



# Draft Genome Sequence of the Necrotrophic Plant-Pathogenic Bacterium *Pectobacterium carotovorum* subsp. *carotovorum* Strain LMG 2410

William M. Rooney,<sup>a,b</sup> Marta Wojnowska,<sup>a\*</sup> Daniel Walker<sup>a</sup>

<sup>a</sup>Institute of Infection, Immunity and Inflammation, College of Medical, Veterinary and Life Sciences, University of Glasgow, Glasgow, United Kingdom

<sup>b</sup>Plant Science Group, Institute of Molecular, Cell and Systems Biology, University of Glasgow, Glasgow, United Kingdom

**ABSTRACT** Here, we report the draft genome sequence of *Pectobacterium carotovorum* subsp. *carotovorum* strain LMG 2410, isolated from cucumber in the United Kingdom. The draft genome is 4,773,000 bp, with a G+C content of 51.9%, and carries a total of 4,536 coding sequences.

*Pectobacterium carotovorum* subsp. *carotovorum* is a Gram-negative bacterial plant pathogen, possesses a broad host range, including many important crop species, and is a causative agent of soft rot (1). Soft rot disease symptoms arise from an arsenal of secreted cell wall-degrading enzymes, such as pectinases, cellulases, and xylanases, which allow the bacterium to necrotize its host (2, 3). One of the *P. carotovorum* subsp. *carotovorum* strains isolated from cucumber in the United Kingdom has been used to investigate the import of the abundant plant protein ferredoxin from which *P. carotovorum* can acquire iron for growth (4–6). Iron acquisition from ferredoxin is mediated by the ferredoxin uptake system (Fus), of which the outer membrane TonB-dependent receptor, FusA, and the periplasmic M16 protease FusC have been demonstrated to be essential for this process (4, 5). This study used this strain to generate a draft genome which can be a tool to further dissect this protein import pathway.

The genomic DNA of *P. carotovorum* subsp. *carotovorum* was extracted from a freshly grown single colony grown in lysogeny broth (5 g liter<sup>-1</sup> peptone, 5 g liter<sup>-1</sup> tryptone, 10 g liter<sup>-1</sup> NaCl [pH 7.5]) using the GenElute bacterial genomic DNA kit (Sigma-Aldrich, Dorset, UK). Library preparation was performed using the TruSeq DNA Nano kit and size selected for the large fragment size (Illumina, CA, USA). Sequencing was performed using the Illumina MiSeq 500 platform with a 2 × 300-bp paired-end protocol with 50× read depth. A total of 504,901 raw reads were trimmed (with a quality score limit of 0.05) and assembled *de novo* using CLC Genomics Workbench version 7.5.2 (CLC bio, Denmark) using the default settings, which generated 132 contigs with a total length of 4,783,145 bp (G+C content, 51.9%), an  $N_{50}$  value of 339,669 bp, and minimum and maximum contig lengths of 140 and 879,473 bp, respectively. Genome annotation of the assembled contigs was performed using the Rapid Annotations using Subsystems Technology (RAST) server, which identified 4,536 coding sequences, 70 tRNAs, and 28 rRNAs (7–9). From the RAST subsystem annotations, 41 genes were identified as being involved in iron acquisition and metabolism. Furthermore, from the keyword searches of the annotated genome, we identified 6 genes encoding TonB-like proteins, 5 sets of *exbBD* genes, and two genes encoding M16 proteases, including FusC (5).

In conclusion, we have reported the draft genome sequence of *P. carotovorum* subsp. *carotovorum*, a bacterium originally isolated from cucumber in the United Kingdom.

**Citation** Rooney WM, Wojnowska M, Walker D. 2019. Draft genome sequence of the necrotrophic plant-pathogenic bacterium *Pectobacterium carotovorum* subsp. *carotovorum* strain LMG 2410. Microbiol Resour Announc 8:e00614-19. <https://doi.org/10.1128/MRA.00614-19>.

**Editor** Kenneth M. Stedman, Portland State University

**Copyright** © 2019 Rooney et al. This is an open-access article distributed under the terms of the [Creative Commons Attribution 4.0 International license](https://creativecommons.org/licenses/by/4.0/).

Address correspondence to Daniel Walker, [Daniel.Walker@glasgow.ac.uk](mailto:Daniel.Walker@glasgow.ac.uk).

\* Present address: Marta Wojnowska, Biomedical Sciences Research Complex, The University of St. Andrews, St. Andrews, United Kingdom.

W.M.R. and M.W. contributed equally to this work.

**Received** 4 June 2019

**Accepted** 28 June 2019

**Published** 1 August 2019

**Data availability.** This draft genome project has been deposited at DDBJ/EMBL/GenBank under the accession number [VBUA00000000](#) (BioProject number [PRJNA543207](#) and BioSample number [SAMN11658211](#)). The version described in this paper is the first version, VBUA01000000.

## ACKNOWLEDGMENTS

The LMG 2410 strain was obtained from the Belgian Coordinated Collections of Microorganisms. Genome sequencing was performed by Glasgow Polyomics (<https://www.polyomics.gla.ac.uk/>).

This work was funded by the University of Glasgow MVLS DTP and the BBSRC (grant BB/L02022X/1). The funders had no role in the study design, data collection and interpretation, or the decision to submit this work for publication.

## REFERENCES

1. Mansfield J, Genin S, Magori S, Citovsky V, Sriariyanum M, Ronald P, Dow M, Verdier V, Beer SV, Machado MA, Toth I, Salmond G, Foster GD. 2012. Top 10 plant pathogenic bacteria in molecular plant pathology. *Mol Plant Pathol* 13:614–629. <https://doi.org/10.1111/j.1364-3703.2012.00804.x>.
2. Pérombelon M. 2002. Potato diseases caused by soft rot erwinias: an overview of pathogenesis. *Plant Pathol* 51:1–12.
3. Toth IK, Bell KS, Holeva MC, Birch P. 2003. Soft rot erwinias: from genes to genomes. *Mol Plant Pathol* 4:17. <https://doi.org/10.1046/j.1364-3703.2003.00149.x>.
4. Grinter R, Josts I, Mosbahi K, Roszak AW, Cogdell RJ, Bonvin A, Milner JJ, Kelly SM, Byron O, Smith BO, Walker D. 2016. Structure of the bacterial plant-ferredoxin receptor FusA. *Nat Commun* 7:13308. <https://doi.org/10.1038/ncomms13308>.
5. Mosbahi K, Wojnowska M, Albalat A, Walker D. 2018. Bacterial iron acquisition mediated by outer membrane translocation and cleavage of a host protein. *Proc Natl Acad Sci U S A* 115:6840–6845. <https://doi.org/10.1073/pnas.1800672115>.
6. Grinter R, Milner J, Walker D. 2012. Ferredoxin containing bacteriocins suggest a novel mechanism of iron uptake in *Pectobacterium* spp. *PLoS One* 7:e33033. <https://doi.org/10.1371/journal.pone.0033033>.
7. Aziz RK, Bartels D, Best A, DeJongh M, Disz T, Edwards RA, Formsma K, Gerdes S, Glass EM, Kubal M, Meyer F, Olsen GJ, Olson R, Osterman AL, Overbeek RA, McNeil LK, Paarmann D, Paczian T, Parrello B, Pusch GD, Reich C, Stevens R, Vassieva O, Vonstein V, Wilke A, Zagnitko O. 2008. The RAST server: Rapid Annotations using Subsystems Technology. *BMC Genomics* 9:75. <https://doi.org/10.1186/1471-2164-9-75>.
8. Overbeek R, Olson R, Pusch GD, Olsen GJ, Davis JJ, Disz T, Edwards RA, Gerdes S, Parrello B, Shukla M, Vonstein V, Wattam AR, Xia F, Stevens R. 2014. The SEED and the Rapid Annotation of microbial genomes using Subsystems Technology (RAST). *Nucleic Acids Res* 42:D206–D214. <https://doi.org/10.1093/nar/gkt1226>.
9. Brettin T, Davis JJ, Disz T, Edwards RA, Gerdes S, Olsen GJ, Olson R, Overbeek R, Parrello B, Pusch GD, Shukla M, Thomason JA, Stevens R, Vonstein V, Wattam AR, Xia F. 2015. RASTtk: a modular and extensible implementation of the RAST algorithm for building custom annotation pipelines and annotating batches of genomes. *Sci Rep* 5:8365. <https://doi.org/10.1038/srep08365>.