	32	MIC ₅₀ ^b	MIC ₉₀ ^b	MIC ^c	
Omacycline	—	0 (0)	0 (0)	0 (0)	0 (0)	3 (4.6)	41 (63.1)	17 (26.1)	0 (0)	2 (3.1)	1 (1.5)	1 (1.5)	NT	0.25/0.5	1	0.67		
Metronidazole	≥2	NT ^d	NT	0 (0)	0 (0)	4 (6.1)	49 (75.3)	8 (12.3)	2 (3.1)	1 (1.5)	NT	NT	NT	0.25/0.5	1	0.55		
Vancomycin	≥2	NT	NT	NT	NT	0 (0)	0 (0)	55 (84.6)	6 (9.2)	4 (6.1)	NT	NT	NT	0.5/1	2	1.16		
Fidaxomicin	—	0 (0)	0 (0)	2 (3.1)	11 (16.9)	24 (36.9)	8 (12.3)	0 (0)	2 (3.07)	NT	NT	NT	NT	0.12/0.25	0.5	0.17		
Tigecycline	≥0.25	NT	NT	NT	55 (84.6)	7 (10.7)	1 (1.5)	0 (0)	1 (1.5)	0 (0)	NT	NT	NT	0.03/0.06	0.12	0.07		
Moxifloxacin	≥4	NT	NT	NT	NT	NT	0 (0)	0 (0)	35 (53.8)	23 (35.3)	7 (10.7)	NT	NT	2	4/8	2.99		

^aECOFFs established by EUCAST were used where available. —, no breakpoint established.

^bA range of concentrations is given for ambiguous MIC₅₀ and MIC₉₀ interpretation.

^cGM, geometric mean.

^dNT, concentration not tested.

TABLE 2 Geometric mean MIC by ribotype

Ribotype or strain (no. of isolates tested)	Geometric mean MIC (mg/liter) (unless otherwise indicated) for:					
	Omadacycline	Metronidazole	Vancomycin	Fidaxomicin	Tigecycline	Moxifloxacin
Total (65)	0.67	0.55	1.16	0.17	0.07	2.99
001 (7)	0.84	0.82	1.48	0.13	0.1	2.60
002 (11)	0.53	0.53	1.21	0.28	0.06	3.24
005 (5)	0.72	0.52	1.44	0.18	0.06	2.76
012 (1)	0.5	0.5	1.58	0.08	0.06	8
014 (20)	0.64	0.53	1.12	0.12	0.07	3.4
020 (4)	0.59	0.74	1.18	0.19	0.06	4.23
023 (8)	0.45	0.35	1.02	0.11	0.06	2.51
027 (1)	0.5	0.5	1	0.5	0.06	2
078 (5)	0.58	0.95	1.66	0.19	0.09	2.40
078/126 (3)	1.31	0.62	1.25	0.24	0.09	4.32
ATCC 43594 ^a	1	0.5	2	0.25	0.06	2

^aMICs are given instead of geometric mean MICs for strain ATCC 43594.

caused by *C. difficile* ribotype 027 are still not common in the Stockholm region (15), and hence, these strains were unavailable for testing. In summary, omadacycline demonstrated *in vitro* activity against a contemporary ribotype collection of *C. difficile* isolates from Sweden. Our results are promising but suggest the need for susceptibility testing of a larger and more internationally diverse group of *C. difficile* strains and further investigation of the mechanism of high MICs observed in selected isolates.

ACKNOWLEDGMENT

We kindly thank Paratek Pharmaceuticals for providing omadacycline powder.

REFERENCES

- Johnson S, Louie TJ, Gerding DN, Cornely OA, Chasan-Taber S, Fitts D, Gelone SP, Broom C, Davidson DM, Polymer Alternative for CDI Treatment (PACT) Investigators. 2014. Vancomycin, metronidazole, or tolevamer for *Clostridium difficile* infection: results from two multinational, randomized, controlled trials. *Clin Infect Dis* 59:345–354. <https://doi.org/10.1093/cid/ciu313>.
- Stevens V, Nelson R, Schwab-Daugherty E, Khader K, Jones M, Brown K, Greene T, Croft LD, Neuhauser M, Glassman P, Goetz MB, Samore MH, Rubin MA. 2017. Comparative effectiveness of vancomycin and metronidazole for the prevention of recurrence and death in patients with *Clostridium difficile*. *JAMA Intern Med* 177:546–553. <https://doi.org/10.1001/jamainternmed.2016.9045>.
- Nelson RL, Suda KJ, Evans CT. 2017. Antibiotic treatment for *Clostridium difficile*-associated diarrhoea in adults. *Cochrane Database Syst Rev* 3: CD004610. <https://doi.org/10.1002/14651858.CD004610.pub5>.
- Kechagias KS, Chorepsima S, Triarides NA, Falagas ME. 2020. Tigecycline for the treatment of patients with *Clostridium difficile* infection: an update of the clinical evidence. *Eur J Clin Microbiol Infect Dis* 39:1053–1058. <https://doi.org/10.1007/s10096-019-03756-z>.
- Villano S, Steenbergen J, Loh E. 2016. Omadacycline: development of a novel aminomethylcycline antibiotic for treating drug-resistant bacterial infections. *Future Microbiol* 11:1421–1434. <https://doi.org/10.2217/fmb-2016-0100>.
- Macone AB, Caruso BK, Leahy RG, Donatelli J, Weir S, Draper MP, Tanaka SK, Levy SB. 2014. *In vitro* and *in vivo* antibacterial activities of omadacycline, a novel aminomethylcycline. *Antimicrob Agents Chemother* 58: 1127–1135. <https://doi.org/10.1128/AAC.01242-13>.
- Pfaller MA, Huband MD, Shortridge D, Flamm RK. 2018. Surveillance of omadacycline activity tested against clinical isolates from the United States and Europe as part of the 2016 SENTRY antimicrobial surveillance program. *Antimicrob Agents Chemother* 62:e02327-17. <https://doi.org/10.1128/AAC.02327-17>.
- Stapert L, Wolfe C, Shinabarger D, Marra A, Pillar C. 2018. *In vitro* activities of omadacycline and comparators against anaerobic bacteria. *Antimicrob Agents Chemother* 62:e00047-18. <https://doi.org/10.1128/AAC.00047-18>.
- Lan SH, Chang SP, Lai CC, Lu LC, Chao CM. 2019. The efficacy and safety of omadacycline in treatment of acute bacterial infection: a systemic review and meta-analysis of randomized controlled trials. *Medicine (Baltimore)* 98:e18426. <https://doi.org/10.1097/MD.00000000000018426>.
- Moura IB, Buckley AM, Ewin D, Shearman S, Clark E, Wilcox MH, Chilton CH. 2019. Omadacycline gut microbiome exposure does not induce *Clostridium difficile* proliferation or toxin production in a model that simulates the proximal, medial, and distal human colon. *Antimicrob Agents Chemother* 63:e01581-18. <https://doi.org/10.1128/AAC.01581-18>.
- Begum K, Basseres E, Miranda J, Lancaster C, Gonzales-Luna AJ, Carlson TJ, Rashid T, Eyre DW, Wilcox MH, Alam MJ, Garey KW. 2020. *In vitro* activity of omadacycline, a new tetracycline analog, and comparators against *Clostridioides difficile*. *Antimicrob Agents Chemother* 64:e00522-20. <https://doi.org/10.1128/AAC.00522-20>.
- Hastey CJ, Dale SE, Nary J, Citron D, Law JH, Roe-Carpenter DE, Chesnel L. 2017. Comparison of *Clostridium difficile* minimum inhibitory concentrations obtained using agar dilution vs broth microdilution methods. *Anaerobe* 44:73–77. <https://doi.org/10.1016/j.anaerobe.2017.02.006>.
- Rodvold KA, Pai MP. 2019. Pharmacokinetics and pharmacodynamics of oral and intravenous omadacycline. *Clin Infect Dis* 69:S16–S22. <https://doi.org/10.1093/cid/ciz309>.
- Zhanell GG, Esquivel J, Zelenitsky S, Lawrence CK, Adam HJ, Golden A, Hink R, Berry L, Schweizer F, Zhanell MA, Bay D, Lagace-Wiens PRS, Walky AJ, Lynch JP, III, Karlowsky JA. 2020. Omadacycline: a novel oral and intravenous aminomethylcycline antibiotic agent. *Drugs* 80:285–313. <https://doi.org/10.1007/s40265-020-01257-4>.
- Sandell S, Rashid MU, Jorup-Ronstrom C, Ellstrom K, Nord CE, Weintraub A. 2016. *Clostridium difficile* recurrences in Stockholm. *Anaerobe* 38:97–102. <https://doi.org/10.1016/j.anaerobe.2016.01.005>.