



Lung function and disease severity in cystic fibrosis patients heterozygous for *p.Arg117His*

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ABSTRACT Expression of *p.Arg117His* cystic fibrosis (CF) transmembrane conductance regulator is influenced by a polythymidine (poly-T) tract and a thymidine–guanine (TG) repeat on intron 9, which vary in length and affect exon 10 skipping.

We compared clinical characteristics and the rate of progression of lung disease of CF patients carrying the *p.Arg117His* mutation with different intron 9 varying sequences (poly-T) and mutation classes *in trans*.

Data were collected from patients in Northern Ireland, UK, including diagnostic features, sweat chloride, nutritional status, sputum microbiology, CF-related complications and lung function. Poly-T and TG repeats were determined by PCR. Forced expiratory volume in 1 s (FEV₁) decline was determined from linear regression of FEV₁ measurements of patients over time.

We identified 62 patients with *p.Arg117His*, 55 with a class I/II mutation *in trans* and six with *p.Arg117His/p.Gly551Asp*. 42 patients had 5T and 13 had 7T. All patients had 12 TG repeats. Patients with *p.Arg117His*-5T had greater lung function decline, sweat chloride concentrations, pancreatic insufficiency and prevalence of *Pseudomonas aeruginosa* infection compared with patients with *p.Arg117His*-7T.

Lung function decline and disease severity in *p.Arg117His* is determined by the poly-T tract length and identity of the mutation *in trans*. Patients with *p.Arg117His*-5T and a second class I/II mutation have a severity similar to *p.Phe508del* homozygous patients, although lung function decline is delayed to an older age. There may be linkage disequilibrium between *p.Arg117His* and 12 TG repeats.



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***p.Arg117His* CFTR with 5T repeats is associated with accelerated lung function decline compared with *p.Arg117His*-7T** <http://ow.ly/yAdS308q3dn>

Cite this article as: Shteinberg M, Downey DG, Beattie D, *et al.* Lung function and disease severity in cystic fibrosis patients heterozygous for *p.Arg117His*. *ERJ Open Res* 2017; 3: 00056-2016 [<https://doi.org/10.1183/23120541.00056-2016>].



Introduction

Cystic fibrosis (CF) is caused by mutations in the gene encoding the CF transmembrane regulator (CFTR) protein, a chloride channel located in the luminal membrane of epithelial cells. Over 2000 *CFTR* mutations have been found, and are classified into five classes (I–V) according to the affected step in protein translation and function [1]. Briefly, these mutation classes are: class I, impaired protein syntheses; class II, protein degradation; class III, altered function, e.g. blocked CFTR channel (altered gating); class IV, reduced conductance of the CFTR; and class V, reduced amount of CFTR. CF genotypes which result in partial functioning CFTR (classes IV and V) are usually associated with late diagnosis and a milder phenotype [2].

The *p.Arg117His* (formerly R117H) mutation results in a substitution of histidine for arginine at position 117 of the protein. The CFTR channel generated from *p.Arg117His* has reduced chloride conductance and altered channel gating [3]. Individuals homozygous for the *p.Arg117His* mutation are rarely described and when identified have very limited manifestations of CFTR dysfunction, typically male infertility with little or no sino-pulmonary disease [4–6]. Compound heterozygotes, carrying one severe CF-causing mutation and *p.Arg117His*, have a less severe phenotype than individuals with class I–III mutations; some have no disease at the time of identification. This has been estimated by calculation of the penetrance of CF among individuals expected to carry the *p.Arg117His/p.Phe508del* mutations [7]. This has led to some debate as to whether the *p.Arg117His* mutation should be included in newborn screening panels for CF [7, 8]. Disease severity may also be influenced by the mutation *in trans* with *p.Arg117His*. COMER *et al.* [9] found that patients with *p.Arg117His* and a class I/II mutation have a more severe phenotype than patients with *p.Arg117His* and a class III mutation.

The variable phenotypes of people with CF and the *p.Arg117His* mutation can in part be explained by alternative splicing. Intron 9 (IVS9; formerly exon 8 by previous nomenclature) of the *CFTR* gene contains a variable sequence of five, seven or nine thymidine bases (“polythymidine (poly-T) repeats”), which are immediately adjacent to a splicing site on exon 10 (formerly exon 9). All poly-T variants result in exon 10 skipping. The 5T allele is associated with the least effective exon 10 retention, resulting in high skipping levels of exon 10 (exon 10–). The 9T allele is associated with the highest levels of exon 10 retention. Hence, the proportion of “exon 10–” mRNA gradually decreases from 9T to 7T and 5T individuals, both CF and non-CF [10]. When found *in trans* with another disease-causing mutation, the poly-T tract variable sequence on intron 9 (IVS9T) can be associated with congenital bilateral absence of the vas deferens [11, 12] or no clinical consequences. However, some studies report patients with significant sino-pulmonary disease and a genotype of 5T *in trans* with a CF-causing mutation [13–15]. More commonly, when IVS9-5T is *in cis* with a mutated allele, such as *p.Arg117His*, and accompanied by a second CF-causing mutation, it may cause CF [16].

CF patients with the combination *p.Phe508del/p.Arg117His*-5T may have severe lung disease. MASSIE *et al.* [16] described 41 Australian CF patients with *p.Phe508del/p.Arg117His* and known poly-T status. *p.Phe508del/p.Arg117His*-5T patients had lung disease consistent with CF, while *p.Phe508del/p.Arg117His*-7T patients did not. Lung function was not significantly different between 5T and 7T patients; however, few patients had lung function data and lung function decline by age group was not assessed.

A second genetic variation that may influence protein levels is a sequence of thymidine–guanine (TG) dinucleotide repeats of varying length. This sequence lies immediately before the poly-T tract in intron 9 and may consist of 11, 12 or 13 repeats [17–19]. When associated with 5T poly-T repeats, longer (12 or 13) repeats are associated with less effective splicing and lower protein product, and a higher disease penetrance [20].

The adult and paediatric CF centres care for all patients with CF in Northern Ireland, UK. It was previously found that 14.6% of these patients carry the *p.Arg117His* mutation [9] with varying degrees of disease severity. The aim of this study was to determine whether IVS9-5T is associated with greater decline in forced expiratory volume in 1 s (FEV₁) than IVS9-7T CF in patients carrying a *p.Arg117His* mutation. Secondary objectives were to determine the decline in FEV₁ of *p.Arg117His* patients compared with *p.Phe508del* homozygous patients and a small number of patients who were *p.Gly551Asp/p.Arg117His* heterozygotes.

Methods

We identified all patients in Northern Ireland carrying *p.Arg117His* and *p.Phe508del* homozygous through our CF patient registry. Ethics approval for data analysis from the registry was given (reference 07/Q0104/2). Retrospective data included diagnostic features, *CFTR* mutations, sweat chloride, nutritional status, CF-related complications and lung function. For patients with *p.Arg117His* who participated in a clinical trial with ivacaftor, all data collected were prior to study inclusion. IVS9T length and TG repeats were determined by PCR as described elsewhere [21] using patients’ stored blood samples. This method is based on a single-step PCR, making use of an allele-specific reverse primer matching the 5T allele plus one additional nucleotide at the 3’ end. Accordingly, the complete 5T stretch is encapsulated within the primer sequence which prevents binding and amplification to the 7T or 9T alleles.

Statistical analysis was performed using IBM SPSS Statistics version 21 (IBM, Armonk, NY, USA). Continuous variables are presented as mean, median and standard deviation. Categorical variables are presented as percentages. Continuous variables were compared using the independent t-test or Mann–Whitney test, as appropriate. Categorical variables were compared using the Chi-squared test. The linear mixed model was used to examine if there were group differences in FEV₁ values with age. Logistic regression was used to compare differences in *Pseudomonas aeruginosa* prevalence between the groups adjusted for age. p-values <0.05 were considered statistically significant.

Results

Patient demographics and genetic variation

In total, 62 patients with *p.Arg117His* were included in the analysis. Of these, 55 had *p.Arg117His* in trans with a class I or II mutation (figure 1). Six patients were compound heterozygotes for *p.Arg117His/p.Gly551Asp*. No patients homozygous for *p.Arg117His* or compound heterozygotes for *p.Arg117His* with a class IV or V mutation were identified.

IVS9T status was determined for 54 out of 61 *p.Arg117His* patients. Of these, 42 patients had 5T and 12 patients had 7T. Determination of TG repeats was attempted for all stored DNA samples. However, TG repeats could not be reliably determined by the PCR method for the patients with 7T. In total, 39 patients with 5T were analysed and all had 12 TG repeats (figure 1).

Clinical characteristics of different genetic variants of patients with the *p.Arg117His* mutation compared with *p.Phe508del* homozygotes

To determine whether different genotypes influence clinical outcomes of severity, we grouped CF patients into the following four groups: 1) 39 patients with *p.Arg117His* and IVS9-5T, along with a second class I or II mutation (“5T”); 2) 10 patients with *p.Arg117His* and IVS9-7T, along with a second class I or II mutation (“7T”); 3) six patients with *p.Arg117His* and *p.Gly551Asp* (5T and 7T were grouped together due to the small number of patients in this group); and 4) 191 patients homozygous for *p.Phe508del*.

Lung function

Spirometry values, determined at every clinic visit and several times during hospitalisation for pulmonary exacerbations, were extracted from the patients’ files. To determine lung function decline over time, we calculated median values of FEV₁ % pred over a calendar year for every patient. We used the linear mixed model to calculate FEV₁ % pred decline from the median FEV₁ values for patients in each of the four groups (figure 2). Patients’ ages in the different patient groups were not statistically different, and FEV₁ decline with age was not different between younger and older patients. FEV₁ % pred decline per year was:

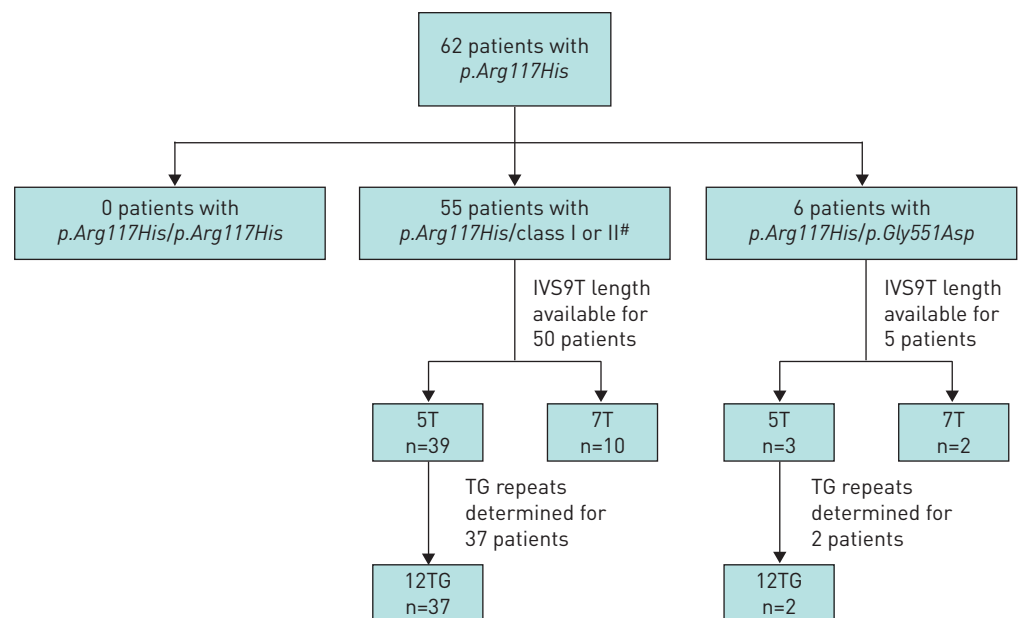


FIGURE 1 Screening of cystic fibrosis database. Patients with *p.Arg117His* and different polythymidine tract variable sequence on intron 9 (IVS9T) length and thymidine–guanine (TG) repeats. #: *p.Phe508del/p.Arg117His* (n=49), *p.Arg560Thr/p.Arg117His* (n=3), *p.Gly542X/p.Arg117His* (n=2) and *p.Glu60X/p.Arg117His* (n=1).

p.Arg117His-5T/class I or II -0.6% , *p.Arg117His*-7T/class I or II $+0.17\%$, *p.Arg117His/p.Gly551Asp* -0.26% and *p.Phe508del* homozygotes -1.02% . From the model it is estimated that *p.Phe508del* homozygotes reach an FEV₁ % pred of 50% at age 36 years, while *p.Arg117His*-5T/class I or II patients reach an FEV₁ % pred of 50% at age 67 years. Patients with *p.Arg117His*-7T/class I or II and *p.Arg117His/p.Gly551Asp* would maintain an FEV₁ % pred of $>50\%$ throughout their lifetime.

Significant differences in the rate of FEV₁ decline were found between 5T and *p.Arg117His/p.Gly551Asp* ($p=0.03$), and between *p.Arg117His/p.Gly551Asp* and *p.Phe508del* homozygotes ($p=0.004$) (table 1). Comparisons between other groups of patients did not reach statistical significance.

Presentation and diagnosis

We compared the different modes of presentation between the four groups of patients, with Bonferroni corrections for multiple comparisons. The mean \pm SD age at diagnosis for the different groups was: *p.Arg117His*-5T 16 ± 21 years, *p.Arg117His*-7T 3.4 ± 10.6 years, *p.Arg117His/p.Gly551Asp* 13.6 ± 19 years and *p.Phe508del* 1.7 ± 5.4 years ($p<0.0001$). For patients diagnosed following a symptomatic clinical presentation (rather than newborn screening), the mean \pm SD age at diagnosis was: *p.Arg117His*-5T 23.9 ± 24 (median 18) years, *p.Arg117His/p.Gly551Asp* 22.4 ± 21 (median 25) years and *p.Phe508del* 1.8 ± 5.2 (median 0.3) years. Sweat chloride values were significantly different between the four groups, with the following mean \pm SD values: *p.Arg117His*-5T 82 ± 14 mEq·L⁻¹, *p.Arg117His*-7T 35 ± 8 mEq·L⁻¹, *p.Arg117His/p.Gly551Asp* 83 ± 38 mEq·L⁻¹ and *p.Phe508del* 110 ± 15 mEq·L⁻¹ ($p<0.0001$ for all comparisons). 14 (35.9%) of the *p.Arg117His*-5T patients were diagnosed following neonatal screening or CF in a sibling, while 18 were diagnosed due to CF-related symptoms: respiratory symptoms (15 patients (38.5%)), failure to thrive or steatorrhoea (two patients (5.1%)), meconium ileus (two patients (5.1%)) or infertility (one patient (2.6%)), some patients had more than one presenting symptom). All nine patients with *p.Arg117His*-7T for whom information regarding diagnosis was available were diagnosed following neonatal screening (table 2).

Metabolic complications

32.4% of the *p.Arg117His*-5T patients were pancreatic insufficient, requiring pancreatic enzyme replacement therapy, compared with 11% of the *p.Arg117His*-7T patients, 50% of the *p.Arg117His/p.Gly551Asp* patients and 100% of *p.Phe508del* homozygotes ($p<0.001$). No patients with *p.Arg117His* had CF-related diabetes versus 11.5% of *p.Phe508del* homozygotes ($p=0.09$). One patient (2.7%) with *p.Arg117His*-5T had liver disease versus 20.8% of *p.Phe508del* homozygotes ($p=0.02$). Mean body mass index (BMI) values are shown in table 2 and were lowest for *p.Phe508del* homozygotes, higher among *p.Arg117His*/class I or II (with no difference between 5T and 7T) and highest among *p.Arg117His/p.Gly551Asp* ($p<0.0001$).

Microbiology

Chronic *P. aeruginosa* colonisation (defined as $>50\%$ positive cultures per year) was highest among *p.Phe508del* homozygotes (52%) and lower in patients with *p.Arg117His*/class I or II and 5T (26.5%), with no *Pseudomonas* colonisation among *p.Arg117His/p.Gly551Asp* or among patients with *p.Arg117His*/class I

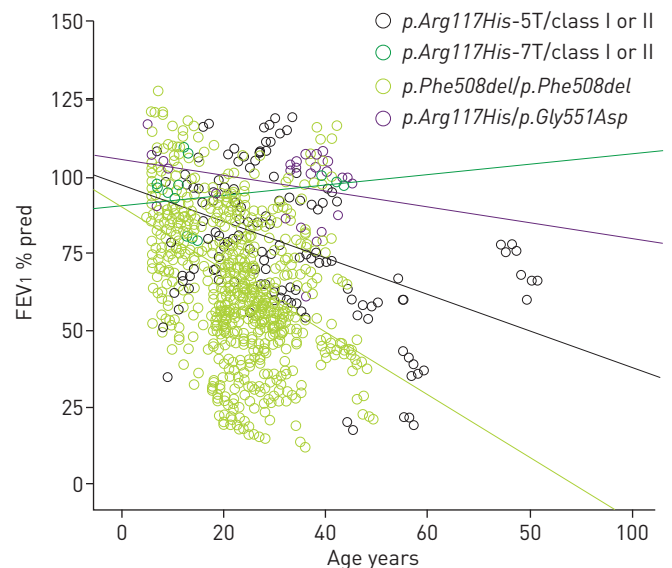


FIGURE 2 Forced expiratory volume in 1 s (FEV₁) % pred values taken as median values of all recordings over a 6-month period for each patient and the FEV₁ per year decline calculated for each of the four groups as shown.

TABLE 1 Estimated rates of lung function decline

	<i>p.Arg117His</i> /class I or II		<i>p.Arg117His</i> / <i>p.Gly551Asp</i>	<i>p.Phe508del</i> homozygotes
	5T [#]	7T [#]		
Patients n	39	10	6	191
Decline in FEV₁ % pred year⁻¹	-0.6	+0.17	-0.26	-1.02
Estimated age when FEV₁ % pred reaches 50% years	67	-	-	36

FEV₁: forced expiratory volume in 1 s. FEV₁ % pred decline as estimated from the median FEV₁ values for patients in each of the four groups. Significant differences were found between 5T and *p.Arg117His/p.Gly551Asp* (p=0.03), and between *p.Arg117His/p.Gly551Asp* and *p.Phe508del* homozygotes (p=0.004). Comparisons between other groups of patients did not reach statistical significance.

or II and 5T (p<0.0001 for comparison). As age was different among the groups, this difference may result from acquired *Pseudomonas* colonisation at older age. To address this, we performed logistic regression for the risk of *Pseudomonas* colonisation. Homozygosity for *p.Phe508del* was found to be associated with a risk of 13.4 (95% CI 4.8–37.7) compared with patients with *p.Arg117His*-5T with no contribution of age. No differences were noted in prevalence of infection with *Staphylococcus aureus* (methicillin sensitive or methicillin resistant), *Haemophilus influenzae*, *Stenotrophomonas maltophilia*, *Burkholderia cepacia* complex, *Achromobacter* or *Aspergillus* species (table 3).

Discussion

The clinical severity of CF is largely determined by the class of the CFTR mutation [22]. Mutations that are associated with residual or partial CFTR function are typically associated with a milder phenotype. The *p.Arg117His* mutation is associated with a poly-T tract and TG repeats which determine the CFTR function. This results in variable clinical consequences. Some patients with *p.Arg117His* can have significant lung disease, while others have minimal or absent disease and are not diagnosed, as estimated by the finding of reduced prevalence [7].

We found that the rate of lung function decline by age is different between four groups of patients: highest for *p.Phe508del* homozygotes, lowest for *p.Arg117His*-7T, and intermediate for *p.Arg117His*-5T and *p.Arg117His*/

TABLE 2 Clinical characteristics of *p.Arg117His* patients

	<i>p.Arg117His</i> /class I or II		<i>p.Arg117His</i> / <i>p.Gly551Asp</i>	<i>p.Phe508del</i> homozygotes	p-value
	5T [#]	7T [#]			
Patients n	39	10	6	191	
Age years	29.5±18	10±13.2	33±13.3	20.2±1	0.001 ^{##}
Male %	62	50	50	60.0	0.893
Age at diagnosis years	16±21	3.4±10.6	13.6±19	1.7±5.4	<0.0001 ^{##,§§}
Age at diagnosis due to symptoms (n) years	(20) 23.9±24	-	(3) 22.4±21	(70) 1.8±5.2	0.01 ^{##}
Diagnostic features[¶] n (%)					
Screening or family history [*]	14 (35.9)	9 (90)	1 (16.7)	98 (50.2)	0.004 ^{¶¶}
Respiratory symptoms	15 (38.5)	0	2 (33.3)	16 (8.4)	<0.0001 ^{##}
Gastrointestinal symptoms	2 (5.1)	0	0	28 (14.7)	0.181
Meconium ileus	2 (5.1)	0	0	29 (15.2)	0.119
Mean sweat chloride mEq·L ⁻¹ (n)	82±14 (28)	35±8 (10)	83±38 (4)	110±15 (72)	<0.0001 ^{##,¶¶,§§}
Current clinical features					
Pancreatic insufficient %	32.4	11.1	50	100	<0.0001 ^{##,+,§§}
BMI kg·m ^{-2§} (n)	25±5 (28)	24.8 (1)	29.4±4 (5)	21.5±3 (100)	<0.0001 ^{##,§§}
BMI percentile ^f (n)	80±21 (7)	62±37 (6)	91 (1)	47.5 (74)	0.011 ^{##}
CFRD %	0	0	0	21 (11.5)	0.089
Liver disease %	2.7	0	0	38 (20.8)	0.018 ^{##}

Data are presented as mean±SD, unless otherwise stated. BMI: body mass index; CFRD: cystic fibrosis (CF)-related diabetes. Clinical features of patients with *p.Arg117His* classified according to IVS9T length and second mutation, and compared with *p.Phe508del* homozygotes. #: IVS9T length was determined for 50 out of 55 patients in this group; ¶: more than one diagnostic feature may be present per patient, information was missing in some patients; *: diagnosis following neonatal screening or a sibling with CF; §: BMI was calculated for adults; f: BMI percentile was calculated for children. After Bonferroni corrections, differences were significant for comparisons: ##: between 5T and *p.Phe508del*; ¶¶: between 7T and *p.Arg117His/p.Gly551Asp*; +: between 7T and *p.Phe508del*; §§: between *p.Arg117His/p.Gly551Asp* and *p.Phe508del*.

p.Gly551Asp compound heterozygotes, with significant differences in rates of lung function decline between patients with *p.Arg117His/p.Gly551Asp* and both 5T and *p.Phe508del* homozygotes. The prediction of the “increase” in lung function among patients with *p.Arg117His-7T* may result from the young age of the patients and inaccurate extrapolation of the change in FEV₁ to older ages. The patients with *p.Arg117His/p.Gly551Asp* had either 5T or 7T (figure 1), so the intermediate phenotype may reflect the average of these two subgroups. This is the first time that the rate of lung function decline by age has been shown to be affected among individuals with *p.Arg117His* and different genetic backgrounds.

We determined IVS9T length by PCR among patients with *p.Arg117His*. All of these patients were compound heterozygotes for a severe mutation and *p.Arg117His*. In most of them (50 out of 56) the second mutation was *p.Phe508del*. We did not check patients' parents for carriage of IVS9 variations and therefore cannot be certain that the “5T” is *in cis* with the *p.Arg117His*. However, it has been previously shown that *p.Arg117His* is in linkage disequilibrium with 5T, whereas the *p.Phe508del* alleles are always on a 9T background [23]. We can therefore assume that when a poly-T is found, a 5T sequence is *in cis* with *p.Arg117His* while the longer sequence is associated with the second mutation, at least in the 50 patients with *p.Phe508del/p.Arg117His*. Among patients with *p.Arg117His*, the most severe phenotype was associated with an IVS9T length of 5 and a class I/II mutation, followed by an IVS9T length of 7 and a class I/II mutation. A small group of patients with *p.Arg117His* and *p.Gly551Asp* had features with inconsistent severity (higher sweat chloride and pancreatic insufficiency but no *Pseudomonas* colonisation and a higher BMI). All patients in whom TG repeats could be determined had 12 TG repeats; therefore, a further classification according to TG repeat number could not be performed. A sequence of 12 TG repeats may be in linkage disequilibrium with the *p.Arg117His* mutation or with an IVS9T length of 5 in our population.

Interestingly, we found fewer patients with *p.Arg117His-7T* than with -5T. This could be explained by reduced penetrance of the *p.Arg117His* mutation, as was hypothesised by THAUVIN-ROBINET *et al.* [7]. Indeed, higher detection through newborn screening (introduced in 1983) may explain the younger average age and age at diagnosis of the patients with *p.Arg117His-7T*, assuming that most of these patients were too mild to have been diagnosed because of symptoms. We found no patients homozygous for *p.Arg117His* or compound heterozygous for *p.Arg117His* and a class IV/V mutation. These classes of mutation are rare, and individuals carrying these combinations of mutations who may either be healthy, have CFTR-related metabolic syndrome or have a very atypical CF phenotype are not diagnosed with CF [7, 24].

However, we have found that 37 patients carrying the *p.Arg117His* mutation with an IVS9T length of 5 and a second class I or II mutation were quite severely affected. Of these patients, 32% were pancreatic insufficient and 25% were chronically colonised with *P. aeruginosa*. Two such patients had meconium ileus at birth; one patient with this genotype was infected with *B. cepacia* complex and another with methicillin-resistant *S. aureus*. The FEV₁ decline was smaller but not significantly different from *p.Phe508del* homozygotes. This could not be attributed to the older age of *p.Phe508del* homozygotes and we were not able to find an age at which there was a change in the slope of lung function decline. These findings argue in favour of the need for early detection and treatment of patients with *p.Arg117His* with an IVS9T length of 5.

TABLE 3 Bacterial isolates in sputum samples from *p.Arg117His* patients versus *p.Phe508del* homozygotes

	<i>p.Arg117His</i> /class I or II		<i>p.Arg117His/p.Gly551Asp</i>	<i>p.Phe508del</i> homozygotes	p-value
	5T	7T			
Patients	34	10	4	173	
<i>Pseudomonas aeruginosa</i> [#]	9 (23.7)	0	0 (0)	108 (57.4)	<0.0001
Mucoid <i>P. aeruginosa</i> [#]	9 (26.5)	0	0	90 (52)	<0.0001
<i>Staphylococcus aureus</i>	20 (60.6)	6 (60)	1 (25)	73 (42.2)	0.157
Methicillin-resistant <i>S. aureus</i>	1 (2.9)	0	0	9 (5.2)	0.848
<i>Haemophilus influenzae</i>	11 (32.4)	0	0	46 (26.6)	0.416
<i>Stenotrophomonas maltophilia</i>	4 (11.8)	0	0	20 (11.6)	0.649
<i>Achromobacter</i> spp.	2 (5.9)	0	1 (25)	1 (2.3)	0.106
<i>Aspergillus</i> spp.	1 (2.6)	0	0	8 (4.3)	0.847
<i>Burkholderia cepacia</i> complex	1 (2.6)	0	0	9 (5.2)	0.573

Data are presented as n or n (%). #: percentages of patients with intermittent (<50% samples positive) or chronic (>50% samples positive) colonisation in the previous year.

This study was a single-centre study, which limited the number of patients as well as longitudinal lung function data. However, patients in this centre had consistent care and we were able to ascertain full details for all patients. As newborn screening has been available since the 1980s, we have ascertainment of almost all CF cases. Rate of FEV₁ decline of *p.Phe508del* homozygotes was 1.02% per year, which is comparable to previous studies [25–27].

Some of the differences between the groups may be attributed to the younger age, and age at diagnosis, of 7T versus 5T patients. However, the differences in sweat chloride values and pancreatic status are typically not influenced by age, and likely represent lower CFTR function in *p.Arg117His*-5T. In patients with the *p.Arg117His* mutation, a class I/II mutation *in trans* and a poly-T sequence of 5 on intron 9 *in cis* convey a more severe phenotype, which has similar rates of decline in FEV₁ compared with *p.Phe508del* homozygotes. These findings support the inclusion of the *p.Arg117His* mutation in newborn screening panels and the potential treatment of such patients with a CFTR potentiator.

Acknowledgements

J.S. Elborn has served as an investigator in Vertex Pharmaceuticals Incorporated clinical studies; has participated in advisory boards or as a consultant for Vertex Pharmaceuticals Incorporated, Novartis, Bayer, Actavis and Boehringer Ingelheim, and has received grant support from Gilead and Novartis.

M. Shteinberg was a visiting research fellow at Queen's University Belfast and received financial support from Novartis Pharmaceuticals.

We acknowledge with gratitude Katherine Lavery, Carina Loughran, Danielle Corey, Joyce Murray, Jane Bell and Jane Carson (Northern Ireland Regional Adult and Paediatric Cystic Fibrosis Centres, Belfast, UK) for assistance with lung function data; Doron Behar (Gene by Gene Ltd, Houston, TX, USA) for assisting with the genetic interpretation; and Elinor Johnston, Susan Martin, Katherine O'Neill and Judy Bradley (Northern Ireland Regional Adult and Paediatric Cystic Fibrosis Centres, Belfast, UK) for assistance with other aspects this project.

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