



Since January 2020 Elsevier has created a COVID-19 resource centre with free information in English and Mandarin on the novel coronavirus COVID-19. The COVID-19 resource centre is hosted on Elsevier Connect, the company's public news and information website.

Elsevier hereby grants permission to make all its COVID-19-related research that is available on the COVID-19 resource centre - including this research content - immediately available in PubMed Central and other publicly funded repositories, such as the WHO COVID database with rights for unrestricted research re-use and analyses in any form or by any means with acknowledgement of the original source. These permissions are granted for free by Elsevier for as long as the COVID-19 resource centre remains active.

Journal Pre-proof

Current understanding on long non-coding RNAs in immune response to COVID-19

Jing Ding , Xude Yin , Jing Chen , Jin zhou

PII: S0168-1702(22)00285-4
DOI: <https://doi.org/10.1016/j.virusres.2022.198956>
Reference: VIRUS 198956



To appear in: *Virus Research*

Received date: 31 May 2022
Revised date: 11 September 2022
Accepted date: 3 October 2022

Please cite this article as: Jing Ding , Xude Yin , Jing Chen , Jin zhou , Current understanding on long non-coding RNAs in immune response to COVID-19, *Virus Research* (2022), doi: <https://doi.org/10.1016/j.virusres.2022.198956>

This is a PDF file of an article that has undergone enhancements after acceptance, such as the addition of a cover page and metadata, and formatting for readability, but it is not yet the definitive version of record. This version will undergo additional copyediting, typesetting and review before it is published in its final form, but we are providing this version to give early visibility of the article. Please note that, during the production process, errors may be discovered which could affect the content, and all legal disclaimers that apply to the journal pertain.

© 2022 Published by Elsevier B.V.

Current understanding on long non-coding RNAs in immune response to COVID-19

Jing Ding¹, Xude Yin¹, Jing Chen^{1*}, Jin zhou^{1*}

¹ Department of Medical Oncology, Sichuan Cancer Hospital & Institute, Sichuan Cancer Center, School of Medicine, University of Electronic Science and Technology of China, Chengdu, Sichuan, China.

*To whom correspondence should be addressed. Tel: +86 28 85420847;

Email: zhoujt521@163.com; chenjingmaomao111@163.com.

Highlights

- The 2019 coronavirus (COVID-19) caused by severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2) has become a global epidemic threatening the lives and health of people worldwide. Currently, there are no effective therapies or available vaccines for COVID-19. In SARS-CoV-2 infection, lncRNAs are demonstrated to be closely related to viral infection, interferon and cytokine storm in COVID-19. LncRNA NEAT1, DANCR, MALAT1, C058791.1, TTTY15 and TPTEP1 played a role in the infection of SARS-CoV-2. LncRNAs also can modulate the expression of interferon. Inversely, interferon can stimulate the expression of lncRNAs. Besides, lncRNAs can regulate the cytokine storm by downregulating the expression of cytokines. Thus, we can speculate that developing drugs targeting some specific sites in the pathway or relation network of lncRNAs may be promising strategy to treat SARS-CoV-2 infection.

Abstract

Coronavirus disease 2019 (COVID-19) caused by severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2) has become a global pandemic threatening the lives and health of people worldwide. Currently, there are no effective therapies or available vaccines for COVID-19. The molecular mechanism causing acute immunopathological diseases in severe COVID-19 is being investigated. Long noncoding RNAs (lncRNAs) have been proven to be involved in many viral infections, such as hepatitis, influenza and acquired immune deficiency syndrome. Many lncRNAs present differential expression between normal tissue and virus-infected tissue. However, the role of lncRNAs in SARS-CoV-2 infection has not been fully elucidated. This study aimed to review the relationship between lncRNAs and viral infection, interferon and cytokine storms in COVID-19, hoping to provide novel insights into promising targets for COVID-19 treatment.

Keywords: lncRNAs, COVID-19, SARS-CoV-2, viral infection, interferon, cytokine storms

Introduction

Coronavirus disease 2019 (COVID-19) is caused by severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2). COVID-19 spread at an alarming rate, threatening the lives and health of the entire human race. SARS-CoV-2, a single-stranded RNA virus, is a member of the

Coronaviridae family (1). Patients with COVID-19 present with fever, dry cough and respiratory failure, combined with gastrointestinal and neurological symptoms (2). It is projected that asymptomatic dissemination is responsible for nearly half of SARS-CoV-2 transmission (3). Respiratory failure accounts for the highest mortality of patients with COVID-19 (4). SARS-CoV-2 can lead to acute respiratory distress syndrome (ARDS) mediated by proinflammatory cytokines (5). Proinflammatory cytokines can cause severe damage to vital organs and lead to multiple organ dysfunction syndrome (MODS) (6). The main therapeutic measures include convalescent plasma, remdesivir and cytokine blocker drugs (7). Specific vaccination represents a key strategy against SARS-CoV-2 infection. According to the World Health Organization (WHO), more than 360 vaccines have been developed, and 45 vaccines have been evaluated in phase III clinical trials to date (8). These COVID-19 vaccines seem to be well tolerated and display a preventive effect on the original strain and variants of SARS-CoV-2 (8). However, the regulatory mechanism of host genetic expression in response to infection remains unclear (9). It is necessary to explore viral pathogenesis and biological reactions against SARS-CoV-2 in the host. Long noncoding RNAs (lncRNAs), a subclass of noncoding RNAs, lack an open reading frame and have a length of more than 200 nucleotides (10).

By competing with endogenous RNA (ceRNA), lncRNA serves as a miRNA sponge, regulating the expression of target genes. LncRNA, miRNA, and mRNA were used to construct a lncRNA–miRNA–mRNA competing endogenous (ceRNA) network (11). LncRNAs are involved in a broad spectrum of biological processes, especially immunity and inflammatory reactions (12, 13). The lncRNA XIST plays a role in inflammation via the NF- κ B/NLRP3 pathway (14). Chen et al. found that the lncRNA MALAT1 functions as a negative factor in inflammation in sepsis (15). It has been revealed that lncRNA SNHG14 enhances the inflammatory response by increasing the level of ROCK1 and downregulating the expression of miR-136-5p (16). Several lncRNAs have been identified in multiple viral infections, such as hepatitis and AIDS (17, 18). Notably, lncRNAs were also identified in the process of SARS-CoV-2 infection. LncRNA H19 regulates spike transcription of SARS-CoV-2 by binding to the genome of SARS-CoV-2, thereby affecting SARS-CoV-2 infection (19). However, the relationship of lncRNAs with viral infection and the antiviral inflammatory response in COVID-19 needs to be clarified.

LncRNAs in SARS-CoV-2 viral infection

Some lncRNAs, such as NEAT1 and EGOT, increase in influenza A virus and HIV infection (20). In addition, NEAT1 suppresses the replication and growth of HIV and Hantaan virus (HTNV) (21-24). Similarly, in

SARS-CoV-2 viral infection, lncRNA NEAT1 and DANCR change the level of inflammatory transcripts by regulating the expression of immune-related genes (25). Approximately 500 lncRNAs were found to be differentially expressed during SARS-CoV-2 infection (26). HOTAIRM1, PVT1 and AL392172.1 can bind to the SARS-CoV-2 genome with high affinity, suggesting the major regulatory role of lncRNAs in COVID-19 (27). Transcriptome analysis of bronchial epithelial cells in patients with SARS-CoV-2 infection showed that the lncRNA MALAT1 had significantly different expression levels. The lncRNA MALAT1 may be a potential biomarker of SARS-CoV-2 infection (28). An *in silico* study detected the expression levels of lncRNAs both in cell lines and lung tissue. *In vitro*, there were a total of 20 overexpressed lncRNAs and 4 downregulated lncRNAs during SARS-CoV-2 infection. Moreover, NEAT1 was upregulated in the lung tissue of patients with SARS-CoV-2 infection (20). Another study found that a total of 898 lncRNAs (414 overexpressed lncRNAs; 484 downregulated lncRNAs) were differentially expressed between healthy individuals and patients with SARS-CoV-2 infection. In addition, these differentially expressed lncRNAs were likely to be associated with exosomes and regulate the inflammatory process (29). Transcriptome analysis of SARS-CoV-2-infected lung tissues showed that three lncRNAs (C058791.1, TTTY15 and TPTEP1) were enriched with

maximum target genes. These three lncRNAs interact with genes via the lncRNA–miRNA–mRNA pathway and release the mRNA for translation. The activity of AC058791.1 has not been identified; TTTY15 is involved in proteolysis and ubiquitin-dependent catabolism (30), and TPTEP1 inhibits STAT3 phosphorylation (31).

Furthermore, lncRNA H19 can bind to the 5'UTR of the SARS-CoV-2 genome and modulate the spike transcript in viral infection (19). LncRNA-based oligosequences can be candidates to combat SARS-CoV-2. Of note, these results need to be experimentally validated at the patient level. Clinically, a comprehensive profile of COVID-19-related lncRNAs in peripheral blood mononuclear cells of patients with SARS-CoV-2 infection and healthy individuals was built. More specifically, 1072 lncRNAs were differentially expressed between the two populations. The top three increased lncRNAs are ENSG00000231412 (AC005392.2), followed by ENSG00000274173 (AL035661.1) and ENSG00000231535 (LINC00278). The top decreased lncRNAs are ENSG00000229807 (XIST) and ENSG00000273160 (AL359962.2) (32). These differentially expressed lncRNAs may be potential biomarkers for the diagnosis and prognosis of patients with SARS-CoV-2 viral infection. Collectively, these COVID-19-related lncRNAs may be possible innovative candidates against SARS-CoV-2 infection. Further studies are needed to fully identify and understand the

functions of lncRNAs in COVID-19.

LncRNAs and interferon in COVID-19

LncRNAs were shown to modulate interferon-stimulated genes (ISGs) in the inflammatory process. LncRNA#32/LUARIS can combat EMCV, HBV and HCV infection by regulating the transcription factor ATF2 to alter the expression of ISG (33). Interestingly, angiotensin I converting enzyme 2 (ACE2), the receptor for COVID-19, is an ISG. SARS-CoV-2 could exploit species-specific interferon-driven upregulation of ACE2 to aggravate the host immune response and enhance infection (5). Additionally, lncRNAs can be stimulated by interferon (IFN). The expression of NRIR/lncCMPK2 can be activated by IFN and then modulated by signal transducer and activator of transcription 2 (STAT2). There was a reliable association between the expression of IFN and SARS-CoV-2 infection both in laboratory and clinical studies (28). It was observed that lncRNAs can modify ISG levels in different viral infections (33-36). In contrast, Laha et al. found that the expression of lncRNAs had no correlation with interferons. Then, they investigated the interaction between lncRNAs and heterogeneous nuclear ribonucleoproteins (hnRNPs), a group of RNA-binding proteins. They found that several lncRNAs could be regulated by interferon regulatory factors (IRFs) and STAT in response to SARS-CoV-2 infection (20). LncRNAs might be involved in the antiviral response by regulating IFN. However, several

host lncRNAs can repress the viral immune response by downregulating type I interferons (IFN-1). In the early stage of SARS-CoV-2 viral infection in asymptomatic patients, the production of IFN, especially IFN- β , was delayed. More importantly, SARS-CoV-2 N protein can inhibit IFN production by disturbing the retinoic acid-induced gene I (RIG-I) pathway (37). Clinically, IFN- α is a common interferon for antiviral therapy in patients with COVID-19. These lncRNAs mediate IFN regulation in COVID-19 and should be validated to explore effective antiviral strategies in future studies.

lncRNAs regulate cytokine storms in COVID-19

The organ damage caused by the inflammatory reaction accounts for the high mortality and morbidity in COVID-19 (38, 39). Studies in human and animal models implied that immunopathological events might lead to the death of patients with SARS-CoV-2 viral infection (40, 41). In addition, the lung tissue of patients with SARS-CoV-2 infection displayed pathological infiltration of immune cells such as macrophages and monocytes (42). It has been validated that an inflammatory cytokine storm, an overwhelming inflammatory immune response with hyperproduction of mainly proinflammatory cytokines, such as IL-1, IL-6, IL-12, IFN- γ , and TNF- α , contributes to the development of SARS-CoV-2 infection and is the cause of severe COVID-19 (43-45). lncRNAs can target cytokines in inflammatory cytokine storms in

COVID-19. LncRNAs with the potential to regulate the inflammatory response showed differential expression in patients with SARS-CoV-2 infection compared with healthy individuals (29). LncRNAs MALAT1 and NEAT1 may contribute to the development of inflammation in SARS-CoV-2-infected cells (27). In contrast, 22 lncRNAs bound to 10 important cytokines, and 8 of 22 lncRNAs targeted multiple cytokines. RAD51-AS1 and lncCXCR4 each can target 3 cytokines. Notably, the lncRNA NORAD, which is activated by DNA damage, can bind with 5 cytokines, including interleukin (IL)-6, IL-10, CSF3, tumor necrosis factor (TNF)- α and CXCL10 (46). LncRNAs may target and bind important cytokine nucleotide sequences and possibly decrease the expression of cytokines, thus reducing the emergence of cytokine storms in the infection. Given the interaction between lncRNAs and inflammatory cytokines, some methods, such as viral gene therapy, RNAi knockdown, viral vectors and antisense oligonucleotides, have been used in clinical practice (47, 48). Agents targeting lncRNAs show promise in enhancing the anti-SARS-CoV-2 response by inhibiting the cytokine storm.

LncRNAs function as potential targets for COVID-19

Several functional lncRNAs involved in viral infection and cytokine storms in COVID-19 are summarized in **Table 1**. In a coexpression network analysis of human lung epithelial cell lines and bronchoalveolar

lavage fluid from patients with SARS-CoV-2 infection, four lncRNAs (WAKMAR2, EGOT, EPB41L4A-AS1, and ENSG00000271646) were upregulated. These four lncRNAs were associated with multiple cytokine pathways and overactivated inflammatory responses (49). LncRNAs can regulate the expression of IL-6 and NLRP3 through epigenetic, transcriptional, and post-transcriptional mechanisms. Agents targeting these signalling pathways have been developed. Tocilizumab, an IL-6 receptor antagonist, effectively inhibits the IL-6 receptor (IL-6R) and reduces the serum levels of C-reactive protein (CRP) and serum amyloid A (SAA) (50, 51). Similarly, BML-111, an IL-6 blocker, can increase the level of lncRNA MALAT1 and then downregulate the expression of inflammatory factors, such as monocyte chemoattractant protein-1 (MCP-1) and IL-6 (52, 53). A drug perturbation analysis found that digoxin and proscillaridin can regulate gene expression levels by increasing or decreasing the expression levels of some lncRNAs. By conducting molecular docking and drug perturbations on gene expression, we found that digoxin and proscillaridin can be used to treat severe COVID-19 infections (54). Moreover, a bioinformatic analysis implied that the TGF-beta signalling pathway is interactive and involved in the network of lncRNAs, human proteins, and miRNAs. The TGF-beta signalling pathway may be a promising target for COVID-19 treatment (55). These studies suggested that lncRNAs can interact with several inflammatory

factors, including inflammatory genetic processes and cytokine release in COVID-19. LncRNAs may serve as potential therapeutic enhancers in combatting SARS-CoV-2.

Table1. A summary of functional lncRNAs involved in viral infection and cytokine storm in COVID-19.

LncRNA	Function
LncRNAs in SARS-CoV-2 viral infection	
NEAT1	Upregulated in the lung tissue of patients with SARS-CoV-2 infection (20)
MALAT1	May be a potential biomarker of SARS-CoV-2 infection(28)
HOTAIRM1/PVT1 /AL392172.1	Bind to the SARS-CoV-2 genome with the high affinity(27)
TTY15	Regulates proteolysis, ubiquitin dependent catabolism(30)
TPTEP1	Inhibits STAT3 phosphorylation(31)
LncRNA H19	Binds to the 5'UTR of SARS-CoV-2 genome and modulates the Spike transcript(19)
LncRNAs regulate cytokine storm in COVID-19	
MALAT1	Induces inflammatory responses and release of IL-6 and TNF- α (52)
DANCR	Regulates ncRNA-mRNA network in inflammation(25)
HOTAIR	Regulates activation of NF- κ B and IL-6 and iNOS expression(56)
NEAT1	Promotes activation of inflammasomes in macrophages(57)
NORAD	Binds with IL-6, IL-10, CSF3, TNF-a and CXCL10(46)
RAD51-AS1	Leads to expression of pro-inflammatory cytokines(46)

Summary

In SARS-CoV-2 infection, lncRNAs have been demonstrated to be related to viral infection, interferon and cytokine storms in COVID-19 (**Figure 1**). These lncRNAs interacted with genes of SARS-CoV-2 via the lncRNA-miRNA-mRNA pathway. LncRNAs can also modulate the expression of interferon. Conversely, interferon can stimulate the expression of lncRNAs. In addition, lncRNAs bind with important or multiple cytokine storm cytokines. LncRNAs identified in the COVID-19

cytokine storm have the potential to serve as disease markers or drug targets. Thus, we can speculate that developing drugs targeting some specific sites in the pathway or network of lncRNAs may be a promising strategy to treat SARS-CoV-2 infection. More experimental studies are needed to further confirm the regulatory mechanism of lncRNAs in COVID-19.

AUTHOR CONTRIBUTION

Jing Ding collected and reviewed the literature and wrote the manuscript. Xude Yin assisted in drawing. Jing Chen revised the manuscript. Jing zhou designed the main study and reviewed this manuscript. All authors have read and approved the final submitted manuscript.

CONFLICT OF INTEREST

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

FUNDING

This work was financially supported by National Natural Science Foundation of China (82002580).

ACKNOWLEDGMENTS

None.

REFERENCES

1. Pontecorvi G, M Bellenghi, E Ortona and A Carè. microRNAs as new possible actors in gender disparities of Covid-19 pandemic. *Acta Physiol (Oxf)*(2020), 230(1), e13538 doi:10.1111/apha.13538
2. Wiersinga W J, A Rhodes, A C Cheng, S J Peacock and H C Prescott. Pathophysiology, Transmission, Diagnosis, and Treatment of Coronavirus Disease 2019 (COVID-19): A Review. *Jama*(2020), 324(8), 782-793 doi:10.1001/jama.2020.12839
3. Al-Tawfiq J A. Asymptomatic coronavirus infection: MERS-CoV and SARS-CoV-2 (COVID-19). *Travel Med Infect Dis*(2020), 35, 101608 doi:10.1016/j.tmaid.2020.101608
4. Ruan Q, K Yang, W Wang, L Jiang and J Song. Clinical predictors of mortality due to COVID-19 based on an analysis of data of 150 patients from Wuhan, China. *Intensive Care Med*(2020), 46(5), 846-848 doi:10.1007/s00134-020-05991-x
5. Ziegler C G K, S J Allon, S K Nyquist, I M Mbanjo, V N Miao, C N Tzouanas, et al. SARS-CoV-2 Receptor ACE2 Is an Interferon-Stimulated Gene in Human Airway Epithelial Cells and Is Detected in Specific Cell Subsets across Tissues. *Cell*(2020), 181(5), 1016-1035.e19 doi:10.1016/j.cell.2020.04.035
6. Xiong Y, Y Liu, L Cao, D Wang, M Guo, A Jiang, et al. Transcriptomic characteristics of bronchoalveolar lavage fluid and peripheral blood mononuclear cells in COVID-19 patients. *Emerg Microbes Infect*(2020), 9(1), 761-770 doi:10.1080/22221751.2020.1747363
7. Chen L, J Xiong, L Bao and Y Shi. Convalescent plasma as a potential therapy for COVID-19. *Lancet Infect Dis*(2020), 20(4), 398-400 doi:10.1016/s1473-3099(20)30141-9
8. Fiolet T, Y Kherabi, C J MacDonald, J Ghosn and N Peiffer-Smadja. Comparing COVID-19 vaccines for their characteristics, efficacy and effectiveness against SARS-CoV-2 and variants of concern: a narrative review. *Clin Microbiol Infect*(2022), 28(2), 202-221 doi:10.1016/j.cmi.2021.10.005
9. Casagrande M, A Fitzek, M Spitzer, K Püschel, M Glatzel, S Krasemann, et al. Detection of SARS-CoV-2 genomic and subgenomic RNA in retina and optic nerve of patients with COVID-19. *Br J Ophthalmol*(2021), doi:10.1136/bjophthalmol-2020-318618
10. Yang G, X Lu and L Yuan. LncRNA: a link between RNA and cancer. *Biochim Biophys Acta*(2014), 1839(11), 1097-109 doi:10.1016/j.bbagr.2014.08.012
11. Liu H, Z Zhang, N Wu, H Guo, H Zhang, D Fan, et al. Integrative Analysis of Dysregulated lncRNA-Associated ceRNA Network Reveals Functional lncRNAs in Gastric Cancer. *Genes (Basel)*(2018), 9(6) doi:10.3390/genes9060303
12. Atianand M K, D R Caffrey and K A Fitzgerald. Immunobiology of Long Noncoding RNAs. *Annu Rev Immunol*(2017), 35, 177-198 doi:10.1146/annurev-immunol-041015-055459
13. Stojic L, A T L Lun, P Mascalchi, C Ernst, A M Redmond, J Mangei, et al. A high-content RNAi screen reveals multiple roles for long noncoding RNAs in cell division. *Nat Commun*(2020), 11(1),

- 1851 doi:10.1038/s41467-020-14978-7
14. Ma M, Y Pei, X Wang, J Feng, Y Zhang and M Q Gao. LncRNA XIST mediates bovine mammary epithelial cell inflammatory response via NF- κ B/NLRP3 inflammasome pathway. *Cell Prolif*(2019), 52(1), e12525 doi:10.1111/cpr.12525
 15. Chen H, X Wang, X Yan, X Cheng, X He and W Zheng. LncRNA MALAT1 regulates sepsis-induced cardiac inflammation and dysfunction via interaction with miR-125b and p38 MAPK/NF κ B. *Int Immunopharmacol*(2018), 55, 69-76 doi:10.1016/j.intimp.2017.11.038
 16. Zhong Y, C Yu and W Qin. LncRNA SNHG14 promotes inflammatory response induced by cerebral ischemia/reperfusion injury through regulating miR-136-5p /ROCK1. *Cancer Gene Ther*(2019), 26(7-8), 234-247 doi:10.1038/s41417-018-0067-5
 17. Imam H, A S Bano, P Patel, P Holla and S Jameel. The lncRNA NRON modulates HIV-1 replication in a NFAT-dependent manner and is differentially regulated by early and late viral proteins. *Sci Rep*(2015), 5, 8639 doi:10.1038/srep08639
 18. Sur S, R Sasaki, P Devhare, R Steele, R Ray and R B Ray. Association between MicroRNA-373 and Long Noncoding RNA NORAD in Hepatitis C Virus-Infected Hepatocytes Impairs Wee1 Expression for Growth Promotion. *J Virol*(2018), 92(20) doi:10.1128/jvi.01215-18
 19. Natarelli L, L Parca, T Mazza, C Weber, F Virgili and D Fratantonio. MicroRNAs and Long Non-Coding RNAs as Potential Candidates to Target Specific Motifs of SARS-CoV-2. *Noncoding RNA*(2021), 7(1) doi:10.3390/ncrna7010014
 20. Laha S, C Saha, S Dutta, M Basu, R Chatterjee, S Ghosh, et al. In silico analysis of altered expression of long non-coding RNA in SARS-CoV-2 infected cells and their possible regulation by STAT1, STAT3 and interferon regulatory factors. *Heliyon*(2021), 7(3), e06395 doi:10.1016/j.heliyon.2021.e06395
 21. Pan Q, Z Zhao, Y Liao, S H Chiu, S Wang, B Chen, et al. Identification of an Interferon-Stimulated Long Noncoding RNA (LncRNA ISR) Involved in Regulation of Influenza A Virus Replication. *Int J Mol Sci*(2019), 20(20) doi:10.3390/ijms20205118
 22. Maarouf M, B Chen, Y Chen, X Wang, K R Rai, Z Zhao, et al. Identification of lncRNA-155 encoded by MIR155HG as a novel regulator of innate immunity against influenza A virus infection. *Cell Microbiol*(2019), 21(8), e13036 doi:10.1111/cmi.13036
 23. Zhang Q, C Y Chen, V S Yedavalli and K T Jeang. NEAT1 long noncoding RNA and paraspeckle bodies modulate HIV-1 posttranscriptional expression. *mBio*(2013), 4(1), e00596-12 doi:10.1128/mBio.00596-12
 24. Ma H, P Han, W Ye, H Chen, X Zheng, L Cheng, et al. The Long Noncoding RNA NEAT1 Exerts Antihantaviral Effects by Acting as Positive Feedback for RIG-I Signaling. *J Virol*(2017), 91(9) doi:10.1128/jvi.02250-16
 25. Meydan C, N Madrer and H Soreq. The Neat Dance of COVID-19: NEAT1, DANCR, and Co-Modulated Cholinergic RNAs Link to Inflammation. *Front Immunol*(2020), 11, 590870 doi:10.3389/fimmu.2020.590870
 26. Peng X, L Gralinski, C D Armour, M T Ferris, M J Thomas, S Proll, et al. Unique signatures of long noncoding RNA expression in response to virus infection and altered innate immune signaling. *mBio*(2010), 1(5) doi:10.1128/mBio.00206-10
 27. Moazzam-Jazi M, H Lanjanian, S Maleknia, M Hedayati and M S Daneshpour. Interplay between SARS-CoV-2 and human long non-coding RNAs. *J Cell Mol Med*(2021), 25(12), 5823-5827 doi:10.1111/jcmm.16596

28. Vishnubalaji R, H Shaath and N M Alajez. Protein Coding and Long Noncoding RNA (lncRNA) Transcriptional Landscape in SARS-CoV-2 Infected Bronchial Epithelial Cells Highlight a Role for Interferon and Inflammatory Response. *Genes (Basel)*(2020), 11(7) doi:10.3390/genes11070760
29. Wu Y, T Zhao, R Deng, X Xia, B Li and X Wang. A study of differential circRNA and lncRNA expressions in COVID-19-infected peripheral blood. *Sci Rep*(2021), 11(1), 7991 doi:10.1038/s41598-021-86134-0
30. Lei F, H Zhang and X Xie. Comprehensive analysis of an lncRNA-miRNA-mRNA competing endogenous RNA network in pulpitis. *PeerJ*(2019), 7, e7135 doi:10.7717/peerj.7135
31. Ding H, J Liu, R Zou, P Cheng and Y Su. Long non-coding RNA TPTEP1 inhibits hepatocellular carcinoma progression by suppressing STAT3 phosphorylation. *J Exp Clin Cancer Res*(2019), 38(1), 189 doi:10.1186/s13046-019-1193-0
32. Cheng J, X Zhou, W Feng, M Jia, X Zhang, T An, et al. Risk stratification by long non-coding RNAs profiling in COVID-19 patients. *J Cell Mol Med*(2021), 25(10), 4753-4764 doi:10.1111/jemm.16444
33. Nishitsuji H, S Ujino, S Yoshio, M Sugiyama, M Mizokami, T Kanto, et al. Long noncoding RNA #32 contributes to antiviral responses by controlling interferon-stimulated gene expression. *Proc Natl Acad Sci U S A*(2016), 113(37), 10388-93 doi:10.1073/pnas.1525022113
34. Kambara H, F Niazi, L Kostadinova, D K Moonka, C T Siegel, A B Post, et al. Negative regulation of the interferon response by an interferon-induced long non-coding RNA. *Nucleic Acids Res*(2014), 42(16), 10668-80 doi:10.1093/nar/gku713
35. Xie Q, S Chen, R Tian, X Huang, R Deng, B Xue, et al. Long Noncoding RNA ITPRIP-1 Positively Regulates the Innate Immune Response through Promotion of Oligomerization and Activation of MDA5. *J Virol*(2018), 92(17) doi:10.1128/jvi.00507-18
36. Carpenter S, D Aiello, M K Atianand, E P Ricci, P Gandhi, L L Hall, et al. A long noncoding RNA mediates both activation and repression of immune response genes. *Science*(2013), 341(6147), 789-92 doi:10.1126/science.1240925
37. Yang Q, F Lin, Y Wang, M Zeng and M Luo. Long Noncoding RNAs as Emerging Regulators of COVID-19. *Front Immunol*(2021), 12, 700184 doi:10.3389/fimmu.2021.700184
38. Tay M Z, C M Poh, L Rénia, P A MacAry and L F P Ng. The trinity of COVID-19: immunity, inflammation and intervention. *Nat Rev Immunol*(2020), 20(6), 363-374 doi:10.1038/s41577-020-0311-8
39. Perlman S and A A Dandekar. Immunopathogenesis of coronavirus infections: implications for SARS. *Nat Rev Immunol*(2005), 5(12), 917-27 doi:10.1038/nri1732
40. Channappanavar R, A R Fehr, R Vijay, M Mack, J Zhao, D K Meyerholz, et al. Dysregulated Type I Interferon and Inflammatory Monocyte-Macrophage Responses Cause Lethal Pneumonia in SARS-CoV-Infected Mice. *Cell Host Microbe*(2016), 19(2), 181-93 doi:10.1016/j.chom.2016.01.007
41. Rockx B, T Baas, G A Zornetzer, B Haagmans, T Sheahan, M Frieman, et al. Early upregulation of acute respiratory distress syndrome-associated cytokines promotes lethal disease in an aged-mouse model of severe acute respiratory syndrome coronavirus infection. *J Virol*(2009), 83(14), 7062-74 doi:10.1128/jvi.00127-09
42. Yao X H, T Y Li, Z C He, Y F Ping, H W Liu, S C Yu, et al. [A pathological report of three COVID-19 cases by minimal invasive autopsies]. *Zhonghua Bing Li Xue Za Zhi*(2020), 49(5), 411-417 doi:10.3760/cma.j.cn112151-20200312-00193
43. Zumla A, D S Hui, E I Azhar, Z A Memish and M Maeurer. Reducing mortality from 2019-nCoV:

- host-directed therapies should be an option. *Lancet*(2020), 395(10224), e35-e36 doi:10.1016/s0140-6736(20)30305-6
44. Zhang C, Z Wu, J W Li, H Zhao and G Q Wang. Cytokine release syndrome in severe COVID-19: interleukin-6 receptor antagonist tocilizumab may be the key to reduce mortality. *Int J Antimicrob Agents*(2020), 55(5), 105954 doi:10.1016/j.ijantimicag.2020.105954
 45. Costela-Ruiz V J, R Illescas-Montes, J M Puerta-Puerta, C Ruiz and L Melguizo-Rodríguez. SARS-CoV-2 infection: The role of cytokines in COVID-19 disease. *Cytokine Growth Factor Rev*(2020), 54, 62-75 doi:10.1016/j.cytogfr.2020.06.001
 46. Morenikeji O B, K Bernard, E Strutton, M Wallace and B N Thomas. Evolutionarily Conserved Long Non-coding RNA Regulates Gene Expression in Cytokine Storm During COVID-19. *Front Bioeng Biotechnol*(2020), 8, 582953 doi:10.3389/fbioe.2020.582953
 47. Fatemi R P, D Velmeshev and M A Faghihi. De-repressing LncRNA-Targeted Genes to Upregulate Gene Expression: Focus on Small Molecule Therapeutics. *Mol Ther Nucleic Acids*(2014), 3(11), e196 doi:10.1038/mtna.2014.45
 48. Roberts T C, R Langer and M J A Wood. Advances in oligonucleotide drug delivery. *Nat Rev Drug Discov*(2020), 19(10), 673-694 doi:10.1038/s41573-020-0075-7
 49. Mukherjee S, B Banerjee, D Karasik and M Frenkel-Morgenstern. mRNA-lncRNA Co-Expression Network Analysis Reveals the Role of lncRNAs in Immune Dysfunction during Severe SARS-CoV-2 Infection. *Viruses*(2021), 13(3) doi:10.3390/v13030402
 50. Nishimoto N, K Yoshizaki, K Maeda, T Kuritani, H Deguchi, B Sato, et al. Toxicity, pharmacokinetics, and dose-finding study of repetitive treatment with the humanized anti-interleukin 6 receptor antibody MRA in rheumatoid arthritis. Phase I/II clinical study. *J Rheumatol*(2003), 30(7), 1426-35
 51. Nishida S, K Hagihara, Y Shima, M Kawai, Y Kuwahara, J Arimitsu, et al. Rapid improvement of AA amyloidosis with humanised anti-interleukin 6 receptor antibody treatment. *Ann Rheum Dis*(2009), 68(7), 1235-6 doi:10.1136/ard.2008.099267
 52. Li H, H Shi, N Ma, P Zi, Q Liu and R Sun. BML-111 alleviates acute lung injury through regulating the expression of lncRNA MALAT1. *Arch Biochem Biophys*(2018), 649, 15-21 doi:10.1016/j.abb.2018.04.016
 53. Gong J, S Guo, H B Li, S Y Yuan, Y Shang and S L Yao. BML-111, a lipoxin receptor agonist, protects haemorrhagic shock-induced acute lung injury in rats. *Resuscitation*(2012), 83(7), 907-12 doi:10.1016/j.resuscitation.2011.12.035
 54. Aishwarya S, K Gunasekaran and A A Margret. Computational gene expression profiling in the exploration of biomarkers, non-coding functional RNAs and drug perturbagens for COVID-19. *J Biomol Struct Dyn*(2020), 1-16 doi:10.1080/07391102.2020.1850360
 55. Yousefi H, A Poursheikhani, Z Bahmanpour, M Vatanmakanian, M Taheri, L Mashouri, et al. SARS-CoV infection crosstalk with human host cell noncoding-RNA machinery: An in-silico approach. *Biomed Pharmacother*(2020), 130, 110548 doi:10.1016/j.biopha.2020.110548
 56. Obaid M, S M N Udden, P Deb, N Shihabeddin, M H Zaki and S S Mandal. LncRNA HOTAIR regulates lipopolysaccharide-induced cytokine expression and inflammatory response in macrophages. *Sci Rep*(2018), 8(1), 15670 doi:10.1038/s41598-018-33722-2
 57. Zhang P, L Cao, R Zhou, X Yang and M Wu. The lncRNA Neat1 promotes activation of inflammasomes in macrophages. *Nat Commun*(2019), 10(1), 1495 doi:10.1038/s41467-019-09482-6

FIGURE CAPTIONS:

Figure1. LncRNAs are related to viral infection, interferon and cytokine storm in COVID-19.

