

Substituent Effects

C-Functionalized Cationic Diazaoxatriangulenes: Late-Stage Synthesis and Tuning of Physicochemical Properties

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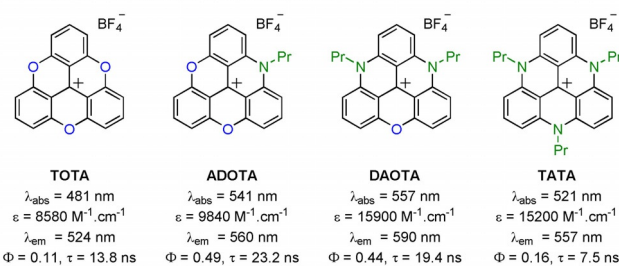
Abstract: A series of nine C-functionalized cationic diazaoxatriangulene (DAOTA) dyes have been successfully synthesized and fully characterized, including X-ray structural analysis of four derivatives. The introduction of electron-withdrawing or -donating functions enables the tuning of both electro- and photochemical properties with, for instance,

two consecutive (reversible) reductions or oxidations observed for nitro or amino derivatives, respectively. The substituents also impacted on the optical properties, with absorption maxima varying from $\lambda = 528$ to 640 nm and fluorescence being shifted from the yellow to the red range, up to $\lambda = 656$ nm.

Introduction

Cationic heterocyclic triangulenes are well-known chromophores studied in a variety of fields from biology,^[1] supramolecular chemistry,^[2] electrogenerated chemiluminescence,^[3] photosensitization,^[4] to materials science.^[5] These derivatives (Figure 1), which differ by the number of bridging oxygen and nitrogen atoms, are commonly named TOTA, ADOTA, DAOTA, and TATA in view of their trioxa, azadioxa, diazoxa, and triaza nature, respectively.^[6] They present 1) exceptionally high chemical stabilities for carbenium ions (pK_{R}^+ up to 23.7),^[7] and 2) intense fluorescence in the visible range characterized by particularly long lifetimes.^[8] Syntheses are straightforward from simple triarylcarbenium precursors (one to three steps in total). In terms of fluorescence, DAOTA moieties are noticeable because they present the most redshifted emission ($\lambda = 590$ nm), and the highest brightness ($\epsilon\Phi \approx 7000 \text{ M}^{-1} \text{ cm}^{-1}$) of the series. These features are particularly relevant for applications in fluo-

Previously reported cationic oxa and aza triangulenes



This work

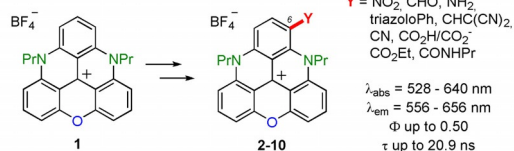


Figure 1. Top: Cationic triangulenes with corresponding optical properties in acetonitrile. Bottom: Scope of the study.

rescence microscopy in the transparency window of biological media.^[9]

So far, molecular engineering of DAOTA structures has focused mainly on the atoms or residues attached to the nitrogen atoms. Specific side chains can be introduced to provide efficient fluorophores for bioconjugation to bovine serum albumin (BSA),^[10] fluorescence lifetime imaging (FLIM) of G-quadruplex,^[11,11] or pH-controlled selective imaging of late endosomes.^[11] The C-functionalized triangulenes have only been scarcely reported and such derivatives can be made from pre-functionalized TOTA and ADOTA precursors.^[12] The N-functionalized triaryl-TATA derivatives react under oxidative chlorination conditions with moderate levels of chemo- and regioselectivity.^[5c] Similar behavior is observed with TOTA moieties.^[13] Recently, the iridium-catalyzed C–H borylation of TATA moieties has been achieved to give access, after cross-coupling chemistry, to C-functionalized triaryl derivatives. This sequence of re-

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actions is, to the best of our knowledge, the only example of regioselective late-stage functionalization of a triangulene skeleton.^[14]

Herein, inspired by studies on the postfunctionalization of related cationic [6]- and [4]helicenes,^[15] we report on the regioselective nitration and formylation of DAOTA **1** that has opened up unprecedented access to various triangulenes functionalized at position 6 (compounds **2–10**). The C-substituents influence the solid-state conformations and packing of the structures. The added functional groups also impact strongly on the physicochemical properties of the DAOTA core in solution. This late-stage functionalization strategy thus allows, for the first time, in the DAOTA series, fine-tuning of both electrochemical and optical properties of this important class of dyes and luminophores.

Results and Discussion

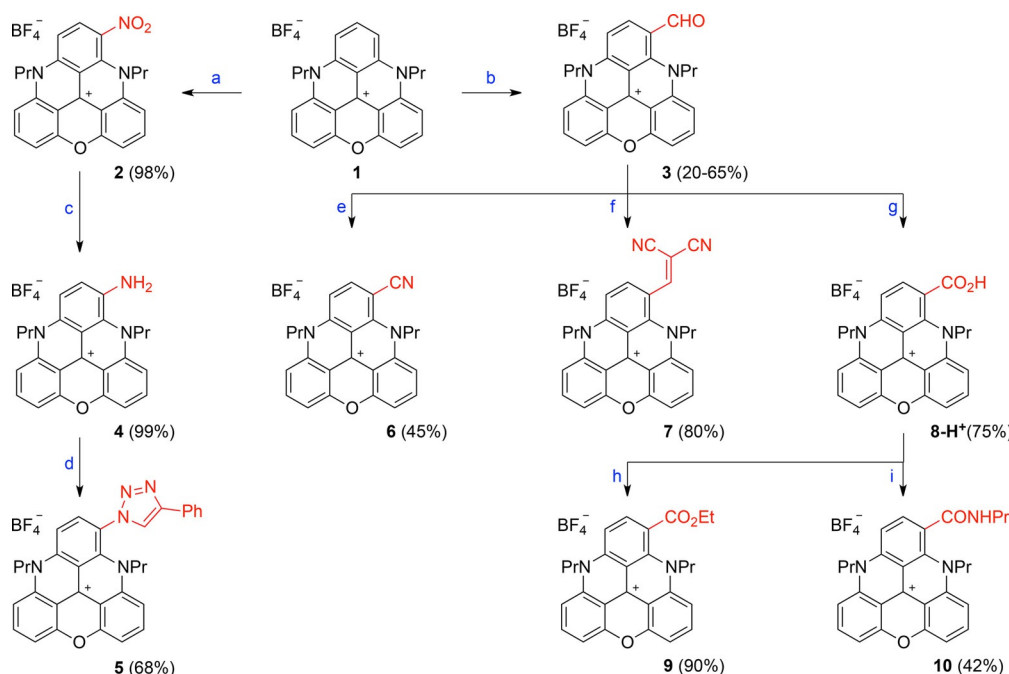
Synthesis

Initial attempts at functionalizing **1** were performed with halogenating reagents, such as *N*-chloro- or *N*-bromosuccinimide. However, as observed for TOTA,^[13] TATA,^[5c] and diaza[4]helicene analogues,^[15b] mixtures of mono- and polyhalogenated species are obtained. With substrate **1**, separation of the various products was impossible. To overcome this hurdle, it was decided to avoid polysubstitution reactions as a whole. The introduction of strong electron-withdrawing groups was considered, such that, after the first substitution, these substituents would

deactivate the triangulene core and stop further reactivity. Upon nitration of **1** under biphasic conditions (CH₂Cl₂/HNO₃ (13:2)) the orange-colored mononitro derivative **2** was obtained in excellent yield of 98% (Scheme 1). Experimentally, due to the low solubility of **1** in dichloromethane, dilution (0.05 M) and prolonged reaction times are required.^[16] Following the same strategy, compound **1** was treated under Vilsmeier–Haack conditions with large excesses of POCl₃ and DMF at 90 °C for 14 h to afford formylated derivative **3**. Recrystallization from a mixture of dichloromethane/toluene was necessary to isolate the product as a pink solid in yields of up to 65%.^[17]

With nitro and aldehyde derivatives **2** and **3**, respectively, in hand, we performed further derivatizations. For instance, compound **2** was converted efficiently into (light green) amino derivative **4** by hydrogenation over Pd/C (99% yield). To prepare **5**, a two-step procedure was necessary. First, amino **4** was transformed into an azido derivative by treatment with *tert*-butyl nitrite and azidotrimethylsilane in acetonitrile for 1 h. This compound was unstable and quickly engaged in a copper-catalyzed azide–alkyne cycloaddition. Treatment with phenylacetylene, CuSO₄, NaHSO₄, and ascorbic acid afforded **5** in 68% overall yield.

Formylated **3** was derivatized into cyano **6** under Schmidt conditions, with NaN₃ and TfOH in acetonitrile (45% yield).^[18,19] A Knoevenagel condensation was achieved by treatment of **3** with malonitrile (3 equiv) under PPh₃ catalysis (20 mol%) in acetonitrile (130 °C, MW irradiation, 1 h) to afford **7** in 80% yield. Compound **8-H⁺** was obtained under Pinnick–Kraus conditions, by treating **3** with NaH₂PO₄ and NaClO₂, in the pres-



Scheme 1. Synthesis of functionalized DAOTAs. Reagents and conditions: a) HNO₃ (60% aqueous solution), CH₂Cl₂, 25 °C, 12 h. b) POCl₃ (24 equiv), DMF (12 equiv), 90 °C, 14 h. c) H₂, Pd/C (20 mol%), CH₂Cl₂/MeOH, 25 °C, 30 min. d) *t*BuONO (1.5 equiv), trimethylsilyl azide (TMSN₃; 2 equiv), CH₃CN, 0 to 25 °C, 1 h, then PhC≡CH, CuSO₄·5 H₂O (0.1 equiv), ascorbic acid (0.2 equiv), NaHCO₃ (0.2 equiv), CH₃CN/H₂O, 25 °C, 30 min. e) NaN₃ (1.5 equiv), trifluoromethanesulfonic acid (TfOH; 3 equiv), CH₃CN, 25 °C, 5 min. f) NCCH₂CN (3 equiv), Ph₃P (20 mol%), CH₃CN, 130 °C (microwave (MW)), 25 °C, 1 h. g) NaH₂PO₄ (1 equiv), NaClO₂ (2 equiv), H₂O₂, CH₃CN, 60 °C, 1 h, then HBF₄ (1 M aqueous solution). h) SOCl₂ (6 equiv), CH₂Cl₂, 25 °C, 10 min, then EtOH (6 equiv), 25 °C, 10 min. i) SOCl₂ (6 equiv), CH₂Cl₂, 25 °C, 10 min, then PrNH₂ (18 equiv), 0 °C then 25 °C, 15 min.

ence of H₂O₂ as a scavenger. After acidic workup, the product was isolated as the carboxylic acid in 75% yield. Finally, compound **8-H**⁺ was converted into ester **9** and amide **10** by using a stepwise protocol. In both cases, the acid was converted into the corresponding acyl chloride derivative by treatment of **8-H**⁺ with SOCl₂ in dichloromethane. Upon reactions with ethanol (excess) or *n*-propylamine, products **9** and **10** were obtained in 90 and 42% yields, respectively.

Overall, in very few steps, a rather large chemical diversity of cationic DAOTA derivatives could be afforded by this late-stage functionalization strategy. The observed regioselectivity, in favor of diaza- rather than azaoxa-substituted phenyl rings, is in agreement with that previously observed on related helices.^[15a,b,20]

Solid-state analysis

Crystals of DAOTA **1**, nitro **2**, aldehyde **3**, and amino **4** derivatives were obtained by slow diffusion of hexane into solutions of the corresponding compounds in dichloromethane. The substituents influenced the stacking of the triangulene cores and the conformations of the N-alkyl side chains in the solid state. The X-ray structure of **1** is presented in Figure S1 in the Supporting Information and relevant values for all compounds are summarized in Table 1; complementary data can be found in the Supporting Information.

Table 1. Selected solid-state parameters for 1–4.

| Molecule ^[a] | Y | Distance ^[a] [Å] | RMSD ^[b] [Å] |
|-------------------------|-----------------|-----------------------------|-------------------------|
| 1 | H | 3.5 | 0.024 |
| 2 | NO ₂ | 3.4 | 0.123 |
| 3 | CHO | 3.4 | 0.041 |
| 4 | NH ₂ | 3.7 | 0.131 |

[a] Interplanar distance of stacking. [b] Root-mean-square deviation (RMSD) of the atomic positions from the calculated least-squares plane.

As already reported for TATA and ADOTA, DAOTA **1** is a quasi-planar system,^[7b] with an average RMSD value of 0.024 Å. Both side chains are disposed in the *syn* conformation, and extensive π – π overlap means that DAOTA stacks in dimers that are slightly shifted, with close interplanar distances of 3.5 Å, in a head to tail disposition (Figure 2a). The dimers stack in layers running along the *a* axis, with a distance of 4.7 Å between the dimers. Hydrogen bonds are observed between the BF₄[–] counterions and side chains (Figure 2b; tables of the hydrogen bonds are reported in the Supporting Information). A similar situation is observed for aldehyde derivative **3**, which also stacks in dimers in a head to tail disposition with an equal in-

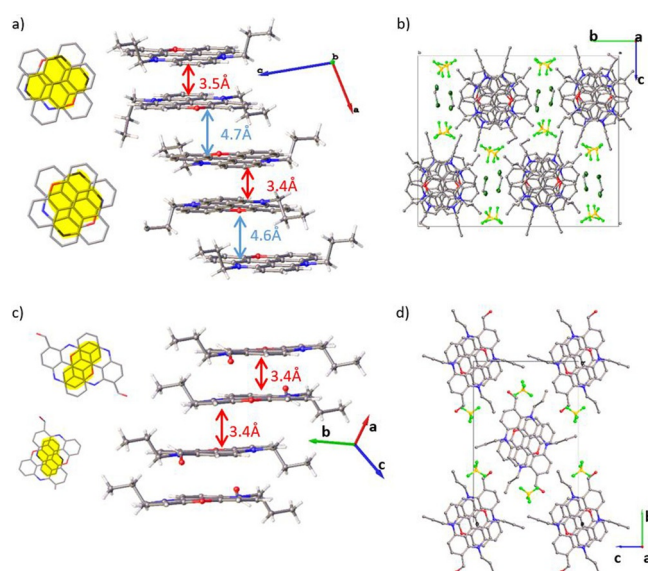


Figure 2. a) Extensive π – π stacking of **1**, and b) view along the *a* axis, showing the column of triangulene molecules formed. c) The π – π stacking of **3** and d) view along the *a* axis, showing the triangulene molecules. Overlap between the π systems of close neighboring molecules is shown in yellow.

terplanar distance of 3.4 Å (Figure 2c). In this case, the propyl side chains are disposed in an *anti* conformation, layers form along the *a* axis, and hydrogen-bonding interactions are observed with the BF₄[–] counterions (Figure 2d). The introduction of the formyl group provokes a slight increase of the RMSD value (0.041 Å).

Somewhat surprisingly, and in contrast to the diaza [4]helicene analogue of **3**,^[15b] the predominant conformation of the CHO group is *s-trans*, rather than *s-cis*. In the solid state, the formyl group and adjacent C–H of the same molecule interact, but with a C–H...O angle of 102°. More linear geometries are observed for intermolecular hydrogen bonds (Figure 3). In so-

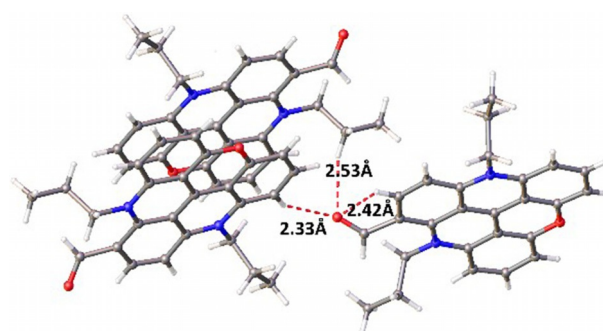


Figure 3. Inter- and intramolecular hydrogen bonds of aldehyde derivative **3**.

lution, both conformations occur, as shown by ¹H NOESY NMR spectroscopy experiments (CD₃CN; Figure 4). Through-space correlation between the H of the formyl group, H⁹, and H¹⁰ indicates the occurrence of the *s-trans* geometry, whereas close proximity with H⁷ and H⁸ demonstrates the *s-cis* conformation.

Nitro derivative **2** presents a greater RMSD value of 0.123 Å. The π – π interactions are still present in this derivative; howev-

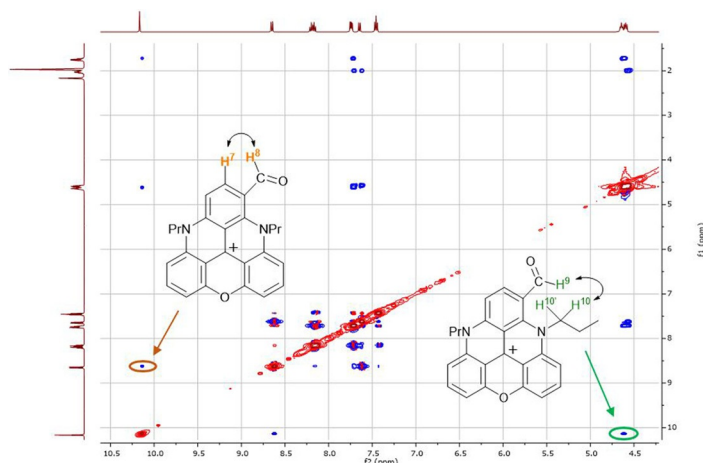


Figure 4. ^1H NOESY map of **3** (CD_3CN , 400 MHz, 298 K) showing the minor (left, *s-cis*) and major (right, *s-trans*) conformers.

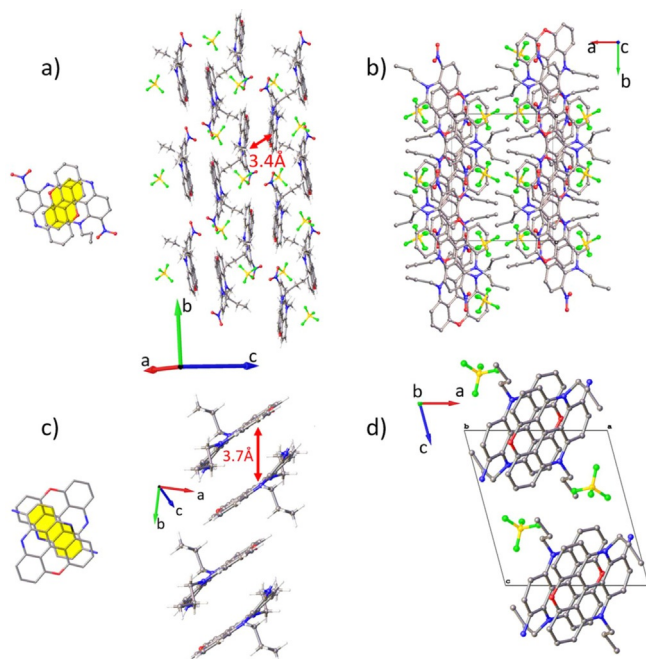


Figure 5. a) The π - π stacking of nitro **2** and b) view along the *c* axis of the columns. c) The π - π stacking of amino **4** and d) view along the *c* axis of the columns. Overlap between the π systems of close neighboring molecules is shown in yellow. Disorder was omitted for clarity.

er, layered arrangements are observed rather than columns, as characterized by a head to tail disposition with an interplane distance of 3.4 Å (Figure 5 a). These layers are held together by hydrogen-bonding interactions with the BF_4^- counterions (Figure 5). Amino derivative **4** also presents a greater RMSD value of 0.131 Å. The dimer is held in head to tail disposition by π - π interactions, forming columns, as observed for **1** and **3**. The distance is slightly increased to 3.7 Å (Figure 5 c). Again, hydrogen bonds are observed with the BF_4^- counterions (Figure 5 d).

Overall, in the four structures, π - π interactions generate dimers in a head to tail disposition. They induce the formation of columns in DAOTA **1**, aldehyde **3**, and amino **4**, or layers in the case of nitro derivative **2**. The BF_4^- counterions act as hydrogen-bond donors, generating interactions between and within the columns (layers for dye **2**).

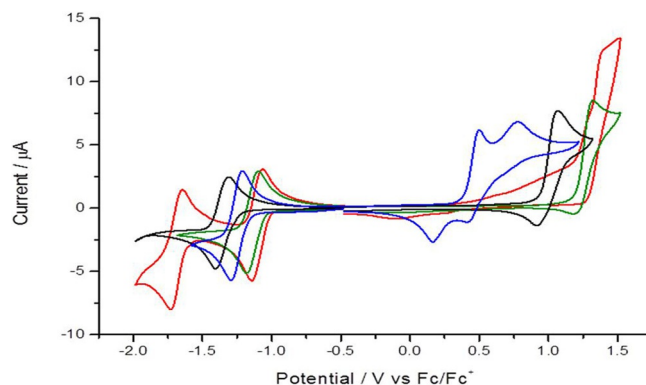


Figure 6. Cyclic voltammograms measured in solutions containing 10^{-3} M of **1** (black), **2** (red), **3** (green), and **4** (blue) in acetonitrile ($[\text{TBA}][\text{PF}_6]$; 10^{-1} M). Data are recorded at the Pt working electrode ($\nu = 0.1 \text{ V s}^{-1}$).

Electrochemical properties

The electrochemical behavior of **1–4** was analyzed by means of cyclic voltammetry (CV). The CV curves presented in Figure 6 were recorded in degassed solutions containing 10^{-3} M of each of the DAOTAs in acetonitrile and 0.1 M tetrabutylammonium hexafluorophosphate, $[\text{TBA}][\text{PF}_6]$, as the supporting electrolyte (electrochemical data are gathered in Table 2). It is noteworthy that repeated cleaning and polishing of the elec-

Table 2. Oxidation and reduction half-wave potential values [mV] measured by CV for **1–4** (10^{-3} M) in acetonitrile ($[\text{TBA}][\text{PF}_6]$; 10^{-1} M) at a Pt electrode ($\varnothing = 1.5$ mm, $\nu = 0.1 \text{ V s}^{-1}$), E , versus the ferrocene/ferrocenium redox couple (Fc/Fc^+). Red_{*n*} and Ox_{*n*} represent the *n* successive reduction and oxidation processes observed in the tested electrochemical window.

| Dye | Y | Reduction | | | Oxidation | | |
|----------|---------------|-----------|-------------------------------|-------|---------------------|------------------------------|--------------------|
| | | Red1 | $\Delta E(\text{Red1})^{[a]}$ | Red2 | Ox1 | $\Delta E(\text{Ox1})^{[a]}$ | Ox2 |
| 1 | H | -1359 | 0 | - | 992 ^[b] | 0 | - |
| 2 | NO_2 | -1109 | -250 | -1690 | 1383 ^[c] | 391 | - |
| 3 | CHO | -1140 | -219 | - | 1258 ^[b] | 266 | - |
| 4 | NH_2 | -1250 | -109 | - | 333 ^[b] | -659 | 600 ^[c] |

[a] Potential difference relative to the first reduction or oxidation waves of reference compound **1**. [b] Not a fully reversible process. [c] Irreversible process.

trode surface is mandatory to record accurate and reproducible cyclic voltammograms because these compounds have a strong tendency to adsorb at the surface of the working electrode.

Unfunctionalized DAOTA **1**, used as a reference, exhibits relatively simple redox behavior. This compound is typically reduced at $E_{1/2}^{\text{red}} = -1.36$ V versus Fc/Fc^+ , with a peak to peak separation of 89 mV (ΔE_{peaks}). This provides evidence of a quasi-reversible mono-electronic transfer that generates a neutral species from the cation. Oxidation shows a slow, but still reversible behavior at the tested scan rate, with $E_{1/2}^{\text{ox}} = 0.99$ V versus Fc/Fc^+ . The difference in intensity between both peaks indicates a deviation from an ideal fully reversible system, which is attributed to a lower stability of the oxidized form. Evaluation of such an oxidative decomposition of the triangulene core was not investigated further, but already observed with several analogues.^[3a,11]

With nitro derivative **2**, a reversible one-electron reduction is also observed first, albeit proving to be an easier reduction process ($E_{1/2}^{\text{red}} = -1.11$ V vs. Fc/Fc^+ and $\Delta E_{\text{peaks}} = 88$ mV). A second reduction wave appeared at more cathodic potential ($E_{1/2}^{\text{red}} = -1.69$ V vs. Fc/Fc^+ and $\Delta E_{\text{peaks}} = 88$ mV). This second wave can be assigned to the mono-electronic reduction of the nitro group of **2**, as already observed for nitro derivatives of [4]helicenes.^[15b] The reduction of nitro compounds to form a stable radical species has been widely studied in nonaqueous media.^[21] On the other hand, a fully irreversible oxidation process is observed, which occurs at a large anodic overpotential ($E_{1/2}^{\text{ox}} = 1.38$ V vs. Fc/Fc^+). With aldehyde derivative **3**, the reductive behavior is apparently unchanged, in comparison to **1**, revealing a single one-electron reversible reduction, which occurs at a more cathodic potential ($E_{1/2}^{\text{red}} = -1.14$ V vs. Fc/Fc^+ and $\Delta E_{\text{peaks}} = 87$ mV). A partially irreversible oxidation process is observed, with a very positive $E_{1/2}^{\text{ox}}$ value (1.26 V vs. Fc/Fc^+). Finally, the electrochemical characterization of amino derivative **4** reveals a slightly easier reversible reduction process than that of **1** ($E_{1/2}^{\text{red}} = -1.25$ V vs. Fc/Fc^+ and $\Delta E_{\text{peaks}} = 82$ mV), and two partially irreversible oxidations at very mild $E_{1/2}^{\text{ox}}$

values (0.33 and 0.60 V). Such an easier reduction is somehow unexpected in comparison with **1** and the corresponding amino derivative **4**.

The introduction of functional groups onto the cationic triangulene core does affect the reversibility of the oxidation pathways because the various products of oxidation do not exhibit the same stability. By comparing the CV data gathered in Figure 6, nitro derivative **2** is the only one that exhibits a fully irreversible oxidation at a scan rate of 0.1 V s^{-1} . On the other hand, the oxidation processes of electron-deficient nitro **2** and aldehyde **3** are above that of DAOTA **1**, with cathodic shifts of 0.39 and 0.27 V, respectively. Amino derivative **4** shows more complex oxidation behavior, with two anodic waves at less positive potentials. This may be attributed to the oxidation of the amino group,^[22] as already observed for amino derivatives of [4]helicenes.^[15b] In terms of the reduction process, most of the compounds feature a single, fully reversible, electron transfer, as seen for aldehyde **3** and amino **4**, and as observed for reference compound DAOTA **1**. Nevertheless, nitro derivative **2** displays two reversible mono-electronic reductions; the second of which is assigned to the NO_2 group.

Photophysical properties

Electronic absorption and fluorescence properties of the functionalized DAOTA derivatives were recorded in acetonitrile (ca. 10^{-5} M) at 20°C . Selected spectra are presented in Figure 7 and a summary of the data is provided in Table 3. The effect of the

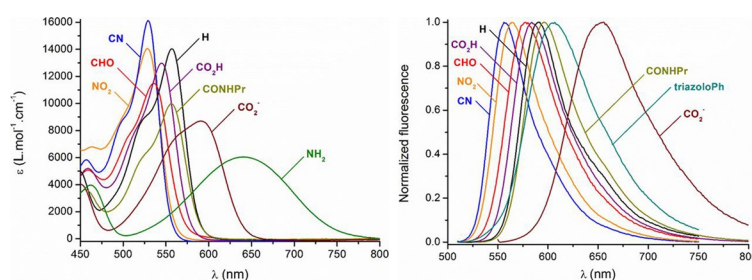


Figure 7. Selected electronic absorption (left) and normalized fluorescence (right) spectra in acetonitrile.

| Table 3. Photophysical properties of 1–10 in acetonitrile. | | | | | | | | | |
|--|---------------------------|-----------------------------|---|----------------------------|-----------------------------------|-----------------------|--------------------|---|--|
| Dye | Y | λ_{abs} [nm] | ϵ [$\text{M}^{-1} \text{cm}^{-1}$] | λ_{em} [nm] | Stokes shift [cm^{-1}] | ϕ ^[a] | τ [ns] | k_{R} (10^6 s^{-1}) ^[b] | k_{NR} (10^6 s^{-1}) ^[b] |
| 1 | H | 557 | 14 000 | 591 | 1030 | 0.42 | 19.6 | 21.4 | 29.6 |
| 2 | NO_2 | 528 | 14 100 | 564 | 1170 | 0.09 | 3.7 | 24.3 | 246 |
| 3 | CHO | 535 | 11 500 | 578 | 1390 | 0.26 | 11.7 | 22.2 | 63.3 |
| 4 | NH_2 | 640 | 6000 | – | – | – | – | – | – |
| 5 | triazolo-Ph | 543 | 11 400 | 607 | 1940 | 0.29 | 18.3 | 15.9 | 38.8 |
| 6 | CN | 529 | 16 100 | 556 | 920 | 0.26 | 16.2 | 16.1 | 45.7 |
| 7 | $\text{CHC}(\text{CN})_2$ | 546 | 11 400 | 590 | 1370 | 0.22 | 13.0 | 16.9 | 60.0 |
| 8-H⁺[c] | CO_2H | 545 | 13 000 | 584 | 1230 | 0.46 | 18.4 | 25.0 | 29.4 |
| 8^[d] | CO_2^- | 591 | 8700 | 656 | 1680 | 0.04 ^[e] | 2.7 ^[f] | 14.8 | 356 |
| 9 | CO_2Et | 547 | 13 200 | 584 | 1160 | 0.49 | 19.2 | 25.5 | 26.6 |
| 10 | CONHPr | 557 | 10 000 | 596 | 1180 | 0.50 | 20.9 | 23.9 | 23.3 |

[a] Reference: rhodamine B ($\phi = 0.49$ in ethanol). [b] With $k_{\text{R}} = \phi/\tau$ and $k_{\text{NR}} = (1 - \phi)/\tau$. [c] 0.1 M trifluoroacetic acid (TFA) in CH_3CN . [d] 0.1 M Et_3N in CH_3CN . [e] Reference: oxazine 725 ($\phi = 0.11$ in ethanol). [f] Biexponential fit: 2.7 (63), 5.5 ns (37%).

nine substituents follows a similar trend to that previously reported for functionalized diaza [4]helicenes.^[15b] Electron-withdrawing functional groups induce a moderate hypsochromic effect of the absorption maximum, up to $\lambda=528$ nm for derivative **2**, corresponding to a 990 cm^{-1} blueshift relative to that of unsubstituted **1**. The intensity of the lower energy transition is weakly influenced, with ϵ between 10000 and $16100\text{ M}^{-1}\text{ cm}^{-1}$. On the contrary, electron-donating substituents give rise to a noticeable low-energy shift of the band with maxima centered at $\lambda=591$ and 640 nm (1030 and 2330 cm^{-1} redshifts relative to **1**) for carboxylate and amino derivatives **8** and **4**, respectively. The broadening of the transition presumably indicates the establishment of intramolecular charge transfer from the substituent towards the cationic skeleton.^[15c]

In term of fluorescence, the compounds carrying electron-withdrawing moieties present fluorescence profiles quasi-similar to that of **1**, centered between $\lambda=556$ (**6**) and 596 nm (**10**), with Stokes shifts in the range of 900 – 1400 cm^{-1} . The fluorescence quantum yields and lifetimes are sensibly equivalent to that of **1**, with, for instance, $\Phi\approx 0.5$ for **9**–**10**, but are noticeably lowered for compounds **3**, **6**, and **7** ($\Phi\approx 0.3$). Surprisingly, nitro derivative **2** is characterized by a dramatic loss of fluorescence intensity and lifetime due to increased nonradiative de-excitation pathways. On the other hand, the dyes featuring triazolo-Ph (**5**) and carboxylate (**8**) presents redshifted emission bands centered at $\lambda=607$ and 656 nm, respectively, with higher Stokes shifts (ca. 1700 – 1900 cm^{-1}). Their diminished intensity and lifetimes, together with the absence of signal for amino derivative **4**, is consistent with previous observations on cationic diaza [4]- and [6]helicenes introducing strong donor groups.^[15b,d] Finally, it is noteworthy to underline that deprotonation of carboxylic derivative **8**-H⁺ to zwitterionic form **8** leads to 46 and 72 nm (1430 and 1880 cm^{-1}) redshifts of absorption and fluorescence maxima, respectively. Such a system may find particular interest as a pH-sensitive probe for biological applications.^[11]

Conclusion

A simple and efficient derivatization strategy has been developed to afford C-functionalized cationic DAOTA derivatives (9 examples). The electronic effects caused by the added substituents are visible by CV, which has revealed a tuning of the redox potentials with, in a general manner, easier reductions of the substituted dyes. The tuning was also effective for the absorption and fluorescence properties, with a cutoff emission reaching the near-infrared region for carboxylate derivative **8**. By analogy with related cationic diaza [4]helicene derivatives, the direct introduction of functional groups onto the DAOTA skeleton paves the way to future chemical engineering of these dyes for bioconjugation or vectorization in biological media.^[23]

Experimental Section

Reagents

Compound **1** was prepared according to a reported procedure.^[7b] Column chromatography purifications were performed by using Siliacflash P60 silica gel (40 – $63\text{ }\mu\text{m}$, $60\text{ }\text{\AA}$).

Analytical methods and instruments

Melting points were measured in open capillary tubes with a Buchi B-550 melting points apparatus and are uncorrected. Absorption spectra were recorded on a JASCO V-650 spectrophotometer at 20°C in analytical-grade solvents (ca. 10^{-5} M). IR spectra were recorded on a PerkinElmer 1650 FTIR spectrometer by using a diamond attenuated total reflectance (ATR) Golden Gate sampling attachment. NMR spectra were recorded on Bruker Advance II+ AMX-500 and AMX-400 spectrometers at room temperature. NMR chemical shifts are given in ppm (δ) relative to Me₄Si with solvent resonances as internal standards (CD₂Cl₂: $\delta=5.32$ ppm for ¹H and $\delta=53.8$ ppm for ¹³C; CD₃CN: $\delta=1.94$ ppm for ¹H, $\delta=118.3$ ppm for ¹³C, $\delta=-164.7$ ppm for ¹⁹F, considering an external calibration with C₆F₆ as a reference).^[24] Electrospray mass spectra were obtained on a Finnigan SSQ 7000 spectrometer QSTAR pulsar *i* (AB/MDS Sciex), ESI (TIS)/nanoESI/APCI-QqToF by the Department of Mass Spectroscopy of the University of Geneva.

Crystallography

All data were collected on an Agilent supernova dual source diffractometer equipped with an Atlas detector, by using Cu_{K α} radiation. Data reduction was carried out in the CrysAlis Pro software.^[25] Structure solution was made by using direct methods (sir2004),^[26] dual-space methods (SHELXT), or charge-flipping (OLEX2). Refinements were carried out in SHELXL^[27] within the OLEX2^[28] software. CCDC-1824707, 1443634, 1824709, and 1824706 contain the supplementary crystallographic data for this paper. These data can be obtained free of charge from The Cambridge Crystallographic Data Centre.

Electrochemistry

Voltammetric experiments were performed with a PGSTAT30 Autolab potentiostat connected to a conventional three-electrode cell, consisting of a silver wire pseudoreference electrode, a platinum wire auxiliary electrode, and a 1.5 mm diameter platinum disk working electrode. Prior to measurements, Pt disks were polished with alumina slurry of different sizes, rinsed thoroughly with Milli-Q water between each polishing step, and sonicated in water and ethanol successively, followed by a final rinse with acetonitrile, and dried with a stream of N₂. All solutions were degassed for 10 min before measurements.

Fluorescence spectroscopy

Steady-state fluorescence spectra were measured by using a Varian Cary 50 Eclipse spectrofluorimeter. All fluorescence spectra were corrected for the wavelength-dependent sensitivity of the detection. Fluorescence quantum yields, Φ , were measured in diluted solutions (at least five different concentrations for each sample) with an optical density lower than 0.1 by using the following equation: $\Phi_x/\Phi_r = [A_r(\lambda)/A_x(\lambda)](n_x^2/n_r^2)(D_x/D_r)$, in which A is the absorbance at the excitation wavelength (λ), n is the refractive index, and D the integrated intensity; r and x indicate reference and sample, respectively. The fluorescence quantum yields were measured rela-

tive to rhodamine B in ethanol ($\Phi=0.49$) for all compounds, except for **8**, which was measured relative to oxazine 725 in ethanol ($\Phi=0.11$). Excitation of reference and sample compounds was performed at the same wavelength ($\lambda=520$ nm, except for **8** and oxazine 725, for which excitation was performed at $\lambda=590$ nm). Short luminescence decay was monitored by using a Horiba-Jobin-Yvon Fluorolog-3 iHR320 fluorimeter equipped with the TC-SPC Horiba apparatus and by using Ludox in distilled water to determine the instrumental response function used for deconvolution. Excitation was performed by using a $\lambda=495$ nm nano-light-emitting diode (NanoLED; peak wavelength: $\lambda=490$ nm; pulse duration: <250 ps), and deconvolution was performed by using the DAS6 fluorescence-decay analysis software.

Compound 2

An aqueous solution of HNO₃ (2 mL, 60%, 19 mmol) was added to a solution of **1** (300 mg, 0.7 mmol) in CH₂Cl₂ (13 mL, 0.05 M). After 12 h, the reaction mixture was quenched by the addition of an aqueous solution of NaOH (1 M), then it was extracted with CH₂Cl₂ (3 × 20 mL) and washed with 1 M HBF₄. The organic layer was dried over Na₂SO₄, filtered, and concentrated under reduced pressure. The desired product was obtained as an orange solid after dissolution in the minimum amount of CH₂Cl₂ and precipitation with Et₂O (320 mg, 98%). *R*_f = 0.2 (SiO₂, CH₂Cl₂/MeOH, 98:2); m.p. 187 °C (decomp); UV/Vis (CH₃CN): λ_{\max} (ϵ_{\max}) = 528 nm (14 100 L mol⁻¹ cm⁻¹); IR (CH₂Cl₂): $\tilde{\nu}$ = 3080, 2974, 2880, 1612, 1588, 1514, 1459, 1313, 1273, 1166, 1042, 822, 753 cm⁻¹; ¹H NMR (400 MHz, CD₂Cl₂): δ = 8.79 (d, *J* = 9.6 Hz, 1 H; CH), 8.30 (t, *J* = 8.5 Hz, 1 H; CH), 8.20 (t, *J* = 8.5 Hz, 1 H; CH), 7.72 (d, *J* = 8.8 Hz, 1 H; CH), 7.64–7.47 (m, 4H; 4 × CH), 4.74–4.58 (m, 2H; CH₂), 4.25 (t, *J* = 7.0 Hz, 2H; CH₂), 2.16–2.00 (m, 2H; CH₂), 1.82–1.68 (m, 2H; CH₂), 1.26 (t, *J* = 7.4 Hz, 3H; CH₃), 0.64 ppm (t, *J* = 7.3 Hz, 3H; CH₃); ¹³C NMR (101 MHz, CD₂Cl₂): δ = 153.0 (C), 152.8 (C), 143.8 (C), 140.0 (CH), 140.0 (C), 139.9 (C), 139.2 (C), 138.9 (CH), 138.1 (C), 136.8 (CH), 131.8 (C), 112.6 (CH), 112.4 (C), 110.8 (CH), 110.8 (CH), 110.1 (CH), 109.4 (C), 108.9 (C), 106.8 (CH), 57.9 (CH₂), 50.8 (CH₂), 21.1 (CH₂), 19.7 (CH₂), 10.7 (CH₃), 10.3 ppm (CH₃); ¹⁹F NMR (282 MHz, CD₃CN): δ = -149.6 (20%), -149.7 ppm (80%); HRMS (ESI+): *m/z* calcd for C₂₅H₂₂N₃O₃⁺ [*M*]⁺: 412.1656; found: 412.1661.

Compound 3

POCl₃ (2.5 mL, 26.42 mmol) was added to a flask containing a solution of salt **1** (500 mg, 1.1 mmol) in DMF (1.0 mL, 13.21 mmol) at 90 °C. After being stirred for 14 h at this temperature, H₂O (10 mL) was added to the reaction mixture at 0 °C. The obtained solution was stirred for 30 min at 25 °C. Then, the reaction mixture was washed with a 5% aqueous solution of LiCl and extracted with CH₂Cl₂ (3 × 20 mL). The organic layer was washed with 1 M HBF₄, dried over Na₂SO₄, filtered, and concentrated under reduced pressure. The crude material was purified by flash chromatography on silica gel (CH₂Cl₂/MeOH 98:2) and recrystallization by slow diffusion in a CH₂Cl₂/toluene system. The product was obtained as a purple solid (20–65%).^[17] *R*_f = 0.3 (SiO₂, CH₂Cl₂/MeOH, 98:2); m.p. 217 °C (decomp); UV/Vis (CH₃CN): λ_{\max} (ϵ_{\max}) = 535 nm (11 500 L mol⁻¹ cm⁻¹); IR (CH₂Cl₂): $\tilde{\nu}$ = 3001, 1741, 1615, 1456, 1369, 1276, 1263, 1217, 837, 755 cm⁻¹; ¹H NMR (400 MHz, CD₂Cl₂): δ = 10.16 (s, 1H; CH), 8.67 (d, *J* = 9.1 Hz, 1H; CH), 8.26–8.15 (m, 2H; 2 × CH), 7.70–7.60 (m, 2H; 2 × CH), 7.57 (d, *J* = 9.1 Hz, 1H; CH), 7.55–7.45 (m, 2H; 2 × CH), 4.71–4.55 (m, 4H; CH₂), 2.17–1.98 (m, 2H; CH₂), 1.85–1.75 (m, 2H; CH₂), 1.26 (t, *J* = 7.4 Hz, 3H; CH₃), 0.67 ppm (t, *J* = 7.3 Hz, 3H; CH₃); ¹³C NMR (101 MHz, CD₂Cl₂): δ = 187.2 (CHO), 153.8 (C), 153.6 (C), 145.2 (C), 144.9 (CH), 144.5 (C), 141.2 (C), 140.9

(C), 140.3 (C), 140.0 (CH), 139.3 (CH), 118.8 (C), 113.6 (CH), 113.4 (C), 111.0 (CH), 110.8 (CH), 110.7 (CH), 110.4 (C), 109.8 (C), 107.9 (CH), 59.9 (CH₂), 51.0 (CH₂), 22.1 (CH₂), 20.1 (CH₂), 11.1 (CH₃), 10.5 ppm (CH₃); ¹⁹F NMR (282 MHz, CD₃CN): δ = -150.1 (20%), -150.2 ppm (80%); HRMS (ESI+): *m/z* calcd for C₂₃H₂₂N₂O₂⁺ [*M*]⁺: 395.1754; found: 395.1753.

Compound 4

Pd/C (12.8 mg, 10 wt%, 20 mol%) was added to solution of **2** (300 mg, 0.6 mmol) in CH₂Cl₂/MeOH (10:10 mL) under a N₂ atmosphere. Then H₂ was bubbled through the reaction mixture for 5 min and the solution was stirred for an additional 30 min at 25 °C under a H₂ atmosphere. Then the reaction mixture was purged with N₂ and filtered through Celite, which was then washed with CH₂Cl₂. The product was concentrated under reduced pressure. The desired product was obtained as a light green solid (280 mg, 99%). *R*_f = 0.2 (SiO₂, CH₂Cl₂/MeOH, 98:2); m.p. 257 °C (decomp); UV/Vis (CH₃CN): λ_{\max} (ϵ_{\max}) = 640 nm (6000 L mol⁻¹ cm⁻¹); IR (CH₂Cl₂): $\tilde{\nu}$ = 3007, 2990, 1611, 1462, 1276, 1262, 1163, 1068, 755 cm⁻¹; ¹H NMR (500 MHz, CD₃CN): δ = 7.96 (t, *J* = 8.2 Hz, 1H; CH), 7.90 (t, *J* = 8.4 Hz, 1H; CH), 7.79 (d, *J* = 9.0 Hz, 1H; CH), 7.49–7.37 (m, 3H; 3 × CH), 7.13 (d, *J* = 8.0 Hz, 1H; CH), 7.08 (d, *J* = 8.0 Hz, 1H; CH), 4.76–4.71 (m, 2H; CH₂), 4.46 (s, 2H; NH₂), 4.40–4.31 (m, 2H; CH₂), 1.93–1.84 (m, 2H; CH₂), 1.65–1.55 (m, 2H; CH₂), 1.16 (t, *J* = 7.3 Hz, 3H; CH₃), 0.67 ppm (t, *J* = 7.3 Hz, 3H; CH₃); ¹³C NMR (126 MHz, CD₃CN): δ = 153.4 (C), 153.3 (C), 144.7 (C), 140.9 (C), 140.3 (CH), 138.9 (CH), 138.8 (CH), 135.0 (C), 132.5 (C), 132.0 (CH), 129.4 (C), 115.4 (C), 112.0 (CH), 109.7 (CH), 109.1 (C), 109.0 (CH), 108.8 (CH), 108.6 (C), 108.2 (CH), 51.0 (CH₂), 50.1 (CH₂), 22.7 (CH₂), 20.1 (CH₂), 11.1 (CH₃), 11.0 ppm (CH₃); ¹⁹F NMR (282 MHz, CD₃CN): δ = -150.1 (20%), -150.2 ppm (80%); HRMS (ESI+): *m/z* calcd for C₂₅H₂₄N₃O⁺ [*M*]⁺: 382.1914; found: 382.1918.

Compound 5

*t*BuONO (8 μ L, 0.08 mmol) and TMSN₃ (13 μ L, 0.10 mmol) were added dropwise to a solution of **4** (24 mg, 0.05 mmol) in CH₃CN (60 μ L) at 0 °C and in the absence of light. The reaction mixture was stirred for 5 min at 25 °C. Then the addition of Et₂O led to the precipitation of azido intermediate, which was separated from the mother liquor by centrifugation. The solid was dissolved in MeOH (40 μ L) and phenylacetylene (5.5 μ L, 0.05 mmol) was added. A solution of CuSO₄·5H₂O (1.0 mg, 8 mol%), ascorbic acid (1.9 mg, 21 mol%), and NaHCO₃ (0.9 mg, 21 mol%) in water (40 μ L) was added dropwise. The reaction mixture was stirred for 30 min. The crude product was dissolved in a minimum amount of CH₂Cl₂ and then the addition of Et₂O led to the precipitation of the product, which was separated from the mother liquor by centrifugation. Then it was purified by flash chromatography on silica gel (CH₂Cl₂/MeOH 98:2) and recrystallized by slow diffusion in a biphasic system of CH₂Cl₂ and hexane. The desired product was obtained as a light pink solid (20.3 mg, 68% yield from **4**). *R*_f = 0.2 (SiO₂, CH₂Cl₂/MeOH, 98:2); m.p. 205 °C (decomp); UV/Vis (CH₃CN): λ_{\max} (ϵ_{\max}) = 543 nm (11 400 L mol⁻¹ cm⁻¹); IR (CH₂Cl₂): $\tilde{\nu}$ = 2988, 2922, 2852, 1616, 1465, 1276, 1262, 1059, 755 cm⁻¹; ¹H NMR (500 MHz, CD₃CN): δ = 8.62 (s, 1H; CH), 8.24–8.13 (m, 2H; 2 × CH), 8.09 (t, *J* = 8.4 Hz, 1H; CH), 8.03–7.98 (m, 2H; 2 × CH), 7.72 (d, *J* = 8.7 Hz, 1H; CH), 7.67 (d, *J* = 9.1 Hz, 1H; CH), 7.59–7.50 (m, 3H; 3 × CH), 7.48–7.42 (m, 2H; 2 × CH), 7.41 (d, *J* = 8.1 Hz, 1H; CH), 4.64–4.52 (m, 2H; CH₂), 3.79–3.65 (m, 2H; CH₂), 2.07–1.97 (m, 2H; CH₂), 1.60–1.47 (m, 3H; CH₂), 1.20 (t, *J* = 7.3 Hz, 3H; CH₃), 0.52 ppm (t, *J* = 7.3 Hz, 3H; CH₃); ¹³C NMR (126 MHz, CD₃CN): δ = 153.7 (C), 153.6 (C), 149.3 (C), 143.5 (C), 142.3 (C), 141.8 (C), 141.1 (CH), 140.5 (C), 140.1 (CH), 139.7 (CH),

138.4 (C), 131.1 (C), 130.2 (CH), 129.7 (CH), 126.7 (CH), 125.3 (CH), 117.7 (C), 114.4 (C), 112.1 (CH), 110.6 (CH), 110.4 (CH), 110.3 (CH), 109.7 (C), 109.6 (C), 108.4 (CH), 53.2 (CH₂), 51.1 (CH₂), 21.7 (CH₂), 20.1 (CH₂), 11.1 (CH₃), 10.7 ppm (CH₃); ¹⁹F NMR (282 MHz, CD₃CN): δ = -150.1 (20%), -150.2 ppm (80%); HRMS (ESI+): *m/z* calcd for C₃₃H₂₈N₅O⁺ [M]⁺: 510.2288; found: 510.2306.

Compound 6

TfOH (55 μL, 0.62 mmol) was added to a flask containing a solution of salt **3** (100 mg, 0.21 mmol) and NaN₃ (20.2 mg, 0.31 mmol) in CH₃CN (414 μL, 0.5 M). The obtained mixture was stirred for 5 min at 25 °C. Then the mixture was extracted with CH₂Cl₂ (3 × 10 mL) and the organic layer was washed with 1 M HBF₄, dried over Na₂SO₄, filtered, and concentrated under reduced pressure. The crude material was dissolved in the minimum amount of CH₂Cl₂ and precipitated with Et₂O. After flash chromatography on silica gel (CH₂Cl₂/MeOH 98:2), the product was obtained as a purple solid (44.7 mg, 45% yield). *R*_f = 0.2 (SiO₂, CH₂Cl₂/MeOH, 98:2); m.p. 183 °C (decomp); UV/Vis (CH₃CN): λ_{max} (ε_{max}) = 529 nm (16 100 L mol⁻¹ cm⁻¹); IR (CH₂Cl₂): ν̄ = 2930, 2210, 1739, 1615, 1530, 1460, 1327, 1266, 1154, 1030, 821, 753 cm⁻¹; ¹H NMR (500 MHz, CD₃CN): δ = 8.42 (d, *J* = 9.2 Hz, 1H; CH), 8.14–8.21 (m, 2H; 2 × CH), 7.71–7.66 (m, 2H; 2 × CH), 7.57 (d, *J* = 9.2 Hz, 1H; CH), 7.46–7.43 (m, 2H; 2 × CH), 5.3–4.6 (m, 2H; CH₂), 4.58–4.47 (m, 2H; CH₂), 2.17–2.11 (m, 2H; CH₂), 2.00–1.95 (m, 2H; CH₂), 1.25–1.15 ppm (m, 6H; 2 × CH₃); ¹³C NMR (126 MHz, CD₃CN): δ = 153.6 (C), 153.4 (C), 146.8 (CH), 145.3 (C), 143.5 (C), 141.2 (C), 140.8 (C), 140.2 (CH), 140.2 (CH), 140.1 (C), 119.9 (C), 118.3 (C), 112.8 (CH), 111.2 (CH), 111.1 (CH), 111.0 (CH), 111.0 (CH), 109.7 (C), 109.5 (C), 108.5 (CH), 53.1 (CH₂), 51.2 (CH₂), 22.2 (CH₂), 20.0 (CH₂), 11.0 (CH₃), 9.7 ppm (CH₃); ¹⁹F NMR (282 MHz, CD₃CN): δ = -150.3 (20%), -150.3 ppm (80%); HRMS (ESI+): *m/z* calcd for C₂₆H₂₂N₃O⁺ [M]⁺: 392.1757; found: 392.1759.

Compound 7

Ph₃P (11 mg, 20 mol%) was added to a flask containing a solution of salt **3** (100 mg, 0.21 mmol) and malononitrile (41.1 mg, 0.62 mmol) in CH₃CN (70 μL, 3 M). The obtained mixture was heated under MW irradiation at 130 °C. The crude material was purified by selective precipitation (Et₂O addition to a CH₂Cl₂ solution). The desired product was obtained as a pink solid (77 mg, 80% yield). *R*_f = 0.3 (SiO₂, CH₂Cl₂/MeOH, 98:2); m.p. 133 °C (decomp); UV/Vis (CH₃CN): λ_{max} (ε_{max}) = 546 nm (11 400 L mol⁻¹ cm⁻¹); IR (CH₂Cl₂): ν̄ = 3364, 2930, 2230, 1612, 1577, 1460, 1344, 1276, 1173, 1113, 930, 894, 781, 752 cm⁻¹; ¹H NMR (500 MHz, CD₃CN): δ = 8.57–8.45 (m, 2H; 2 × CH), 8.22–8.12 (m, 2H; 2 × CH), 7.76–7.70 (m, 2H; 2 × CH), 7.67 (d, *J* = 9.2 Hz, 1H; CH), 7.46–7.40 (m, 2H; 2 × CH), 4.67–7.53 (m, 4H; 2 × CH₂), 2.03–1.96 (m, 2H; CH₂), 1.86–1.77 (m, 2H; CH₂), 1.19 (t, *J* = 7.3 Hz, 3H; CH₃), 0.71 ppm (t, *J* = 7.3 Hz, 3H; CH₃); ¹³C NMR (126 MHz, CD₃CN): δ = 159.6 (CH), 153.8 (C), 153.6 (C), 145.1 (C), 144.7 (C), 141.5 (CH), 141.4 (C), 140.9 (C), 140.2 (C), 140.2 (CH), 139.6 (CH), 115.0 (C), 113.9 (C), 113.5 (CH), 113.4 (C), 112.7 (C), 111.1 (CH), 111.0 (CH), 110.8 (CH), 110.1 (C), 110.0 (C), 108.6 (CH), 83.7 (C), 58.6 (CH₂), 51.2 (CH₂), 22.9 (CH₂), 20.3 (CH₂), 11.1 (CH₃), 10.5 ppm (CH₃); ¹⁹F NMR (282 MHz, CD₃CN): δ = -150.2 (20%), -150.4 ppm (80%); HRMS (ESI+): *m/z* calcd for C₂₉H₂₃N₄O⁺ [M]⁺: 443.1866; found: 443.1860.

Compound 8-H⁺

Compound **3** (200 mg, 0.37 mmol) was dissolved in CH₃CN (8 mL, 0.05 M). NaH₂PO₄ (44 mg, 0.37 mmol), H₂O₂ (83 μL, 0.74 mmol), and NaClO₄ (90 mg, 0.74 mmol) were added separately to this solution.

The reaction was stirred for 1 h at 60 °C, then concentrated under reduced pressure. The residue was purified by column chromatography on silica gel with CH₂Cl₂/MeOH (90/10) as the eluent. The obtained solid was dissolved in CH₂Cl₂, washed with an aqueous solution of HBF₄ (1 M), dried over Na₂SO₄, filtered, and concentrated under reduced pressure, to afford the pure product as a pink solid (154 mg, 75%). *R*_f = 0.2 (SiO₂, CH₂Cl₂/MeOH, 98:2); m.p. 146 °C (decomp); UV/Vis (CH₃CN): λ_{max} (ε_{max}) = 545 nm (13 000 L mol⁻¹ cm⁻¹); IR (neat): ν̄ = 2927, 1708, 1616, 1460, 1347, 1276, 1261, 1223, 1169, 1112, 840, 755 cm⁻¹; ¹H NMR (400 MHz, CD₃CN): δ = 8.51 (d, *J* = 9.1 Hz, 1H; CH), 8.04–7.96 (m, 2H; 2 × CH), 7.57–7.51 (m, 2H; 2 × CH), 7.46 (d, *J* = 9.2 Hz, 1H; CH), 7.21–7.17 (m, 2H; 2 × CH), 4.47–4.40 (m, 4H; 2 × CH₂), 1.98–1.87 (m, 2H; CH₂), 1.73–1.60 (m, 2H; CH₂), 1.18 (t, *J* = 7.4 Hz, 3H; CH₃), 0.56 ppm (t, *J* = 7.3 Hz, 3H; CH₃); ¹³C NMR (101 MHz, CD₃CN): δ = 167.7 (C), 153.4 (C), 153.2 (C), 143.7 (C), 143.3 (C), 142.4 (CH), 141.1 (C), 140.4 (C), 140.1 (C), 139.9 (CH), 139.3 (CH), 113.7 (C), 113.3 (C), 113.2 (CH), 110.6 (CH), 110.2 (CH), 110.1 (CH), 109.5 (C), 109.1 (C), 107.3 (CH), 58.1 (CH₂), 50.8 (CH₂), 21.5 (CH₂), 20.1 (CH₂), 11.1 (CH₃), 10.6 ppm (CH₃); ¹⁹F NMR (282 MHz, CD₃CN): δ = -150.2 (20%), -150.4 ppm (80%); HRMS (ESI+): *m/z* calcd for C₂₆H₂₃N₂O₄ [M]⁺: 411.1703; found: 411.1675.

Compound 9

Compound **8-H⁺** (25 mg, 0.04 mmol) was dissolved in anhydrous CH₂Cl₂ (1 mL, 0.05 M). SOCl₂ (27 μL, 0.22 mmol) was added to this solution, and the reaction was stirred for 10 min at 25 °C. Then distilled ethanol (13 μL, 0.22 mmol) was added and, after 10 min of stirring at 25 °C, the mixture was concentrated under reduced pressure. The residue was diluted with CH₂Cl₂ and washed with an aqueous diluted solution of HBF₄ (1 M), dried over Na₂SO₄, filtered, and finally evaporated. Purification by column chromatography on silica gel with CH₂Cl₂/MeOH (95/5) as the eluent afforded the pure product as a pink solid (23 mg, 90%). *R*_f = 0.3 (SiO₂, CH₂Cl₂/MeOH, 98:2); m.p. 186 °C; UV/Vis (CH₃CN): λ_{max} (ε_{max}) = 547 nm (13 200 L mol⁻¹ cm⁻¹); IR (neat): ν̄ = 3625, 2979, 2880, 1710, 1613, 1527, 1461, 1345, 1262, 1222, 1171, 1105, 1046, 820, 756 cm⁻¹; ¹H NMR (500 MHz, CD₃CN): δ = 8.49 (d, *J* = 9.1 Hz, 1H; CH), 8.13–8.06 (m, 2H; 2 × CH), 7.65–7.60 (m, 2H; 2 × CH), 7.52 (d, *J* = 9.1 Hz, 1H; CH), 7.36–7.31 (m, 2H; 2 × CH), 4.54–4.47 (m, 4H; 2 × CH₂), 4.44–4.37 (m, 2H; CH₂), 2.01–1.95 (m, 2H; CH₂), 1.74–1.65 (m, 2H; CH₂), 1.45 (t, *J* = 7.1 Hz, 3H; CH₃), 1.18 (t, *J* = 7.4 Hz, 3H; CH₃), 0.58 ppm (t, *J* = 7.3 Hz, 3H; CH₃); ¹³C NMR (126 MHz, CD₃CN): δ = 167.5 (C), 153.7 (C), 153.6 (C), 143.8 (C), 143.2 (C), 142.2 (CH), 141.3 (C), 140.9 (C), 140.4 (C), 139.9 (CH), 139.3 (CH), 114.1 (C), 113.6 (C), 113.1 (CH), 110.6 (CH), 110.2 (CH), 110.2 (CH), 109.8 (C), 109.4 (C), 109.3 (C), 107.4 (CH), 63.3 (CH₂), 57.9 (CH₂), 50.8 (CH₂), 21.4 (CH₂), 20.1 (CH₂), 14.5 (CH₃), 11.1 (CH₃), 10.6 ppm (CH₃); ¹⁹F NMR (282 MHz, CD₃CN): δ = -150.1 (20%), -150.2 ppm (80%); HRMS (ESI+): *m/z* calcd for C₂₄H₂₇N₂O₃⁺ [M]⁺: 439.2016; found: 439.2012.

Compound 10

Compound **8-H⁺** (25 mg, 0.04 mmol) was dissolved in anhydrous CH₂Cl₂ (1 mL, 0.05 M). SOCl₂ (27 μL, 0.22 mmol) was added to this solution, and the reaction was stirred for 10 min at 25 °C. Then distilled propylamine (40 μL, 0.7 mmol) was added at 0 °C and, after 15 min of stirring at 25 °C, the mixture was quenched with water. The organic layer was extracted and washed with a 1 M aqueous diluted solution of HBF₄ (1 M), dried over Na₂SO₄, filtered, and concentrated under reduced pressure. Purification by column chromatography on silica gel with CH₂Cl₂/MeOH (95/5) as the eluent afforded the pure product as a pink solid (11 mg, 42%). *R*_f = 0.4 (SiO₂,

CH₂Cl₂/MeOH, 98:2); m.p. 146 °C; UV/Vis (CH₃CN): λ_{max} (ε_{max}) = 557 nm (10000 L mol⁻¹ cm⁻¹); IR (neat): ν̄ = 3409, 2969, 2980, 1615, 1526, 1461, 1344, 1275, 1171, 1119, 1077, 967, 839, 755 cm⁻¹; ¹H NMR (CD₂Cl₂, 500 MHz): δ = 8.17 (d, J = 8.8 Hz, 1H; CH), 8.13–8.05 (m, 2H; 2 × CH), 7.65–7.55 (m, 2H; 2 × CH), 7.49 (d, J = 9.1, 1H; CH), 7.38–7.30 (m, 3H; 3 × CH), 4.55–4.45 (m, 4H; 2 × CH₂), 3.44–3.38 (m, 2H; CH₂), 12.0–1.90 (m, 2H; CH₂), 1.86–1.74 (m, 2H; CH₂), 1.74–1.65 (m, 2H; CH₂), 1.18 (t, J = 7.3 Hz, 3H; CH₃), 1.03 (t, J = 7.4 Hz, 3H; CH₃), 0.78 ppm (t, J = 7.3 Hz, 3H; CH₃); ¹³C NMR (126 MHz, CD₃CN): δ = 169.1 (C), 153.7 (C), 153.6 (C), 142.7 (C), 141.7 (C), 141.4 (C), 141.2 (CH), 140.6 (C), 139.7 (CH), 139.3 (CH), 119.3 (C), 113.8 (C), 112.1 (CH), 110.3 (CH), 109.9 (CH), 109.7 (CH), 109.6 (C), 109.2 (C), 107.0 (CH), 54.4 (CH₂), 50.6 (CH₂), 42.7 (CH₂), 23.2 (CH₂), 21.4 (CH₂), 19.9 (CH₂), 12.0 (CH₃), 11.1 (CH₃), 10.8 ppm (CH₃); ¹⁹F NMR (CD₃CN, 282 MHz): δ = -150.0 (20%), -150.1 (80%); HRMS (ESI+): m/z calcd for C₂₉H₃₀N₃O₂⁺ [M]⁺: 452.2333; found: 452.2340.

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Conflict of interest

The authors declare no conflict of interest.

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- [1] a) S. Dileesh, K. R. Gopidas, *Chem. Phys. Lett.* **2000**, *330*, 397–402; b) A. Pothukuchy, S. Ellapan, K. R. Gopidas, M. Salazar, *Bioorg. Med. Chem. Lett.* **2003**, *13*, 1491–1494; c) J. Reynisson, G. B. Schuster, S. B. Howerton, L. D. Williams, R. N. Barnett, C. L. Cleveland, U. Landman, N. Harrit, J. B. Chaires, *J. Am. Chem. Soc.* **2003**, *125*, 2072–2083; d) S. Dileesh, K. R. Gopidas, *J. Photochem. Photobiol. A* **2004**, *162*, 115–120; e) A. Pothukuchy, C. L. Mazzitelli, M. L. Rodriguez, B. Tuesuwan, M. Salazar, J. S. Brodbelt, S. M. Kerwin, *Biochemistry* **2005**, *44*, 2163–2172; f) B. P. Maliwal, R. Fudala, S. Raut, R. Kokate, T. J. Sørensen, B. W. Laursen, Z. Gryczynski, I. Gryczynski, *PLoS One* **2013**, *8*, e63043; g) T. J. Sørensen, E. Thyrhaug, M. Szabelski, R. Luchowski, I. Gryczynski, Z. Gryczynski, B. W. Laursen, *Methods Appl. Fluoresc.* **2013**, *1*, 025001; h) A. Shivalingam, M. A. Izquierdo, A. Le Marois, A. Vysniauskas, K. Suhling, M. K. Kuimova, R. Vilar, *Nat. Commun.* **2015**, *6*, 8178; i) A. Wallabregue, D. Moreau, P. Sherin, P. M. Lorente, Z. Jarolímová, E. Bakker, E. Vauthey, J. Gruenberg, J. Lacour, *J. Am. Chem. Soc.* **2016**, *138*, 1752–1755.
- [2] a) T. J. Sørensen, B. W. Laursen, R. Luchowski, T. Shtoyko, I. Akopova, Z. Gryczynski, I. Gryczynski, *Chem. Phys. Lett.* **2009**, *476*, 46–50; b) Y. Haketa, S. Sasaki, N. Ohta, H. Masunaga, H. Ogawa, N. Mizuno, F. Araoka, H. Takezoe, H. Maeda, *Angew. Chem. Int. Ed.* **2010**, *49*, 10079–10083; *Angew. Chem.* **2010**, *122*, 10277–10281; c) M. Folmar, T. Shtoyko, R. Fudala, I. Akopova, Z. Gryczynski, S. Raut, I. Gryczynski, *Chem. Phys. Lett.* **2012**, *531*, 126–131; d) J. Hamacek, C. Besnard, N. Mehanna, J. Lacour, *Dalton Trans.* **2012**, *41*, 6777–6782; e) B. Dong, H. Maeda, *Chem. Commun.* **2013**, *49*, 4085–4099; f) Y. Haketa, Y. Tamura, N. Yasuda, H. Maeda, *Org. Biomol. Chem.* **2016**, *14*, 8035–8038; g) B. Qiao, B. E. Hirsch, S. Lee, M. Pink, C.-H. Chen, B. W. Laursen, A. H. Flood, *J. Am. Chem. Soc.* **2017**, *139*, 6226–6233.
- [3] a) C. Adam, A. Wallabregue, H. Li, J. Gouin, R. Vanel, S. Grass, J. Bosson, L. Bouffier, J. Lacour, N. Sojic, *Chem. Eur. J.* **2015**, *21*, 19243–19249; b) H. Li, S. Voci, A. Wallabregue, C. Adam, G. M. Labrador, R. Duwald, I. H. Delgado, S. Pascal, J. Bosson, J. Lacour, L. Bouffier, N. Sojic, *ChemElectroChem* **2017**, *4*, 1750–1756.
- [4] R. Gueret, L. Poulard, M. Oshinowo, J. Chauvin, M. Dahmane, G. Dupeyre, P. P. Lainé, J. Fortage, M.-N. Collomb, *ACS Catal.* **2018**, *8*, 3792–3802.
- [5] a) B. Baisch, D. Raffa, U. Jung, O. M. Magnussen, C. Nicolas, J. Lacour, J. Kubitschke, R. Herges, *J. Am. Chem. Soc.* **2009**, *131*, 442–443; b) S. L. Raut, R. Rich, T. Shtoyko, I. Bora, B. W. Laursen, T. J. Sørensen, J. Borejdo, Z. Gryczynski, I. Gryczynski, *Nanoscale* **2015**, *7*, 17729–17734; c) X.-M. Hu, Q. Chen, Z.-Y. Sui, Z.-Q. Zhao, N. Bovet, B. W. Laursen, B.-H. Han, *RSC Adv.* **2015**, *5*, 90135–90143.
- [6] Recently, carbon-bridged triangulenium dyes were reported, see: M. Rosenberg, K. R. Rostgaard, Z. Liao, A. O. Madsen, K. L. Martinez, T. Vosch, B. W. Laursen, *Chem. Sci.* **2018**, *9*, 3122–3130.
- [7] a) B. W. Laursen, F. C. Krebs, *Angew. Chem. Int. Ed.* **2000**, *39*, 3432–3434; *Angew. Chem.* **2000**, *112*, 3574–3576; b) B. W. Laursen, F. C. Krebs, *Chem. Eur. J.* **2001**, *7*, 1773–1783; c) J. Bosson, J. Gouin, J. Lacour, *Chem. Soc. Rev.* **2014**, *43*, 2824–2840; d) J. F. Hitzemberger, T. Drewello, A. Uhl, J. Schatz, *J. Mass Spectrom.* **2017**, *52*, 174–181.
- [8] a) E. Thyrhaug, T. J. Sørensen, I. Gryczynski, Z. Gryczynski, B. W. Laursen, *J. Phys. Chem. A* **2013**, *117*, 2160–2168; b) S. A. Bogh, M. Simmermacher, M. Westberg, M. Bregnhøj, M. Rosenberg, L. De Vico, M. Veiga, B. W. Laursen, P. R. Ogilby, S. P. A. Sauer, T. J. Sørensen, *ACS Omega* **2017**, *2*, 193–203.
- [9] a) J. L. Bricks, A. D. Kachkovskii, Y. L. Slominskii, A. O. Gerasov, S. V. Popov, *Dyes Pigm.* **2015**, *121*, 238–255; b) Y. Ni, J. Wu, *Org. Biomol. Chem.* **2014**, *12*, 3774–3791; c) Z. Guo, S. Park, J. Yoon, I. Shin, *Chem. Soc. Rev.* **2014**, *43*, 16–29; d) L. Yuan, W. Lin, K. Zheng, L. He, W. Huang, *Chem. Soc. Rev.* **2013**, *42*, 622–661.
- [10] I. Bora, S. A. Bogh, M. Rosenberg, M. Santella, T. J. Sørensen, B. W. Laursen, *Org. Biomol. Chem.* **2016**, *14*, 1091–1101.
- [11] A. Shivalingam, A. Vyšniauskas, T. Albrecht, A. J. P. White, M. K. Kuimova, R. Vilar, *Chem. Eur. J.* **2016**, *22*, 4129–4139.
- [12] a) B. W. Laursen, F. C. Krebs, M. F. Nielsen, K. Bechgaard, J. B. Christensen, N. Harrit, *J. Am. Chem. Soc.* **1998**, *120*, 12255–12263; b) B. W. Laursen, T. J. Sørensen, *J. Org. Chem.* **2009**, *74*, 3183–3185; c) T. J. Sørensen, B. W. Laursen, *J. Org. Chem.* **2010**, *75*, 6182–6190; d) F. Westerlund, C. B. Hildebrandt, T. J. Sørensen, B. W. Laursen, *Chem. Eur. J.* **2010**, *16*, 2992–2996.
- [13] M. Loftthagen, R. VernonClark, K. K. Baldrige, J. S. Siegel, *J. Org. Chem.* **1992**, *57*, 61–69.
- [14] a) T. Hirose, K. Sasatsuki, H. Noguchi, S. Yokoyama, K. Matsuda, *Chem. Lett.* **2016**, *45*, 1090–1092; b) H. Noguchi, T. Hirose, S. Yokoyama, K. Matsuda, *CrystEngComm* **2016**, *18*, 7377–7383.
- [15] a) F. Torricelli, J. Bosson, C. Besnard, M. Chekini, T. Bürgi, J. Lacour, *Angew. Chem. Int. Ed.* **2013**, *52*, 1796–1800; *Angew. Chem.* **2013**, *125*, 1840–1844; b) I. H. Delgado, S. Pascal, A. Wallabregue, R. Duwald, C. Besnard, L. Guénee, C. Nancoz, E. Vauthey, R. C. Tovar, J. L. Lunkley, G. Muller, J. Lacour, *Chem. Sci.* **2016**, *7*, 4685–4693; c) S. Pascal, C. Besnard, F. Zinna, L. Di Bari, B. Le Guennic, D. Jacquemin, J. Lacour, *Org. Biomol. Chem.* **2016**, *14*, 4590–4594; d) J. Bosson, G. M. Labrador, S. Pascal, F.-A. Miannay, O. Yushchenko, H. Li, L. Bouffier, N. Sojic, R. C. Tovar, G. Muller, D. Jacquemin, A. D. Laurent, B. Le Guennic, E. Vauthey, J. Lacour, *Chem. Eur. J.* **2016**, *22*, 18394–18403.
- [16] In the [6]helicene series, bis- rather than mono-nitration is routinely achieved; see ref. [15a].
- [17] A second regioisomer of formylation of the triangulene is observed. Substitution occurs on azaoxa- rather than diaza-substituted phenyl rings. This minor derivative can only be removed by recrystallizing desired product **3** in a mixture of toluene/dichloromethane. This recrystallization is somewhat capricious; hence, the wide variation in yields.
- [18] B. V. Rokade, K. R. Prabhu, *J. Org. Chem.* **2012**, *77*, 5364–5370.
- [19] A formaniide byproduct (Y = NHCHO) can be observed as a minor component. It presents a characteristic green color.
- [20] On cationic triangulenes and related helicenes, the position of the substituents is important for subsequent control of the physicochemical

- properties; refs. [6,15d], see also: R. Duwald, S. Pascal, J. Bosson, S. Grass, C. Besnard, T. Bürgi, J. Lacour, *Chem. Eur. J.* **2017**, *23*, 13596–13601.
- [21] a) C. Kraiya, P. Singh, D. H. Evans, *J. Electroanal. Chem.* **2004**, *563*, 203–212; b) B. G. Chauhan, W. R. Fawcett, A. Lasia, *J. Phys. Chem.* **1977**, *81*, 1476–1481.
- [22] K. Sasaki, W. J. Newby, *J. Electroanal. Chem. Interfacial Electrochem.* **1969**, *20*, 137–165.
- [23] a) A. Babič, S. Pascal, R. Duwald, D. Moreau, J. Lacour, E. Allémann, *Adv. Funct. Mater.* **2017**, *27*, 1701839; b) C. Bauer, R. Duwald, G. M. Labrador, S. Pascal, P. M. Lorente, J. Bosson, J. Lacour, J.-D. Rochaix, *Org. Biomol. Chem.* **2018**, *16*, 919–923.
- [24] A value of +1.7 ppm was added to all measured ¹⁹F signals in consideration of the external calibration.
- [25] CrysAlisPro Software system, Agilent Technologies UK Ltd., Oxford, UK.
- [26] M. C. Burla, R. Caliandro, M. Camalli, B. Carrozzini, G. L. Casciaro, L. De Caro, C. Giacovazzo, G. Polidori, R. Spagna, *J. Appl. Crystallogr.* **2005**, *38*, 381–388.
- [27] a) G. Sheldrick, *Acta Crystallogr. Sect. C* **2015**, *71*, 3–8; b) G. Sheldrick, *Acta Crystallogr. Sect. A* **2008**, *64*, 112–122.
- [28] O. V. Dolomanov, L. J. Bourhis, R. J. Gildea, J. A. K. Howard, H. Puschmann, *J. Appl. Crystallogr.* **2009**, *42*, 339–341.

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