



Working in "NK Mode": Natural Killer Group 2 Member D and Natural Cytotoxicity Receptors in Stress-Surveillance by $\gamma\delta$ T Cells

Bruno Silva-Santos1* and Jessica Strid2*

¹ Instituto de Medicina Molecular – João Lobo Antunes, Faculdade de Medicina, Universidade de Lisboa, Lisboa, Portugal, ² Division of Immunology and Inflammation, Department of Medicine, Imperial College London, London, United Kingdom

OPEN ACCESS

Edited by:

Pierre Vantourout, King's College London, United Kingdom

Reviewed by:

Olivier Toutirais, University of Caen Normandy, France Paul Austin Moss, University of Birmingham, United Kingdom Daniela Wesch, Christian-Albrechts University (CAU) of Kiel, Germany

*Correspondence:

Bruno Silva-Santos bssantos@medicina.ulisboa.pt; Jessica Strid j.strid@imperial.ac.uk

Specialty section:

This article was submitted to T Cell Biology, a section of the journal Frontiers in Immunology

Received: 23 February 2018 Accepted: 06 April 2018 Published: 24 April 2018

Citation:

Silva-Santos B and Strid J (2018) Working in "NK Mode": Natural Killer Group 2 Member D and Natural Cytotoxicity Receptors in Stress-Surveillance by γδ T Cells. Front. Immunol. 9:851. doi: 10.3389/fimmu.2018.00851 Natural killer cell receptors (NKRs) are germline-encoded transmembrane proteins that regulate the activation and homeostasis of NK cells as well as other lymphocytes. For $\gamma\delta$ T cells, NKRs play critical roles in discriminating stressed (transformed or infected) cells from their healthy counterparts, as proposed in the "lymphoid stress-surveillance" theory. Whereas the main physiologic role is seemingly fulfilled by natural killer group 2 member D, constitutively expressed by $\gamma\delta$ T cells, enhancement of their therapeutic potential may rely on natural cytotoxicity receptors (NCRs), like NKp30 or NKp44, that can be induced selectively on human V δ 1⁺ T cells. Here, we review the contributions of NCRs, NKG2D, and their multiple ligands, to $\gamma\delta$ T cell biology in mouse and human.

Keywords: $\gamma\delta$ T cells, natural killer cell receptors, natural killer group 2 member D, natural cytotoxicity receptors, immunotherapy

INTRODUCTION

Natural killer cell receptors (NKRs) comprise various germline-encoded transmembrane proteins characterized for their capacity to regulate NK cell activation and homeostasis. This large family includes lectin-type receptors, natural cytotoxicity receptors (NCRs), and killer immunoglobulin receptors. The balance between activating and inhibitory signals derived from these receptors controls NK cell functionality. Besides their roles for NK cells, some NKRs, most notably natural-killer group 2 member D (NKG2D), have been known for long to be expressed by some subsets of T cells (1), including $\gamma\delta$ T cells (2). In fact, nearly all human $\gamma\delta$ T cells, and most mouse $\gamma\delta$ T cells, express NKG2D. Importantly, we, and others, have shown that NKG2D is a key determinant of tumor cell recognition by murine intraepithelial $\gamma\delta$ T cells (3, 4), as well as human peripheral blood (2, 5) and tumor-infiltrating (6) $\gamma\delta$ T cells.

By contrast to NKG2D, NCRs were initially thought to be NK cell-specific (7), although this has changed particularly with the discovery of innate lymphoid cells (ILCs) (8). In fact, the acquisition of an NK-like phenotype and functionality was earlier reported on human intestinal intraepithelial lymphocytes (IELs), particularly in celiac disease (9, 10). The NCRs expressed on $\alpha\beta$ IELs triggered interferon- γ (IFN- γ) secretion and degranulation (10), thus suggesting that IEL activation under inflammatory conditions favored the differentiation of "NK-like" effectors performing type 1 cytotoxic functions in an NKR-mediated (and TCR-independent) manner. We have recently built on this to show that human $\gamma\delta$ T cells, specifically of the V δ 1⁺ subset, can be induced to express NCRs upon TCR plus IL-15 (or IL-2) stimulation *in vitro*, and these NCRs enhanced the capacity to target tumor cells of multiple origins, both *in vitro* and *in vivo* [(11, 12), and unpublished data]. In this mini-review, we will focus on the roles of NCRs, NKG2D, and their ligands for $\gamma\delta$ T cell biology in mouse and human.

NKG2D AND ITS LIGANDS

The best-characterized activating NKR is NKG2D. NKG2D is a C-type lectin-like transmembrane receptor, which recognizes a range of different major histocompatibility complex class (MHC) I-related self-ligands induced or upregulated by a variety of cellular stress events, and notably on infected or transformed epithelial cells (ECs) (13). In mice, two isoforms of NKG2D exist, NKG2D-short (S) or NKG2D-long (L), while only the counterpart to the NKG2D-L isoform is expressed in human. The receptor functions as an activating receptor only through its association with signaling adaptor proteins, which are determined by the isoform of NKG2D expressed. NKG2D-S can associate with both DAP10 (recruits phosphatidylinositol 3-kinase) and DAP12 (activates tyrosine kinases Syk and ZAP70) while NKG2D-L is structurally incapable of associating with DAP12 and NKG2D-mediated signaling is mediated solely through DAP10 (14–16).

Engagement of NKG2D can trigger degranulation, cytotoxicity, and/or cytokine production—the distinct outcome of the receptor ligation may be explained by differential isoform and adaptor protein expression. Whereas, mouse CD8⁺ $\alpha\beta$ T cells do not express DAP12 (and the exclusive NKG2D-DAP10 association serves as a costimulatory receptor), mouse epidermal $\gamma\delta$ IELs constitutively express NKG2D-S, NKG2D-L, DAP10, and DAP12, and NKG2D ligation may trigger activity without TCR engagement (17).

Despite the different isoforms of NKG2D, the receptor is highly conserved with the receptors being 70% homologous between human and mouse, for example. NKG2D from one species can bind ligands from another (18). This is curious as the ligands are multiple and are both highly diverse in their amino-acid sequence, domain structure, membrane anchoring as well as exhibiting considerable allelic variation, and a wide range of receptor-binding affinities (Figure 1A). NKG2D ligands identified so far in humans include the MHC class I-chain-related proteins A and B (MICA and MICB) and six different UL16-binding proteins. In mice, three subgroups of NKG2D ligands have been identified: five isoforms of retinoic acid early-inducible 1 (Rae-1) proteins, one murine UL16-binding protein-like transcript 1 (MULT1), and three different isoforms of H60 proteins (Figure 1A). Why the NKG2D receptor is so promiscuous and engaging with so many ligands is not know, however, there are indications that not all ligands are functionally equivalent and that the diversity may allow for unique tissue-specific and contextual functions (1).

NKG2D AS A CRITICAL DETERMINANT OF MOUSE $\gamma\delta$ T CELL ACTIVATION

Study of the NKG2D receptor is not only of huge academic interest, but clearly has therapeutic importance both within cancer, infection, and autoimmunity. Study of this receptor has also given us fundamental insight into $\gamma\delta$ T cell biology. The capacity of murine tissue $\gamma\delta$ T cells to act solely on alterations of autologous stress-antigens, such as those of the NKG2D receptor, and thus survey the "health-status" of a given EC has been termed lymphoid stress-surveillance (LSS) (4, 20, 21) (Figure 1Bi,ii). LSS highlights an important function of $\gamma\delta$ T cells as afferent sensors of cellular dysregulations and as initiators of local and systemic immunity-a clear distinction from conventional $\alpha\beta$ T cell biology. The activation of tissue γδ T cells during LSS in situ occurs seemingly without TCR stimulation (4). However, an alternative explanation could be that the TCR is constitutively engaged in the tissue as was suggested by an elegant study visualizing the $\gamma\delta$ TCR continually signaling in the skin epidermis (22). The self-ligand recognized by the Vy5V81 TCR was not identified in this study, however, the implication of a possible constitutive TCR engagement in the tissue could be a predisposition to respond rapidly to stress-induced ligands recognized by co-stimulatory receptors, such as NKG2D. Whether substituting or synergizing with TCR signaling, these (co)stimulators are of pivotal importance in initiating and tuning the yo T cell response. Interestingly, while epidermal $\gamma\delta$ IEL activate and respond rapidly *in vivo* to transgenic acute upregulation of Rae-1 on ECs (4) (Figure 1Bi), prolonged constitutive expression of Rae-1 (19) renders them hyporesponsive and they remain in a resting state (Figure 1Biii). It is not clear how this tuning of the $\gamma\delta$ IEL responsiveness is regulated according to the length of NKG2D-ligand expression, but the "beneficial autoimmunity" displayed during acute stress responses may be detrimental in chronicity and could be regulated by negative signals mediated by inhibitory NKRs, such as Ly49E and CD94-NKG2A (23). It would be of therapeutic value to understand when and how NKG2D-ligand hyporesponsiveness may occur as solid tumors, for example, can have long-term display of these ligands, which paradoxically could switch off resident tissue immune surveillance.

Natural killer group 2 member D clearly is a cytotoxic receptor and its engagement on $\gamma\delta$ T cells has been shown in many murine studies to induce degranulation, cytotoxicity, and sometimes cytokine production (3, 17, 24). These studies all assessed $\gamma\delta$ T cell function in isolation *in vitro* using $\gamma\delta$ T celllines. However, NKG2D ligands may not always function to generate cytotoxic responses in situ and the outcome of NKG2D receptor ligation is almost certainly context dependent. The different ligands have variable affinity for the receptor and may invoke differential responses. For example, the relatively low affinity ligand H60c is constitutively (and exclusively) expressed in the skin without evoking apparent cytotoxicity in vivo although it could induce cytotoxicity in γδ IEL cell lines in vitro (25). Moreover, while inducible expression of the high affinity Rae-1 ligand in the epidermis in vivo clearly activates the epidermal γδ IEL (Figure 1Bii) no overt cytotoxicity of the ECs was observed (4). Rather, a possible role for NKG2Dligation in EC repair was indicated as this pathway was shown to induce potent expression of type 2 cytokines, particularly IL-13, from the $\gamma\delta$ IEL, which functions to potentiate EC turnover, maintain an intact barrier, and thereby enhance resistance to carcinogenesis (26, 27). Further, Rae-1 transcripts were initially reported in mouse embryonic tissues and NKG2D ligands are also expressed in cells of the bone marrow (28, 29). Interestingly, MICA is also expressed by trophoblasts during normal human



FIGURE 1 | NKG2D ligands and a timely response to alteration in their expression by epidermal γδTCR⁺ intraepithelial lymphocytes (IELs). (A) Human and mouse NKG2D ligands, their cell surface anchor and their affinity to NKG2D are shown. (B) Representative confocal images of murine epidermal Vγ5Vδ1⁺ IELs in whole epidermal sheets following transgenic upregulation of Rae-1 under the involucrin promoter. (i) Single-transgenic and (ii) bi-transgenic mice were fed with doxycycline for 72 h, inducing expression of Rae-1 only in bi-transgenic mice (4). (iii) Mice with sustained expression of Rae-1 under the involucrin promoter (19). The images depict how acute expression of Rae-1 on epithelial cells induces morphological and activational changes in the neighboring IELs, whereas constitutive expression of Rae-1 renders them hyporesponsive. Abbreviations: *allele-dependent NKG2D, natural killer group 2 member D; MIC, MHC class I-chain-related protein; ULBP, cytomegalovirus UL16-binding protein; Rae-1, retinoic acid early-inducible 1; MULT1, murine UL16-binding protein-like transcript 1; al, a2, and a3, analogous to the a1, a2, and a3 domains of MHC 1a proteins; TM, transmembrane protein; GPI, glycosylphosphatidylinositol-linked protein; ND, not determined.

pregnancy, which may be sensed by uterine NK cells (which are not cytolytic). Together these observations suggest that NKG2Dligand expression does not always evoke cytotoxicity *in vivo*, but may have an additional and relatively unexplored role in development and/or tissue repair.

NKG2D-DEPENDENT ACTIVATION OF HUMAN $\gamma\delta$ T CELLS

Most (60–95%) human peripheral blood $\gamma\delta$ T cells express V γ 9V δ 2 TCRs that are uniquely activated by non-peptidic prenyl pyrophosphate antigens (phosphoantigens, PAg) such as isopenthenyl pyrophosphate, which is abundant in tumor cells; or (E)-4-hydroxy-3-methyl-but-enyl pyrophosphate, that is produced by bacteria and parasites [reviewed in Ref. (30)]. PAg-activated V γ 9V δ 2 T cells play important protective roles in infections, such as tuberculosis (31–34); and can kill a variety of tumor cell lines (35, 36). Recent research has clarified how PAg may be "sensed" by V γ 9V δ 2 TCRs. This involves butyrophilin 3A1 (BTN3A1; also known as CD277), a B7 superfamily member that binds PAg in its intracellular B30.2 domain, which leads to significant conformational changes in the extracellular domains of the protein (37–43). Importantly, the effects of both agonist and blocking anti-BTN3A1 mAbs on V γ 9V δ 2 TCR transductants indicated that the TCR is necessary for the activation process (44, 45).

Besides TCR-dependent sensing of intracellular PAg accumulation, the discrimination between tumor and healthy cells by V γ 9V δ 2 T cells seemingly also involves NKG2D, which is expressed on the cell surface of nearly all V γ 9V δ 2 T cells, as it is on peripheral CD8⁺ $\alpha\beta$ T cells. We have observed that NKG2D blockade reduces by circa 50%, the capacity of (PAg-activated) V γ 9V δ 2 T cells to target leukemic cells (as measured by apoptosis induction *in vitro*) (5). Moreover, when we looked for NKG2D ligands whose expression could account for leukemia cell recognition, we found ULBP1 to be the strongest candidate (5, 35). Consistent with this, the

downregulation of ULBP1 impaired, whereas its overexpression enhanced, Vy9V82 T cell-mediated killing of leukemia/lymphoma cells (5). In independent studies, other NKG2D ligands have emerged as major determinants of tumor cell targeting by $\gamma\delta$ T cells: ULBP4 in ovarian and colon carcinomas (46); and ULBP3 in B-cell chronic lymphocytic leukemia (CLL) (47). In the latter report, the critical $\gamma\delta$ T cell subpopulation were V δ 1⁺ (rather than V δ 2⁺) T cells and V δ 1⁺ T cell counts as well as detectable/inducible ULBP3 expression both associated positively with disease control in CLL patients (47). Along the same lines, a recent study showed that ULBP1 and NKG2D expression associated (positively) with longer overall survival of gastric cancer patients (48). Thus, enhancement of NKG2D ligand expression, as achieved by bortezomib or temozolomide treatment of multiple myeloma (49) and glioblastoma multiforme (50) cells, respectively, may have important therapeutic potential, even if NKG2D ligand shedding may constitute an important immune evasion mechanism (51). On the other hand, tumor-directed recombinant ligands that engage NKG2D may also enhance tumor cell targeting, as recently documented against malignant B cells for bispecifics composed of CD20-binding and NKG2D ligand (MICA or ULBP2) domains (52).

The relative importance of NKG2D versus TCR stimulation of $\gamma\delta$ T cells in human is still debated (53). Some studies reported the ability of Vγ9Vδ2 T cells to trigger effector responses through NKG2D stimulation alone (i.e., similarly to NK cells) (36, 54, 55). However, others have failed to show NKG2D-induced activation without simultaneous TCR stimulation (56). In this later case, NKG2D would function in $\gamma\delta$ T cells like in CD8⁺ $\alpha\beta$ T cells, i.e., as a costimulatory receptor accessory to the TCR. Future research should clarify whether the capacity to deploy NKG2D independently of the TCR varies between $V\delta 2^+$ versus $V\delta 1^+$ $\gamma\delta$ T cells. The latter are preferentially found in mucosal tissues and can often be more abundant than $V\delta 2^+$ T cells within solid tumors (57). Unlike Vγ9Vδ2 T cells, Vδ1⁺ T cells do not recognize PAg; instead, intestinal epithelial V δ 1⁺ T cells were shown to bind MICA (and MICB) via a diverse set of Vδ1⁺ TCRs (58, 59), including on transfectants lacking NKG2D (60). Interestingly, our own data have identified a key difference between $V\delta 2^+$ and Vδ1⁺ T cells with regards to the ability to enhance their cytotoxic potential through the upregulation of a distinct class of NK cell receptors-the NCRs.

INDUCED NCRs ON HUMAN V $\delta1^+$ T CELLS

In contrast with NKG2D, NCRs were until recently thought to be NK-specific. However, reports on the acquisition of NCR expression by activated IELs (9, 10), and the subsequent identification of ILCs constitutively expressing NCRs, particularly NKp46 (8), have clearly demonstrated that NCR expression is not an exclusive property of NK cells. Although mouse $\gamma\delta$ T cells seemingly do not express NKp46, we reported that the continued (>2 weeks) activation by TCR agonists (or mitogens-like PHA) in the presence of IL-15 or IL-2 induced NCR expression in a large fraction (>50%) of human $\gamma\delta$ T cells (11). Interestingly, NCR induction was mostly restricted to V δ 1⁺ T cells, as V δ 2⁺ T cells failed to express any of the NCRs above background levels. And, in V δ 1⁺ T cells, the main induced NCRs were NKp30 and NKp44, with NKp46 limited to a smaller fraction (<20%) of activated cells. Antibody-mediated modulation and redirected lysis assays demonstrated the capacity of NKp30 and, to a lesser extent, NKp44, but not NKp46, to enhance V δ 1+ T cell cytotoxicity against tumor cell targets (11). Furthermore, NCR triggering increased IFN- γ expression in V δ 1⁺ T cells, consistent with their role in NK cells (61). Based on these findings, we have developed and established the pre-clinical proof-of-concept for a new (NCR⁺ V\delta1⁺) cellular product, delta one T (DOT) cells, for adoptive immunotherapy of cancer (12) (Figure 2). Moreover, we also demonstrated that this NKp30mediated activation of $V\delta 1^+T$ cells was able to inhibit HIV viral replication, through the production of CCL3/MIP-1a, CCL4/ MIP-1β, and CCL5/RANTES (62). These three CC-chemokines can inhibit viral replication by binding to CCR5, one of the primary co-receptors that HIV-1 uses for entry into CD4⁺ T cells. Collectively, these studies showed that the induced expression of NCRs on V\delta1⁺ T cells enhances their anti-tumor and anti-viral functions.

The acquisition of NCR expression by peripheral blood $V\delta 1^+$ T cells strictly requires strong TCR activation (11). This is consistent with previous work on human IELs, since the ability to upregulate NKp46 and NKp44 in the gut environment only occurred in effector T cells, and the expanded population of NCR⁺ $\alpha\beta$ IELs in celiac disease displayed a highly restricted TCR repertoire indicative of oligoclonal expansion (10). Thus, the major current working hypothesis is that the expression of NCRs in T cells may derive from chronic activation *via* the TCR. This may allow pre-activated T lymphocytes to circumvent antigen-restricted responses, instead employing NCRs to respond continuously against "danger" or "stress," which clearly fits the concept of LSS by $\gamma\delta$ T cells (4, 20, 21).

In this context, it will be critical to identify the NCR ligands that are relevant to NCR⁺ V δ 1⁺ T cell biology, presumably associated with the processes of cellular transformation or viral infection. Thus, the hemagglutinin (HA) protein of the influenza and vaccinia virus binds NKp46 to stimulate NK cells to lyse virus-infected cells (63, 64). Conversely, a HCMV protein, pp65, was reported to inhibit NK cytotoxicity by dissociating the signaling chain, CD3 ζ , from its complex with NKp30 (65).

Self-derived molecules have also been identified as ligands for NCRs, which underlies their capacity to target tumor and/or stressed cells. One of the first self-molecules identified to interact with NKp30 is the human leukocyte antigen-B-associated transcript 3 (BAT3) (66). The expression of this molecule on the tumor cell surface triggers NKp30-mediated killing and the production of IFN- γ and TNF- α . The anti-tumor role of BAT3 was confirmed *in vivo* by showing that peripheral blood NK cells were less efficient at clearing tumors when an anti-BAT3 blocking antibody was administered in mice.

A member of the B7 receptor family, B7-H6, was also shown to bind NKp30 (67). B7-H6 is present on cell surface of both primary tumor and tumor cell lines, while neither healthy nor stressed cells seemingly expressing it. Interestingly, while the presence of B7-H6 on the surface of tumor cells makes them



susceptible to NKp30-mediated killing (67), the binding of B7-H6 to the alternative splice variant, NKp30c, reduces cytotoxicity and IFN- γ production, instead inducing NK cells to produce the immunoregulatory cytokine IL-10 (68). Moreover, the expression of the NKp30c isoform associated with poor clinical prognosis in gastrointestinal sarcoma.

As for NKp44, until recently its best-characterized ligand was an inhibitory molecule, proliferating cell nuclear antigen, that is commonly expressed by tumor cells, and strongly inhibited by NK cell cytotoxicity and IFN- γ production, likely *via* the inhibitory ITIM motif atypically present in the intracellular domain of NKp44 (69). However, a very recent study showed that NKp44 binds to platelet-derived growth factor-DD, a key promoter of tumor cell proliferation, epithelial–mesenchymal transition, and angiogenesis (70). The interaction provoked NK cell activation and the secretion of IFN- γ and TNF- α , which induced tumor cell growth arrest, including in *in vivo* cancer models. This may be a seminal finding in our understanding of how immune cells can recognize tumor cells. Future research should clarify the repertoire of NCR ligands that underlie the enhanced anti-tumor functions of NCR⁺ V\delta1⁺ T cells (**Figure 2**).

CONCLUSION AND PERSPECTIVES

Our current working model includes two stages of yo T cell activation and target cell recognition: first, yo T cells are activated by $\gamma\delta$ TCR ligands (many of which are still unknown); but then NKRs may play the key role in identifying stressed (transformed or infected) targets. This role is physiologically fulfilled by NKG2D (25), constitutively expressed by $\gamma\delta$ T cells (3, 4); but therapeutically it can rely on induced NCRs, particularly for the human V δ 1⁺ T cell subset whose clinical potential is still to be realized (57). For these, the expression of both NKG2D and NCRs provides two functional layers of innate stress-surveillance, particularly of tumors. From a clinical perspective, we have established a clinical-grade protocol to differentiate NCR⁺ V δ 1⁺ T cells *in vitro*, from peripheral blood of cancer patients, toward the development of a new adoptive cell immunotherapy ("DOT cells") (12). Of note, concomitant with TCR stimulation, IL-15 seems to be the key cytokine for NCR induction on V δ 1⁺ T cells (12), which is consistent with previous data on NK cells and IELs (61). Given that the gut is an IL-15-rich environment, it will be interesting to investigate the expression of NCRs on intestinal $\gamma\delta$ T cells, especially since these compose a

large fraction of the IEL compartment; and are highly enriched in V δ 1⁺ (compared to V δ 2⁺) T cells. Such future research should also clarify the enigmatic role of NKp46, since it was not evident from our studies on blood-derived V δ 1⁺ T cells (12); and address the potential relevance of NCRs for the reported regulatory functions of V δ 1⁺ TILs (71, 72).

Besides cancer, another potential application for NCR⁺ V δ 1⁺ T cells is the control of viral infection. Our demonstration that NKp30 engagement on *in vitro* activated V δ 1⁺ T cells is able to suppress HIV-1 replication through the production of CCL3/ MIP-1 α , CCL4/MIP-1 β , and CCL5/RANTES, opens new avenues for the manipulation of $\gamma\delta$ T cells in HIV-1 disease. This is particularly interesting because HIV-1 infection is characterized by a marked expansion of V δ 1⁺ T cells (73–75). This may be even more relevant in mucosal tissues, namely intestinal and cervical mucosa that are sites of HIV-1 entry, where V δ 1⁺ T cells are particularly abundant (76), while CD4⁺ T cells are strongly depleted (77). Of additional great potential is the use of NCR⁺ V δ 1⁺ T cells in CMV infection, given the well-established anti-CMV activity and long-term expansion of V δ 1⁺ T cells (78–81), including postallogeneic stem cell transplantation (80–81).

In conclusion, we propose that NK cell receptor expression by $\gamma\delta$ T cells contributes decisively to their role as a "bridge" between

REFERENCES

- Eagle RA, Trowsdale J. Promiscuity and the single receptor: NKG2D. Nat Rev Immunol (2007) 7:737–44. doi:10.1038/nri2144
- Bauer S, Groh V, Wu J, Steinle A, Phillips JH, Lanier LL, et al. Activation of NK cells and T cells by NKG2D, a receptor for stress-inducible MICA. *Science* (1999) 285:727–9. doi:10.1126/science.285.5428.727
- Girardi M, Oppenheim DE, Steele CR, Lewis JM, Glusac E, Filler R, et al. Regulation of cutaneous malignancy by gammadelta T cells. *Science* (2001) 294:605–9. doi:10.1126/science.1063916
- Strid J, Roberts SJ, Filler RB, Lewis JM, Kwong BY, Schpero W, et al. Acute upregulation of an NKG2D ligand promotes rapid reorganization of a local immune compartment with pleiotropic effects on carcinogenesis. *Nat Immunol* (2008) 9:146–54. doi:10.1038/ni1556
- Lanca T, Correia DV, Moita CF, Raquel H, Neves-Costa A, Ferreira C, et al. The MHC class Ib protein ULBP1 is a nonredundant determinant of leukemia/ lymphoma susceptibility to gammadelta T-cell cytotoxicity. *Blood* (2010) 115:2407–11. doi:10.1182/blood-2009-08-237123
- Groh V, Rhinehart R, Secrist H, Bauer S, Grabstein KH, Spies T. Broad tumor-associated expression and recognition by tumor-derived gamma delta T cells of MICA and MICB. *Proc Natl Acad Sci U S A* (1999) 96:6879–84. doi:10.1073/pnas.96.12.6879
- Moretta A, Bottino C, Vitale M, Pende D, Cantoni C, Mingari MC, et al. Activating receptors and coreceptors involved in human natural killer cellmediated cytolysis. *Annu Rev Immunol* (2001) 19:197–223. doi:10.1146/annurev. immunol.19.1.197
- Satoh-Takayama N, Vosshenrich CAJ, Lesjean-Pottier S, Sawa S, Lochner M, Rattis F, et al. Microbial flora drives interleukin 22 production in intestinal NKp46+ cells that provide innate mucosal immune defense. *Immunity* (2008) 29:958–70. doi:10.1016/j.immuni.2008.11.001
- Meresse B, Chen Z, Ciszewski C, Tretiakova M, Bhagat G, Krausz TN, et al. Coordinated induction by IL15 of a TCR-independent NKG2D signaling pathway converts CTL into lymphokine-activated killer cells in celiac disease. *Immunity* (2004) 21:357–66. doi:10.1016/j.immuni.2004.06.020
- Meresse B, Curran SA, Ciszewski C, Orbelyan G, Setty M, Bhagat G, et al. Reprogramming of CTLs into natural killer-like cells in celiac disease. *J Exp Med* (2006) 203:1343–55. doi:10.1084/jem.20060028
- Correia DV, Fogli M, Hudspeth K, da Silva MG, Mavilio D, Silva-Santos B. Differentiation of human peripheral blood Vdelta1+ T cells expressing the

innate and adaptive immunity, both physiologically (*via* constitutive NKG2D) and therapeutically (through induced NCRs). We believe this entails great potential for future $\gamma\delta$ T cell-based immunotherapies against viral infections or cancer.

AUTHOR CONTRIBUTIONS

BS-S and JS contributed equally to the inception and writing of the manuscript.

ACKNOWLEDGMENTS

We thank Adrian Hayday, Domenico Mavilio, Kelly Hudspeth, and Daniel Correia for insightful discussions on this topic; and Natacha Gonçalves-Sousa for her help to write the manuscript. We acknowledge funding from the Wellcome Trust (100999/Z/13/Z) and Cancer Research UK (C21010/A19788) (to JS) and Fundação para a Ciência e a Tecnologia (PTDC/DTP-PIC/4931/2014) (to BS-S). This publication was sponsored by LISBOA-01-0145-FEDER-007391, project cofunded by FEDER, through POR Lisboa 2020—Programa Operacional Regional de Lisboa, PORTUGAL 2020, and Fundação para a Ciência e a Tecnologia.

natural cytotoxicity receptor NKp30 for recognition of lymphoid leukemia cells. *Blood* (2011) 118:992–1001. doi:10.1182/blood-2011-02-339135

- Almeida AR, Correia DV, Fernandes-Platzgummer A, Da Silva CL, Da Silva MG, Anjos DR, et al. Delta one T cells for immunotherapy of chronic lymphocytic leukemia: clinical-grade expansion/differentiation and preclinical proof of concept. *Clin Cancer Res* (2016) 22:5795–804. doi:10.1158/1078-0432.CCR-16-0597
- Raulet DH, Gasser S, Gowen BG, Deng W, Jung H. Regulation of ligands for the NKG2D activating receptor. *Annu Rev Immunol* (2013) 31:413–41. doi:10.1146/annurev-immunol-032712-095951
- Diefenbach A, Tomasello E, Lucas M, Jamieson AA, Hsia JK, Vivier E, et al. Selective associations with signaling proteins determine stimulatory versus costimulatory activity of NKG2D. *Nat Immunol* (2002) 3:1142–9. doi:10.1038/ ni858
- Wu J, Song Y, Bakker AB, Bauer S, Spies T, Lanier LL, et al. An activating immunoreceptor complex formed by NKG2D and DAP10. *Science* (1999) 285:730–2. doi:10.1126/science.285.5428.730
- Rosen DB, Araki M, Hamerman JA, Chen T, Yamamura T, Lanier LL. A Structural basis for the association of DAP12 with mouse, but not human, NKG2D. *J Immunol* (2004) 173:2470–8. doi:10.4049/jimmunol.173.4.2470
- Nitahara A, Shimura H, Ito A, Tomiyama K, Ito M, Kawai K. NKG2D ligation without T cell receptor engagement triggers both cytotoxicity and cytokine production in dendritic epidermal T cells. *J Invest Dermatol* (2006) 126:1052–8. doi:10.1038/sj.jid.5700112
- Lilienfeld BG, Garcia-Borges C, Crew MD, Seebach JD. Porcine UL16-binding protein 1 expressed on the surface of endothelial cells triggers human NK cytotoxicity through NKG2D. *J Immunol* (2006) 177:2146–52. doi:10.4049/ jimmunol.177.4.2146
- Oppenheim DE, Roberts SJ, Clarke SL, Filler R, Lewis JM, Tigelaar RE, et al. Sustained localized expression of ligand for the activating NKG2D receptor impairs natural cytotoxicity in vivo and reduces tumor immunosurveillance. *Nat Immunol* (2005) 6:928–37. doi:10.1038/ni1239
- Strid J, Tigelaar RE, Hayday AC. Skin immune surveillance by T cells-a new order? Semin Immunol (2009) 21:110–20. doi:10.1016/j.smim.2009.03.002
- Hayday AC. Gammadelta T cells and the lymphoid stress-surveillance response. *Immunity* (2009) 31:184–96. doi:10.1016/j.immuni.2009.08.006
- Chodaczek G, Papanna V, Zal MA, Zal T. Body-barrier surveillance by epidermal gammadelta TCRs. *Nat Immunol* (2012) 13:272–82. doi:10.1038/ ni0612-621d

- Van Beneden K, De Creus A, Stevenaert F, Debacker V, Plum J, Leclercq G. Expression of inhibitory receptors Ly49E and CD94/NKG2 on fetal thymic and adult epidermal TCR V gamma 3 lymphocytes. *J Immunol* (2002) 168:3295–302. doi:10.4049/jimmunol.168.7.3295
- Ibusuki A, Kawai K, Yoshida S, Uchida Y, Nitahara-Takeuchi A, Kuroki K, et al. NKG2D triggers cytotoxicity in murine epidermal γδ T cells via PI3Kdependent, Syk/ZAP70-independent signaling pathway. *J Invest Dermatol* (2014) 134:396–404. doi:10.1038/jid.2013.353
- Whang MI, Guerra N, Raulet DH. Costimulation of dendritic epidermal T cells by a new NKG2D ligand expressed specifically in the skin. *J Immunol* (2009) 182:4557–64. doi:10.4049/jimmunol.0802439
- Strid J, Sobolev O, Zafirova B, Polic B, Hayday A. The intraepithelial T cell response to NKG2D-ligands links lymphoid stress surveillance to atopy. *Science* (2011) 334:1293–7. doi:10.1126/science.1211250
- Dalessandri T, Crawford G, Hayes M, Castro Seoane R, Strid J. IL-13 from intraepithelial lymphocytes regulates tissue homeostasis and protects against carcinogenesis in the skin. *Nat Commun* (2016) 7:12080. doi:10.1038/ ncomms12080
- Nomura M, Zou Z, Joh T, Takihara Y, Matsuda Y, Shimada K. Genomic structures and characterization of Rae1 family members encoding GPI-anchored cell surface proteins and expressed predominantly in embryonic mouse brain. *J Biochem* (1996) 120:987–95. doi:10.1093/oxfordjournals.jbchem.a021517
- Ogasawara K, Benjamin J, Takaki R, Phillips JH, Lanier LL. Function of NKG2D in natural killer cell-mediated rejection of mouse bone marrow grafts. *Nat Immunol* (2005) 6:938–45. doi:10.1038/ni1236
- Morita CT, Jin C, Sarikonda G, Wang H. Nonpeptide antigens, presentation mechanisms, and immunological memory of human Vgamma2Vdelta2 T cells: discriminating friend from foe through the recognition of prenyl pyrophosphate antigens. *Immunol Rev* (2007) 215:59–76. doi:10.1111/j.1600-065X.2006.00479.x
- Shen Y, Zhou D, Qiu L, Lai X, Simon M, Shen L, et al. Adaptive immune response of Vgamma2Vdelta2+ T cells during mycobacterial infections. *Science* (2002) 295:2255–8. doi:10.1126/science.1068819
- Qaqish A, Huang D, Chen CY, Zhang Z, Wang R, Li S, et al. Adoptive transfer of phosphoantigen-specific γδ T cell subset attenuates *Mycobacterium tuberculosis* infection in nonhuman primates. *J Immunol* (2017) 198:4753–63. doi:10.4049/jimmunol.1602019
- Chen ZW. Protective immune responses of major Vγ2Vδ2 T-cell subset in *M. tuberculosis* infection. *Curr Opin Immunol* (2016) 42:105–12. doi:10.1016/j. coi.2016.06.005
- Kabelitz BYD, Bender A, Schondelmaier S, Schoel B, Kaufmann SHE. A large fraction of human peripheral blood gamma/delta + T cells is activated by *Mycobacterium tuberculosis* but not by its 65-kD heat shock protein. *J Exp Med* (1990) 171:667–79.
- 35. Gomes AQ, Correia DV, Grosso AR, Lança T, Ferreira C, Lacerda JF, et al. Identification of a panel of ten cell surface protein antigens associated with immunotargeting of leukemias and lymphomas by peripheral blood γδ t cells. *Haematologica* (2010) 95:1397–404. doi:10.3324/haematol.2009.020602
- 36. Wrobel P, Shojaei H, Schittek B, Gieseler F, Wollenberg B, Kalthoff H, et al. Lysis of a broad range of epithelial tumour cells by human gamma delta T cells: involvement of NKG2D ligands and T-cell receptor- versus NKG2D-dependent recognition. *Scand J Immunol* (2007) 66:320–8. doi:10.1111/j.1365-3083.2007.01963.x
- Sandstrom A, Peigné C-M, Léger A, Crooks JE, Konczak F, Gesnel M-C, et al. The intracellular B30.2 domain of butyrophilin 3A1 binds phosphoantigens to mediate activation of human Vγ9Vδ2 T cells. *Immunity* (2014) 40:490–500. doi:10.1016/j.immuni.2014.03.003
- Gu S, Sachleben JR, Boughter CT, Nawrocka WI, Borowska MT, Tarrasch JT, et al. Phosphoantigen-induced conformational change of butyrophilin 3A1 (BTN3A1) and its implication on Vγ9Vδ2 T cell activation. *Proc Natl Acad Sci* U S A (2017) 114:E7311–20. doi:10.1073/pnas.1707547114
- Peigné C-M, Léger A, Gesnel M-C, Konczak F, Olive D, Bonneville M, et al. The juxtamembrane domain of butyrophilin BTN3A1 controls phosphoantigenmediated activation of human Vγ9Vδ2 T cells. *J Immunol* (2017) 198:4228–34. doi:10.4049/jimmunol.1601910
- Salim M, Knowles TJ, Baker AT, Davey MS, Jeeves M, Sridhar P, et al. BTN3A1 discriminates γδ T cell phosphoantigens from nonantigenic small molecules via a conformational sensor in its B30.2 domain. ACS Chem Biol (2017) 12:2631–43. doi:10.1021/acschembio.7b00694

- Nguyen K, Li J, Puthenveetil R, Lin X, Poe MM, Hsiao CHC, et al. The butyrophilin 3A1 intracellular domain undergoes a conformational change involving the juxtamembrane region. *FASEB J* (2017) 31:4697–706. doi:10.1096/ fj.201601370RR
- Sebestyen Z, Scheper W, Vyborova A, Gu S, Rychnavska Z, Schiffler M, et al. RhoB mediates phosphoantigen recognition by Vγ9Vδ2 T cell receptor. *Cell Rep* (2016) 15:1973–85. doi:10.1016/j.celrep.2016.04.081
- Wang H, Morita CT. Sensor function for butyrophilin 3A1 in prenyl pyrophosphate stimulation of human Vγ2Vδ2 T cells. *J Immunol* (2015) 195:4583–94. doi:10.4049/jimmunol.1500314
- Harly C, Guillaume Y, Nedellec S, Peigne CM, Monkkonen H, Monkkonen J, et al. Key implication of CD277/butyrophilin-3 (BTN3A) in cellular stress sensing by a major human gammadelta T cell subset. *Blood* (2012) 120(11):2269– 79. doi:10.1182/blood-2012-05-430470
- Gu S, Nawrocka W, Adams EJ. Sensing of pyrophosphate metabolites by Vγ9Vδ2 T cells. Front Immunol (2015) 5:688. doi:10.3389/fimmu.2014.00688
- Kong Y, Cao W, Xi X, Ma C, Cui L, He W. The NKG2D ligand ULBP4 binds to TCRgamma9/delta2 and induces cytotoxicity to tumor cells through both TCRgammadelta and NKG2D. *Blood* (2009) 114:310–7. doi:10.1182/ blood-2008-12-196287
- Poggi A, Venturino C, Catellani S, Clavio M, Miglino M, Gobbi M, et al. Vdelta1 T lymphocytes from B-CLL patients recognize ULBP3 expressed on leukemic B cells and up-regulated by trans-retinoic acid. *Cancer Res* (2004) 64:9172–9. doi:10.1158/0008-5472.CAN-04-2417
- Kamei R, Yoshimura K, Yoshino S, Inoue M, Asao T, Fuse M, et al. Expression levels of UL16 binding protein 1 and natural killer group 2 member D affect overall survival in patients with gastric cancer following gastrectomy. *Oncol Lett* (2018) 15:747–54. doi:10.3892/ol.2017.7354
- Niu C, Jin H, Li M, Zhu S, Zhou L, Jin F, et al. Low-dose bortezomib increases the expression of NKG2D and DNAM-1 ligands and enhances induced NK and γδ T cell-mediated lysis in multiple myeloma. *Oncotarget* (2017) 8:5954–64. doi:10.18632/oncotarget.13979
- 50. Chitadze G, Lettau M, Luecke S, Wang T, Janssen O, Fürst D, et al. NKG2Dand T-cell receptor-dependent lysis of malignant glioma cell lines by human $\gamma\delta$ T cells: modulation by temozolomide and A disintegrin and metalloproteases 10 and 17 inhibitors. *Oncoimmunology* (2016) 5:1–13. doi:10.1080/ 2162402X.2015.1093276
- Chitadze G, Bhat J, Lettau M, Janssen O, Kabelitz D. Generation of soluble NKG2D ligands: proteolytic cleavage, exosome secretion and functional implications. *Scand J Immunol* (2013) 78:120–9. doi:10.1111/sji.12072
- Peipp M, Wesch D, Oberg HH, Lutz S, Muskulus A, van de Winkel JGJ, et al. CD20-specific immunoligands engaging NKG2D enhance γδ T cell-mediated lysis of lymphoma cells. *Scand J Immunol* (2017) 86:196–206. doi:10.1111/ sji.12581
- Correia DV, Lopes AC, Silva-Santos B. Tumor cell recognition by γδ T lymphocytes. Oncoimmunology (2013) 2:e22892. doi:10.4161/onci.22892
- Das H, Groh V, Kuijl C, Sugita M, Morita CT, Spies T, et al. MICA engagement by human Vgamma2Vdelta2 T cells enhances their antigen-dependent effector function. *Immunity* (2001) 15:83–93. doi:10.1016/S1074-7613(01) 00168-6
- Rincon-Orozco B, Kunzmann V, Wrobel P, Kabelitz D, Steinle A, Herrmann T. Activation of V gamma 9V delta 2 T cells by NKG2D. *J Immunol* (2005) 175:2144–51. doi:10.4049/jimmunol.175.4.2144
- Nedellec S, Sabourin C, Bonneville M, Scotet E. NKG2D costimulates human V gamma 9V delta 2 T cell antitumor cytotoxicity through protein kinase C theta-dependent modulation of early TCR-induced calcium and transduction signals. J Immunol (2010) 185:55–63. doi:10.4049/jimmunol.1000373
- Silva-Santos B, Serre K, Norell H. gammadelta T cells in cancer. Nat Rev Immunol (2015) 15:683–91. doi:10.1038/nri3904
- Groh V, Steinle A, Bauer S, Spies T. Recognition of stress-induced MHC molecules by intestinal epithelial γδ T cells. *Science* (1998) 279:1737–40. doi:10.1126/science.279.5357.1737
- Xu B, Pizarro JC, Holmes MA, McBeth C, Groh V, Spies T, et al. Crystal structure of a gammadelta T-cell receptor specific for the human MHC class I homolog MICA. *Proc Natl Acad Sci U S A* (2011) 108:2414–9. doi:10.1073/ pnas.1015433108
- Wu J, Groh V, Spies T. T cell antigen receptor engagement and specificity in the recognition of stress-inducible MHC class I-related chains by human epithelial gd T cells. *J Immunol* (2002) 169:1236–40. doi:10.4049/jimmunol.169.3.1236

- Hudspeth K, Silva-Santos B, Mavilio D. Natural cytotoxicity receptors: Broader expression patterns and functions in innate and adaptive immune cells. *Front Immunol* (2013) 4:69. doi:10.3389/fimmu.2013.00069
- Hudspeth K, Fogli M, Correia DV, Mikulak J, Roberto A, Della Bella S, et al. Engagement of NKp30 on Vdelta1 T-cells induces the production of CCL3, CCL4 and CCL5 and suppresses HIV-1 replication. *Blood* (2012) 119(17):4013–6. doi:10.1182/blood-2011-11-390153
- Mandelboim O, Lieberman N, Lev M, Paul L, Arnon TI, Bushkin Y, et al. Recognition of haemagglutinins on virus-infected cells by NKp46 activates lysis by human NK cells. *Nature* (2001) 409:1055–60. doi:10.1038/ 35059110
- Jarahian M, Fiedler M, Cohnen A, Djandji D, Hämmerling GJ, Gati C, et al. Modulation of nkp30- and nkp46-mediated natural killer cell responses by poxviral hemagglutinin. *PLoS Pathog* (2011) 7:e1002195. doi:10.1371/journal. ppat.1002195
- Arnon TI, Achdout H, Levi O, Markel G, Saleh N, Katz G, et al. Inhibition of the NKp30 activating receptor by pp65 of human cytomegalovirus. *Nat Immunol* (2005) 6:515–23. doi:10.1038/ni1190
- 66. Pogge von Strandmann E, Simhadri VR, von Tresckow B, Sasse S, Reiners KSS, Hansen HP, et al. Human leukocyte antigen-B-associated transcript 3 is released from tumor cells and engages the NKp30 receptor on natural killer cells. *Immunity* (2007) 27:965–74. doi:10.1016/j.immuni.2007.10.010
- Brandt CS, Baratin M, Yi EC, Kennedy J, Gao Z, Fox B, et al. The B7 family member B7-H6 is a tumor cell ligand for the activating natural killer cell receptor NKp30 in humans. *J Exp Med* (2009) 206:1495–503. doi:10.1084/ jem.20090681
- Delahaye NF, Rusakiewicz S, Martins I, Menard C, Roux S, Lyonnet L, et al. Alternatively spliced NKp30 isoforms affect the prognosis of gastrointestinal stromal tumors. *Nat Med* (2011) 17:700–7. doi:10.1038/nm.2366
- Rosental B, Brusilovsky M, Hadad U, Oz D, Appel MY, Afergan F, et al. Proliferating cell nuclear antigen is a novel inhibitory ligand for the natural cytotoxicity receptor NKp44. *J Immunol* (2011) 187:5693–702. doi:10.4049/ jimmunol.1102267
- Barrow AD, Edeling MA, Trifonov V, Luo J, Goyal P, Bohl B, et al. Natural killer cells control tumor growth by sensing a growth factor. *Cell* (2018) 172:534–48. doi:10.1016/j.cell.2017.11.037
- Ye J, Ma C, Wang F, Hsueh EC, Toth K, Huang Y, et al. Specific recruitment of γδ regulatory T cells in human breast cancer. *Cancer Res* (2013) 73:6137–48. doi:10.1158/0008-5472.CAN-13-0348
- Ye J, Ma C, Hsueh EC, Eickhoff CS, Zhang Y, Varvares MA, et al. Tumorderived regulatory T cells suppress innate and adaptive immunity through the induction of immunosenescence. *J Immunol* (2013) 190:2403–14. doi:10.4049/ jimmunol.1202369
- Poccia F, Boullier S, Lecoeur H, Cochet M, Poquet Y, Colizzi V, et al. Peripheral V gamma 9/V delta 2 T cell deletion and anergy to nonpeptidic

mycobacterial antigens in asymptomatic HIV-1-infected persons. *J Immunol* (1996) 157:449–61.

- 74. De Paoli P, Gennari D, Martelli P, Basaglia G, Crovatto M, Battistin S, et al. Subset of gamma delta lymphocytes is increased during HIV-1 infection. *Clin Exp Immunol* (1991) 83:187–91. doi:10.1111/j.1365-2249.1991.tb05612.x
- Hinz T, Wesch D, Friese K, Reckziegel A, Arden B, Kabelitz D. T cell receptor gamma delta repertoire in HIV-1-infected individuals. *Eur J Immunol* (1994) 24:3044–9. doi:10.1002/eji.1830241219
- Poles MA, Barsoum S, Yu W, Yu J, Sun P, Daly J, et al. Human immunodeficiency virus type 1 induces persistent changes in mucosal and blood gammadelta T cells despite suppressive therapy. J Virol (2003) 77:10456–67. doi:10.1128/ JVI.77.19.10456-10467.2003
- Brenchley JM, Price DA, Schacker TW, Asher TE, Silvestri G, Rao S, et al. Microbial translocation is a cause of systemic immune activation in chronic HIV infection. *Nat Med* (2006) 12:1365–71. doi:10.1038/nm1511
- 78. Halary F, Pitard V, Dlubek D, Krzysiek R, de la Salle H, Merville P, et al. Shared reactivity of V $\delta 2^{neg} \gamma \delta$ T cells against cytomegalovirus-infected cells and tumor intestinal epithelial cells. *J Exp Med* (2005) 201:1567–78. doi:10.1084/jem.20041851
- 79. Pitard V, Roumanes D, Lafarge X, Couzi L, Garrigue I, Lafon ME, et al. Long-term expansion of effector/memory V{delta}2- {gamma}-{delta} T cells is a specific blood signature of CMV infection. *Blood* (2008) 112:1317–24. doi:10.1182/blood-2008-01-136713
- Ravens S, Schultze-Florey C, Raha S, Sandrock I, Drenker M, Oberdörfer L, et al. Human γδ T cells are quickly reconstituted after stem-cell transplantation and show adaptive clonal expansion in response to viral infection. *Nat Immunol* (2017) 18:393–401. doi:10.1038/ni.3686
- Knight A, Madrigal AJ, Grace S, Sivakumaran J, Kottaridis P, Mackinnon S, et al. The role of Vdelta2-negative gammadelta T cells during cytomegalovirus reactivation in recipients of allogeneic stem cell transplantation. *Blood* (2010) 116:2164–72. doi:10.1182/blood-2010-01-255166

Conflict of Interest Statement: BS-S is a co-founder and share holder of Lymphact— Lymphocyte Activation Technologies S.A. Jessica Strid declares that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Copyright © 2018 Silva-Santos and Strid. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.