

Review

Of Flies and Men—The Discovery of TLRs

Hauke Johannes Weiss *  and Luke Anthony John O'Neill

School of Biochemistry and Immunology, Trinity Biomedical Sciences Institute, Trinity College Dublin, D02 R590 Dublin, Ireland

* Correspondence: weishh@tcd.ie

Abstract: In 2011, the Nobel Prize in Physiology or Medicine was awarded to three immunologists: Bruce A. Beutler, Jules A. Hoffmann, and Ralph M. Steinman. While Steinman was honored for his work on dendritic cells and adaptive immunity, Beutler and Hoffman received the prize for their contributions to discoveries in innate immunity. In 1996, Hoffmann found the *toll* gene to be crucial for mounting antimicrobial responses in fruit flies, first implicating this developmental gene in immune signaling. Two years later, Beutler built on this observation by describing a Toll-like gene, *tlr4*, as the receptor for the bacterial product LPS, representing a crucial step in innate immune activation and protection from bacterial infections in mammals. These publications spearheaded research in innate immune sensing and sparked a huge interest regarding innate defense mechanisms in the following years and decades. Today, Beutler and Hoffmann's research has not only resulted in the discovery of the role of multiple TLRs in innate immunity but also in a much broader understanding of the molecular components of the innate immune system. In this review, we aim to collect the discoveries leading up to the publications of Beutler and Hoffmann, taking a close look at how early advances in both developmental biology and immunology converged into the research awarded with the Nobel Prize. We will also discuss how these discoveries influenced future research and highlight the importance they hold today.

Keywords: Toll-like receptors (TLRs); innate immunity; immunology; *Drosophila*; LPS; innate immune signaling; Nobel Prize; TIR domain



Citation: Weiss, H.J.; O'Neill, L.A.J.

Of Flies and Men—The Discovery of TLRs. *Cells* **2022**, *11*, 3127. <https://doi.org/10.3390/cells11193127>

Academic Editor: Dieter Kabelitz

Received: 26 August 2022

Accepted: 2 October 2022

Published: 5 October 2022

Publisher's Note: MDPI stays neutral with regard to jurisdictional claims in published maps and institutional affiliations.



Copyright: © 2022 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (<https://creativecommons.org/licenses/by/4.0/>).

1. Introduction

The 2011 Nobel Prize to Hoffmann, Beutler, and Steinmann was a testament to the advances in research on innate immunity made in the 1990s and early 2000s. While concepts of adaptive immunity were already well described at the time, the function of innate immune cells was mostly thought to be reduced to the activation of the adaptive immune system, the nonspecific elimination of microbes and materials by phagocytosis, and inflammation. While Steinmann bridged the gap between the two with his work on dendritic cells [1–3], it was the work of Hoffmann and Beutler that elevated the field of innate immune recognition. Hoffmann published on the antimicrobial function of the developmental gene *toll* in *Drosophila melanogaster* in 1996 [4], and Beutler built on these data by identifying the Toll-like receptor TLR4 as the receptor for bacterial lipopolysaccharide (LPS) in mammals in 1998 [5], sparking a revolution in the field. These publications gave rise to the identification of the TLR family of receptors and allowed for the discovery of many other pattern recognition receptors (PRRs) as well. While ground-breaking, the findings of Hoffmann and Beutler were built on decades worth of discoveries in the fields of immunology and developmental biology, the most important of which we will discuss in this review.

2. Toll and Pathways in *Drosophila*

The origin of Toll in immunity can be traced to development biology research. The fruit fly *Drosophila melanogaster* has been a key model organism in studying developmental

biology and genetics for over a century, and, besides Hoffmann, four more Nobel Prizes have been awarded to scientists for their work with this animal. The first experiments involving *Drosophila* were conducted as early as 1901 by William Castle at Harvard University [6]; however, it was Thomas Hunt Morgan who achieved the first breakthrough results with *Drosophila*, confirming Mendel's theory of inheritance and establishing the concepts of genes and chromosomes [7]. This discovery earned him the 1933 Nobel Prize in Physiology or Medicine. Morgan's student, Hermann Muller, later found that radiation mutates genes and that the offspring of irradiated *Drosophila* have mutated genes too [8], earning him a Nobel Prize in physiology or medicine as well. Easy to breed and inexpensive to house, *Drosophila* quickly became the gold standard animal model in genetic research.

Modern *Drosophila* research emerged in the 1970s, when new techniques allowed for the genetic mapping of embryonic development. It emerged that larvae contain pre-determined groups of cells—so-called imaginal discs—that would give rise to the various features and body parts of the adult *Drosophila* yet hold no function in the larvae themselves [9,10]. These imaginal discs could be transplanted [11,12], allowing for the investigation of their individual functions. Clonal analysis of these discs and the resulting labelling techniques [10,13] helped in understanding the fate of these individual cell clusters, resulting in the discovery of anterior (A) and posterior (P) compartments within various structures of the adult fly, being based on different imaginal discs [14–16]. Several morphogens that organize the development of distinct compartments were discovered during this time [17]; among them is *dorsal*, a mutation which caused a fully dorsalized phenotype [18]. Other mutants displaying similar phenotypes, including *pelle*, *tube*, *toll*, *spätzle*, and *cactus* [19,20], which soon emerged one after the other, revealed the genetic blueprint of the animal. A key factor in the discovery of these genes was the utilization of new molecular techniques in DNA cloning [21], and these findings were later awarded with another Nobel Prize to Edward B. Lewis, Christiane Nüsslein-Volhard, and Eric F. Wieschaus in 1995 for their discoveries regarding “the genetic control of early embryonic development” [22].

Toll was identified as one of the most important genes in embryonic development and the establishment of embryonic polarity. While most mutations of genes from the *dorsal* group retain some residual polarization, *toll* mutants lack any organization, and dorsoventral polarization by the reintroduction of wildtype (WT) cytoplasm is fully dependent on where the cytoplasm is injected [23]. It was suggested that the *toll* protein product is constantly expressed but would exist in an activated or an inactivated state and that members of the *dorsal* group, including *toll*, might be part of a signaling pathway [24], hypotheses that should later be confirmed on a biochemical level. The *toll* gene product Toll was soon identified as a membrane-bound receptor [25], capable of signal transduction through an extracellular stimulus [26]. This ligand was then identified as Spätzle, another member of the *dorsal* group [27], and Dorsal itself was shown to be a transcription factor recruited to the nucleus upon Spätzle binding to Toll. As stated above, Toll was then shown to be homologous to the mammalian type 1 IL-1 receptor (IL1R1) [28], while Dorsal was closely related to the IL-1-induced transcription factor NF- κ B [29]. More similarities between the Toll pathway and the inflammation-inducing anti-microbial IL-1 receptor-induced signaling cascades soon became apparent, laying the groundwork for the discoveries of Hoffmann. Most notably, *cactus* was shown to be a homologue of the inhibitor of NF- κ B (I- κ B) in its function to retain Dorsal or NF- κ B, respectively, in the cytoplasm [30,31], and both pathways involve structurally related protein kinases, such as the *pelle* gene product (pI1) in the Toll pathway [32] and the interleukin-receptor associated kinase (IRAK) in the IL-1 pathway, respectively [33]. In addition, both *dorsal* itself as well as the closely related *Dorsal-related immunity factor* (*dif*) had been shown to exert antimicrobial functions in fat body cells from *Drosophila* [34,35].

Interestingly, a protein structurally related to both Toll and IL1R1 was discovered in a different kingdom of life. The N protein from the tobacco plant shared a cytoplasmic domain with Toll and IL1R1, a domain termed the Toll-IL-1 receptor resistance (TIR) domain

by Barbara Baker, who uncovered the role of the N protein in the antiviral responses against *Tobacco mosaic virus* [36]. The developmental role of Toll and its homology to both IL1R1 and the N protein were therefore important precursors to the discovery of the role of Toll in *Drosophila* innate immunity.

3. Toll Has Antimicrobial Functions

In 1996, Jules Hoffmann published on the antimicrobial function of three different genes, *spätzle*, *cactus*, and *toll* (*tl*), in the fruit fly *Drosophila melanogaster* [4]. Jules Hoffmann was born in Luxembourg on 2 August 1942. From an early age, he had a fascination for insects, an interest that would carry over into his research career. He obtained his doctoral degree in biology in 1969 from the University of Strasbourg, becoming a research associate in the same year. He went on to pursue postdoctoral training at the Philipps University of Marburg, Germany in the area of biochemistry before returning to Strasbourg in 1974 to set up his own lab and establish a research unit on the immune response in insects in 1978. Here, he laid the groundwork for the research that would result in him being awarded the Nobel Prize decades later. After initial success working on grasshoppers [37], he turned towards *Drosophila melanogaster* later, identifying antimicrobial polypeptides such as dipterin [38] or defensin [39] as being part of the immune system of these insects. His groundbreaking and Nobel Prize-awarded work, however, was for the 1996 paper on Toll in *Drosophila* immunity, where he and Bruno Lemaitre were the first to describe the Toll pathway as being responsible for the production of the antifungal peptide drosomycin [4], a discovery sparking a huge interest in similar proteins in mammals and ultimately leading to the discovery of the role of TLRs in innate immunity.

Spätzle, *cactus*, and *toll* are part of the dorsoventral signaling pathway and, curiously, the activation of the pathway had first been shown to lead to the activation of the dorsoventral morphogen *dorsal*, which is key to the establishment of dorsoventral polarity in the developing *Drosophila* embryo [40]. Hoffmann found that all three of these genes are involved in mounting an antifungal response, showing a decreased induction of the antifungal peptide gene *drosomycin* in animals with mutations in *spätzle*, *cactus*, or *toll*. *Dorsal*, however, was not involved. Drawing parallels to the components of the NF- κ B pathway, Hoffmann described how the activation of the Toll receptor by its ligand *spätzle* leads to a similar signaling cascade as the activation of IL1R1 by IL-1 [41]. As stated above, IL1R1 had been shown to be a homologue to Toll, as well as the N protein in tobacco [36], which led to the concept of the 'TIR' domain. The N protein had been shown to be required for resistance to *Tobacco mosaic virus*. That Toll had been shown to be involved in host defense made biological sense, since IL1R1 and the N protein were already known to be similarly involved in immunity in mammals and plants, respectively. The TIR domain was therefore a very ancient signaling domain for innate immunity stretching back billions of years to the common ancestor of plants and animals.

Toll drove an antifungal response but not antibacterial response. Antibacterial responses are facilitated by the *immune deficiency* (*imd*) gene [42], with the antibacterial peptides cecropin, attacin, and defensin being partly dependent on Toll and dipterin and drosocin being independent of Toll. These results were achieved by the mutation of components of either the *imd* or *tl* pathways and the subsequent investigation of the downstream activation of components of these antimicrobial pathways or the challenge of the mutated flies with bacterial or fungal infections. This paper was therefore the first to describe the Toll pathway as a component of the innate immune system, a finding that will later be expanded to mammals and lead to the discovery of a multitude of receptors and pathways in innate immunity common to plants and animals.

4. Innate Immune Signaling and LPS

The idea of innate immune sensing by so-called pattern recognition receptors was proposed by Charles Janeway in 1989 [43]. TLR4 emerged as a prototypical PRR. It is worth going back a bit further in time to fully understand the motivations that drove Beutler

in his pursuit of the LPS receptor. LPS was first described as an “endotoxin” derived from *Vibrio cholerae*, the bacterium responsible for cholera, by Richard Pfeiffer in 1892, inducing fever and death even when the bacterium was killed [44]. Soon after, Eugenio Centanni isolated the substance and proposed it to be non-proteinous due to its remarkable heat stability [45]. Around the same time, physician William B. Coley started treating his cancer patients at the Memorial Hospital in New York with heat-inactivated bacteria, observing tumor remission in some cases [46], which, today, we know was due to the strong activation of the immune system by the bacterial LPS [47]. The endotoxin was identified as lipopolysaccharide in 1943 [48] but was not fully structurally identified until the 1980s [49], and while the detailed composition and the strength of the response may vary depending on the bacteria in question, they all induced the same inflammatory symptoms [50]—that is, until the 1960s. Suddenly a mouse strain was mentioned that was resistant to LPS-induced inflammation [51], indicating how this susceptibility to endotoxin may be down to very distinct cellular processes. It was later discovered that the resistance stemmed from a mutation in a distinct locus, appropriately named *lps* [52,53]. These mice, as well as another resistant strain identified in 1977 [54], termed C57BL/10ScCr, were important pillars for Beutler’s work, as the genetic analysis of the *lps* locus would provide the key for the discovery of the LPS receptor.

With emerging knowledge about the individual cells acting within inflammation and the immune system, it became clear that macrophages were the main cell type responsible for LPS-induced inflammation [55], raising the question of whether such an intense reaction to LPS was a good thing or a bad thing. As a strong inducer of sepsis, it might be viewed as a purely toxic substance; however, Coley’s work showed how LPS responses could keep malignancies at bay. In addition, by the end of the 1960s, it was clear that LPS could be beneficial in pathogen clearance, acting as an adjuvant [56–58]. In the 1980s, it was also shown that mice of the above-mentioned strains that lack an LPS response are more susceptible to infection with gram-negative bacteria [59,60], the very organisms that carry LPS on their surface. It was therefore clear that a sensing mechanism for LPS must exist to detect and respond to LPS-bearing bacteria. One of these responses included the production of cytokines such as tumor necrosis factor (TNF) [61], the induction of which was found to be at least partly responsible for LPS-induced toxicity [62], and TNF seemed to be beneficial in resolving the same type of microbial infections as LPS [63,64].

The work on cytokines such as IL-1 and TNF, which overlap in many of their pro-inflammatory effects, in turn provided the first ideas as to how the signal transduction from LPS to the nucleus might work. In the years before the discovery of TLR4, it had already been established that IL-1 binds IL1R1, resulting in the activation of the transcription factor NF- κ B, as does LPS [65]. Two LPS-binding proteins could be identified prior to the discovery of TLR4. The LPS-binding protein (LBP) was first described in 1986 [66] and was found to enhance LPS-induced signaling events [67,68]. This protein, however, was not membrane-bound and could thus not act as a signal transducer. The second receptor, CD14, is a membrane-bound protein [69,70], but it lacks cytoplasmic domains [71]. Then, in 1996, Hoffmann published his work on the antimicrobial role of Toll in *Drosophila* [4], and, suddenly, the relevance of this receptor family in immunity became apparent and gave Beutler a reason to pursue TLR4 once it came up as a candidate in the unravelling of the *lps* locus. Importantly, Charles Janeway and Ruslan Medzhitov had shown in 1997 that TLR4, described as human Toll in their paper, could activate immune cells [72]. While other TLR members had been described before TLR4, their function and ligands remained unknown until after the identification of TLR4 as the LPS receptor [73,74], further highlighting the significance of Beutler’s discovery. The most important milestones leading up to Beutler and Hoffmann’s discoveries are listed in a timeline in Figure 1.

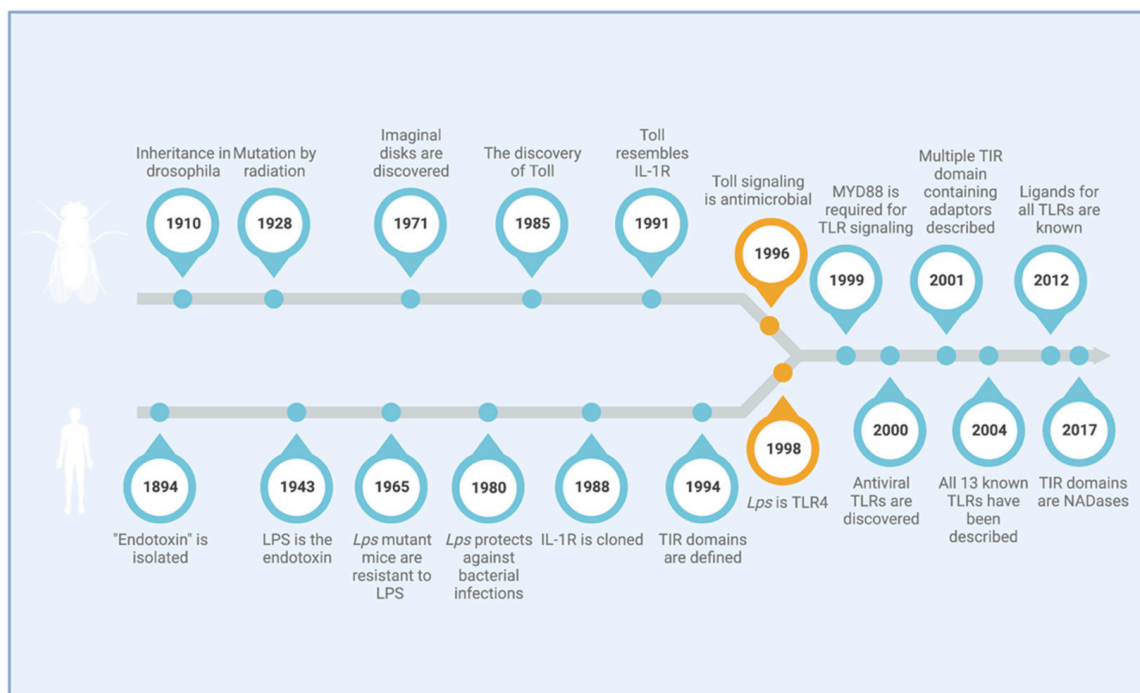


Figure 1. Milestones along the way to TLR discovery. The highlighted publications contributed significantly to today's knowledge on TLRs and innate immunity.

5. TLR4 Is the Receptor for Bacterial LPS

The search for the LPS receptor started with a thorough investigation of the so-called *lps* locus after it was reported decades previously that C3H/HeJ mice, a strain with mutated *lps*, are unresponsive to LPS [75,76]. LPS resistance was also shown in mice of the C57BL/10ScCr strain [54,77]. While an LPS-binding protein termed cluster of differentiation 14 (CD14) had already been identified on the surface of macrophages, this protein is not able to transduce a signal due to a lack of a cytoplasmic domain, as described above [70]. CD14 augments LPS-induced signaling [71]; however, it was likely to act as a co-receptor for the then-unknown TLR4. Utilizing genetic and physical mapping data, Beutler characterized the *lps* locus and identified *tlr4* in the area mapped. Bruce A. Beutler was born on 29 December 1957 in Chicago, Illinois. Growing up in California, he graduated from the University of California, San Diego in 1976 at only 18 years old with a degree in biology. Beutler then pursued an M.D. degree at the University of Chicago, which he obtained in 1981. After postdoctoral training at Rockefeller University in New York, he became Assistant Professor in 1985 before moving to Dallas a year later. It was in New York where he made one of his first important discoveries: isolating tumor necrosis factor-alpha (TNF- α) in mice [61]. At the University of Texas (UT) Southwestern Medical Center in Dallas, he obtained an associate professorship in 1990 and, finally, a full professorship in 1996. Here, he continued working on TNF, generating Immunoglobulin G (IgG) heavy chain-based TNF antagonists [78], eventually leading him to become interested in one particular TNF-inducing agent, LPS. Setting out to explain the induction of TNF and other cytokines by LPS, together with Alexander Poltorak, he was able to identify Toll-like receptor 4 (TLR4) as the receptor for LPS in 1998 [5], earning him his Nobel Prize.

Tlr4 was a promising target, as it was by then known to be part of the IL1R/TLR family. Comparing the macrophage mRNA and genomic DNA from an LPS-responsive vs. C3H/HeJ mouse strain, Beutler and colleagues identified a mutation in the *tlr4* gene, resulting in a proline to histidine substitution in the *tlr4* gene from the LPS-unresponsive mice. In the C57BL/10ScCr strain, *tlr4* mRNA was entirely absent, further supporting the importance of this gene in LPS-induced signaling. Finally, TLR4 was shown to be downregulated in response to LPS treatment, possibly to facilitate tolerance to LPS, a well-known

phenomenon. In the context of the discoveries made by Hoffmann two years prior [37], Beutler therefore concluded that TLR4, like the *Drosophila* homologue Toll, launched a response to microbial infection. In the case of TLR4, however, this pathway has evolved to detect gram-negative bacteria, with developmental aspects that had been described for Toll in insects being lost. The homologies between Toll and TLR4 are highlighted in Figure 2. In this work, Beutler expands the discovery made by Hoffmann in insects to mammals, revealing one of the most important antimicrobial receptors in mammals in the process.

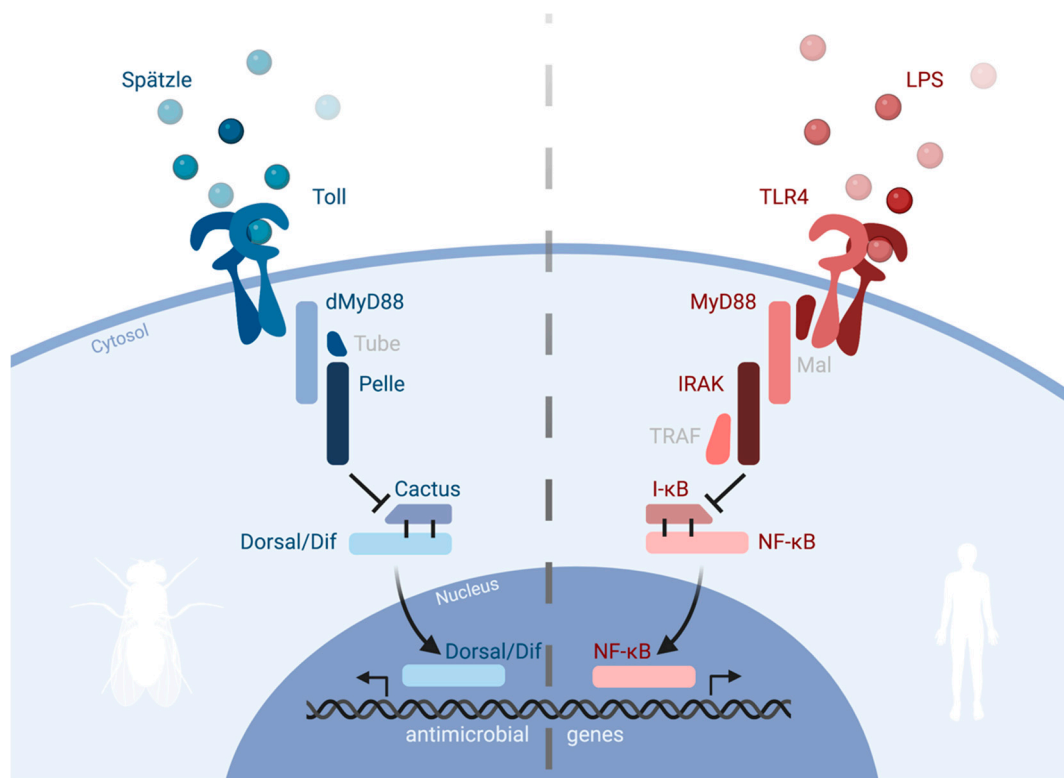


Figure 2. Simplified illustration of the Toll and TLR4 signaling pathways. Homologue components of both pathways are depicted in blue and red, respectively.

The Nobel Prize was awarded to Hoffmann and Beutler in recognition of their contribution to the discovery of Toll and TLRs in innate immunity. While years of research are represented in this award, these two papers are particularly important.

6. Scientific Impact

After the function of TLR4 was determined, the field of innate immune recognition exploded. Suddenly, the innate immune system was no longer seen as a crude, unspecific system with the simple task of activating adaptive immunity. It proved to be a sophisticated detection system capable of responding to and selecting between distinct microbial threats.

Soon after TLR4, the functions of many other TLRs were defined, each with another specific microbial ligand. Shizuo Akira found that TLR2 and TLR6 recognize gram-positive bacteria by binding to lipid structures [79,80], and TLR9 was shown to bind to bacterial DNA [81]. TLR3 was found to bind to double-stranded RNA (dsRNA), as found after viral infection [82], TLR5 was found to bind bacterial flagellin [83], and TLR7 and 8 were found to recognize viral single-stranded RNA (ssRNA) [84]. TLR10 binds triacylated lipopeptides [85] and is expressed in humans but not in mice. Conversely, mice express TLR11, which binds bacterial profilin and flagellin [86,87], TLR12, which binds profilin [86], and TLR13, which binds bacterial ribosomal RNA [88], which are all missing in humans. In addition, TLR2 was found to heterodimerize with TLR1 or TLR6 [89], while TLR11

dimerizes with TLR12 [90]. In Figure 3, all currently known TLRs are displayed, with their ligands and primary adaptors.

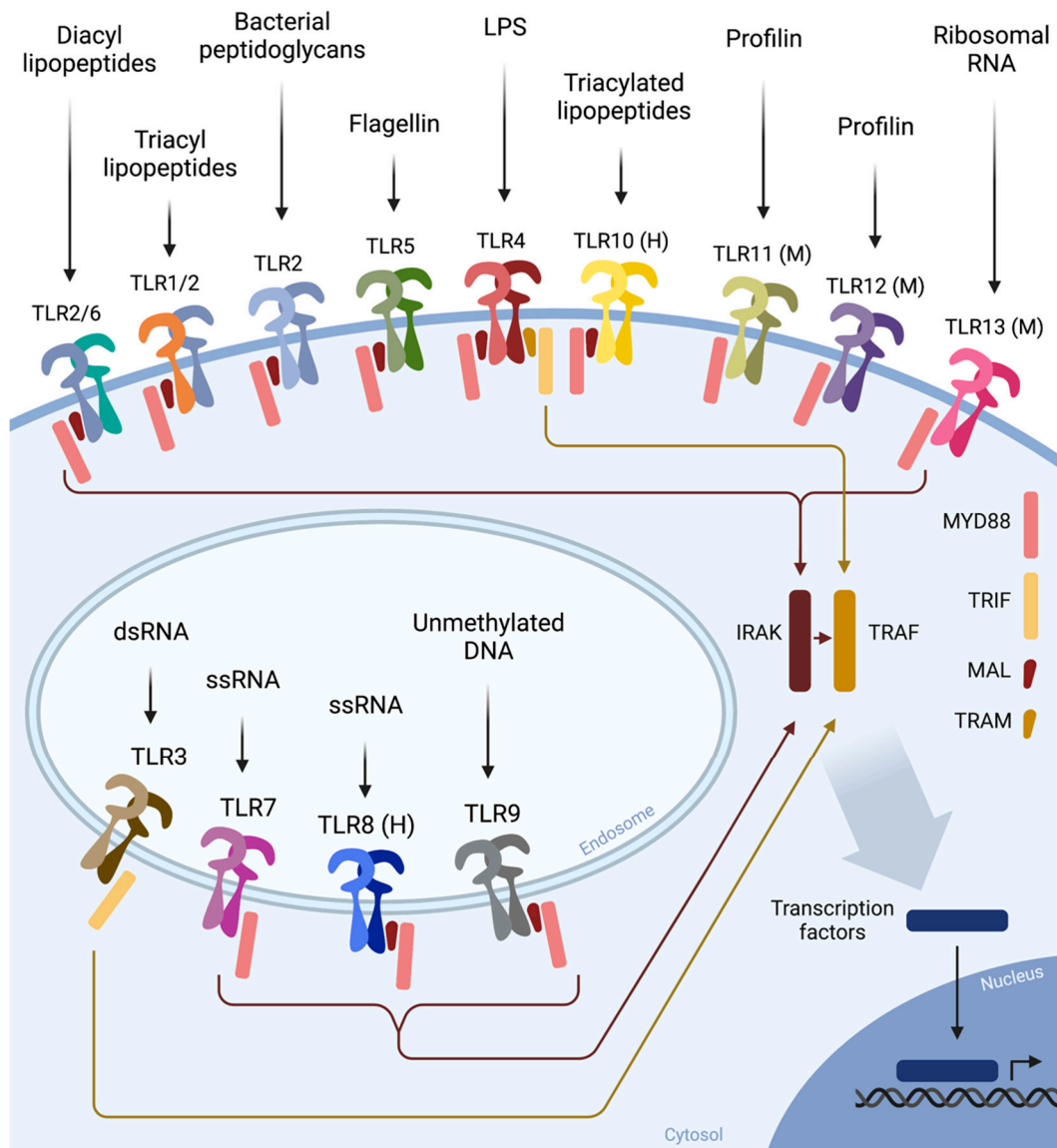


Figure 3. Illustration of all known TLRs and their ligands and adaptors, including those exclusive to mice (M) and humans (H).

Besides TLRs, other PRRs were also discovered in the following years—most notably, NOD-like receptors (NLRs) [91], the first of which was discovered in 1999 [92], RIG-like receptors (RLRs) [93], first described in 2004 [94], and C-type lectin receptors (CLRs) [95], which, although being considered part of the immune system for a long time, were shown to bind microbial products in 2000 [96].

There were also remarkable advances in determining the signaling pathways initiated by TLRs. Adaptor proteins involved in signaling were shown to have TIR domains. The first to be described was Myeloid differentiation primary-response protein 88 (MYD88). Discovered before the TLRs, it was found to bind to IL1R1, facilitating downstream signaling events [97], and, subsequently, it was found to mediate TLR signaling in TLRs as well [98]. Signaling was therefore initiated by TIR–TIR interactions from the receptor to adaptors. An exception was TLR3, which signals through TIR domain-containing adaptor protein inducing IFN β (TRIF) [99,100]. TLR4 is also able to signal independently of MYD88

through TRIF when endocytosed [101,102]. TRIF-related adaptor molecule (TRAM) recruits TRIF to the TLR signaling complex [103,104], while MYD88-adaptor-like protein (MAL) recruits MYD88 to TLR4 and all other TLRs that use MYD88 [105]. From here on, TLR signaling gets more complex (described in more depth elsewhere) [106,107]. Even today, new aspects of TLR signaling are still being uncovered, especially revolving around the functionality of TIR domains. The TIR domain containing protein sterile alpha and the TIR motif containing 1 (SARM1), for example, were found to have enzymatic activity, depleting nicotinamide adenine dinucleotide (NAD⁺) [108]. Indeed, several TIR domains have been found to have NADase activities, across various species as well [109]. While TIR domains from TLR signaling components other than SARM1 in animals do not seem to have NADase activity, the discovery of the enzymatic activity of this domain gives this aspect of TLR-signaling molecules a whole new angle and contributes to the burgeoning field of immunometabolism.

7. Concluding Remarks

Overall, it cannot be denied that the discoveries that resulted in the Nobel Prize being awarded to Hoffmann and Beutler are among the most important immunological milestones of the outgoing 20th century and are a testament to the tenacity of both scientists, along with the members of their research groups. They reshaped the way innate immunity was perceived and our understanding of how the first line of defense in our body actually works. While the Nobel Prize was well deserved, other pioneers in the field deserve recognition too. Without the groundbreaking discoveries of Nüsslein-Volhard in the field of Toll signaling or the emphasis placed on the innate immune system by Charles Janeway, these discoveries would not have been possible. Additionally, other brilliant scientists, such as Ruslan Medzhitov and Shizuo Akira, should be mentioned alongside Hoffmann and Beutler for their many contributions. The two key publications described here truly elevated the field, the consequences of which are still being extensively explored by many scientists.

Author Contributions: Conceptualization, H.J.W. and L.A.J.O.; investigation, H.J.W.; resources, H.J.W.; writing—original draft preparation, H.J.W.; writing—review and editing, L.A.J.O.; visualization, H.J.W.; supervision, L.A.J.O.; funding acquisition, L.A.J.O. All authors have read and agreed to the published version of the manuscript.

Funding: The O'Neill laboratory acknowledges grant support from the European Research Council Metabinate (834370), the Wellcome Trust (205455) and Science Foundation Ireland (19/FFP/6507). H.J.W. is supported by the European Union's Horizon 2020 research and innovation program under the Marie Skłodowska-Curie grant agreement No 813343.

Institutional Review Board Statement: Not applicable.

Informed Consent Statement: Not applicable.

Data Availability Statement: Not applicable.

Acknowledgments: Figures were created with BioRender.com.

Conflicts of Interest: The authors declare no conflict of interest.

References

1. Steinman, R.M.; Cohn, Z.A. Identification of a Novel Cell Type in Peripheral Lymphoid Organs of Mice. *J. Exp. Med.* **1973**, *137*, 1142–1162. [[CrossRef](#)] [[PubMed](#)]
2. Steinman, R.M.; Witmer, M.D. Lymphoid dendritic cells are potent stimulators of the primary mixed leukocyte reaction in mice. *Proc. Natl. Acad. Sci. USA* **1978**, *75*, 5132–5136. [[CrossRef](#)] [[PubMed](#)]
3. Schuler, G.; Steinman, R.M. Murine epidermal Langerhans cells mature into potent immunostimulatory dendritic cells in vitro. *J. Exp. Med.* **1985**, *161*, 526–546. [[CrossRef](#)] [[PubMed](#)]
4. Lemaitre, B.; Nicolas, E.; Michaut, L.; Reichhart, J.-M.; Hoffmann, J.A. The Dorsalventral Regulatory Gene Cassette *spätzle/Toll/cactus* Controls the Potent Antifungal Response in *Drosophila* Adults. *Cell* **1996**, *86*, 973–983. [[CrossRef](#)]
5. Poltorak, A.; He, X.; Smirnova, I.; Liu, M.-Y.; Huffel, C.V.; Du, X.; Birdwell, D.; Alejos, E.; Silva, M.; Galanos, C.; et al. Defective LPS Signaling in C3H/HeJ and C57BL/10ScCr Mice: Mutations in *Tlr4* Gene. *Science* **1998**, *282*, 2085–2088. [[CrossRef](#)]

6. Jennings, B.H. *Drosophila*—A versatile model in biology & medicine. *Mater. Today* **2011**, *14*, 190–195. [[CrossRef](#)]
7. Morgan, T.H. Sex Limited Inheritance in *Drosophila*. *Science* **1910**, *32*, 120–122. [[CrossRef](#)]
8. Muller, H.J. The Production of Mutations by X-Rays. *Proc. Natl. Acad. Sci. USA* **1928**, *14*, 714–726. [[CrossRef](#)]
9. Mandaravally Madhavan, M.; Schneiderman, H.A. Histological analysis of the dynamics of growth of imaginal discs and histoblast nests during the larval development of *Drosophila melanogaster*. *Wilhelm Roux's Arch. Dev. Biol.* **1977**, *183*, 269–305. [[CrossRef](#)]
10. Garcia-Bellido, A.; Merriam, J.R. Parameters of the wing imaginal disc development of *Drosophila melanogaster*. *Dev. Biol.* **1971**, *24*, 61–87. [[CrossRef](#)]
11. Gehring, W. The Stability of the Determined State in Cultures of Imaginal Disks in *Drosophila*. *Develop. Biol.* **1967**, *16*, 438–456. [[CrossRef](#)]
12. Beadle, G.W.; Ephrussi, B. The differentiation of eye pigments in *drosophila* as studied by transplantation. *Genetics* **1936**, *21*, 225–247. [[CrossRef](#)] [[PubMed](#)]
13. Bryant, P.J. Cell lineage relationships in the imaginal wing disc of *Drosophila melanogaster*. *Dev. Biol.* **1970**, *22*, 389–411. [[CrossRef](#)]
14. Steiner, E. Establishment of compartments in the developing leg imaginal discs of *Drosophila melanogaster*. *Wilhelm Roux's Arch. Dev. Biol.* **1976**, *180*, 9–30. [[CrossRef](#)]
15. MORATA, G.; LAWRENCE, P.A. Anterior and posterior compartments in the head of *Drosophila*. *Nature* **1978**, *274*, 473–474. [[CrossRef](#)]
16. Lawrence, P.A.; Morata, G. Compartments in the wing of *Drosophila*: A study of the engrailed gene. *Dev. Biol.* **1976**, *50*, 321–337. [[CrossRef](#)]
17. Nüsslein-Volhard, C.; Wieschaus, E. Mutations affecting segment number and polarity in *Drosophila*. *Nature* **1980**, *287*, 795–801. [[CrossRef](#)]
18. Santamaria, P.; Nüsslein-Volhard, C. Partial rescue of dorsal, a maternal effect mutation affecting the dorso-ventral pattern of the *Drosophila* embryo, by the injection of wild-type cytoplasm. *EMBO J.* **1983**, *2*, 1695–1699. [[CrossRef](#)]
19. Anderson, K.V.; Nüsslein-Volhard, C. Information for the dorsal–ventral pattern of the *Drosophila* embryo is stored as maternal mRNA. *Nature* **1984**, *311*, 223–227. [[CrossRef](#)]
20. Schüpbach, T.; Wieschaus, E. Female sterile mutations on the second chromosome of *Drosophila melanogaster*. I. Maternal effect mutations. *Genetics* **1989**, *121*, 101–117. [[CrossRef](#)]
21. Morata, G.; Lawrence, P. An exciting period of *Drosophila* developmental biology: Of imaginal discs, clones, compartments, parasegments and homeotic genes. *Dev. Biol.* **2022**, *484*, 12–21. [[CrossRef](#)]
22. Press Release: The Nobel Prize in Physiology or Medicine. 1995. Available online: <https://www.nobelprize.org/prizes/medicine/1995/press-release/> (accessed on 10 September 2022).
23. Anderson, K.V.; Bokla, L.; Nüsslein-Volhard, C. Establishment of dorsal-ventral polarity in the *drosophila* embryo: The induction of polarity by the Toll gene product. *Cell* **1985**, *42*, 791–798. [[CrossRef](#)]
24. Nüsslein-Volhard, C. The Toll gene in *Drosophila* pattern formation. *Trends Genet.* **2022**, *38*, 231–245. [[CrossRef](#)]
25. Hashimoto, C.; Hudson, K.L.; Anderson, K.V. The Toll gene of *drosophila*, required for dorsal-ventral embryonic polarity, appears to encode a transmembrane protein. *Cell* **1988**, *52*, 269–279. [[CrossRef](#)]
26. Stein, D.; Roth, S.; Vogelsang, E.; Nüsslein-Volhard, C. The polarity of the dorsoventral axis in the *drosophila* embryo is defined by an extracellular signal. *Cell* **1991**, *65*, 725–735. [[CrossRef](#)]
27. Schneider, D.S.; Jin, Y.; Morisato, D.; Anderson, K.V. A processed form of the Spatzle protein defines dorsal-ventral polarity in the *Drosophila* embryo. *Development* **1994**, *120*, 1243–1250. [[CrossRef](#)]
28. Schneider, D.S.; Hudson, K.L.; Lin, T.Y.; Anderson, K.V. Dominant and recessive mutations define functional domains of Toll, a transmembrane protein required for dorsal-ventral polarity in the *Drosophila* embryo. *Genes Dev.* **1991**, *5*, 797–807. [[CrossRef](#)]
29. Steward, R. Dorsal, an Embryonic Polarity Gene in *Drosophila*, Is Homologous to the Vertebrate Proto-Oncogene, *c-rel*. *Science* **1987**, *238*, 692–694. [[CrossRef](#)]
30. Kidd, S. Characterization of the *Drosophila* cactus locus and analysis of interactions between cactus and dorsal proteins. *Cell* **1992**, *71*, 623–635. [[CrossRef](#)]
31. Geisler, R.; Bergmann, A.; Hiromi, Y.; Nüsslein-Volhard, C. cactus, a gene involved in dorsoventral pattern formation of *Drosophila*, is related to the κ B gene family of vertebrates. *Cell* **1992**, *71*, 613–621. [[CrossRef](#)]
32. Shelton, C.A.; Wasserman, S.A. pelle encodes a protein kinase required to establish dorsoventral polarity in the *Drosophila* embryo. *Cell* **1993**, *72*, 515–525. [[CrossRef](#)]
33. Cao, Z.; Henzel, W.J.; Gao, X. IRAK: A Kinase Associated with the Interleukin-1 Receptor. *Science* **1996**, *271*, 1128–1131. [[CrossRef](#)]
34. Reichhart, J.M.; Georgel, P.; Meister, M.; Lemaitre, B.; Kappler, C.; Hoffmann, J.A. Expression and nuclear translocation of the *rel/NF-kappa B*-related morphogen dorsal during the immune response of *Drosophila*. *C. R. Acad. Sci. III.* **1993**, *316*, 1218–1224.
35. Ip, Y.T.; Reach, M.; Engstrom, Y.; Kadalayil, L.; Cai, H.; González-Crespo, S.; Tatei, K.; Levine, M. Dif, a dorsal-related gene that mediates an immune response in *Drosophila*. *Cell* **1993**, *75*, 753–763. [[CrossRef](#)]
36. Whitham, S.; Dinesh-Kumar, S.P.; Choi, D.; Hehl, R.; Corr, C.; Baker, B. The product of the tobacco mosaic virus resistance gene *N*: Similarity to toll and the interleukin-1 receptor. *Cell* **1994**, *78*, 1101–1115. [[CrossRef](#)]

37. Hoffmann, D.; Brehelin, M.; Hoffmann, J.A. Modifications of the hemogram and of the hemocytopoietic tissue of male adults of *Locusta migratoria* (Orthoptera) after injection of *Bacillus thuringiensis*. *J. Invertebr. Pathol.* **1974**, *24*, 238–247. [[CrossRef](#)]
38. Dimarcq, J.-L.; Keppi, E.; Dunbar, B.; Lambert, J.; Reichart, J.-M.; Hoffmann, D.; Rankine, S.M.; Fothergill, J.E.; Hoffmann, J.A. Insect immunity. Purification and characterization of a family of novel inducible antibacterial proteins from immunized larvae of the dipteran *Phormia terranova* and complete amino-acid sequence of the predominant member, dipteracin A. *Eur. J. Biochem.* **1988**, *171*, 17–22. [[CrossRef](#)]
39. Dimarcq, J.-L.; Hoffmann, D.; Meister, M.; Bulet, P.; Lanot, R.; Reichart, J.-M.; Hoffmann, J.A. Characterization and transcriptional profiles of a *Drosophila* gene encoding an insect defensin. A study in insect immunity. *Eur. J. Biochem.* **1994**, *221*, 201–209. [[CrossRef](#)]
40. Morisalo, D.; Anderson, K.V. Signaling pathways that establish the dorsal-ventral pattern of the *drosophila* embryo. *Annu. Rev. Genet.* **1995**, *29*, 371–399. [[CrossRef](#)]
41. GAY, N.J.; KEITH, F.J. *Drosophila* Toll and IL-1 receptor. *Nature* **1991**, *351*, 355–356. [[CrossRef](#)]
42. Lemaitre, B.; Kromer-Metzger, E.; Michaut, L.; Nicolas, E.; Meister, M.; Georgel, P.; Reichart, J.M.; Hoffmann, J.A. A recessive mutation, immune deficiency (*imd*), defines two distinct control pathways in the *Drosophila* host defense. *Proc. Natl. Acad. Sci. USA* **1995**, *92*, 9465–9469. [[CrossRef](#)]
43. Janeway, C.A. Approaching the Asymptote? Evolution and Revolution in Immunology. *Cold Spring Harb. Symp. Quant. Biol.* **1989**, *54*, 1–13. [[CrossRef](#)]
44. Pfeiffer, R. Untersuchungen über das Cholera Gift. *Z. Hyg. Infekt.* **1892**, *11*, 393–412. [[CrossRef](#)]
45. Centanni, E.; Bruschetti, A. Untersuchungen über das Infektionsfieber. *DMW-Dtsch. Med. Wochenschr.* **1894**, *20*, 270–272. [[CrossRef](#)]
46. Coley, W.B. The Treatment of Inoperable Sarcoma by Bacterial Toxins (the Mixed Toxins of the *Streptococcus erysipelas* and the *Bacillus prodigiosus*). *Proc. R. Soc. Med.* **1910**, *3*, 1–48. [[CrossRef](#)]
47. Starnes, C.O. Coley's toxins in perspective. *Nature* **1992**, *357*, 11–12. [[CrossRef](#)]
48. Shear, M.J.; Turner, F.C.; Perrault, A.; Shovelton, T. Chemical Treatment of Tumors. V. Isolation of the Hemorrhage-Producing Fraction from *Serratia marcescens* (*Bacillus prodigiosus*) Culture Filtrate. *JNCI J. Natl. Cancer Inst.* **1943**, *4*, 81–97. [[CrossRef](#)]
49. Strain, S.M.; Fesik, S.W.; Armitage, I.M. Characterization of lipopolysaccharide from a heptoseless mutant of *Escherichia coli* by carbon 13 nuclear magnetic resonance. *J. Biol. Chem.* **1983**, *258*, 2906–2910. [[CrossRef](#)]
50. Beutler, B.; Rietschel, E.T. Innate immune sensing and its roots: The story of endotoxin. *Nat. Rev. Immunol.* **2003**, *3*, 169–176. [[CrossRef](#)]
51. Heppner, G.; Weiss, D.W. High Susceptibility of Strain A Mice to Endotoxin and Endotoxin-Red Blood Cell Mixtures. *J. Bacteriol.* **1965**, *90*, 696–703. [[CrossRef](#)]
52. Watson, J.; Riblet, R. Genetic control of responses to bacterial lipopolysaccharides in mice. *J. Exp. Med.* **1974**, *140*, 1147–1161. [[CrossRef](#)] [[PubMed](#)]
53. Watson, J.; Riblet, R. Genetic control of responses to bacterial lipopolysaccharides in mice. II. A gene that influences a membrane component involved in the activation of bone marrow-derived lymphocytes by lipopolysaccharides. *J. Immunol.* **1975**, *114*, 1462–1468. [[PubMed](#)]
54. Coutinho, A.; Forni, L.; Melchers, F.; Watanabe, T. Genetic defect in responsiveness to the B cell mitogen lipopolysaccharide. *Eur. J. Immunol.* **1977**, *7*, 325–328. [[CrossRef](#)] [[PubMed](#)]
55. Moore, R.N.; Goodrum, K.J.; Berry, L.J. Mediation of an endotoxic effect by macrophages. *J. Reticuloendothel. Soc.* **1976**, *19*, 187–197.
56. SIBAL, L.R. Effect of endotoxin on antibody production by chicken spleen cells transferred to chick chorioallantois. *J. Immunol.* **1961**, *87*, 362–366.
57. Davies, A.M.; Gery, I.; Rosenmann, E.; Laufer, A. Endotoxin as Adjuvant in Autoimmunity to Cardiac Tissue. *Exp. Biol. Med.* **1963**, *114*, 520–523. [[CrossRef](#)]
58. Neter, E. Endotoxins and the Immune Response. In *Current Topics in Microbiology and Immunology*; Springer: Berlin/Heidelberg, Germany, 1969; pp. 82–124.
59. O'Brien, A.D.; Rosenstreich, D.L.; Scher, I.; Campbell, G.H.; MacDermott, R.P.; Formal, S.B. Genetic control of susceptibility to *Salmonella typhimurium* in mice: Role of the LPS gene. *J. Immunol.* **1980**, *124*, 20–24.
60. Hagberg, L.; Hull, R.; Hull, S.; McGhee, J.R.; Michalek, S.M.; Svanborg Edén, C. Difference in susceptibility to gram-negative urinary tract infection between C3H/HeJ and C3H/HeN mice. *Infect. Immun.* **1984**, *46*, 839–844. [[CrossRef](#)]
61. Beutler, B.; Greenwald, D.; Hulmes, J.D.; Chang, M.; Pan, Y.-C.E.; Mathison, J.; Ulevitch, R.; Cerami, A. Identity of tumour necrosis factor and the macrophage-secreted factor cachectin. *Nature* **1985**, *316*, 552–554. [[CrossRef](#)]
62. Beutler, B.; Milsark, I.W.; Cerami, A.C. Passive Immunization Against Cachectin/Tumor Necrosis Factor Protects Mice from Lethal Effect of Endotoxin. *Science* **1985**, *229*, 869–871. [[CrossRef](#)]
63. Havell, E.A. Production of tumor necrosis factor during murine listeriosis. *J. Immunol.* **1987**, *139*, 4225–4231. [[PubMed](#)]
64. Kindler, V.; Sappino, A.-P.; Grau, G.E.; Pigué, P.-F.; Vassalli, P. The inducing role of tumor necrosis factor in the development of bactericidal granulomas during BCG infection. *Cell* **1989**, *56*, 731–740. [[CrossRef](#)]

65. Shakhov, A.N.; Collart, M.A.; Vassalli, P.; Nedospasov, S.A.; Jongeneel, C. V Kappa B-type enhancers are involved in lipopolysaccharide-mediated transcriptional activation of the tumor necrosis factor alpha gene in primary macrophages. *J. Exp. Med.* **1990**, *171*, 35–47. [[CrossRef](#)] [[PubMed](#)]
66. Tobias, P.S.; Soldau, K.; Ulevitch, R.J. Isolation of a lipopolysaccharide-binding acute phase reactant from rabbit serum. *J. Exp. Med.* **1986**, *164*, 777–793. [[CrossRef](#)] [[PubMed](#)]
67. Wright, S.D.; Tobias, P.S.; Ulevitch, R.J.; Ramos, R.A. Lipopolysaccharide (LPS) binding protein opsonizes LPS-bearing particles for recognition by a novel receptor on macrophages. *J. Exp. Med.* **1989**, *170*, 1231–1241. [[CrossRef](#)] [[PubMed](#)]
68. Schumann, R.R.; Leong, S.R.; Flaggs, G.W.; Gray, P.W.; Wright, S.D.; Mathison, J.C.; Tobias, P.S.; Ulevitch, R.J. Structure and Function of Lipopolysaccharide Binding Protein. *Science* **1990**, *249*, 1429–1431. [[CrossRef](#)]
69. Wright, S.D.; Ramos, R.A.; Tobias, P.S.; Ulevitch, R.J.; Mathison, J.C. CD14, a Receptor for Complexes of Lipopolysaccharide (LPS) and LPS Binding Protein. *Science* **1990**, *249*, 1431–1433. [[CrossRef](#)]
70. Tobias, P.S.; Soldau, K.; Ulevitch, R.J. Identification of a Lipid A Binding Site in the Acute Phase Reactant Lipopolysaccharide Binding Protein. *J. Biol. Chem.* **1989**, *264*, 10867–10871. [[CrossRef](#)]
71. Haziot, A.; Ferrero, E.; Köntgen, F.; Hijiya, N.; Yamamoto, S.; Silver, J.; Stewart, C.L.; Goyert, S.M. Resistance to Endotoxin Shock and Reduced Dissemination of Gram-Negative Bacteria in CD14-Deficient Mice. *Immunity* **1996**, *4*, 407–414. [[CrossRef](#)]
72. Medzhitov, R.; Preston-Hurlburt, P.; Janeway, C.A. A human homologue of the Drosophila Toll protein signals activation of adaptive immunity. *Nature* **1997**, *388*, 394–397. [[CrossRef](#)]
73. Nomura, N.; Miyajima, N.; Sazuka, T.; Tanaka, A.; Kawarabayasi, Y.; Sato, S.; Nagase, T.; Seki, N.; Ishikawa, K.-i.; Tabata, S. Prediction of the Coding Sequences of Unidentified Human Genes. I. The Coding Sequences of 40 New Genes (KIAA0001-KIAA0040) Deduced by Analysis of Randomly Sampled cDNA Clones from Human Immature Myeloid Cell Line KG-1. *DNA Res.* **1994**, *1*, 27–35. [[CrossRef](#)]
74. Rock, F.L.; Hardiman, G.; Timans, J.C.; Kastelein, R.A.; Bazan, J.F. A family of human receptors structurally related to Drosophila Toll. *Proc. Natl. Acad. Sci. USA* **1998**, *95*, 588–593. [[CrossRef](#)] [[PubMed](#)]
75. SULTZER, B.M. Genetic Control of Leucocyte Responses to Endotoxin. *Nature* **1968**, *219*, 1253–1254. [[CrossRef](#)] [[PubMed](#)]
76. Rosenstreich, D.L.; Michael Glade, L.; Mergenhagen, S.E. Action of Endotoxin on Lymphoid Cells. *J. Infect. Dis.* **1977**, *136*, S239–S245. [[CrossRef](#)] [[PubMed](#)]
77. Coutinho, A.; Meo, T. Genetic basis for unresponsiveness to lipopolysaccharide in C57BL/10Cr mice. *Immunogenetics* **1978**, *7*, 17–24. [[CrossRef](#)]
78. Peppel, K.; Crawford, D.; Beutler, B. A tumor necrosis factor (TNF) receptor-IgG heavy chain chimeric protein as a bivalent antagonist of TNF activity. *J. Exp. Med.* **1991**, *174*, 1483–1489. [[CrossRef](#)]
79. Takeuchi, O.; Hoshino, K.; Kawai, T.; Sanjo, H.; Takada, H.; Ogawa, T.; Takeda, K.; Akira, S. Differential Roles of TLR2 and TLR4 in Recognition of Gram-Negative and Gram-Positive Bacterial Cell Wall Components. *Immunity* **1999**, *11*, 443–451. [[CrossRef](#)]
80. Takeuchi, O.; Kawai, T.; Mühlradt, P.F.; Morr, M.; Radolf, J.D.; Zychlinsky, A.; Takeda, K.; Akira, S. Discrimination of bacterial lipoproteins by Toll-like receptor 6. *Int. Immunol.* **2001**, *13*, 933–940. [[CrossRef](#)]
81. Hemmi, H.; Takeuchi, O.; Kawai, T.; Kaisho, T.; Sato, S.; Sanjo, H.; Matsumoto, M.; Hoshino, K.; Wagner, H.; Takeda, K.; et al. A Toll-like receptor recognizes bacterial DNA. *Nature* **2000**, *408*, 740–745. [[CrossRef](#)]
82. Alexopoulou, L.; Holt, A.C.; Medzhitov, R.; Flavell, R.A. Recognition of double-stranded RNA and activation of NF- κ B by Toll-like receptor 3. *Nature* **2001**, *413*, 732–738. [[CrossRef](#)]
83. Hayashi, F.; Smith, K.D.; Ozinsky, A.; Hawn, T.R.; Yi, E.C.; Goodlett, D.R.; Eng, J.K.; Akira, S.; Underhill, D.M.; Aderem, A. The innate immune response to bacterial flagellin is mediated by Toll-like receptor 5. *Nature* **2001**, *410*, 1099–1103. [[CrossRef](#)] [[PubMed](#)]
84. Heil, F.; Hemmi, H.; Hochrein, H.; Ampenberger, F.; Kirschning, C.; Akira, S.; Lipford, G.; Wagner, H.; Bauer, S. Species-Specific Recognition of Single-Stranded RNA via Toll-like Receptor 7 and 8. *Science* **2004**, *303*, 1526–1529. [[CrossRef](#)] [[PubMed](#)]
85. Guan, Y.; Ranoa, D.R.E.; Jiang, S.; Mutha, S.K.; Li, X.; Baudry, J.; Tapping, R.I. Human TLRs 10 and 1 Share Common Mechanisms of Innate Immune Sensing but Not Signaling. *J. Immunol.* **2010**, *184*, 5094–5103. [[CrossRef](#)] [[PubMed](#)]
86. Yarovsky, F.; Zhang, D.; Andersen, J.F.; Bannenberg, G.L.; Serhan, C.N.; Hayden, M.S.; Hiemy, S.; Sutterwala, F.S.; Flavell, R.A.; Ghosh, S.; et al. TLR11 activation of dendritic cells by a protozoan profilin-like protein. *Science* **2005**, *308*, 1626–1629. [[CrossRef](#)]
87. Hatai, H.; Lepelley, A.; Zeng, W.; Hayden, M.S.; Ghosh, S. Toll-Like Receptor 11 (TLR11) Interacts with Flagellin and Profilin through Disparate Mechanisms. *PLoS ONE* **2016**, *11*, e0148987. [[CrossRef](#)]
88. Oldenburg, M.; Krüger, A.; Ferstl, R.; Kaufmann, A.; Nees, G.; Sigmund, A.; Bathke, B.; Lauterbach, H.; Suter, M.; Dreher, S.; et al. TLR13 Recognizes Bacterial 23 S rRNA Devoid of Erythromycin Resistance-Forming Modification. *Science* **2012**, *337*, 1111–1115. [[CrossRef](#)]
89. Jin, M.S.; Kim, S.E.; Heo, J.Y.; Lee, M.E.; Kim, H.M.; Paik, S.-G.; Lee, H.; Lee, J.-O. Crystal Structure of the TLR1-TLR2 Heterodimer Induced by Binding of a Tri-Acylated Lipopeptide. *Cell* **2007**, *130*, 1071–1082. [[CrossRef](#)]
90. Andrade, W.A.; do Carmo Souza, M.; Ramos-Martinez, E.; Nagpal, K.; Dutra, M.S.; Melo, M.B.; Bartholomeu, D.C.; Ghosh, S.; Golenbock, D.T.; Gazzinelli, R.T. Combined Action of Nucleic Acid-Sensing Toll-like Receptors and TLR11/TLR12 Heterodimers Imparts Resistance to *Toxoplasma gondii* in Mice. *Cell Host Microbe* **2013**, *13*, 42–53. [[CrossRef](#)]
91. Rosenstiel, P.; Till, A.; Schreiber, S. NOD-like receptors and human diseases. *Microbes Infect.* **2007**, *9*, 648–657. [[CrossRef](#)]

92. Inohara, N.; Koseki, T.; del Peso, L.; Hu, Y.; Yee, C.; Chen, S.; Carrio, R.; Merino, J.; Liu, D.; Ni, J.; et al. Nod1, an Apaf-1-like Activator of Caspase-9 and Nuclear Factor- κ B. *J. Biol. Chem.* **1999**, *274*, 14560–14567. [[CrossRef](#)]
93. Rehwinkel, J.; Gack, M.U. RIG-I-like receptors: Their regulation and roles in RNA sensing. *Nat. Rev. Immunol.* **2020**, *20*, 537–551. [[CrossRef](#)] [[PubMed](#)]
94. Yoneyama, M.; Kikuchi, M.; Natsukawa, T.; Shinobu, N.; Imaizumi, T.; Miyagishi, M.; Taira, K.; Akira, S.; Fujita, T. The RNA helicase RIG-I has an essential function in double-stranded RNA-induced innate antiviral responses. *Nat. Immunol.* **2004**, *5*, 730–737. [[CrossRef](#)] [[PubMed](#)]
95. Kutikhin, A.; Yuzhalin, A. C-type lectin receptors and RIG-I-like receptors: New points on the oncogenomics map. *Cancer Manag. Res.* **2012**, *4*, 39–53. [[CrossRef](#)] [[PubMed](#)]
96. Neth, O.; Jack, D.L.; Dodds, A.W.; Holzel, H.; Klein, N.J.; Turner, M.W. Mannose-Binding Lectin Binds to a Range of Clinically Relevant Microorganisms and Promotes Complement Deposition. *Infect. Immun.* **2000**, *68*, 688–693. [[CrossRef](#)]
97. Wesche, H.; Henzel, W.J.; Shillinglaw, W.; Li, S.; Cao, Z. MyD88: An Adapter That Recruits IRAK to the IL-1 Receptor Complex. *Immunity* **1997**, *7*, 837–847. [[CrossRef](#)]
98. O’Neill, L.A.J.; Bowie, A.G. The family of five: TIR-domain-containing adaptors in Toll-like receptor signalling. *Nat. Rev. Immunol.* **2007**, *7*, 353–364. [[CrossRef](#)]
99. Yamamoto, M.; Sato, S.; Mori, K.; Hoshino, K.; Takeuchi, O.; Takeda, K.; Akira, S. Cutting Edge: A Novel Toll/IL-1 Receptor Domain-Containing Adapter That Preferentially Activates the IFN- β Promoter in the Toll-Like Receptor Signaling. *J. Immunol.* **2002**, *169*, 6668–6672. [[CrossRef](#)]
100. Oshiumi, H.; Matsumoto, M.; Funami, K.; Akazawa, T.; Seya, T. TICAM-1, an adaptor molecule that participates in Toll-like receptor 3-mediated interferon- β induction. *Nat. Immunol.* **2003**, *4*, 161–167. [[CrossRef](#)]
101. Hoebe, K.; Du, X.; Georgel, P.; Janssen, E.; Tabeta, K.; Kim, S.O.; Goode, J.; Lin, P.; Mann, N.; Mudd, S.; et al. Identification of Lps2 as a key transducer of MyD88-independent TIR signalling. *Nature* **2003**, *424*, 743–748. [[CrossRef](#)]
102. Yamamoto, M.; Sato, S.; Hemmi, H.; Hoshino, K.; Kaisho, T.; Sanjo, H.; Takeuchi, O.; Sugiyama, M.; Okabe, M.; Takeda, K.; et al. Role of Adaptor TRIF in the MyD88-Independent Toll-Like Receptor Signaling Pathway. *Science* **2003**, *301*, 640–643. [[CrossRef](#)]
103. Fitzgerald, K.A.; Rowe, D.C.; Barnes, B.J.; Caffrey, D.R.; Visintin, A.; Latz, E.; Monks, B.; Pitha, P.M.; Golenbock, D.T. LPS-TLR4 Signaling to IRF-3/7 and NF- κ B Involves the Toll Adapters TRAM and TRIF. *J. Exp. Med.* **2003**, *198*, 1043–1055. [[CrossRef](#)] [[PubMed](#)]
104. Yamamoto, M.; Sato, S.; Hemmi, H.; Uematsu, S.; Hoshino, K.; Kaisho, T.; Takeuchi, O.; Takeda, K.; Akira, S. TRAM is specifically involved in the Toll-like receptor 4-mediated MyD88-independent signaling pathway. *Nat. Immunol.* **2003**, *4*, 1144–1150. [[CrossRef](#)] [[PubMed](#)]
105. Fitzgerald, K.A.; Palsson-McDermott, E.M.; Bowie, A.G.; Jefferies, C.A.; Mansell, A.S.; Brady, G.; Brint, E.; Dunne, A.; Gray, P.; Harte, M.T.; et al. Mal (MyD88-adaptor-like) is required for Toll-like receptor-4 signal transduction. *Nature* **2001**, *413*, 78–83. [[CrossRef](#)] [[PubMed](#)]
106. Kawasaki, T.; Kawai, T. Toll-Like Receptor Signaling Pathways. *Front. Immunol.* **2014**, *5*, 461. [[CrossRef](#)] [[PubMed](#)]
107. O’Neill, L.A.J.; Golenbock, D.; Bowie, A.G. The history of Toll-like receptors—Redefining innate immunity. *Nat. Rev. Immunol.* **2013**, *13*, 453–460. [[CrossRef](#)] [[PubMed](#)]
108. Essuman, K.; Summers, D.W.; Sasaki, Y.; Mao, X.; DiAntonio, A.; Milbrandt, J. The SARM1 Toll/Interleukin-1 Receptor Domain Possesses Intrinsic NAD⁺ Cleavage Activity that Promotes Pathological Axonal Degeneration. *Neuron* **2017**, *93*, 1334–1343.e5. [[CrossRef](#)]
109. Essuman, K.; Milbrandt, J.; Dangl, J.L.; Nishimura, M.T. Shared TIR enzymatic functions regulate cell death and immunity across the tree of life. *Science* **2022**, *377*, 6614. [[CrossRef](#)]