

Review

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Allopregnanolone-induced rise in intracellular calcium in embryonic hippocampal neurons parallels their proliferative potential

Jun Ming Wang^{1,2} and Roberta Diaz Brinton*¹

Address: ¹Department of Pharmacology and Pharmaceutical Sciences and Program in Neuroscience, University of Southern California, Los Angeles, CA 90089, USA and ²Department of Pathology, University of Mississippi Medical Center, Jackson, MS 39216, USA

Email: Jun Ming Wang - jwang@pathology.umsmed.edu; Roberta Diaz Brinton* - rbrinton@hsc.usc.edu

* Corresponding author

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Abstract

Background: Factors that regulate intracellular calcium concentration are known to play a critical role in brain function and neural development, including neural plasticity and neurogenesis. We previously demonstrated that the neurosteroid allopregnanolone (AP α ; 5 α -pregnan-3 α -ol-20-one) promotes neural progenitor proliferation *in vitro* in cultures of rodent hippocampal and human cortical neural progenitors, and *in vivo* in triple transgenic Alzheimer's disease mice dentate gyrus. We also found that AP α -induced proliferation of neural progenitors is abolished by a calcium channel blocker, nifedipine, indicating a calcium dependent mechanism for the proliferation.

Methods: In the present study, we investigated the effect of AP α on the regulation of intracellular calcium concentration in E18 rat hippocampal neurons using ratiometric Fura2-AM imaging.

Results: Results indicate that AP α rapidly increased intracellular calcium concentration in a dose-dependent and developmentally regulated manner, with an EC₅₀ of 110 \pm 15 nM and a maximal response occurring at three days *in vitro*. The stereoisomers 3 β -hydroxy-5 α -hydroxy-pregnan-20-one, and 3 β -hydroxy-5 β -hydroxy-pregnan-20-one, as well as progesterone, were without significant effect. AP α -induced intracellular calcium concentration increase was not observed in calcium depleted medium and was blocked in the presence of the broad spectrum calcium channel blocker La³⁺, or the L-type calcium channel blocker nifedipine. Furthermore, the GABA_A receptor blockers bicuculline and picrotoxin abolished AP α -induced intracellular calcium concentration rise.

Conclusion: Collectively, these data indicate that AP α promotes a rapid, dose-dependent, stereospecific, and developmentally regulated increase of intracellular calcium concentration in rat embryonic hippocampal neurons via a mechanism that requires both the GABA_A receptor and L-type calcium channel. These data suggest that AP α -induced intracellular calcium concentration increase serves as the initiation mechanism whereby AP α promotes neurogenesis.

Background

Allopregnanolone (AP α ; 3 α -hydroxy-5 α -hydroxy-pregnan-20-one; also known as tetrahydroprogesterone) is a derivative of progesterone that is produced in both the periphery and the central nervous system via enzymatic conversions of progesterone [1-3]. In mature neurons, AP α is known to act as an allosteric modulator of the γ -aminobutyric acid type A (GABA $_A$) receptor, binding to a specific site within the GABA $_A$ receptor at physiological concentrations (6–35 nM) [4,5] to increase chloride influx, thereby hyperpolarizing the neuronal membrane potential, and decreasing neuron excitability [6-11]. In marked contrast, the flux of chloride in developing neurons is opposite to that of mature neurons. Because of the high intracellular chloride content in immature neurons, AP α provokes an efflux of chloride through the GABA $_A$ receptor, depolarization of the membrane, opening voltage dependent L-type calcium channels, leading to an influx of calcium from the extracellular medium [12-16].

Calcium signalling plays a key role in neural function and neural development [17-20]. Increases in intracellular calcium concentration ($[Ca^{2+}]_i$) also control cell cycle protein expression and promote cell proliferation [21-26]. Therefore, GABA $_A$ receptor-mediated depolarization may be the trigger that leads to activity-independent $[Ca^{2+}]_i$ rise in early precursor cells, or neural progenitors and stem cells, and consequently may influence early developmental events, including neurogenesis and synaptogenesis [16,27-29].

Previously we found that AP α rapidly induced neurite regression in cultured hippocampal neurons [30], which we later identified as a prelude to entry into the cell cycle and mitosis [29]. Recently, we demonstrated that AP α regulates the expression of genes encoding cell cycle-related molecules and enhances human cortical neural progenitor, and rat hippocampal neuronal progenitor cell proliferation *in vitro* [29], and *in vivo* in triple transgenic Alzheimer's disease mice dentate gyrus [31-33]. Moreover, the L-type calcium blocker nifedipine abolished the AP α -induced cell proliferation. We therefore hypothesized that the AP α -induced neural progenitor cell proliferation is mediated by calcium influx via GABA $_A$ receptor-activated L-type calcium channels. To test this hypothesis, we investigated the impact of AP α on calcium dynamics using Fura2 fluorescent ratio calcium imaging in rat E18 hippocampal neurons.

Methods

Animals and primary hippocampal neuron culture

Timed-pregnant Sprague-Dawley rats were purchased from Harlan Sprague Dawley, Inc. (Indianapolis, IN, USA). Rats were housed under controlled conditions of temperature (22°C), humidity (30–50%), and light (14

hour light:10 hour dark); water and food were available *ad libitum*. All experiments conformed to the Animal Welfare Act, Guide to Use and Care of Laboratory Animals, and the US Government Principles of the Utilization and Care of Vertebrate Animals Used in Testing, Research, and Training guidelines on the ethical use of animals. In addition, the minimal number of required animals was used for these experiments and suffering was minimized. Primary cultures of dissociated hippocampal neurons were performed as previously described [29,30]. Briefly, hippocampi were dissected from the brains of E18 rat fetuses, treated with 0.02% trypsin in Hank's balanced salt solution (HBSS, Invitrogen, Grand Island, NY, USA) for 5 minutes at 37°C and dissociated by repeated passage through a series of constricted, fire-polished Pasteur pipettes. Cells were plated onto poly-D-lysine-coated 22 mm diameter cover slips at a density of 2–4 \times 10⁴ cells per cm², and grown in Neurobasal medium without phenol red (NBM; Gibco/Life Technologies, St. Petersburg, FL, USA) supplemented with 2% B27 (Gibco/Life Technologies), 10 U/ml penicillin, 10 μ g/ml streptomycin, 0.5 mM glutamine and 25 μ M glutamate. Cultures were maintained at 37°C in a humidified 5% CO₂ atmosphere until the day of imaging.

Steroids

All steroids used in this study were purchased from Steraloids, Inc (Newport, Rhode Island, USA). They were: allopregnanolone (AP α ; 5 α -pregnan-3 α -ol-20-one); pregnanolone (3 α 5 β AP; 5 β -pregnan-3 α -ol-20-one); epipregnanolone (3 β 5 β AP; 5 β -pregnan-3 β -ol-20-one); epiallopregnanolone (3 β 5 α AP; 5 α -pregnan-3 β -ol-20-one); and progesterone (P₄; 4-pregnane-3,20-dione).

[Ca²⁺] microfluorimetry and imaging

[Ca²⁺] in hippocampal neurons was determined by ratio-metric imaging of the Ca²⁺-sensitive fluorescent dye, Fura-2 acetoxyethyl ester (Fura-2 AM, Molecular Probes, Eugene, OR, USA), as previously described [34,35]. Briefly, cells were loaded with 2 μ M Fura-2 AM in HEPES-buffered solution (HBS; 100 mM NaCl, 2.0 mM KCl, 1.0 mM CaCl₂, 1.0 mM MgCl₂, 1.0 mM NaH₂PO₄, 4.2 mM NaHCO₃, 12.5 mM HEPES and 10.0 mM glucose, pH 7.4) for 45 minutes at 37°C, 5% CO₂. Cells were then washed in HBS to remove excess Fura-2 AM and incubated in HBS for another 30 minutes to equilibrate. The cover slip with cells loaded with Fura-2 AM was then mounted in a perfusion chamber and placed on an inverted microscope (Olympus IMT-2). Baseline [Ca²⁺] was obtained over 1 minute prior to the initiation of stimulus, which was maintained for the duration of the imaging. Stimulation was initiated by perfusion (2 ml/minute), or bolus addition (200 μ l in total of 1,000 μ l), to a static bath of the indicated compounds. Fura-2 was successively excited by a xenon light source at 340 nm and 380 nm by means of

two narrow beam band-pass filters selected by a computer-controlled filter wheel. The emitted fluorescence was filtered through a 520 nm filter, captured with an intensified CCD camera (COHU, San Diego, CA, USA), and analyzed with InCyt Im2 software (Intracellular Imaging, Inc., Cincinnati, OH, USA). The $[Ca^{2+}]_i$ was calculated by comparing the ratio of fluorescence at 340 nm and 380 nm against a standard curve. The curve was made from five different $[Ca^{2+}]_i$ standards available from Molecular Probes. Data from regions of interest were displayed in real-time and logged to hard disk. Data are presented as representative traces averaged from at least 10 cells per experiment. Responses to treatments were quantified by determining the difference between the average $[Ca^{2+}]_i$ for the period of time of maximal response during the drug exposure and the average $[Ca^{2+}]_i$ for 30 seconds prior to exposure. Changes in $[Ca^{2+}]_i$ are presented as mean \pm standard error of the mean (SEM) from three or more independent experiments with ≥ 10 cells per experiment. Statistical comparisons utilized one-way ANOVA followed by Newman-Keul's *post hoc* analysis.

Results

AP α induces a rapid and transient change in $[Ca^{2+}]_i$ in hippocampal neurons

AP α induced a rapid (within seconds) and transient (over the course of minutes) increase in $[Ca^{2+}]_i$ in E18 rat hippocampal neurons (Figure 1A) as observed by ratiometric imaging of the Ca^{2+} -sensitive fluorescent dye Fura-2 AM (Figure 1A). $[Ca^{2+}]_i$ was calculated using a linear regression standard curve of the 340 nm/380 nm ratios of fluorescence generated by a standard series of calcium concentrations (Figure 1B). The multiple correlation coefficient (R-square = 0.9942) indicated a linear relationship between fluorescence ratio and $[Ca^{2+}]_i$ (Figure 1B). Based on this linear relationship, it was possible to generate an approximate $[Ca^{2+}]_i$ value. Three distinct 'calcium responses' in neurons exposed to AP α emerged from this analysis: high response, neurons exhibiting a ≥ 0.78 of 340 nm/380 nm ratio; low response, neurons exhibiting ≤ 0.73 of 340 nm/380 nm ratio; and no response (Figure 1C). In the low response population, AP α (250 nM) induced a $[Ca^{2+}]_i$ rise of 38 ± 2.6 nM, whereas in the high response population, the same concentration of AP α induced a $[Ca^{2+}]_i$ rise of 118 ± 32 nM (Figure 1D; $p < 0.05$ compared to control neurons).

AP α -induced $[Ca^{2+}]_i$ rise is dependent upon dose and days in vitro and is stereoisomer specific

To characterize AP α -induced $[Ca^{2+}]_i$ signalling in embryonic hippocampal neurons, three issues were addressed: dose-response; stereospecificity; and developmental profile of response during days *in vitro* (DIV). Analysis for dose-response and stereospecificity were conducted in neurons at 3 DIV. Application of 10 nM AP α showed an

insignificant increase in $[Ca^{2+}]_i$. A linear and significant increase was observed from 50 nM, (13 ± 7 nM $[Ca^{2+}]_i$, $p < 0.05$ versus vehicle control), and 100 nM AP α (62 ± 12 $[Ca^{2+}]_i$, $p < 0.01$). Maximal response was observed when 250 nM AP α was applied (135 ± 14 nM $[Ca^{2+}]_i$, $p < 0.01$), which could not be further increased using 500–1,000 nM AP α (Figure 2A). The estimated EC₅₀ value for AP α was 124 ± 15 nM.

To determine the specificity of AP α on the increase of $[Ca^{2+}]_i$, the effects of several AP α isomers, and its parent molecule, progesterone, were compared at a concentration of 250 nM. AP α induced an $[Ca^{2+}]_i$ increase with an average of 133 ± 11 nM. No significant $[Ca^{2+}]_i$ response was observed for 3 β 5 α AP, 3 β 5 β AP, or progesterone at the same concentration. Only 3 α 5 β AP induced a significant, but lower, $[Ca^{2+}]_i$ rise (73 ± 8 nM, $p < 0.01$; Figure 2B,C). These data indicate that the AP α -induced $[Ca^{2+}]_i$ increase is stereospecific for AP α , and its 5 β -, but not 3 β -isomers.

To determine the proportion of hippocampal neurons responsive to AP α , and the duration of responsiveness, the number of AP α -induced $[Ca^{2+}]_i$ -responsive neurons were assessed each day for 11 days (216 neurons/day). The percentage of AP α -induced $[Ca^{2+}]_i$ rise in neurons increased linearly from day 0 to day 4 (DIV), followed by a stepwise decrease in the number of neurons that, by day 11, had returned to day 0 levels (Figure 2D). At DIV 0, $25 \pm 5\%$ of the neurons exhibited a AP α -induced $[Ca^{2+}]_i$ increase; at DIV 1, $61 \pm 7\%$; at DIV 2, $84 \pm 4\%$. By DIV 3, the number of AP α -responding neurons reached an asymptote at $90 \pm 6\%$ (Figure 2D; $*p < 0.01$ and $**p < 0.001$) and remained at that level until DIV 4. From DIV 7–10, the number of responding neurons declined and remained stable at approximately 63%. By DIV 11, the number of neurons responding to AP α ($30 \pm 4\%$) declined to the level observed at DIV 0.

Different types of $[Ca^{2+}]_i$ responses

To determine the developmental profile for low and high responders, we determined the change in the magnitude of AP α -induced $[Ca^{2+}]_i$ in low and high responders, and the portion of the population of responders across DIV. We first investigated whether the magnitude of the AP α -induced $[Ca^{2+}]_i$ changed with time in culture. High-response, low-response, and non-responsive neurons were consistently observed from DIV 0 to DIV 11. As summarized in Figure 3A, the magnitude of AP α -induced $[Ca^{2+}]_i$ in low responders (340/380 nm fluorescence ratio lower than 0.73 and reflect < 45 nM increase in free calcium; Figure 1B) showed no significant change during the entire 11-day experimental period. In contrast, the magnitude of AP α -induced $[Ca^{2+}]_i$ in high-response neurons (340/380 nm fluorescent ratio > 0.78 , reflecting a > 65 nM increase in free calcium, $p < 0.01$) increased from 65 to

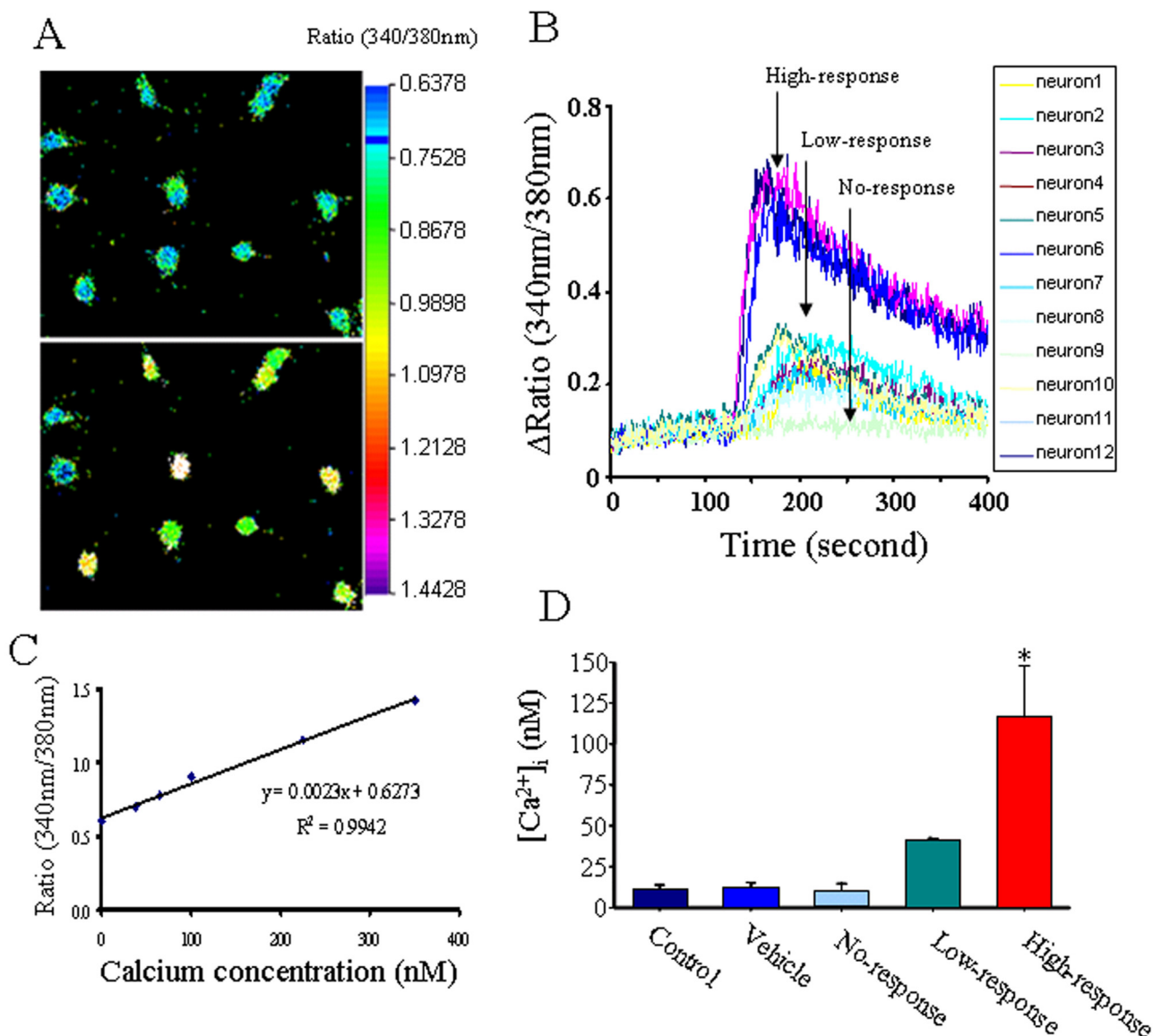


Figure 1

AP α induces a rapid and transient [Ca²⁺]_i rise in rat E18 hippocampal neurons in primary culture. Cells grown on glass cover slips were loaded with Fura-2 AM as described in the Methods. The [Ca²⁺]_i was determined on single cells in HBSS medium. **(A)** Images represent calcium fluoro-2 fluorescence in rat hippocampal neurons under vehicle control (top) and 500 nM AP α (bottom). A fluorescent gradient of fluorescence ratio of 340 nm/380 nm is presented at the right side of the images. **(B)** A linear regression and the multiple correlation coefficient R-square value indicate the perfect correlation between fluorescence ratio of 340 nm/380 nm and [Ca²⁺]_i. **(C)** AP α induced rapid and transient responses of [Ca²⁺]_i recorded in a time course and expressed as the fluorescence ratio between 340 nm/380 nm, which is a representative of three different experiments. **(D)** Bar graph represents the summary of [Ca²⁺]_i in response to control, vehicle control, and 500 nM AP α . Data are mean \pm SEM (**p* < 0.05 compared to control neurons; N = 3 experiments with 36 neurons/condition/experiment). No changes in [Ca²⁺]_i were observed under control or vehicle control conditions. With the addition of 500 nM AP α , three types of calcium responses were observed in neurons: high (defined as an increase in [Ca²⁺]_i >65 nM over the baseline); low (defined as an increase in [Ca²⁺]_i <45 nM over the baseline); and no response (defined as neurons that did not show a measurable increase in [Ca²⁺]_i over their baseline).

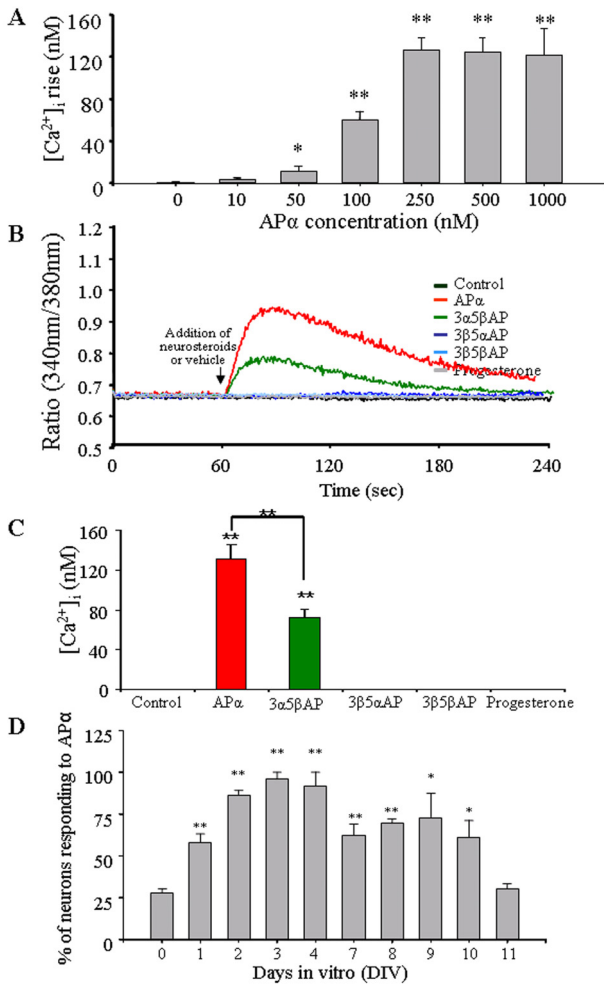


Figure 2
 APα-induced [Ca²⁺]_i rise shows dose- and DIV-dependent manner and is stereoisomer-specific. **(A)** Bar graph shows dose responses of APα on [Ca²⁺]_i rise in 3 DIV neurons. Data are mean ± SEM (N = 3–6 experiments with 36 neurons/condition/experiment, *p < 0.05, **p < 0.01). **(B)** Stereospecificity of APα-induced [Ca²⁺]_i. Neurons were exposed to 250 nM of the stereoisomers APα, 3α5βAP, 3β5αAP, 3β5βAP or progesterone (250 nM) at the time-point indicated by the arrow. The graph is a representative of four different experiments (N = 36 neurons/condition/experiment). **(C)** Bar graph shows the summary of data collected from stereoisomer experiments and represented as [Ca²⁺]_i. **p < 0.01 APα or 3α5βAP compared to control, or APα and 3α5βAP compared to each other; N = 4 experiments with 36 neurons/condition/experiment. **(D)** Bar graph indicates the percentage of neurons that exhibit a [Ca²⁺]_i rise in response to 500 nM APα at different days *in vitro* (DIV). DIV 0 is defined as the day the neurons are seeded. The greatest response, around 90% of cells responding to APα, is observed on 3 DIV. Data are mean ± SEM (N = 3 independent experiments with at least 36 neurons/condition/experiment). *p < 0.01, **p < 0.001 versus DIV 0.

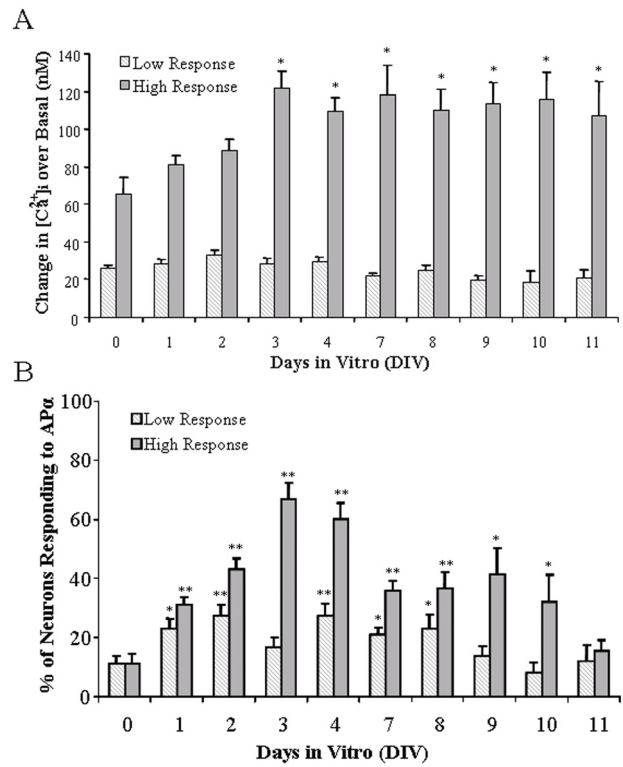


Figure 3
 The magnitude of the APα-induced intracellular calcium response changes at different DIV. **(A)** Bar graph indicates the magnitude of the [Ca²⁺]_i increases, high response (defined as >65 nM [Ca²⁺]_i rise over the baseline) or low response (defined as <45 nM [Ca²⁺]_i rise over the baseline), in [Ca²⁺]_i over the baseline in hippocampal neuron cultures at different DIV treated with 500 nM APα. **(B)** Bar graph indicates the percentage of hippocampal neurons that respond to 500 nM APα with a high or a low response at different DIV. Data are mean ± SEM (N = 3 independent experiments with at least 36 neurons/condition/experiment). *p < 0.01, **p < 0.001 versus DIV 0.

125 ± 10 nM in the first 3 days, and reached a plateau for the remaining days of culture. We then determined whether the number of high-response neurons versus low-response neurons changed during time in culture and present the data as a percentage of total cells (Figure 3B; *p < 0.01 and **p < 0.001). The number of high-response neurons increased rapidly and linearly, and reached the highest percentage of 65 ± 7% on DIV 3, gradually decreasing to less than 40% on DIV 7. The number of low-response neurons also increased early in the course of culturing and reached the greatest percentage of 27 ± 5% on DIV 2, decreasing to 17 ± 3% on DIV 3 (no significant difference with DIV 0). However, on DIV 4, the percentage of low-response neurons returned to a level similar to that observed on DIV 2 and then gradually decreased. DIV 3 was the time-point at which the highest number of high-

response neurons, and the lowest number of low-response neurons, were observed. These data indicate that the magnitude of calcium responses, and the numbers of AP α -induced $[Ca^{2+}]_i$ -responsive neurons, vary with the days of culture and that DIV 3 represents a pivotal time-point in the response to AP α .

AP α -induced $[Ca^{2+}]_i$ rise reflects an influx of extracellular Ca^{2+} and is regulated by an L-type calcium channel

Free $[Ca^{2+}]_i$ increase could result from either an influx of extracellular calcium or a release of calcium from an intracellular pool stored in organelles, including the endoplasmic reticulum and mitochondria. To investigate whether the AP α -induced $[Ca^{2+}]_i$ rise required an influx of extracellular Ca^{2+} , neurons at DIV 3–4, when 84% of neurons should be responsive to AP α (65% high responders), were treated with 500 nM AP α in a calcium-free medium. To sustain comparable osmolarity and ionic strength, 2 mM NaCl was substituted for 1 mM $CaCl_2$. In the absence of extracellular calcium, AP α did not induce a rise in $[Ca^{2+}]_i$ (Figure 4A). This result suggested that the AP α -induced $[Ca^{2+}]_i$ rise could be attributed mainly, if not completely, to the influx of extracellular Ca^{2+} . To discern which calcium channel was required for the calcium influx, neurons were exposed to the inorganic-calcium transport inhibitor La^{3+} (10 μ M) [36,37]. Exposure to La^{3+} reduced the AP α -induced $[Ca^{2+}]_i$ rise from 78 ± 5 nM to 42 ± 7 nM (Figure 4B; $p < 0.05$). When nifedipine, a more specific blocker to L-type calcium channels was used, the AP α -induced $[Ca^{2+}]_i$ rise decreased from 82 ± 14 nM to 20 ± 4 nM (Figure 4C; $p < 0.01$). These data indicate that the AP α -induced $[Ca^{2+}]_i$ rise is dependent upon extracellular Ca^{2+} and is regulated by the L-type calcium channel.

GABA $_A$ receptor mediates the $[Ca^{2+}]_i$ rise induced by AP α

It is well known that AP α acts as an allosteric modulator of the GABA $_A$ receptor/chloride channel by altering chloride flux and thereby hyperpolarizing (in mature neurons) or depolarizing (in immature neurons) the neuronal membrane [15,16,38-42]. To determine whether the AP α -induced $[Ca^{2+}]_i$ rise in hippocampal neurons required the GABA $_A$ receptor, two GABA $_A$ receptor blockers, picrotoxin (a non-competitive antagonist) and bicuculline (the prototypical competitive antagonist that directly competes with GABA for binding to the receptor complex), were applied at 100 μ M and 30 μ M, respectively. Both GABA $_A$ receptor blockers completely abolished the AP α -induced $[Ca^{2+}]_i$ rise (Figure 5A,B). These data indicate that the AP α -induced $[Ca^{2+}]_i$ rise requires activation of GABA $_A$ receptor and the L-type calcium channel in rat hippocampal neuron cultures.

Discussion

Analyses using intracellular ratiometric Fura2 calcium imaging demonstrated that AP α specifically induced a

rapid, transient, and dose-dependent $[Ca^{2+}]_i$ rise in E18 rat hippocampal neurons in primary culture. The AP α -induced $[Ca^{2+}]_i$ rise was not observed in calcium-free medium and was blocked by the L-type calcium channel blockers La^{3+} and nifedipine. In addition, the GABA $_A$ receptor inhibitors picrotoxin and bicuculline completely abolished the AP α -induced $[Ca^{2+}]_i$ rise. These findings in hippocampal neurons parallel those described by Dayanithi and Tapia-Arancibia [43] in fetal rat hypothalamic neurons, suggesting that the AP α -induced rise in $[Ca^{2+}]_i$ is a generalized effect in developing neurons.

The AP α -induced rapid response in $[Ca^{2+}]_i$ rise was specific to AP α as the β -isomers of AP α , namely 3 β 5 β AP and 3 β 5 α AP, and its parent molecule, progesterone, were without effect, whereas 3 α 5 β AP induced a $[Ca^{2+}]_i$ rise that was 50% of the effect of AP α . These data suggest the structure with the most important impact on stereospecificity may reside at the 3 position of the neurosteroid. Indeed, both an extracellular application, and 3 α -substitutions of the A ring of the steroid nucleus, have been reported to be prerequisites for many of the interactions of the neurosteroid with GABA $_A$ receptor/Cl $^-$ channels, because neither intracellular applications, nor 3 β -substituted steroids applied extracellularly, show agonistic effects of GABA [4,44,45]. Recent analyses by Hosie *et al.* [10] indicate that AP α can bind to two sites on the GABA $_A$ receptor, one that potentiates, and one that directly activates the GABA $_A$ receptor. The potentiating binding site of AP α resides in a cavity formed by the α -subunit transmembrane domains. The direct activating binding site of AP α located among interfacial residues between the α and β subunits and is enhanced by steroid binding to the potentiation site [10]. Data presented here demonstrate that the AP α -induced $[Ca^{2+}]_i$ rise can be abolished by two GABA $_A$ receptor blockers, namely bicuculline [46-49] and picrotoxin [50-52], strongly supporting the notion that the AP α -induced $[Ca^{2+}]_i$ rise is a GABA $_A$ receptor-mediated process and most likely through the direct activating binding site.

GABA $_A$ receptor is an ion channel that allows either influx or efflux of chloride ions (Cl $^-$), depending upon the prevailing transmembrane [Cl $^-$] gradient. Because immature neurons have a higher intracellular [Cl $^-$], activation of the GABA $_A$ receptor by GABA, or other agonists, for example, muscimol, causes efflux of Cl $^-$ and thus membrane depolarization [15,53]. This depolarization is sufficient to open L-type voltage-gated calcium channels, leading to calcium influx [54-56]. GABA and GABA-induced calcium influx have been linked to trophic actions important for developmental processes, including the expression of brain-derived neurotrophic factor [20]. Thus, GABA $_A$ receptor-mediated depolarization may be the trigger for a spontaneous, activity-independent $[Ca^{2+}]_i$ rise in early precursor cells, or subventricular zone radial precursor

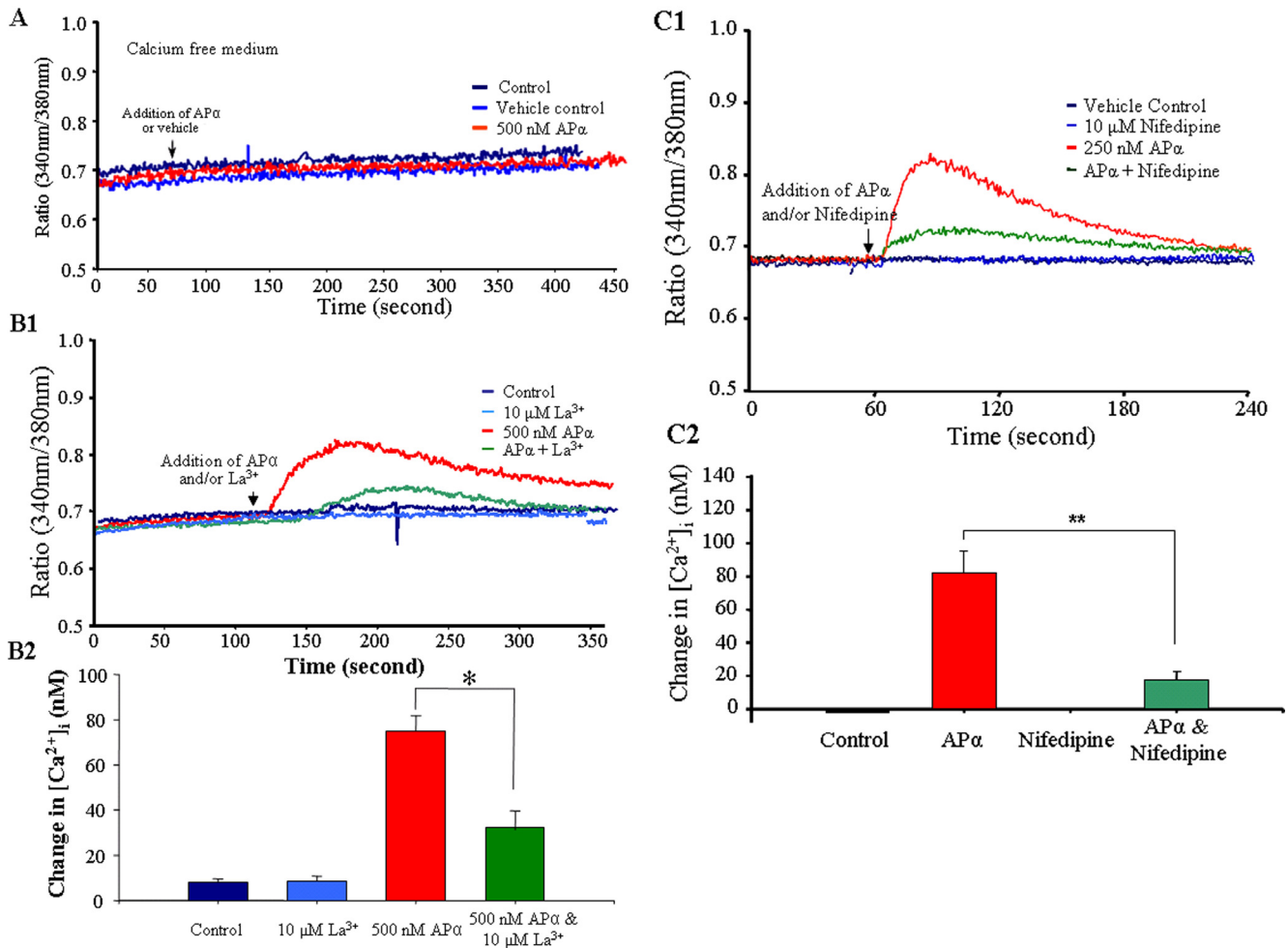


Figure 4
 AP α -induced $[Ca^{2+}]_i$ increases result from influx of extracellular calcium and are mediated by the L-type calcium channel. **(A)** Neurons (3–4 DIV) were exposed to 500 nM AP α or vehicle control in Ca^{2+} -free medium at the time-point indicated by the arrow. In the medium without calcium, AP α showed no effect on intracellular calcium concentration. A representative graph of four independent experiments is shown (N = 36 neurons/condition/experiment). **(B)** La^{3+} diminished AP α -induced $[Ca^{2+}]_i$ rises. Neurons (3–4 DIV) were incubated with or without 10 μM La^{3+} (a calcium channel blocker) for 30 minutes prior to and continuously throughout imaging, and 500 nM AP α or vehicle was added at the time-point indicated by the arrow. **(B1)** Representative graph of three different experiments (N = 36 neurons/condition/experiment). **(B2)** Bar graph represents the summary of $[Ca^{2+}]_i$ rise in response to control, 10 μM La^{3+} , 500 nM AP α , and AP α + La^{3+} . Data are mean \pm SEM, * p < 0.05 compared to AP α treated neurons; N = 3 experiments with 36 neurons/condition/experiment. **(C)** Nifedipine blocked AP α induced $[Ca^{2+}]_i$ rises. Neurons were incubated with or without 10 μM nifedipine (an L-type calcium channel blocker) for 30 minutes prior to and continuously throughout imaging, and 250 nM AP α or vehicle was added at the time-point indicated by the arrow. **(C1)** The graph is representative of three different experiments (N = 36 neurons/condition/experiment). **(C2)** The bar graph represents quantitative changes in $[Ca^{2+}]_i$ in response to control, 10 μM nifedipine, AP α , and AP α + 10 μM nifedipine. Data are mean \pm SEM (** p < 0.01 compared to AP α treated neurons; N = 36 neurons/condition/experiment).

cells, thereby influencing early developmental events, including neurogenesis and synaptogenesis [16,28,29]. In the present study, we have demonstrated that the AP α -induced $[Ca^{2+}]_i$ rise can be blocked by a voltage-gated calcium channel blocker, La^{3+} , as well as a more specific L-type calcium channel blocker, nifedipine. Further, we recently demonstrated that the L-type calcium channel

blocker nifedipine specifically abolished the AP α -induced proliferation of rat hippocampal neuronal progenitor cells [29]. Taken together, these data suggest that the AP α -induced $[Ca^{2+}]_i$ rise, regulated by the L-type calcium channel and evoked by GABA $_A$ receptor, may be the signalling initiation mechanism for AP α -induced neuroprogenitor cell proliferation and cell cycle gene expression.

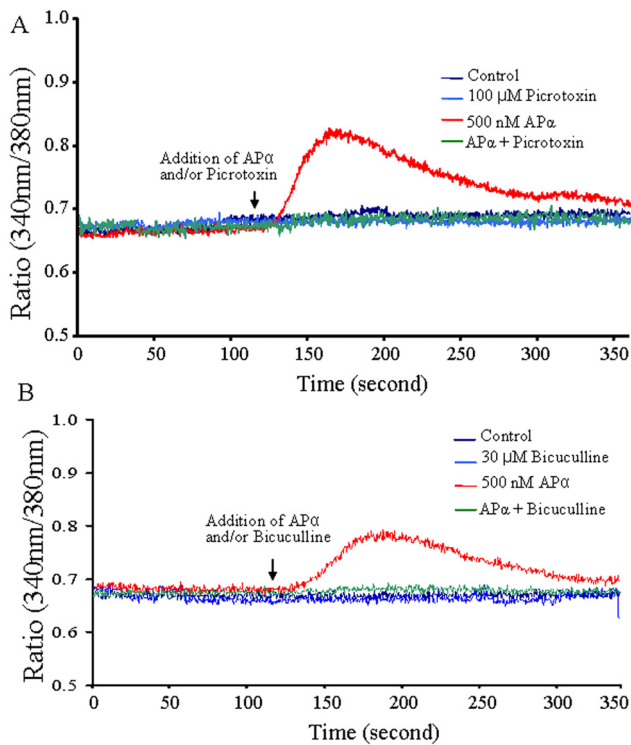


Figure 5
 AP α -induced $[Ca^{2+}]_i$ rises are mediated by the GABA $_A$ receptor. **(A)** Picrotoxin completely abolished AP α -induced $[Ca^{2+}]_i$ rises. Fluorescent Fura2-AM imaging of 3 DIV hippocampal neurons incubated with or without 100 μ M picrotoxin (a non-competitive GABA $_A$ receptor antagonist) 30 minutes prior to and continuously throughout imaging. AP α (500 nM) or vehicle was added at the time-point indicated by the arrow. The graph is representative of the four experiments conducted (N = 36 neurons/condition/experiment). **(B)** Bicuculline completely abolished AP α -induced $[Ca^{2+}]_i$ rises. Fluorescent Fura2-AM imaging of 3 DIV hippocampal neurons incubated with or without 30 μ M bicuculline (a competitive GABA $_A$ receptor antagonist) for 30 minutes prior to and continuously throughout imaging. AP α (500 nM) or vehicle was added at the time-point indicated by the arrow. The graph is representative of the four different experiments conducted (N = 36 neurons/condition/experiment).

As development progresses, the effect of AP α associated with GABA $_A$ receptor binding gradually switches from excitatory to inhibitory [15,16,57,58]. The timing of the shift from depolarizing to hyperpolarizing via GABA $_A$ receptor varies across brain regions, but is generally complete by the second week of life in the rat and mouse [12,59-61]. Remarkably, AP α induction of $[Ca^{2+}]_i$ rise in cultured hippocampal neurons closely parallels the developmental time course of Na $^+$ -K $^+$ -2Cl $^-$ co-transporter expression [62,63]. In primary cultures of the E18 rat hippocampal neurons, a mixture of E16-E18 neurons with

differing phenotypes will exist. Therefore, it is not surprising that high and low AP α -induced calcium responses were observed in this study. The high and low calcium responses may be indicative of the developmental stage of the neuron in culture.

The GABA $_A$ receptor is composed of a pentamer of structurally homologous subunits that may be drawn from the α 1-6, β 1-3, γ 1-3, δ , ϵ , θ , ρ 1-3, and π subunit families. The precise subunit composition of different GABA $_A$ receptor isoforms is an important determinant of their pharmacological and biophysical properties [4,64-69], but the exact combination heterogeneity exist. For example, a study indicated that the subunit combinations comprising α 1 β 1 γ 2 and α 3 β 1 γ 2 required a three- to seven-fold lower concentration for AP α to enhance GABA-evoked current to the same degree as other combinations [70], while another study demonstrated that the potencies of AP α to enhance GABA response were significantly higher in the α 5 β 2 γ 2 receptor versus α 1 β 2 γ 2 [71]. Another study suggested that the efficacy of AP α to enhance GABA response depended on the γ subunit subtype: α 1 β 1 γ 3 > α 1 β 1 γ 2 = α 1 β 1 γ 1 [72]. Interestingly, during migration from the subventricular zone to the cortical plate, neurons became predominantly GABAergic, and their dominant GABA $_A$ receptor subunit expression pattern changed from α 4 β 1 γ 1 to α 3 β 3 γ 2 or α 3 β 3 γ 3, coincident with an increasing potency of GABA on GABA $_A$ receptor-mediated depolarization [73]. Although heterogeneity of the subunit combination exists, all these studies suggest that the effects of AP α depend on subunit combinations of the GABA $_A$ receptor and the combination is changing during development. Therefore, the different GABA $_A$ receptor subunit combinations might be the underlying mechanism for the variety of AP α -induced $[Ca^{2+}]_i$ responses in cultured hippocampal neurons. Along with DIV, the combination of the GABA $_A$ receptor subunits is changing and a possible time-point at which one GABA $_A$ receptor subunit combination switches to another is 3-4 DIV. However, exact matching of neuronal high and low calcium responses to the expression of specific GABA $_A$ receptor subunit combinations, and investigation of the role of each combination, needs to be further addressed.

Conclusion

These data demonstrate that AP α induced a dose and developmentally regulated rise in $[Ca^{2+}]_i$ that was stereospecific, rapid, and transient in E18 rat hippocampal neurons in primary culture. Moreover, our data indicated that the AP α -induced rise in $[Ca^{2+}]_i$ requires both GABA $_A$ receptor and L-type calcium channel. Together with earlier findings, we propose that the AP α -induced rise in $[Ca^{2+}]_i$ provides a mechanism whereby AP α can promote proliferation of rodent, and also possible of human, neural progenitor cells.

List of abbreviations used

3 α 5 β AP: pregnanolone; 3 β 5 α AP: epiallopregnanolone; 3 β 5 β AP: epipregnanolone; AP α : allopregnanolone; [Ca²⁺]_i: intracellular calcium concentration; DIV: days *in vitro*; Fura-2 AM: Fura-2 acetoxymethyl ester; GABA_A: γ -aminobutyric acid type A; HBS: HEPES-buffered solution; HBSS: Hank's balanced salt solution; SEM: standard error of the mean.

Competing interests

The authors declare that they have no competing interests.

Authors' contributions

JMW analyzed the data and drafted the manuscript. RDB designed the experiments, coordinated the study and edited the manuscript.

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