



Diverse Functions of $\gamma \delta$ T Cells in the Progression of Hepatitis B Virus and Hepatitis C Virus Infection

Wen Hou^{1,2} and Xiaoli Wu^{3*}

¹ Key Laboratory for Critical Care Medicine of the Ministry of Health, Tianjin First Central Hospital, Tianjin, China, ² State Key Laboratory of Medicinal Chemical Biology, Nankai University, Tianjin, China, ³ School of Life Sciences, Tianjin University, Tianjin, China

Hepatitis B virus (HBV) and hepatitis C virus (HCV) infections are primary risk factors for a wide spectrum of liver diseases that severely affect human health. The liver is an immunological organ that has an abundance of immune cells. Thus, various innate or adaptive immune cells are involved in the progression of HBV or HCV infection. Among those cells, a unique kind of immune cell, the $\gamma\delta$ T cell, contributes to promoting or inhibiting the progression of liver diseases. To reveal the diverse roles of $\gamma\delta$ T cells in HBV or HCV infection, the properties and functions of these cells in human and mouse models are analyzed. Here, we briefly describe the characteristics and functions of $\gamma\delta$ T cells in the progression of HBV or HCV infection, including stages of acute infection, chronic infection, liver cirrhosis, and hepatocellular carcinoma. Finally, the functions and existing problems of $\gamma\delta$ T cells in HBV or HCV infection are summarized. A better understanding of the function of $\gamma\delta$ T cells during the progression of HBV and HCV infection will be helpful for the treatment of virus infection.

Keywords: $\gamma\delta$ T cells, hepatitis B virus, hepatitis C virus, progression, cytokines

*Correspondence: Xiaoli Wu wuxiaoli@tju.edu.cn

Argentina

OPEN ACCESS

Jinan University, China

Carolina Cristina Jancic.

Université libre de Bruxelles, Belgium

Hannover Medical School, Germany

Consejo Nacional de Investigaciones

Científicas y Técnicas (CONICET),

Edited by:

Jianlei Hao.

Reviewed by: David Vermijlen,

Sarina Ravens.

Specialty section:

This article was submitted to T Cell Biology, a section of the journal Frontiers in Immunology

Received: 21 October 2020 Accepted: 17 December 2020 Published: 01 February 2021

Citation:

Hou W and Wu X (2021) Diverse Functions of γδ T Cells in the Progression of Hepatitis B Virus and Hepatitis C Virus Infection. Front. Immunol. 11:619872. doi: 10.3389/fimmu.2020.619872 INTRODUCTION

Hepatitis B virus (HBV) and hepatitis C virus (HCV) infections are major risk factors for a wide spectrum of liver diseases. Although most adults recover from HBV infection, about 5% of patients are unable to clear HBV and thus develop chronic HBV infection (1) and experience virus flares and long-term morbidity. Similarly, acute HCV infection can easily convert into chronic HCV infection (2). The persistent inflammatory environment in chronic HBV (CHB) or chronic HCV (CHC) infection patients is associated with the elevated expression of α -smooth muscle actin and collagen fibers in hepatic stellate cells (HSCs), which then develop into liver cirrhosis (2–4). Hepatocellular carcinoma (HCC) is a common cancer and is mainly caused by HBV or HCV infection. HCV patients show a higher probability of developing HCC than HBV patients (5).

The liver is known as an immune tolerance organ. Aside from hepatocytes and stellate cells, there are various hepatic residential immune cells, including Küpffer cells (hepatic macrophages), T cells, natural killer (NK) cells, and dendritic cells (6). These cells play crucial roles in the pathogenesis of HBV or HCV infection. During acute HBV or HCV infection, innate immune cells such as NK cells

1

are activated and further induce antiviral function of adaptive immune cells (7). In chronic HBV and HCV infections, the liver is infiltrated with impaired antiviral T cells and activated inflammatory cells such as IL-17-producing CD4⁺ T cells that further exacerbate liver inflammation (8, 9). Moreover, other hepatic immune cells, including regulatory T cells and myeloidderived suppressor cells (MDSC), prompt the pathogenesis of chronic HBV or HCV infection, liver cirrhosis, or even liver cancer (10). The proportion of hepatic $\gamma\delta$ T cells in hepatic T cells in humans and mice is found to be 15%–25% and 4.5%, respectively (6, 11), indicating the crucial role of these cells in liver diseases. However, the current understanding of the function of $\gamma\delta$ T cells compared with other immune cells in HBV or HCV infection is limited.

 $\gamma \delta$ T cells, as the bridge of innate and adaptive immunity, play critical roles in various diseases, including liver diseases, infections, and cancer. yo T cells can be divided into different subsets through γ and δ TCR chains. Based on δ TCR chains, human $\gamma\delta$ T cells can mainly be separated into V δ 1 (in peripheral blood or organs), V δ 2 (peripheral blood dominant $\gamma\delta$ T cells, usually combined with $V\gamma 9$), and $V\delta 3$ (in intestine and lamina propria) T cell subsets. Based on γ TCR chains, mouse $\gamma\delta$ T cells can be divided into V γ 1, Vy4, Vy5, Vy6, and Vy7 T cell subsets (12). In liver diseases, hepatic $\gamma\delta$ T cells usually include V γ 1, V γ 4, and V γ 6 in mice and V δ 1, V δ 2, and V δ 3 in humans (13–15). These cells can produce cytokines such as IFN-y, TNF-a, IL-17, and IL-22, as well as express cytotoxic and regulatory molecules such as Granzyme B (GrB), perforin, NK receptor, and Toll-like receptors (16). γδ T cells play different roles in the pathogenesis of HBV and HCV infections. In acute HBV infection, human $\gamma\delta$ T cells are activated and exhibit antiviral functions by secreting IFN- γ and TNF- α . During other stages of HBV and HCV infections (chronic infection, liver cirrhosis, and HCC), these cells can inhibit or promote progression of the diseases. Surprisingly, different subsets of $\gamma\delta$ T cells play contradictory roles in the same stage of liver infection. For example, in chronic HBV infection, human Vδ2 T cell subsets inhibit HBV infection progression by inhibiting Th17induced liver damage (17). However, human CD4⁻CD8⁻ $\gamma\delta$ T cell (18) and mouse IL-17-producing Vy4 T cell (19) subsets are found to inhibit the function of T cells and promote HBV infection in CHB patients and an HBV mouse model. Similar contradictory functions are also observed in other stages. In HCC, human V82 T cells, which can be activated and proliferate in vitro (20), are used in the clinic to prolong the survival time of HCC patients (21).

To determine the precise role of these cells, we summarize the functions of different human and mouse $\gamma\delta$ T cells subsets in the different stages of HBV and HCV infections. Moreover, we indicate the opportunities and challenges in clinical application of $\gamma\delta$ T cells.

ROLE OF $\gamma\delta$ T CELLS IN ACUTE AND CHRONIC HBV INFECTION

During human acute HBV infection, about 5% of adult patients progress to chronic hepatitis B infection, whereas the rest go

through a self-limited process that results in recovery (1). Accumulating data have demonstrated that different outcomes of HBV infection are associated with the intensity of antiviral immune responses (22). As shown in our previous study, the numbers of $\gamma\delta$ T cells increase in liver tissue, but decrease in the peripheral blood of acute hepatitis B (AHB) patients (3). These peripheral $\gamma\delta$ T cells are highly activated and terminally differentiated into memory phenotype, which has increased cytotoxic capacity and enhanced antiviral activity. Interestingly, in asymptomatic HBV infection patients, the frequencies of peripheral Vo1 and Vo2 T cells are higher, and the level of peripheral IFN- γ^+ V δ 2 T cells is also significantly elevated compared to healthy controls (23). Furthermore, in an AHB infection mouse model, the number of hepatic $\gamma\delta$ T cells significantly increases with the upregulation of HBV markers and exhibits elevated expression of the activation marker CD69, IFN- γ production, and IFN- β mRNA abundance in liver tissues (24). The above studies indicate that the antiviral function of $\gamma\delta$ T cells in AHB patients can inhibit the progression of AHB infection.

 $\gamma\delta$ T cells display contradictory roles in CHB infection. Several studies have shown that these cells are impaired and exhibit liver protective functions to inhibit the progression of CHB infection (17). Our study and others show that the frequency of human peripheral and hepatic V δ 2 T cells is significantly lower in severe CHB patients with impaired chemotaxis (17) or degranulation (25). Although they display an active effector-memory phenotype (17), the IFN- γ or TNF- α induced cytotoxicity of V δ 2 T cells is impaired (26) and can be reversed by IFN- α treatment in vitro and in vivo (27). In addition, in vitro proliferated human V82 T cells can inhibit inflammatory cytokines production in pathogenic Th17 cells (17), which contributes to significant liver damage and pathology. However, a recent study indicates that the frequency of human $\gamma\delta$ T cells and their subsets barely change and antiviral function of V δ 2 T cells is enhanced in CHB patients (28). This opposite result maybe because of the different applied standard for patient enrollment, including age, gender, and race, which would interfere the characteristics of $\gamma\delta$ T cells (29).

However, other studies report that $\gamma\delta$ T cells promote the progression of chronic HBV infection. By suppressing the secretion of HBV core peptide-stimulated IFN- γ and TNF- α by CD8⁺ T cells, human CD4⁻CD8⁻ $\gamma\delta$ T cells limit T cell responses to HBV partially through NKG2A and may impede HBeAg seroconversion during antiviral therapy of CHB patients (18). Moreover, in HBV-associated acute-on-chronic liver failure (CHB-ACLF) patients, more human peripheral $\gamma\delta$ T cells exhibit upregulation of TNF- α or IL-17 and GrB or CD107, demonstrating the participation of $\gamma\delta$ T cells in liver injury which in turn promote the progression of liver diseases (30). Meanwhile, in an immune tolerance chronic HBV infection mouse model, IL-17-producing V γ 4 T cells recruit MDSCs into the liver and induce CD8⁺ T cell exhaustion (19).

In conclusion, IFN- γ - or TNF- α -producing $\gamma\delta$ T cells can inhibit AHB and CHB infection, while human CD4⁻CD8⁻ $\gamma\delta$ T cells and mouse IL-17-producing V $\gamma4$ T cell subsets promote the

progression of chronic HBV infection. The opposite roles of these cells can be attributed to the different subsets of $\gamma\delta$ T cells and their variable cytokine production (IFN- γ , TNF- α , or IL-17).

ROLE OF $\gamma\delta$ T CELLS IN CHRONIC HCV INFECTION

Numerous researchers have focused on the function of $\gamma\delta$ T cells in chronic HCV (CHC) infection. The number of hepatic $\gamma\delta$ T cells is higher in CHC patients, and V δ 1 T cells are the predominant subset of hepatic $\gamma\delta$ T cells (31, 32). However, the number of peripheral V γ 9V δ 2 and V δ 1 T cells decrease in CHC patients compared with healthy control and asymptomatic HCV carriers (33). Moreover, in mice, the level of hepatic $\gamma\delta$ T cells is significantly higher in HCV transgenic mice compared with wild-type mice (34). It is assumed that peripheral $\gamma\delta$ T cells are recruited into the liver and contribute to the pathogenesis of HCV infection.

 $\gamma\delta$ T cells play different roles in the pathogenesis of CHC infection. In some studies, $\gamma\delta$ T cells manifest their antiviral role and inhibit the progression of CHC infection. In CHC patients, the cytotoxicity of hepatic $\gamma\delta$ T cells is higher than that of hepatic $\alpha\beta$ T cells. This is attributable to their elevated secretion of IFN- γ , TNF- α , and IL-8 (31) and their expression of activation marker (human leukocyte antigen-DR) and memory/effector (CD62L⁻CD45RO⁺ CD95⁺) marker (32). In particular, the frequency of human hepatic IFN- γ^+ V δ 1 T cells is positively correlated with the degree of liver necroinflammation, indicating their involvement in liver pathogenesis and liver damage (32). Furthermore, the expression of CD56 and CD16 (markers of natural killer cells) increase in peripheral V γ 9V δ 2 T cells and is further enhanced in hepatic V γ 9V δ 2 T cells of CHC patients (35). In humans, after stimulation by non-peptide antigen-isopentenyl diphosphate (IPP), activated peripheral Vγ9Vδ2 T cells are associated with a dramatic reduction in HCV RNA levels. Neutralizing experiments have further revealed the function of IFN- γ in HCV clearance (36). Moreover, in a mouse model, the number of hepatic $\gamma\delta$ T cells increases and activated CD69⁺ $\gamma\delta$ T cells produce more IFN- γ and TNF- α during MHV (mouse hepatitis virus) infection than controls. Interestingly, those activated hepatic $\gamma\delta$ T cells can kill MHV-infected hepatocytes in vitro by secreting IFN-y and TNFα (37).

However, several studies have indicated that human peripheral $\gamma\delta$ T cells exhibit impaired function in CHC patients even after antiviral treatment. Human peripheral V γ 9V δ 2 T cells are activated and differentiate into effector cells with upregulated GrB and perforin expression, but have a markedly impaired capacity to produce IFN- γ in CHC patients (38). Furthermore, IFN- α treatments result in the upregulation of cytotoxic markers such as GrB, perforin, and CD107a, but not the IFN- γ production capacity of peripheral V γ 9V δ 2 T cells in CHC patients (35, 38). The above results suggest a functional dichotomy of V γ 9V δ 2 T cells in chronic HCV infections that contribute to both liver inflammation and HCV persistence. Moreover, dysfunction of $\gamma\delta$ T cells in CHC patients has also been observed in antiviral therapy. Direct-active antiviral agents (DAAs) are widely used in the treatment of chronic HCV infection. In clinical trials, DAAs have induced minor changes in $\gamma\delta$ T cells both in terms of numbers and in alterations of TRG and TRD repertoires 1 year after treatment (39). Although human peripheral $V\gamma9V\delta2$ T cells display an elevated effector phenotype in sustained virologic-response HCV patients, recent DAA treatment research demonstrates that these cells show poor cytokine response and proliferative responses to antigens (40).

In summary, human and mouse hepatic $\gamma\delta$ T cells as well as *in vitro* stimulated human peripheral V γ 9V δ 2 T cells can inhibit HCV pathogenesis. However, impaired cytokine response of peripheral V γ 9V δ 2 T cells in CHC patients contributes to HCV infection progression, even after DAA treatment. Further studies on recovery from the cytokine response impairment of V γ 9V δ 2 T cells is very important for CHC treatment.

ROLE OF $\gamma\delta$ T CELLS IN LIVER CIRRHOSIS AND HCC

Persistent inflammation of HBV or HCV can lead to liver fibrosis and liver cirrhosis. HSCs are critical cells in the pathogenesis of liver cirrhosis. Activation of these cells promote the progression of liver cirrhosis (41). A liver cirrhosis mouse model shows different relationships between HSCs and hepatic $\gamma\delta$ T cells. IL-17-producing CCR6⁺ $\gamma\delta$ T cells induce apoptosis of HSCs in a FasL-dependent manner to inhibit the progression of liver cirrhosis (42). Moreover, IFN- γ -producing $\gamma\delta$ T cells can directly kill activated HSCs and increase NK cell-mediated cytotoxicity against activated HSCs partially through a 4-1BB dependent manner (43). However, hepatocyte-secreted exosomes can activate HSCs via Toll-like receptor 3. These HSCs further enhance the activity of IL-17-producing $\gamma\delta$ T cells, which exacerbates liver fibrosis and promotes the progression of liver cirrhosis (4). In view of the contradictory roles of IL-17-producing $\gamma\delta$ T cells in the same mouse model, further studies involving patients and a virus-induced liver cirrhosis mouse model should be performed to elucidate the exact role of $\gamma\delta$ T cells.

A recent study has shown that the increased peritumor ratio in human $\gamma\delta$ T cells contributes to the progression and recurrence of HCC, indicating the important role of $\gamma\delta$ T cells in HCC (44). Interestingly, $\gamma\delta$ T cells play different roles in the pathogenesis of HCC. In several studies, $\gamma\delta$ T cells display cytotoxicity and inhibit proliferation of tumor cells *in vivo* and *in vitro*. In HCC patients, the number of human peritumoral $\gamma\delta$ T cells is positively related to better prognosis of HCC curative resection (45). A recent biostatistics study has shown that the increase of human tumor-infiltrated $\gamma\delta$ T cells, which is driven by the accumulation of chemokines such as CCL4 and CCL5, is significantly positively correlated with the survival rate and negatively correlated with HCC recurrence. $\gamma\delta$ T cells play protective roles by regulating the infiltration and differentiation of CD8⁺ T cells in HCC procession (46). Furthermore, human $\gamma\delta$ T cells can induce the death of HCC cell lines and reverse the immune escape of HCC *in vitro* (47). Moreover, the anti-HCC function of peripheral $\gamma\delta$ T cells, especially V γ 9V δ 2 T cells, can be further enhanced by activating agents, including histone deacetylase inhibitors (48), pyrophosphate (49), zoledronate (20), CD226 (50), and even the Chinese herb artesunate (51).

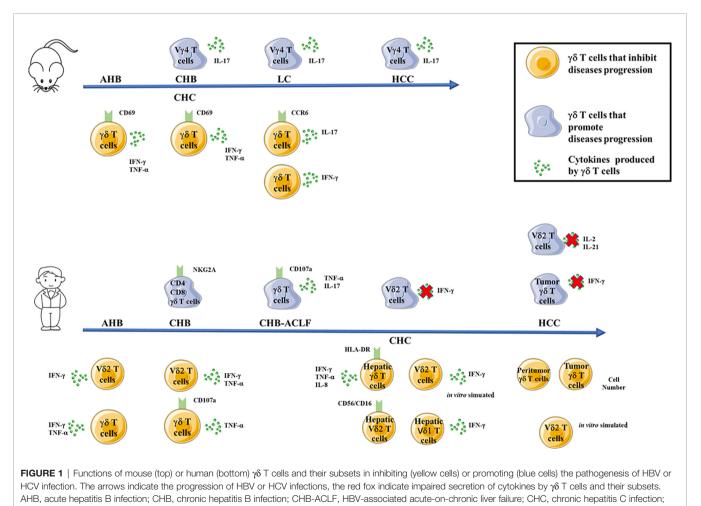
However, other studies reveal that impaired human $\gamma\delta$ T cells or mouse $\gamma\delta$ T cells can also contribute to the progression of HCC. In an immunosuppressed tumor microenvironment, $\gamma\delta$ T cells show impaired IFN- γ production and degranulation (perforin and CD107a) capacity, which is attributed to the secretion of TGF- β and IL-10 by tumor-infiltrating Tregs (52). In addition, a decrease in the number and cytotoxicity of peripheral V δ 2 T cells is observed in HCC patients and possibly associated with the lack of IL-2 and IL-21 (53). The total number of $\gamma\delta$ T cells and effector $\gamma\delta$ T cells is significantly lower in tumors than in peritumoral tissues and non-tumor livers (52, 54). In addition, in an HCC mouse model, IL-17producing V γ 4 T cells recruit MDSCs in a CXCL5/CXCR2dependent manner and further suppress the anti-tumor function of CD8⁺ T cells (55). Human peripheral V δ 2 T cells can proliferate *in vitro* and kill HCC and thus have been used in clinical immunotherapy of HCC patients. Zoledronate induces the proliferation of $\gamma\delta$ T cells in HCC patients who exhibit upregulated expression of IFN- γ , TNF- α , GrB, perforin, and lysosome-associated membrane protein 1 (47). A clinical trial has shown that the combined use of $\gamma\delta$ T cells, NK cells, and cytokine-induced killer (CIK) therapy significantly inhibits virus replication and prolongs the survival rate of HCV-positive HCC patients (21).

In conclusion, $\gamma\delta$ T cells and their subsets play opposite roles in liver cancer, and their underlying mechanisms require further investigation.

CONCLUSIONS AND PERSPECTIVES

Different subsets of $\gamma\delta$ T cells play various roles in pathogenesis of HBV or HCV infection. Most of the mouse and human studies are summarized in **Figure 1**.

In mouse model, IL-17-producing Vy4 T cells subsets promote the progression of CHB, LC and HCC. However, in



LC, liver cirrhosis; HCC, hepatocellular carcinoma.

other studies, IFN-γ and TNF-α-producing CD69⁺ mouse γδ T cells can inhibit the progression of AHB and CHC. Furthermore, IL-17-producing CCR6⁺ mouse γδ T cells or IFN-γ producing mouse γδ T cells inhibit the progression of LC. (**Figure 1**, top).

In human studies (Figure 1, bottom), $CD4^{-}CD8^{-}\gamma\delta$ T cells subsets and IL-17/TNF-a⁺ $\gamma\delta$ T cells promote the progression of CHB and CHB-ACLF patients, respectively. Impairment secretion of IFN- γ by peripheral V δ 2 T cells contributes to the progression of CHC. Moreover, impairment secretions of IL-2 and IL-21 by peripheral V δ 2 T cells and IFN- γ by tumorinfiltrating $\gamma\delta$ T cells contribute to the progression of HCC. Contradictorily, in AHB patients, IFN-y-producing peripheral V δ 2 T cells and IFN- γ and TNF- α -producing peripheral $\gamma\delta$ T cells can inhibit AHB infection. In addition, IFN-y and TNF-aproducing peripheral V δ 2 T cells and TNF- α -producing CD107a⁺ peripheral $\gamma\delta$ T cells inhibit the progression of CHB infection. Furthermore, hepatic $\gamma\delta$ T cells as well as *in vitro* activated peripheral V\delta2 T cells inhibit the progression of CHC infection. Furthermore, increased number of peritumor and tumor $\gamma\delta$ T cells as well as *in vitro* activated peripheral V $\delta2$ T cells inhibit the progression of HCC (Figure 1, bottom).

Although functions of $\gamma\delta$ T cells are summarized above, some of their roles in virus infection remain obscure. For instance, IL-17-producing V γ 4 T cells display diverse roles to influence the development of liver cirrhosis in the same mouse model. Furthermore, the role of human peripheral $\gamma\delta$ T cells but not hepatic $\gamma\delta$ T cells has been extensively studied. Thus, the impact of cytokine production and the functions of hepatic $\gamma\delta$ T cell subsets in the pathogenesis of HBV and HCV infections require further investigation. The frequency and function of $\gamma\delta$ T cells

REFERENCES

- Rajoriya N, Fergusson JR, Leithead JA, Klenerman P. Gamma Delta Tlymphocytes in Hepatitis C and Chronic Liver Disease. *Front Immunol* (2014) 5:400. doi: 10.3389/fimmu.2014.00400
- Lee HM, Banini BA. Updates on Chronic HBV: Current Challenges and Future Goals. Curr Treat Options Gastroenterol (2019) 17:271–91. doi: 10.1007/s11938-019-00236-3
- Jia ZH, Li YY, Wang JY, Zhang JY, Huang A, Guo XD, et al. Activated gammadelta T cells exhibit cytotoxicity and the capacity for viral clearance in patients with acute hepatitis B. *Clin Immunol* (2019) 202:40–8. doi: 10.1016/ j.clim.2019.03.005
- Seo W, Eun HS, Kim SY, Yi HS, Lee YS, Park SH, et al. Exosome-mediated activation of toll-like receptor 3 in stellate cells stimulates interleukin-17 production by gammadelta T cells in liver fibrosis. *Hepatology* (2016) 64:616– 31. doi: 10.1002/hep.28644
- Lafaro KJ, Demirjian AN, Pawlik TM. Epidemiology of hepatocellular carcinoma. Surg Oncol Clinics North America (2015) 24(1):1–17. doi: 10.1016/j.soc.2014.09.001
- Racanelli V, Rehermann B. The liver as an immunological organ. *Hepatology* (2006) 43:S54–62. doi: 10.1002/hep.21060
- Shin EC, Sung PS, Park SH. Immune responses and immunopathology in acute and chronic viral hepatitis. Nat Rev Immunol (2016) 16:509–23. doi: 10.1038/nri.2016.69
- Saeidi A, Zandi K, Cheok YY, Saeidi H, Wong WF, Lee CYQ, et al. T-Cell Exhaustion in Chronic Infections: Reversing the State of Exhaustion and Reinvigorating Optimal Protective Immune Responses. *Front Immunol* (2018) 9:2569. doi: 10.3389/fimmu.2018.02569
- Zhang JY, Zhang Z, Lin F, Zou ZS, Xu RN, Jin L, et al. Interleukin-17producing CD4(+) T cells increase with severity of liver damage in patients with chronic hepatitis B. *Hepatology* (2010) 51:81–91. doi: 10.1002/hep.23273

can be distinguished based on human race, age, and gender, thus these factors have to be considered in related researches (28, 29). Asian Americans display two- to three-fold higher number of peripheral V δ 2 T cells compared to non-Asian Americans (28), which in turn may partially contribute to the immune responses and outcome of virus infection. Moreover, the fate of transferred $\gamma\delta$ T cells in the human body as well as the indication and race of liver cancer patients should be assessed to achieve better therapeutic outcomes during treatment. Last but not least, in view of their antiviral function, IFN- γ -producing $\gamma\delta$ T cell-based therapies should be developed for patients in stages of virus infection other than HCC. Understanding the roles of $\gamma\delta$ T cells in relation to the pathogenesis of HBV and HCV infections may facilitate in the development of $\gamma\delta$ T cell-based therapy or $\gamma\delta$ T cell-based targets for the treatment of virus infections.

AUTHOR CONTRIBUTIONS

WH wrote the main part of the review. XW wrote the Introduction and revised the manuscript. All authors contributed to the article and approved the submitted version.

FUNDING

This work was supported by the Natural Science Foundation of Tianjin City (18JCZDJC34500), the State Key Laboratory of Medicinal Chemical Biology, Nankai University (2019003), and Tianjin First Central Hospital Spring Funding (CM201813).

- Li TY, Yang Y, Zhou G, Tu ZK. Immune suppression in chronic hepatitis B infection associated liver disease: A review. World J Gastroenterol (2019) 25:3527–37. doi: 10.3748/wjg.v25.i27.3527
- Kohlgruber AC, Gal-Oz ST, LaMarche NM, Shimazaki M, Duquette D, Koay HF, et al. γδ T cells producing interleukin-17A regulate adipose regulatory T cell homeostasis and thermogenesis. *Nat Immunol* (2018) 19:464–74. doi: 10.1038/s41590-018-0094-2
- Pellicci DG, Koay HF, Berzins SP. Thymic development of unconventional T cells: how NKT cells, MAIT cells and gammadelta T cells emerge. *Nat Rev Immunol* (2020) 20:756–70. doi: 10.1038/s41577-020-0345-y
- Wang X, Tian Z. gammadelta T cells in liver diseases. Front Med (2018) 12:262–8. doi: 10.1007/s11684-017-0584-x
- Papadopoulou M, Sanchez SG, Vermijlen D. Innate and adaptive γδ T cells: How, when, and why. *Immunol Rev* (2020) 00:1–18. doi: 10.1111/ imr.12926
- Hunter S, Willcox CR, Davey MS, Kasatskaya SA, Jeffery HC, Chudakov DM, et al. Human liver infiltrating γδ T cells are composed of clonally expanded circulating and tissue-resident populations. *J Hepatol* (2018) 69(3):654–65. doi: 10.1016/j.jhep.2018.05.007
- Benova K, Hanckova M, Koci K, Kudelova M, Betakova T. T cells and their function in the immune response to viruses. *Acta Virol* (2020) 64:131–43. doi: 10.4149/av_2020_203
- Wu X, Zhang JY, Huang A, Li YY, Zhang S, Wei J, et al. Decreased Vdelta2 gammadelta T cells associated with liver damage by regulation of Th17 response in patients with chronic hepatitis B. J Infect Dis (2013) 208:1294– 304. doi: 10.1093/infdis/jit312
- Lai Q, Ma S, Ge J, Huang Z, Huang X, Jiang X, et al. TCR gammadelta(+)CD4 (-)CD8(-) T cells suppress the CD8(+) T-cell response to hepatitis B virus peptides, and are associated with viral control in chronic hepatitis B. *PloS One* (2014) 9:e88475. doi: 10.1371/journal.pone.0088475

- Kong X, Sun R, Chen Y, Wei H, Tian Z. gammadeltaT cells drive myeloidderived suppressor cell-mediated CD8+ T cell exhaustion in hepatitis B virusinduced immunotolerance. *J Immunol* (2014) 193:1645–53. doi: 10.4049/ jimmunol.1303432
- Sugai S, Yoshikawa T, Iwama T, Tsuchiya N, Ueda N, Fujinami N, et al. Hepatocellular carcinoma cell sensitivity to Vgamma9Vdelta2 T lymphocytemediated killing is increased by zoledronate. *Int J Oncol* (2016) 48:1794–804. doi: 10.3892/ijo.2016.3403
- Qian L, Wang N, Tian H, Jin H, Zhao H, Niu C, et al. Dual Effects of Cellular Immunotherapy in Inhibition of Virus Replication and Prolongation of Survival in HCV-Positive Hepatocellular Carcinoma Patients. J Immunol Res (2016) 2016:6837241. doi: 10.1155/2016/6837241
- Ferrari C. HBV and the immune response. *Liver Int* (2015) 35(Suppl 1):121–8. doi: 10.1111/liv.12749
- Conroy MJ, Mac Nicholas R, Taylor M, O'dea S, Mulcahy F, Norris S, et al. Increased Frequencies of Circulating IFN-gamma-Producing Vdelta1(+) and Vdelta2(+) gammadelta T Cells in Patients with Asymptomatic Persistent Hepatitis B Virus Infection. *Viral Immunol* (2015) 28:201–8. doi: 10.1089/ vim.2014.0133
- 24. Chang L, Wang L, Ling N, Peng H, Chen M. Increase in liver gammadelta T cells with concurrent augmentation of IFN-beta production during the early stages of a mouse model of acute experimental hepatitis B virus infection. *Exp Ther Med* (2020) 19:67–78. doi: 10.3892/etm.2019.8197
- 25. Cannizzo ES, Tincati C, Binda F, Ronzi P, Cazzaniga FA, Antinori S, et al. Unconventional T cells in chronic hepatitis B patients on long-term suppressive therapy with tenofovir followed by a Peg-IFN add-on strategy: A randomized study. J Viral Hepat (2018) 25:381–90. doi: 10.1111/jvh.12820
- 26. Chen M, Zhang D, Zhen W, Shi Q, Liu Y, Ling N, et al. Characteristics of circulating T cell receptor gamma-delta T cells from individuals chronically infected with hepatitis B virus (HBV): an association between V(delta)2 subtype and chronic HBV infection. J Infect Dis (2008) 198:1643–50. doi: 10.1086/593065
- 27. Chen M, Hu P, Ling N, Peng H, Lei Y, Hu H, et al. Enhanced functions of peripheral gammadelta T cells in chronic hepatitis B infection during interferon alpha treatment in vivo and in vitro. *PLoS One* (2015) 10: e0120086. doi: 10.1371/journal.pone.0120086
- Chang KM, Traum D, Park JJ, Ho S, Ojiro K, Wong DK, et al. Distinct phenotype and function of circulating Vdelta1+ and Vdelta2+ gammadeltaTcells in acute and chronic hepatitis B. *PLoS Pathog* (2019) 15:e1007715. doi: 10.1371/journal.ppat.1007715
- Cairo C, Armstrong CL, Cummings JS, Deetz CO, Tan M, Lu C, et al. Impact of age, gender, and race on circulating γδ T cells. *Hum Immunol* (2010) 71 (10):968–75. doi: 10.1016/j.humimm.2010.06.014
- Chen M, Hu P, Peng H, Zeng W, Shi X, Lei Y, et al. Enhanced peripheral gammadeltaT cells cytotoxicity potential in patients with HBV-associated acuteon-chronic liver failure might contribute to the disease progression. J Clin Immunol (2012) 32:877–85. doi: 10.1007/s10875-012-9678-z
- 31. Tseng CT, Miskovsky E, Houghton M, Klimpel GR. Characterization of liver T-cell receptor gammadelta T cells obtained from individuals chronically infected with hepatitis C virus (HCV): evidence for these T cells playing a role in the liver pathology associated with HCV infections. *Hepatology* (2001) 33:1312–20. doi: 10.1053/jhep.2001.24269
- 32. Agrati C, D'offizi G, Narciso P, Abrignani S, Ippolito G, Colizzi V, et al. Vdelta1 T lymphocytes expressing a Th1 phenotype are the major gammadelta T cell subset infiltrating the liver of HCV-infected persons. *Mol Med* (2001) 7:11–9. doi: 10.1007/BF03401834
- 33. Par G, Rukavina D, Podack ER, Horanyi M, Szekeres-Bartho J, Hegedus G, et al. Decrease in CD3-negative-CD8dim(+) and Vdelta2/Vgamma9 TcR+ peripheral blood lymphocyte counts, low perforin expression and the impairment of natural killer cell activity is associated with chronic hepatitis C virus infection. *J Hepatol* (2002) 37:514–22. doi: 10.1016/S0168-8278(02)00218-0
- Alonzi T, Agrati C, Costabile B, Cicchini C, Amicone L, Cavallari C, et al. Steatosis and intrahepatic lymphocyte recruitment in hepatitis C virus transgenic mice. J Gen Virol (2004) 85:1509–20. doi: 10.1099/vir.0.19724-0
- 35. Yin W, Tong S, Zhang Q, Shao J, Liu Q, Peng H, et al. Functional dichotomy of Vdelta2 gammadelta T cells in chronic hepatitis C virus infections: role in cytotoxicity but not for IFN-gamma production. *Sci Rep* (2016) 6:26296. doi: 10.1038/srep26296

- 36. Agrati C, Alonzi T, De Santis R, Castilletti C, Abbate I, Capobianchi MR, et al. Activation of Vgamma9Vdelta2 T cells by non-peptidic antigens induces the inhibition of subgenomic HCV replication. *Int Immunol* (2006) 18:11–8. doi: 10.1093/intimm/dxh337
- 37. Wu D, Yan WM, Wang HW, Huang D, Luo XP, Ning Q. gammadelta T Cells Contribute to the Outcome of Murine Fulminant Viral Hepatitis via Effector Cytokines TNF-alpha and IFN-gamma. *Curr Med Sci* (2018) 38:648–55. doi: 10.1007/s11596-018-1926-x
- Lee JW, Kim W, Kwon EK, Kim Y, Shin HM, Kim DH, et al. Immunological dynamics associated with rapid virological response during the early phase of type I interferon therapy in patients with chronic hepatitis C. PLoS One (2017) 12:e0179094. doi: 10.1371/journal.pone.0179094
- Ravens S, Hengst J, Schlapphoff V, Deterding K, Dhingra A, Schultze-Florey C, et al. Human gammadelta T Cell Receptor Repertoires in Peripheral Blood Remain Stable Despite Clearance of Persistent Hepatitis C Virus Infection by Direct-Acting Antiviral Drug Therapy. *Front Immunol* (2018) 9:510. doi: 10.3389/fimmu.2018.00510
- Ghosh A, Mondal RK, Romani S, Bagchi S, Cairo C, Pauza CD, et al. Persistent gamma delta T-cell dysfunction in chronic HCV infection despite directacting antiviral therapy induced cure. *J Viral Hepat* (2019) 26:1105–16. doi: 10.1111/jvh.13121
- Dewidar B, Meyer C, Dooley S, Meindl-Beinker AN. TGF-beta in Hepatic Stellate Cell Activation and Liver Fibrogenesis-Updated 2019. *Cells* (2019) 8 (11):1419. doi: 10.3390/cells8111419
- Hammerich L, Bangen JM, Govaere O, Zimmermann HW, Gassler N, Huss S, et al. Chemokine receptor CCR6-dependent accumulation of gammadelta T cells in injured liver restricts hepatic inflammation and fibrosis. *Hepatology* (2014) 59:630–42. doi: 10.1002/hep.26697
- Liu M, Hu Y, Yuan Y, Tian Z, Zhang C. gammadeltaT Cells Suppress Liver Fibrosis via Strong Cytolysis and Enhanced NK Cell-Mediated Cytotoxicity Against Hepatic Stellate Cells. *Front Immunol* (2019) 10:477. doi: 10.3389/ fimmu.2019.00477
- 44. Zhou BY, Gong JH, Cai XY, Wang JX, Luo F, Jiang N, et al. An imbalance between stellate cells and gammadeltaT cells contributes to hepatocellular carcinoma aggressiveness and recurrence. *Hepatol Int* (2019) 13:631–40. doi: 10.1007/s12072-019-09969-w
- 45. Cai XY, Wang JX, Yi Y, He HW, Ni XC, Zhou J, et al. Low counts of gammadelta T cells in peritumoral liver tissue are related to more frequent recurrence in patients with hepatocellular carcinoma after curative resection. *Asian Pac J Cancer Prev* (2014) 15:775–80. doi: 10.7314/APJCP.2014.15.2.775
- 46. Zhao N, Dang H, Ma L, Martin SP, Forgues M, Ylaya K, et al. Intratumoral gammadelta T-cell infiltrates, CCL4/5 protein expression and survival in patients with hepatocellular carcinoma. *Hepatology* (2020). doi: 10.1002/ hep.31412
- Tian W, Ma J, Shi R, Ren C, He J, Zhao H. gammadelta T cell-mediated individualized immunotherapy for hepatocellular carcinoma considering clinicopathological characteristics and immunosuppressive factors. *Oncol Lett* (2018) 15:5433–42. doi: 10.3892/ol.2018.8026
- Hoh A, Dewerth A, Vogt F, Wenz J, Baeuerle PA, Warmann SW, et al. The activity of gammadelta T cells against paediatric liver tumour cells and spheroids in cell culture. *Liver Int* (2013) 33:127–36. doi: 10.1111/liv.12011
- 49. Tanaka Y, Kobayashi H, Terasaki T, Toma H, Aruga A, Uchiyama T, et al. Synthesis of pyrophosphate-containing compounds that stimulate Vgamma2Vdelta2 T cells: application to cancer immunotherapy. *Med Chem* (2007) 3:85–99. doi: 10.2174/157340607779317544
- Toutirais O, Cabillic F, Le Friec G, Salot S, Loyer P, Le Gallo M, et al. DNAX accessory molecule-1 (CD226) promotes human hepatocellular carcinoma cell lysis by Vgamma9Vdelta2 T cells. *Eur J Immunol* (2009) 39:1361–8. doi: 10.1002/eji.200838409
- 51. Qian P, Zhang YW, Zhou ZH, Liu JQ, Yue SY, Guo XL, et al. Artesunate enhances gammadelta T-cell-mediated antitumor activity through augmenting gammadelta T-cell function and reversing immune escape of HepG2 cells. *Immunopharmacol Immunotoxicol* (2018) 40:107–16. doi: 10.1080/08923973.2017.1386212
- 52. Yi Y, He HW, Wang JX, Cai XY, Li YW, Zhou J, et al. The functional impairment of HCC-infiltrating gammadelta T cells, partially mediated by regulatory T cells in a TGFbeta- and IL-10-dependent manner. *J Hepatol* (2013) 58:977–83. doi: 10.1016/j.jhep.2012.12.015

- 53. Jiang H, Yang Z, Song Z, Green M, Song H, Shao Q. gammadelta T cells in hepatocellular carcinoma patients present cytotoxic activity but are reduced in potency due to IL-2 and IL-21 pathways. *Int Immunopharmacol* (2019) 70:167–73. doi: 10.1016/j.intimp.2019.02.019
- Di Blasi D, Boldanova T, Mori L, Terracciano L, Heim MH, De Libero G. Unique T-Cell Populations Define Immune-Inflamed Hepatocellular Carcinoma. *Cell Mol Gastroenterol Hepatol* (2020) 9:195–218. doi: 10.1016/ j.jcmgh.2019.08.004
- 55. Ma S, Cheng Q, Cai Y, Gong H, Wu Y, Yu X, et al. IL-17A produced by gammadelta T cells promotes tumor growth in hepatocellular carcinoma. *Cancer Res* (2014) 74:1969–82. doi: 10.1158/0008-5472.CAN-13-2534

Conflict of Interest: The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Copyright © 2021 Hou and Wu. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.