



Therapeutic Anabolic and Anticatabolic Benefits of Natural Chinese Medicines for the Treatment of Osteoporosis

Jianbo He^{1,2,3†}, Xiaojuan Li^{3,4†}, Ziyi Wang^{2†}, Samuel Bennett², Kai Chen², Zhifeng Xiao³, Jiheng Zhan^{1,3}, Shudong Chen³, Yu Hou³, Junhao Chen², Shaofang Wang⁵, Jiake Xu^{2*} and Dingkun Lin^{3*}

¹ Guangzhou University of Chinese Medicine, Guangzhou, China, ² The School of Biomedical Sciences, University of Western Australia, Perth, WA, Australia, ³ The Second Affiliated Hospital of Guangzhou University of Chinese Medicine, Guangdong Provincial Hospital of Chinese Medicine, Guangzhou, China, ⁴ Formula-Pattern Research Center, School of Traditional Chinese Medicine, Jinan University, Guangzhou, China, ⁵ Centre for Legumes in Mediterranean Agriculture, University of Western Australia, Perth, WA, Australia

OPEN ACCESS

Edited by:

Rong-Rong He, Jinan University, China

Reviewed by:

Xiaogang Wang, Beihang University, China Xinluan Wang, Shenzhen Institutes of Advanced Technology (CAS), China

*Correspondence:

Jiake Xu jiake.xu@uwa.edu.au Dingkun Lin lindingkuntcm@126.com

[†]These authors have contributed equally to this work

Specialty section:

This article was submitted to Ethnopharmacology, a section of the journal Frontiers in Pharmacology

Received: 20 July 2019 Accepted: 24 October 2019 Published: 25 November 2019

Citation:

He J, Li X, Wang Z, Bennett S, Chen K, Xiao Z, Zhan J, Chen S, Hou Y, Chen J, Wang S, Xu J and Lin D (2019) Therapeutic Anabolic and Anticatabolic Benefits of Natural Chinese Medicines for the Treatment of Osteoporosis. Front. Pharmacol. 10:1344. doi: 10.3389/fphar.2019.01344 Osteoporosis is a bone disease characterized by increasing osseous fragility and fracture due to the reduced bone mass and microstructural degradation. Primary pharmacological strategies for the treatment of osteoporosis, hormone replacement treatment (HRT), and alendronate therapies may produce adverse side-effects and may not be recommended for long-term usage. Some classic and bone-specific natural Chinese medicine are very popularly used to treat osteoporosis and bone fracture effectively in clinical with their potential value in bone growth and development, but with few adverse side-effects. Current evidence suggests that the treatments appear to improve bone metabolism and attenuate the osteoporotic imbalance between bone formation and bone resorption at a cellular level by promoting osteoblast activity and inhibiting the effects of osteoclasts. The valuable therapies might, therefore, provide an effective and safer alternative to primary pharmacological strategies. Therefore, the purpose of this article is to comprehensively review these classic and bone-specific drugs in natural Chinese medicines for the treatment of osteoporosis that had been deeply and definitely studied and reported with both bone formation and antiresorption effects, including Gynochthodes officinalis (F.C.How) Razafim. & B.Bremer (syn. Morinda officinalis F.C.How), Curculigo orchioides Gaertn., Psoralea corylifolia (L.) Medik Eucommia ulmoides Oliv., Dipsacus inermis Wall. (syn. Dipsacus asperoides C.Y.Cheng & T.M.Ai), Cibotium barometz (L.) J. Sm., Velvet Antler, Cistanche deserticola Ma, Cuscuta chinensis Lam., Cnidium monnieri (L.) Cusson, Epimedium brevicomum Maxim, Pueraria montana (Lour.) Merr. and Salvia militiorrhiza Bunge., thus providing evidence for the potential use of alternative Chinese medicine therapies to effectively treat osteoporosis.

Keywords: osteoporosis, natural Chinese medicine, osteogenesis, osteoclastogenesis, antibone resorption, therapeutic, osteoblasts, osteoclasts

Abbreviations: ALP, alkaline phosphatase; BMC, bone mineral content; BMD, bone mineral density; BMP-2, bone morphogenetic protein 2; CTSK, cathepsin K; HRT, hormone replacement therapy; hAFSCs, human amniotic fluid-derived stem cells; IGF-I, insulin-like growth factor-I; NFATc1, nuclear factor of activated T cells cytoplasmic 1; OBs, osteoblasts; OCs, osteocalcin; OPG, osteoprotegerin; OPN, osteopontin; Osx, osterix; RANKL, receptor activator of NF-κB ligand; ROS, reactive oxygen species; Runx2, runt-related transcription factor 2; TRACP, tartrate-resistant acid phosphatase.

INTRODUCTION

Osteoporosis may result from imbalanced bone metabolism leading to a systemic deterioration in bone mass and bone microstructure, characterized by skeletal fragility and an increased risk of bone fracture (Albright, 1947; Rachner et al., 2011). Almost one in three women and one in five men would experience one bone fracture in their life after 50 years of age, resulting from osteoporosis (Sozen et al., 2017). Furthermore, the risk of additional fractures will rise exponentially with each incidence of fracture (Lorentzon and Cummings, 2015). Osteoporosis is, therefore, a debilitating disease for sufferers, leading to reduced quality of life, and places a large economic burden on society (Curtis et al., 2016). Thus, medical intervention is imperative to provide adequate care for patients and improve societal health. Moreover, it is necessary to continue research leading to the development of a medical treatment that may effectively treat and potentially prevent osteoporosis (Wang et al., 2017b).

Currently, there are numerous pharmacological products used for the treatment of osteoporosis in the clinic (Pavone et al., 2017). Hormone replacement therapy (HRT) and bisphosphonates are the primary therapeutic strategies for bone loss diseases including osteoporosis (He et al., 2017). Long-term HRT may significantly increase the risk of endometrial and mammary cancer, and coronary heart disease and other cardiovascular diseases (Dinger et al., 2016a; Dinger et al., 2016b). While bisphosphonates may lead to osteonecrosis of the long bones and jaws (Spivakovsky, 2017; Lungu et al., 2018). These adverse side-effects limit the clinical use of HRT and bisphosphonates. Therefore, alternative therapeutic agents are required to develop medicines for the treatment of osteoporosis that are less likely to have adverse side-effects.

Traditional Chinese medicine (TCM) has become increasingly popular due to its effectiveness in treating diseases, with fewer side-effects. Natural Chinese medicine has been widely and effectively used to treat a variety of orthopaedic diseases, including bone fractures, rheumatism, and osteoporosis (Mukwaya et al., 2014; He et al., 2017; Suvarna et al., 2018). Some TCMs are the most classical and bone-specific of drugs when applied to the treatment of bone loss and bone fracture diseases, with the effects on the growth and development of skeleton tissue (Shu et al., 2015; Wang et al., 2019a). Recent scientific reports suggest that these natural Chinese medicine therapies appear to have both the anabolic and anticatabolic effects for the treatment of osteoporosis by promoting bone formation and attenuating imbalanced bone resorption, leading to improved bone mineral density and biomechanical properties, and reduced bone microstructural degradation (Figure 1) (He et al., 2017; Wang et al., 2017b; Suvarna et al., 2018). Further, in vitro findings indicate that these natural medicines may enhance the proliferation and survival of osteoblasts, and they could induce the differentiation of osteoblast cells from bone mesenchymal stem cells (MSCs). While the bone catabolic effects of osteoclastogenesis and bone resorption were effectively inhibited (Figure 2).





Therefore, we selected the natural Chinese medicines, which are the classical bone-specific drugs for the bone loss and fracture diseases in clinic, including Gynochthodes officinalis (F.C.How) Razafim. & B.Bremer (syn. Morinda officinalis F.C.How), Curculigo orchioides Gaertn., Psoralea corylifolia (L.) Medik, Eucommia ulmoides Oliv., Dipsacus inermis Wall. (syn. Dipsacus asperoides C.Y.Cheng & T.M.Ai), Cibotium barometz (L.) J. Sm., Velvet Antler, and so on. Then, according to the literature search of PubMed, each natural Chinese medicine was searched with the keywords of bone, osteoporosis, bone loss, osteolysis, bone formation, skeleton, osteoclastogenesis, antibone osteogenesis, resorption, bone resorption, bone absorption, therapeutic, osteoblasts, osteoclasts, bone mesenchymal stem cell, differentiation, apoptosis, formula, and combination, accompanied with the Boolean connectors of "AND", "OR", and "NOT". After careful reading, the medicines with both anabolic and anticatabolic effects were included and reviewed. Medicines without osteoporosis-related research, or without both anabolic and anticatabolic benefits, or with flaws in experimental design were excluded. Despite the massive amount of experimental data regarding antiosteoporotic effects by the natural Chinese medicines from the in vivo and in vitro studies, high-quality clinical trials are lacking. Additionally, this review highlights

and evaluates the scientific evidence for the potential use of natural Chinese medicines in the treatment of osteoporosis with both therapeutic anabolic and anticatabolic benefits, and their potential mechanisms of action.

NATURAL CHINESE MEDICINES

Gynochthodes officinalis (F.C.How) Razafim. & B.Bremer (syn. Morinda officinalis F.C.How)

Gynochthodes officinalis (F.C.How) Razafim. & B.Bremer (syn. *Morinda officinalis* (F.C.How) (MO, "Ba-Ji-Tian") is a famous natural herb in Chinese medicine, containing many bioactive components including oligosaccharides, iridoid glycosides, and bajijiasu (Chen et al., 2014; Wu et al., 2015b; Li et al., 2016a). Recent studies have found that it could enhance sexual and reproductive function; and may ameliorate Alzheimer's disease *via* the microbiota-gut-brain axis, providing benefits including improved memory and learning abilities (Wu et al., 2015b; Chen et al., 2017a). Additionally, the components of polysaccharides, monotropein and bajijiasu could act as potential agents to treat osteoporosis.

The protective effects of MO root extract on ovariectomyinduced bone loss have been reported (Li et al., 2009). Findings



FIGURE 2 | Natural Chinese medicine could promote the proliferation (Pr) and differentiation of osteoblasts and bone mesenchymal stem cells, enhance the osteogenesis ability, and inhibit the apoptosis (Ap) of osteoblasts induced by oxidative stress. While the osteoclastogenesis and bone-resorption function of osteoclasts are inhibited by their beneficial effects.

indicate that MO root extract could enhance the bone mineral content (BMC) and bone mineral density (BMD) of the tibia in ovariectomized (OVX) rats (Li et al., 2009). The levels of serum phosphorus (P), calcium(Ca), and osteoprotegerin (OPG) increased, and the levels of serum deoxypyridinoline crosslinks to creatinine ratio (DPD/Cr), tartrate-resistant acid phosphatase (TRAcP), adrenocorticotropin (ACTH), and corticosterone decreased, but did not reverse the levels of serum alkaline phosphatase (ALP), tumor necrosis factor-a (TNF-a), or interleukin-6 (IL-6) (Li et al., 2009). Correspondingly, MO capsules attenuated the ovariectomy-induced loss of bone mass by enhancing bone strength, and reducing further deterioration of the trabecular microarchitecture of the femurs in rats, which was associated with increased serum osteocalcin (OC) levels (Li et al., 2014b). Polysaccharides are the extract isolated from MO that could significantly elevate whole femoral BMD as compared with the osteoporosis group. The mineral levels of Ca, P, Mg, Zn, Mn, Cu, and Fe in the femur were enhanced dose-dependently (Zhu et al., 2008). Monotropein is a natural compound extracted from MO that appears to exhibit antiosteoporotic effects by increasing BMC, BMD, bone volume fraction (BVF), and attenuating trabecular microstructural degradation in OVX rodents (Zhu et al., 2008; Zhang et al., 2016c). In vivo findings from a disuse-model of osteoporosis indicate that MO driedroot extracts exhibit both bone formation activity and appear to suppress bone resorption. Mornidae Radix (MR), the dried-root of MO, was found to inhibit the osteoporosis-induced decrease of hind limb thickness, tibia failure load, BMD, and tibia Ca and P levels (Seo et al., 2005). Histomorphometry results indicated that it could remarkably protect tibiae's bone parameters, including volume, length, and cortical and trabecular thickness. Furthermore, both preventive and therapeutic studies indicated that the effects of MR appear to be favourable for both the prevention and treatment of osteoporosis (Seo et al., 2005).

Consistently, in vitro studies have found that MO and its bioactive components appear to provide potentially therapeutic treatment for osteoporosis, and that the molecular mechanisms appear to act via receptor activator of nuclear factor kappa-B ligand (NF-kB, or RANKL) signalling pathways. For example, rubiadin-1-methyl ether (RBM, a natural anthraquinone compound isolated from the root of MO) may effectively inhibit osteoclastic bone resorption by blocking the NF-KB pathway (He et al., 2018). Bajijiasu was found to inhibit osteoclast formation and bone resorption via the mediation of RANKL signalling pathways (Hong et al., 2017a). Expression of RANKL-induced marker genes, including cathepsin K (CTSK), nuclear factor of activated T cells cytoplasmic 1 (NFATc1), TRAcP, vacuolar-type H+-ATPase V0 subunit D2 (V-ATPase d2), and matrix metalloproteinase-2 (MMP-2) was inhibited by Bajijiasu (Hong et al., 2017a). Further *in vitro* studies of anthraquinones isolated from MO have identified these molecules as potential inhibitors of osteoclastic bone resorption and enhancers of osteoblastic bone-forming activity (Wu et al., 2009; Bao et al., 2011). MO-anthraquinones have demonstrated the ability to decrease the formation of bone resorption pits, the number of multinucleated osteoclasts, and the activity of tartrate resistant acid phosphates (TRAP) in an osteoblastbone marrow coculture system, in addition to expediting the apoptosis of osteoclasts (Bao et al., 2011). MO-anthraquinones also appear to promote osteoblastic bone-forming activity by increasing osteoblast proliferation and ALP activity (Wu et al., 2009). Investigation of the polysaccharides extracted from MO revealed that osteogenic inulins of MO promote the proliferation, differentiation, and mineralization of osteoblast cells, as confirmed by the up-regulation of marker genes, including runt-related transcription factor 2, osterix, osteocalcin, bone sialoprotein, and OPG (Jiang et al., 2018).

Taken together, these findings suggest that MO and its bioactive components might provide an important therapeutic role for the treatment and prevention of osteoporosis. MO appears to provide potentially effective treatment for osteoporosis by the attenuation of bone loss and trabecular microstructural degradation, the enhancement of bone mineral density and bone mass, the promotion of osteoblastic bone-forming activity, and the inhibition of osteoclastic bone resorption by direct effects on osteoclasts and effects mediated via known signalling pathways. In the in vitro research, the effective concentration of Bajijiasu compound was found to be 0.8 mM (Hong et al., 2017a), which may be higher than compounds in other in vitro experiments, and the clinical usage may be increased to get better curative effects, therefore, a careful caution of the safety should be observed. Further research is needed to develop the therapeutic potential of this natural medicine.

Curculigo orchioides Gaertn

There are more than 20 species of *Curculigo* plants in the world. They are native to the tropical and subtropical regions of Asia, Africa, South America, and Oceania (Nie et al., 2013). *Curculigo orchioides* Gaertn (CO, "Xian Mao"), one of curculigo species, is used to relieve the symptoms of limb weakness, lumbar and knee joint arthritis, and diarrhea in Chinese medicine (Tan et al., 2019). Recent studies have found that CO also appears to have antioxidant properties and anticancer potential, and may be used as an antiosteoporotic herb (Cao et al., 2008; Ramchandani et al., 2014; Hejazi et al., 2018).

CO is used in Chinese medicine for the treatment of postmenopausal osteoporosis and the antiosteoporotic effects of CO have been investigated *in vivo* (Cao et al., 2008). Administration of CO prevented trabecular bone loss in the tibia of ovariectomized rats by inhibiting bone resorption and increasing serum phosphorus, calcium, and OPG levels, without affecting the body or uterus mass (Cao et al., 2008). The serum levels of bone resorption related makers, DPD/Cr, TRAcP, ACTH, and corticosterone were decreased following CO administration (Cao et al., 2008). CO may also enhance bone formation upon induction during wound healing (Wong et al., 2007a).

Curculigoside (CCG, a phenolic glycoside) is the main bioactive compound of CO and appears to have both osteogenic and antiosteoclastic effects (Liu et al., 2014a). A recent *in vitro* study evaluated the effect of CCG on osteogenic differentiation of human amniotic fluid-derived stem cells (hAFSCs) and found that CCG up-regulated osteogenic activity in a dose-dependent manner, including increased expression of osteopontin (OPN) and Collagen I, increased ALP activity and calcium deposition (Liu et al., 2014a). Furthermore, the ratio of OPG to RANKL increased, indicating that osteoclastogenesis was inhibited. The simultaneous up-regulation of β -catenin and Cyclin-D1 indicate that these effects appear to be mediated *via* the Wnt/ β -catenin signalling pathway (Liu et al., 2014a). The *in vitro* effect of CCG on osteogenic differentiation has also been investigated in relation to BMSCs, and the data indicate that CCG may promote the proliferation and osteogenic differentiation of BMSCs (Shen et al., 2013).

The antioxidant properties of CO are also thought to be attributable to CCG, and may play a critical role in attenuating osteoporosis pathophysiology (Wang et al., 2012b). The antioxidant protective effects of CCG have therefore been investigated in rat calvarial osteoblasts exposed to hydrogen peroxide (H_2O_2) , and findings indicate that CCG significantly reduce the cytotoxic effects of H₂O₂ by reducing the production of reactive oxygen species (ROS) by osteoblasts, and recovering the levels of osteoblast differentiation markers, including ALP, calcium deposition, and runt-related protein 2 (Runx2) (Wang et al., 2012b). CCG has also been shown to protect rat calvarial osteoblasts from dexamethasone (DEX) induced cytotoxicity by regulating osteoblast proliferation, differentiation, and proinflammatory cytokine levels (Zhu et al., 2015). The effects of DEX on the levels of osteoblast differentiation markers, including ALP, OPG, β-catenin, and RANKL was reversed, indicating that CCG may be a suitable compound for the treatment of osteoporosis (Zhu et al., 2015). Accordingly, metabolic analysis indicates that CCG via its main metabolite, M2, could produce these antiosteoporotic effects (Wang et al., 2017a).

Further *in vitro* and rat calvarial studies have shown that compounds isolated from CO appear to have antiosteoporotic properties, including the promotion of osteoblast proliferation and differentiation, increased bone-forming activity, and the inhibition of osteoclastic bone resorption. The bioactive compounds isolated from MO include novel homogeneous polysaccharides, COP90-1 and COP70-3, and phenolic glycosides (Jiao et al., 2009; Wang et al., 2013b; Wang et al., 2017c; Wang et al., 2019b).

Briefly, the promotion on osteogenesis of *C. orchioides* might be the beneficial effect leading to attenuation of osteoporosis, and the inhibition of reactive oxygen species is a great property of this medicine. Additionally, some compounds isolated from CO exhibit the suppression of osteoclastic bone resorption. Further research is necessary to discover the molecular mechanisms, signalling pathways, and bioactive constituents of CO; and to determine the molecular basis of the potential relationship between the antioxidant effects of CCG and osteoporosis pathophysiology in osteoclasts and osteoblasts.

Psoralea corylifolia (L.) Medik

Psoralea corylifolia (L.) Medik (PC, "Bu Gu Zhi") is widely used in Asia for medicinal purposes; the dried ripe seeds are the active component of the plant. PC is used in Chinese pharmacopoeia and TCM formulas for the treatment of skin diseases, cardiovascular disease, nephritis, and osteoporosis (Zhang et al., 2016a). The primary bioactive compounds of PC are coumarins (e.g., psoralen), flavonoids (e.g., neobavaisoflavone, bavachalcone, bavachin, and corylin), and meroterpenes (e.g., bakuchiol) (Zhang et al., 2016a). Additional components of PC have immunoregulatory, antioxidant, and neuroprotective properties for the treatment of Parkinson's disease (Jing et al., 2017). PC may also have therapeutic potential for the treatment of prostate cancer (Lin et al., 2018).

The potential beneficial effects of PC in relation to osteoporosis treatment has been evaluated in animal models (Miura et al., 1996; Tsai et al., 2007). Administration of PC was found to decrease urinary calcium secretion and decrease serum calcium in OVX rats, resulting in improvements in BMD and bone formation, and may have therapeutic potential for the treatment of postmenopausal osteoporosis (Tsai et al., 2007). Complementary investigation indicates that PC may have therapeutic benefits for the broader treatment of bone pathological conditions, including bone fracture, osteomalacia, and osteoporosis (Miura et al., 1996). Furthermore, two compounds of PC, bavachin (BA, a flavonoid), and bakuchiol (BK, a meroterpene) have been shown to have osteoanabolic activity by inducing osteoblast differentiation. And maybe bioactive components of PC that could provide protection against post-menopausal bone loss (Lim et al., 2009; Weng et al., 2015). Bavachin appears to be more effective in stimulating cell proliferation, whereas bakuchiol appears to have a greater effect on osteoblast differentiation (Li et al., 2014a). The molecular structures of bavachin and bakuchiol that hypothetically produced variation in the osteogenic effects between these two and additional compounds of PC depending on its position, is a prenyl group side chain (Li et al., 2014a). Further research is needed to investigate the effect of changing the position of the prenyl group on the strength of osteoblast activity of PC compounds (Li et al., 2014a). The osteogenic effects of BA and BK appear to be produced by up-regulation of the Wnt signalling pathway, and produce increased ALP levels, Ca serum concentration, and BMD (Lim et al., 2009; Weng et al., 2015). Further in vitro findings suggest that BK inhibits RANKL-induced osteoclast differentiation and bone resorption by disrupting the AKT and AP-1 signalling pathways (Chai et al., 2018).

Psoralen (PSO), an active coumarin-compound of PC, is reportedly a promoter of bone mass and has been tested in a rodent ovariectomized model of osteoporosis (Yang et al., 2012). Findings showed that PSO produced an increase in trabecular thickness, and up-regulated osteogenic markers, including osteocalcin and ALP (Yang et al., 2012). Furthermore, the *OVX*induced gene expression profile was reversed by PSO treatment, appearing to be mediated *via* the Notch signalling pathway, and resulted in regulation of the differentiation of bone mesenchymal stem cell, thus indicating the osteogenic potential of PSO for the treatment of postmenopausal osteoporosis (Yang et al., 2012). In vitro findings indicate that PSO may also attenuate osteoclast differentiation and bone resorption *via* inhibition of the AKT and AP-1 signalling pathways (Chai et al., 2018).

Neobavaisoflavone (NBIF), an isoflavone compound of PC, may also have an osteogenic activity that could be applied for the

treatment of bone fractures, osteomalacia, and osteoporosis (Don et al., 2012). In vitro findings demonstrated that NBIF enhanced osteogenesis in a concentration-dependent manner by increasing ALP activity, increasing the expression of bone-specific matrix proteins (including type I collagen, Col-I), increasing osteocalcin and bone sialoprotein levels, up-regulating the expression of Runx2 and osterix (OSX), and by the formation of bone nodules (Don et al., 2012). Further experiments indicate that inhibition of the p38 receptor results in the inhibition of NBIF osteogenic activity, and, that NBIF also increases the active phosphorylated level of p38 in a concentration-dependent manner. Therefore, the osteogenic activity of NBIF appears to be mediated by activation of the p38-dependent signalling pathway (Don et al., 2012).

The flavonoid compound, bavachalcone, isolated from PC may be an effective herbal compound to inhibit osteoclast activity and attenuate osteoporosis-induced bone loss (Park et al., 2008). Bavachalcone was found to inhibit osteoclast formation of primary culture osteoclast precursor cells, *in vitro* (Park et al., 2008). In the presence of bavachalcone, the activity of the osteoclast differentiation factor, NFkB (RANKL), was reduced, leading to inhibition of osteoclastogenesis *via* suppression of transcription factors, c-Fos, NFATc1, and by blocking MEK, ERK, and Akt signalling (Park et al., 2008).

Taken together, these findings suggest that *P. corylifolia* appears to be an important therapeutic agent in Chinese medicine for the treatment of osteoporosis. The compound of bakuchiol should be studied more deeply for it appears to have dual properties of promoting osteogenic activity and ameliorating osteoclastic bone resorption to attenuate osteoporosis, and it has a higher promotion on osteoblast differentiation than bavachin (Li et al., 2014a; Chai et al., 2018). Further research is needed to isolate, characterize, and investigate the effect of variation of the molecular structure of the active compounds of PC on osteoblast and osteoclast activity, including their biochemical effects and signalling pathways.

Eucommia ulmoides Oliv.

Eucommia ulmoides Oliv. (EU, "Du Zhong") is a plant, and the dried bark of EU is used in TCM and as a food source (Hussain et al., 2016). EU has pharmacological properties as an antioxidant, anti-inflammatory, antimicrobial, anticancer, cardioprotective, and neuroprotective agent that have been applied for the treatment of cardiovascular disease, sexual dysfunction, cancer, metabolic disease, neurological disease, rheumatoid arthritis, osteoarthritis, and diabetes (Xie et al., 2015; Hussain et al., 2016; Wang et al., 2016a; Do et al., 2018). The bioactive compounds of EU include lignans, iridoids, phenolics, steroids, and flavonoids (Hussain et al., 2016). Additional studies have found that the EU could also be effective in the treatment of osteoporosis.

Du Zhong cortex extract (DZCE) has been evaluated to investigate the potential protective benefits against lead-induced, estrogen deficiency-induced, and disuse-induced osteoporosis (Zhang et al., 2009; Pan et al., 2014; Qi et al., 2019). In a rodent model of lead acetate-induced bone loss, DZCE attenuated the loss of BMD of the lumbar spine and femur, and restored serum calcium, phosphorous, ALP, osteocalcin, and RANKL to normal levels (Qi et al., 2019). Furthermore, antiosteoclastic activity was indicated by the effect of DZCE on adjusting the serum OPG/ RANKL ratio to normal values (Qi et al., 2019). In an OVX-rat model of estrogen deficiency-induced osteoporosis, higher doses of DZCE prevented further deterioration of the biomechanical properties of the femur, including maximum stress and Young's modulus, which was accompanied by attenuation of loss of BMD (Zhang et al., 2009). These findings were supported by micro-CT analysis indicating that the parameters of BMD and bone thickness were improved by higher doses of DZCE, and because the levels of bone turnover markers, osteocalcin, ALP, and deoxypyridinoline (DPD) were decreased (Zhang et al., 2009). Total lignans (TL) extracted from EU-cortex, was also found to inhibit the loss of bone mass due to estrogen deficiencyinduced osteoporosis in a rodent model (Zhang et al., 2014). In vivo findings demonstrated improvements in the biomechanical quality of the femur, in terms of maximum stress and Young's modulus, and micro-CT analysis showed prevention of further trabecular microstructural degradation (Zhang et al., 2014). In vitro findings showed that TL promoted the proliferation and differentiation of osteoblasts; and inhibited osteoclastogenesis, by an increase in OPG and decrease in RANKL expression (Zhang et al., 2014). In a rodent disuse-induced osteoporosis model, DZCE treatment improved bone strength and prevented trabecular microstructural degradation, and reduced the levels of bone turnover markers, including TRAcP, DPD, and OC (Pan et al., 2014). Additional findings in rodents indicate that EU could promote longitudinal bone growth by increasing chondrogenesis of the tibial growth plate, and increasing levels of BMP-2 and IGF-1 (Kim et al., 2015). Taken together, these results suggest that DZCE may provide effective treatment for osteoporosis by attenuating bone loss, both by the formation of bone, and by inhibiting osteoclastic bone resorption; and may promote longitudinal growth of the long bones.

5-(hydroxymethyl)-2-furaldehyde (5-HMF) is a bioactive compound isolated from Eucommiae Cortex that has been shown to promote osteogenesis and inhibit adipogenesis (Tan et al., 2014). In vitro findings demonstrated that 5-HMF induction of BMSCs in normal medium up-regulated the expression of osteogenesis-markers (ALP, COL1alpha1, OC, and OPN), and that 5-HMF decreased the expression of adipogenesis-markers (PPARy, FABP4, C/EBPa, and LPL) by BMSCs in adipogenic induction medium (Tan et al., 2014). Furthermore, mineralized nodule formations were produced by BMSCs induced by 5-HMF cultured in both normal and adipogenic-inducing medium, indicating the potent osteogenic, antiadipogenic, and antiosteoporotic properties of 5-HMF (Tan et al., 2014). Further research is necessary to investigate the molecular mechanisms involved with the osteogenic, and potential pro-osteoblastic and antiosteoclastic effects of 5-HMF and additional bioactive components of EU-cortex (Ha et al., 2003).

The leaves and seeds of EU may also be of potential benefits for the treatment of osteoporosis (Li et al., 2011; Zhang et al., 2012). In a combined estrogen deficiency-induced osteoporosis and obesity rodent model, Eucommia leaf extract (ELE) treatment decreased body weight and BMI, and increased tibial and femoral BMD, and increased bone strength, which appeared to be the effects of restoring bone metabolism, respective of bone formation, and adsorption (Zhang et al., 2012). Further rodent study indicates that administration of total glycosides from Eucommia ulmoides seed (TGEUS) could enhance the BMD and the microarchitecture parameters of the femur in healthy rats (Li et al., 2011). Iridoid compounds isolated from the leaves of the EU may be the bioactive components that render the therapeutic effects (Takamura et al., 2007).

Taken together, we think that the herb of E. ulmoides is a great medicine to treat bone loss diseases, for many parts of EU including the cortex, leaf, and seed, might have potential therapeutic benefits for the treatment of osteoporosis, as depicted above (Li et al., 2011; Zhang et al., 2012; Qi et al., 2019). Furthermore, in vivo and in vitro findings show that they provide effective treatment for osteoporosis by increasing bone strength and preventing trabecular microstructural degradation, promoting osteogenesis, and inhibiting bone resorption. Usually, the cortex is the major part of the EU that is used to treat bone, cardiovascular, and sexual diseases. Based on the positive evidence and effects of the leaf and seed, more research is needed to isolate and characterize the bioactive molecules of EU, and compare the effects between cortex, leaf, and seed for the treatment of osteoporosis, including the mechanisms that produce the potential therapeutic effects among them.

Dipsacus inermis Wall.

Radix Dipsaci (RD, "Xu Duan") is the dried roots of *Dipsacus inermis* Wall., and it is used in Chinese medicine to strengthen bone ("Xu Duan" means "to promote the growth of bones to correct bone fractures"). Recently, studies have reported that RD may be a beneficial bone formation agent to treat osteoporosis (Liu et al., 2009; Niu et al., 2012; Liu et al., 2012b).

The bone formation properties that may provide therapeutic benefit for the treatment of osteoporosis have been investigated in recent animal studies. In ovariectomy-induced postmenopausal rodent models, RD treatment prevented the loss of bone mass and trabecular microstructural degradation, and improved bone strength (Liu et al., 2009; Liu et al., 2012b). These results may be attributed to altered bone remodeling, as evidenced by a decrease in the level of bone turnover markers, including ALP, OC, and DPD. Furthermore, the effects of ovariectomy on OPG and RANKL levels of osteoblasts and BMSCs were reversed by treatment with RD (OPG levels increased and RANKL levels decreased), indicating the osteogenic effect of RD on molecular biomarkers (Liu et al., 2012b). The osteogenic effect of the dichloromethane fraction of RD on BMSCs has been demonstratedin vitro by the formation of calcified nodules, increased ALP activity, increased expression of bone sialoprotein (BSP) and OC (Kim et al., 2011). An active single compound isolated from the dichloromethane fraction of RD, hederagenin 3-O-(2-O-acetyl)-a-L-arabinopyranoside, significantly increased ALP, OC, and BSP levels (Kim et al., 2011). A hindlimb unloading rodent model of osteoporosis investigated the effect of RD treatment on BMD and bone microarchitecture, and showed that the biomechanical properties were enhanced (Niu et al., 2015a). Oral administration of RD improved the mechanical strength of bone, BMD, BMC, and bone turnover markers, including urinary calcium and phosphorus excretion. Micro-CT analysis showed that RD prevented trabecular microstructural degradation, improved the bone volume fraction, and improved tissue mineral density and content (Niu et al., 2015a). Healthy rats fed with Radix Dipsaci extract (RDE) showed an anabolic systemic skeletal effect by increased bone density and altered bone histomorphology (Wong et al., 2007b). An increase of bone trabeculae of 11.82% was measured, indicating that bone density was increased, and the bone histomorphology has also been improved by the enhancement of BV/TV (increased 4.5%), and improvements in BS/TV, Tb.N, and the reduction in Tb.Sp (Wong et al., 2007b). Taken together, these findings suggest that RD may provide beneficial therapeutic effects for the treatment of osteoporosis.

Further studies of the bioactive components of RD indicate that several compounds, including saponins and iridoid glycosides extracted from RD and may produce biological effects (Kim et al., 2011; Tao et al., 2019). The antiosteoporotic effect of RD total saponins (RTS) may be related to its effect on osteoblast and osteoclast cells (Niu et al., 2012; Niu et al., 2015b). Oral administration of RTS has been shown to prevent OVX-induced loss of bone mass in rats, indicated by decreased levels of bone turnover markers, including urinary calcium, and phosphorous excretion, and by increasing the biomechanical strength of bone and preventing trabecular microstructural degradation (Niu et al., 2012). In vitro findings determined that RTS promoted osteoblastic cell maturation and differentiation, increased ALP and OC levels, and increased the synthesis of BMP-2, leading to increased bone formation (Niu et al., 2012; Niu et al., 2015b). The positive effect of RTS on osteoblast cells appears to be mediated via the BMP2/mitogen-activated protein kinase (MAPK)/Smad1/5/8-dependent Runx2 signalling pathway (Niu et al., 2015b). Furthermore, RTS inhibited osteoclastogenesis by increasing the expression of OPG and decreasing the expression of NF-KB (RANKL) (Niu et al., 2012). Additionally, asperosaponin VI is a saponin isolated from RD that has been shown to promote the proliferation, differentiation and mineralization of osteoblastic cells, and these effects appear to be mediated by BMP-2 synthesis, and activation of p38 and ERK1/2 signalling (Niu et al., 2011).

Collectively, these findings suggest that RD appears to provide beneficial therapeutic effects for the treatment of osteoporosis, including attenuating bone loss and increasing bone strength and quality, that may be attributed to their improved physiological bone remodelling in vivo. At cellular and molecular levels, the RD and its bioactive components appear to promote osteoblast differentiation, proliferation, maturation, and mineralization, and may inhibit osteoclastogenesis. The bone research-related amimal models were rich in these studies including ovariectomyinduced postmenopausal rodent models, hindlimb unloading model, and healthy rats to detect the bone metabolism. While some treatment ingredients of Radix Dipsaci, such as ethyl alcohol or crude extractions, are not well studied. It is necessary for further research to identify the efficient compounds and to develop the therapeutic potential and mechanism of RD in the treatment of osteoporosis.

Cibotium barometz (L.) J.Sm

Cibotium barometz (L.) J.Sm (CB, "Gou Ji") is well-known in TCM and its hairs are a staple ingredient used in ointments, such as an antihaemorrhagic agent for wound healing poultices (Wu and Yang, 2009). The rhizome of CB contains anti-inflammatory properties and is used in the treatment of diseases including rheumatism, lumbago, sciatica, and dysuria in the aged (Wu and Yang, 2009). CB is thought to have properties that nourish bone and improve gonadal function. And it is frequently used in herbal remedies for the treatment of osteoporosis (Zhao et al., 2011).

Due to the potential benefits of CB for the treatment of osteoporosis, research has been performed to determine the effects of CB in vivo and in vitro. In a rodent model of post-menopausal estrogen-deficiency-induced osteoporosis, daily administration of the extract of CB (CBE) was shown to prevent femur total BMD loss in OVX-rats, which appeared to be associated with a decrease in skeletal remodeling, as evidenced by decreased levels of bone turnover markers OC, ALP, DPD, and urinary excretions of calcium and phosphorous (Zhao et al., 2011). Furthermore, the CBE treatment also appeared to enhance bone strength and prevent trabecular microarchitectural degradation as determined by improvements in micro-CT microstructural parameters. When compared with the untreated model rats, CBE treatment significantly increased the bone maximum stress, energy, and Young's modulus of OVX rats (Zhao et al., 2011). CBE treatment performed favourably in comparison with estrogen-therapy in terms of its effect on body weight and uterine weight (Zhao et al., 2011). Additional in vitro experiments have been performed to evaluate the potential effectiveness of the constituents of the rhizome of CB in the treatment of osteoporosis (Nguyen et al., 2009; Xu et al., 2014). Findings from these experiments indicate that these CB-rhizome constituents could promote the proliferation and differentiation of rat osteoblasts, and thus be potential therapeutic agents for the treatment of osteoporosis (Xu et al., 2014). Furthermore, compounds isolated from CB-rhizome appeared to inhibit osteoclastogenesis from primary bone marrow macrophages (BMMs) when cultured in an inductive medium, without any adverse effects on the viability of precursor BMM cells (Nguyen et al., 2009).

Therefore, evidence suggests that CB could be used as an effective therapeutic agent for the treatment of osteoporosis. The ingredients of CB in some research were still not very clear or definite, which are obtained by ethanol extraction from the dried and crude *C. barometz* (Zhao et al., 2011). Further research is needed to isolate and characterize the bioactive constituents of CB, and to determine the molecular mechanisms and signalling pathways by which they produce therapeutic effects.

Velvet Antler

Velvet Antler (VA, "Lu Rong") is the precalcified cartilaginous antler, in TCM commonly obtained from the Silka deer and Red deer, that is used as an effective agent to strengthen bone and for the improvement of immune health, physical strength, and sexual function (Zhang et al., 2013; Sui et al., 2014). Velvet antlers regenerate annually in mammals, and ingestion of VA is thought to confer similar benefits upon the consumer (Gilbey and Perezgonzalez, 2012). Both VA and its close relative, deer antler base, are designated medicinal products in the Chinese Pharmacopoeia (Wu et al., 2013). The bone-health promoting properties of VA has been investigated in relation to the treatment of bone diseases including osteoporosis and osteonecrosis (Zhang et al., 2013; Wei et al., 2016).

The effectiveness of VA for the treatment of postmenopausal estrogen deficiency-induced osteoporosis has been investigated using rodent models (Yang et al., 2010a; Tseng et al., 2012; Zhang et al., 2013). Total velvet antler polypeptides (TVAP) administered to OVX-rats were found to prevent bone loss and improve BMD, BMC, and bone microarchitecture, which appeared to be associated with inhibition of IL-1 and Il-6 (Zhang et al., 2013). Furthermore, natural velvet antler polypeptides (nVAP) and synthetic-VAP (sVAP) promoted the proliferation of cartilage and osteoblast-like cells, and inhibited the activity of IL-1a secreted from THP-1 monocytic cells in vitro, indicating that TVAPL and sVAP may be potential therapeutic agents for the treatment of postmenopausal osteoporosis (Zhang et al., 2013). Both VA and VA-blood can be used in TCM to tonify and invigorate bones and tendons, and the combination of middle sections of VA and VA-blood (VAM-B) is thought to have superior pharmacological effects (Tseng et al., 2012). In OVX-rats, treatment with VAM-B improved body weight, and increased the strength of the vertebra and femur, and improved tibial trabecular microarchitecture, which was accompanied by a decrease in ALP levels. The results supported the therapeutic use of VAM-B for the treatment of postmenopausal osteoporosis (Tseng et al., 2012). Additional research has shown that oral administration of VA-blood to OVX-rats could attenuate reduced BMD of the lumbar vertebra and femur, and normalize serum levels of insulin-like growth factor -1 (IGF-1) and testosterone (Yang et al., 2010a).

Interestingly, the effect of fermentation on the bone formation capability of VA was investigated in vitro (Lee et al., 2011). Findings indicated that both non-fermented antler (NFA) and fermented antler (FA) treatment increased the proliferation of osteoblasts and bone matrix proteins, including type I collagen and BSP (Lee et al., 2011). Moreover, FA showed enhanced osteoblast proliferation, increased ALP activity, and increased mineralization compared to NFA and, therefore, the fermentation process may enhance the bone-forming effects of antler (Lee et al., 2011). Furthermore, the effect of fermentation on antiresorptive activity was investigated in vitro, and it appears that the fermentation process may improve the capability of VA to inhibit osteoclast differentiation and signalling activity (Choi et al., 2013). Findings indicate that the extract of FA inhibited RANKL-induced osteoclast differentiation from BMMs by downregulating the expression and activity of NFATc1, which was associated with inhibition of phospholipase Cy2 (PLCy2), a signalling molecule known to affect NFATc1 transcriptional activity (Choi et al., 2013). Thus, research suggests that FA extract may inhibit osteoclastic activity via disrupting PLCy2-NFATc1 signalling, and may provide therapeutic benefit for the treatment of osteoclast-related bone diseases, including osteoporosis (Choi et al., 2013). Furthermore, in vitro studies have shown

that the chloroform extract (CE-C) of deer antler appears to inhibit osteoclast differentiation by suppressing the activation of extracellular signal-regulated kinase (ERK), protein kinase B (PKB/Akt), the inhibitor of kappa B (I- κ B), which would be increased by RANKL under osteoporosis conditions (Li et al., 2007). CE-C also appears to inhibit the bone resorption activity of osteoclasts and disrupt the actin rings, leading to osteoclast apoptosis (Li et al., 2007). Taken together, these findings suggest that VA and CE-C may be effective compounds for the treatment of osteoporosis.

Further research has compared the osteogenic capacity of VA from different sections of antler on longitudinal bone growth and osteoporosis (Tseng et al., 2014; Kim et al., 2016). In vivo and *in vitro* findings indicate that VA promotes longitudinal bone growth in adolescent rats through enhanced BMP-2 expression, osteogenic gene expression, including collagen, ALP, and OC, and by promoting the proliferation, differentiation, and mineralization of osteoblast-cells (Kim et al., 2016). The osteogenic effects of VA appear to decrease from the upper or distal sections to the basal sections of antler (Tseng et al., 2014; Kim et al., 2016).

Taken together, these findings indicate that VA appears to contain properties to strengthen bone, promote bone growth, and may provide therapeutic benefit for the treatment of osteoporosis by the bone formation and antiresorptive activity. Interestingly, research by Kim and Tseng et al. indicates that the basal sections of antler should be the prior selection for the better osteogenic effects when applied for the treatment of osteprorosis clinically. Further research is needed to identify and characterize the bioactive components of different sections of VA and the molecular signalling mechanisms that mediate their therapeutic effects, under the influence of varying preparation conditions.

Cistanche deserticola Y.C.Maf

Cistanche deserticola Y.C.Maf (CD, "Rou Cong Rong") is used in TCM for the treatment of kidney deficiency, sexual dysfunction, female infertility, and constipation (Fu et al., 2018). Recent reports suggest that CD could promote male fertility, and may attenuate tinnitus in patients with chronic nephritis, in addition to providing effective treatment for osteoporosis (Jiang et al., 2016; Fu et al., 2018; Fan et al., 2019).

In vivo studies using rodent models of postmenopausal estrogen deficiency-induced osteoporosis have shown that Cistanches deserticola-extract (CDE) could provide therapeutic benefits for the treatment of osteoporosis (Liang et al., 2011; Liang et al., 2013). Findings indicate that CDE could dose-dependently enhance the femoral BMD and BMC in OVX rats, and improve biomechanical femur parameters of loading and autobreak including maximum load, maximum displacement, and stress (Liang et al., 2011). In comparison with OVX rats, CDE-treatment rats showed biochemical differences including decreased blood calcium, zinc, and copper levels (Liang et al., 2011). Further *in vivo* investigation of the molecular mechanisms behind the antiosteoporosis effect of CDE indicate that the attenuation of bone degeneration is associated with the regulation of genes involved with bone metabolism, including *Smad1*,

Smad5, TGF- β 1, and TIEG1 (Liang et al., 2013). Additionally, *in vivo* investigation of the effects of CD compound, Cistanoside, on OVX rats indicates that CD may contain both osteogenic and antiosteoclastic properties (Xu et al., 2017). In addition to increasing bone strength, BMD, and improving trabecular microstructure, Cistanoside may also decrease the activity of bone resorption markers including TRAP, DPD, and cathepsin K (Xu et al., 2017). These effects appear to be mediated by down-regulation of TNF-receptor associated factor 6 (TRAF6), which downstream mediates both the inactivation of NF- κ B, to inhibit osteoclast activity, and the stimulation of the PI3K/Akt osteogenic pathway (Xu et al., 2017).

In vitro studies have demonstrated that CD may induce bone formation for the treatment of osteoporosis (Li et al., 2012a; Li et al., 2012b). CD extract increased the expression of ALP, BMP-2, and OPN, and increased bone mineralization by cultured osteoblast cells (Li et al., 2012b). Echinacoside (ECH), a phenolic compound isolated from CD, appears to promote bone formation activity by cultured osteoblast cells, including increased cell proliferation, ALP activity, OC levels, and mineralization, which may be associated with increasing the OPG/RANKL ratio (Li et al., 2012a). Correspondingly, in vivo findings support the positive dose-dependent effects of ECH for the treatment of osteoporosis in OVX rats (Li et al., 2013a). Following ECH treatment for 12 weeks at the doses of 90 mg/kg and 270 mg/kg, the femur BMD was significantly enhanced (Li et al., 2013a). Furthermore, the histomorphological and micro-CT analysis indicated that the deteriorated microarchitecture and biomechanical parameters had been reversed, and that the cortical bone thickness, osteoblasts number, and trabecular thickness were enhanced (Li et al., 2013a). Further research suggests that CD derived compounds may also decrease osteoclastic bone resorption by inhibiting osteoclastogenesis, attenuating RANKL activity, and disabling NFAT and MAPK (Song et al., 2018).

Collectively, these findings suggest that CD may contain properties that are potentially effective for the treatment of postmenopausal osteoporosis. Though the compound of Cistanoside may act as a good representative of *Cistanches deserticola* with the interesting study that it could possess both osteogenic and antiosteoclastogenesis properties, further research is necessary to isolate and characterize the bioactive constituents of CD and to investigate their potential osteogenic and antiresorptive effects, and the molecular signalling pathways affected by Cistanches herb in relation to osteoporosis.

Cuscuta chinensis Lam.

The seed of *C. chinensis* Lam. (CC, "Tu Si Zi") is one of the commonly used herbs in Chinese medicine for the enhancement of sexual function, vision, and birth control (Donnapee et al., 2014). The biological activity of CC includes skin and liver protection, immune regulation, neuroprotection, antioxidant, and anti-inflammatory properties (Donnapee et al., 2014). Current research is investigating the effectiveness of CC for the treatment of osteoporosis (Donnapee et al., 2014).

In vivo and *in vitro* experiments indicate that the bioactive compounds of CC that appear to be effective against osteoporosis

The Antiosteoporotic Effects of Natural Chinese Drugs

are kaempferol, hyperoside, and campesterol (Yao et al., 2005; Yang et al., 2011; Nowak et al., 2017). In a rodent model of postmenopausal estrogen deficiency-induced osteoporosis, kaempferol (isolated from the seeds of CC), increased the femoral BMD and Young's modulus of OVX rats as compared to untreated controls, which was accompanied by reduced bone turnover, increased bone tissue volume ratio and increased trabecular bone perimeter (Nowak et al., 2017).

In vitro studies showed that the extract of CC could dosedependently increase osteoblast-like cell proliferation and mineralization, and that this was accompanied by enhancing ALP activity, increased collagen synthesis, and BMP-2 expression (Yang et al., 2009). Furthermore, CC appears to protect osteoblast cells from tertiary butyl hydroperoxide (TBHP) oxidative stress-induced apoptosis, possibly due to its antioxidant activity and function *via* mitochondria-dependent pathways (Gao et al., 2013). Additional *in vitro* evidence has demonstrated that CC may promote the proliferation and differentiation of osteoblast from their precursor cells to induce mineralized nodule formation and decrease osteoclastic activity as indicated by TRAcP (Yao et al., 2005).

Taken together, these findings indicate that the osteogenic, antioxidant, and antiosteoclastic properties of CC may provide therapeutic benefit for the treatment of osteoporosis. Although the evidence has indicated the promotion on osteogenesis and the inhibition on bone resorption by the compounds, kaempferol, and campesterol, their respective functional mechanisms were still not well studied, not like the researches in Yang et al. (2009) and Gao et al. (2013). However, the compounds in Yang and Gao *et al.* were the aqueous extraction, the certain compound were not well known, therefore, further research is necessary to isolate and characterize the bioactive components of *C. chinensis* including their therapeutic potential and molecular mechanisms for the treatment of bone diseases, including osteoporosis.

Cnidium monnieri (L.) Cusson

Cnidium monnieri (L.) Cusson (CM, "She Chuang Zi") is a commonly used herb in Chinese medicine to alleviate pain and inflammation, improve sexual potency, for the treatment of skin-related diseases and to improve bone strength (Li et al., 2015c). Over 300 compounds have been isolated from CM and the main bioactive constituents appear to be coumarins compounds (Li et al., 2015c). Further research has reported the potential application of CM for the treatment of Parkinson's disease, cancer, and osteoporosis (Zhang et al., 2007b; Hong et al., 2017b; Wang et al., 2019c).

Coumarins extracted from CM have been shown to produce potentially beneficial therapeutic effects for the treatment of osteoporosis (Meng et al., 2004; Zhang et al., 2007b; Ming et al., 2013). The chloroform fraction of CM was found to promote osteoblast activity and three coumarins (osthole, bergapten, and imperatorin) were subsequently isolated for further analysis (Meng et al., 2004). In neonatal rodent calvaria culture, osthole was shown to promote osteoblast proliferation and ALP activity, and to inhibit bone resorption by decreasing the formation, differentiation, and TRACP activity of rat marrow osteoclasts, thus indicating that CM may have potential therapeutic benefits for the treatment of osteoporosis (Meng et al., 2004).

Further in vivo and in vitro investigations of the potential effectiveness of osthole for the treatment of osteoporosis have been performed (Zhang et al., 2017b). Analysis of findings from a mouse femur fracture model indicated that osthole enhanced fracture repair, bone regeneration, and increased bone strength as compared to untreated controls (Zhang et al., 2017b). In vitro findings determined that osthole promoted osteogenesis by osteoblast cells in a time- and concentrationdependent relationship by increasing osteogenic differentiation, ALP activity, and calcium nodule formation (Ming et al., 2013; Zhang et al., 2017b). In addition, osthole appeared to induce osteogenesis via the BMP-dependent signalling pathway as determined by the increased expression of osteogenic-related genes BMP-2, Runx2, osterix (Osx), and OCN, and by the observed inhibition of osteogenesis resulting from antagonization by the BMP-antagonist, noggin (Ming et al., 2013; Zhang et al., 2017b). Further mechanistic analysis demonstrated that the osteogenic effects of osthole appear to be mediated via activation of the cAMP/CREB signalling pathway, which appeared to target the transcription factor, Osx (Zhang et al., 2017b). Additional in vivo evidence suggests that osthole may provide therapeutic benefit for the treatment of postmenopausal osteoporosis, that may be as effective as 17β -estradiol (Li et al., 2002). In OVX rats, both osthole and 17β-estradiol inhibited estrogen deficiencyinduced cancellous bone loss, and increased the biomechanical maximal load of the femoral neck (Li et al., 2002). Osthole could also suppress the urinary deoxypyridinoline (DPD) level of OVX rats. However, osthole did not appear to reduce serum OC levels, nor effects on body or uterus weight as was observed in the 17β -estradiol group, thus indicating that the therapeutic benefits of osthole for the treatment of postmenopausal osteoporosis may not be mediated by the estrogen pathway (Li et al., 2002). Additionally, in healthy rats, osthole was shown to increase total body and femur BMD, trabecular microstructural parameters, and biomechanical properties of maximum load and Young's modulus, which were accompanied by increased serum OC and TRAcP levels (Cheng et al., 2014). Taken together, these findings suggest that osthole may provide potential therapeutic benefit for the treatment of osteoporosis by promoting osteogenic activity and attenuating bone resorption. Further research is necessary to characterize the antiosteoporosis effects of osthole more extensively.

In vitro evidence indicates that imperatorin may also have anabolic potential by stimulating the proliferation and osteogenic activity of MCF-7 cells, and ALP activity of Saos-2 osteoblast cells, possibly by exerting estrogenic properties *via* the estrogenreceptor pathway (Jia et al., 2016). Further research is necessary to investigate the potential therapeutic effects for the treatment of osteoporosis *in vivo*.

Bergapten has also been tested for the potential biopharmacological properties that may be of therapeutic value for the treatment of bone disease, including osteoporosis (Chen et al., 2019a). In vivo investigation of the potential effects of bergapten against ovariectomy-induced osteoporosis in mice, and *in vitro* analysis of RANKL-induced osteoclastogenesis

was performed (Chen et al., 2019a). Findings indicated that bergapten appears to inhibit osteoclastic bone resorption and attenuates RANKL-induced osteoclastogenesis, which is mediated *via* disruption of the NF- κ B and JNK signalling pathways (Chen et al., 2019a). Furthermore, bergapten effectively reduced the activity of NFATc1 and c-fos (osteoclastogenesis associated transcription factors) which decreased the expression of osteoclast differentiation-related genes, and attenuated osteoclastogenesis by BMMs and RAW264.7 cells without any cytotoxic side-effects. Moreover, bergapten disrupted the formation of F-actin rings, which are implicated in bone resorption activity (Chen et al., 2019a).

Overall, *in vivo* and *in vitro* evidence suggests that coumarin compounds isolated from CM, mainly osthole and bergapten, may produce bio-pharmacological effects for the treatment of osteoporosis by acting to increase bone formation and to decrease bone resorption. Interestingly, the level of serum OC was not affected in OVX rats, while it was enhanced by osthole in the healthy rats, indicating that more studies and attention should be paid to the effects of osthole on serum OC. Further research is necessary to extensively characterize the bioactive effects of CM that render potential therapeutic benefit for the treatment of osteoporosis.

Epimedium brevicornum Maxim

Epimedium brevicornum Maxim (EBM, "Yin Yang Huo") is a very popular natural drug been traditionally used to treat bone diseases, pregnancy, and gonad dysfunction in Chinese medicine for thousands of years. It could relieve postmenopausal symptoms and inhibit osteoporosis and other bone loss diseases, while few hyperplastic effects on the uterus were found. These antiosteoporotic effects may be related with the estrogenic properties by the intrinsic phytoestrogens including some of the flavonoids, lignans, sterols, etc. (Wang et al., 2007; Xu et al., 2016). In the systems pharmacology study, there are 77 components in Epimedium possessing the analogous structure to estrogen (Xu et al., 2016). Many of these phytoestrogenic compounds have the beneficial effects to inhibit osteoporosis, including icariin, epimedin A, epimedin B, epimedin C, icariside II and icaritin, epimedoside C, baohuoside I, baohuoside II, etc. (Meng et al., 2005; Huang et al., 2007; Zhai et al., 2013; Liu et al., 2014b; Wang et al., 2016b; Liu et al., 2017b). Among these ingredients, icariin is the main compound of Epimedium brevicornum Maxim. Now, there have been many studies and reviews focusing on its anabolic and anticatabolic effects. Certain studies found that icariin has better antiosteoporotic effects than other compounds (Ma et al., 2011; Wang et al., 2018). This review would emphatically introduce the potential effects of icariin to treat osteoporosis, being represented for Epimedium brevicornum Maxim.

Icariin, a prenylated flavonol glycoside, was one of the main effective compounds in *Epimedium*. With the instinct estrogen biosynthetic effect (Yang et al., 2013), it had potential osteogenic and antiosteoclastogenic effects *in vitro* and *in vivo*, and antiosteoporotic effects in clinical.

Recent *in vitro* studies have demonstrated that icariin could enhance the ALP activity, osteogenic differentiation and improve

the maturation and mineralization of MSCs and osteoblasts including hFOB 1.19 cells, MC3T3-E1, UMR 106 cells (Chen et al., 2007b; Mok et al., 2010; Fan et al., 2011; Cao et al., 2012; Liang et al., 2012). Icariin could also have a pronounced ability to promote the differentiation of osteoblast even with the absence of dexamethasone (Ma et al., 2013). Correspondingly, the mRNA expression of osteogenesis-related genes including COL1a2, OSX, RUNX-2, BMP-2, Smad4, Notch2, and OPG/RANKL ratio were significantly increased (Xiao et al., 2005; Zhao et al., 2008; Hsieh et al., 2010; Ma et al., 2011; Bian et al., 2012; Cao et al., 2012; Liang et al., 2012; Li et al., 2013b). Extra studies found that icariin treatment could significantly induce the activation of ERK, JNK, and p38 kinase, and their respective inhibitors would dramatically attenuate icariin-stimulated osteogenic effects. Ye et al. found that TAZ (the transcriptional coactivator with PDZ-binding motif) depletion could significantly block the promoting proliferation and osteogenic differentiation induced by icariin treatment (Ye et al., 2017). These studies indicated the involvement of Wnt/β-catenin-BMP2, Notch, MAPK, and RhoA-TAZ signalling pathways in the osteogenic effects by icariin (Song et al., 2013; Wu et al., 2015a; Ye et al., 2017). Additionally, the osteogenic differentiation ability of BMSCs from OVX rats would be significantly decreased compared with that in the sham operation group. While icariin treatment could act to protect and increase the osteogenic differentiation and mineralization via the estrogen pathway (Luo et al., 2015). Icariin could also protect osteoblasts cell cycle and suppress their apoptosis induced by oxidative stress. There was less production of reactive oxygen species and malondialdehyde, and more superoxide dismutase activity with the treatment of icariin (Liu et al., 2012a). Therefore, icariin could effectively preserve potential osteogenic differentiation of the cells in hypoxic condition, with the increased levels of RUNX-2, OSX, and BMP-2 gene expression, and the functions of ALP activity, and mineralized nodules (Liu et al., 2012a).

Icariin not only stimulated osteogenic differentiation but also suppressed the osteoclastogenesis and inhibited bone resorption activity in vivo. It was found that icariin could effectively control the proliferation and differentiation of hemopoietic cells which could develop into osteoclasts at the concentration of 10 mM. With the exposure of icariin, the TRAP-positive multinuclear cells appeared to be less. The formed bone resorption pits were inhibited and the osteoclastogenesis-related expressions of TRAP, RANK, and CTR genes were controlled by icariin (Chen et al., 2007a). Huang reported that icariin could suppress the bone resorption functions of osteoclasts via the affection on cytosolic free calcium, actin rings, and superoxide generation (Huang et al., 2007). The positive activities of TRAcP, and the activities of osteoclasts formation and bone resorption stimulated by LPS were diminished by icariin. Correspondingly, the synthesis of cyclo-oxygenase type-2, prostaglandin E2, hypoxia-inducible factor-1, and the activation of p38 and JNK were inhibited (Hsieh et al., 2011). Additionally, icariin could inhibit Ti particles-stimulated increase of RNA expressions of the RANKL, CTSK, TRAcP, and MMP9 in RAW264.7 cells. The expressions of IL-1 β and TNF- α were increased induced by Ti particles of RAW264.7 cells had also been inhibited (Cui et al., 2014). These experiments indicate the potential inhibitory effects of icariin on the prevention of inflammatory bone loss diseases.

In vivo studies with OVX rats, the flavonoids treatment of Epimedium Brevicornum could increase the level of serum osteocalcin and decrease the TRAcP with the comparison to untreated rats. The micro-CT result indicated that the parameters of BMD, BV/TV, Conn.D, and other similar indicators in flavonoids-treated OVX rats were obviously better. The bone histomorphometric parameters of OS/BS, MAR, and BFR/BS were improved. In the mechanical testing, the OVX would induce the reduction of the failure force. However, it was effectively inhibited by flavonoids treatment. While no increase of uterus weight was found during the treatment progress (Zhang et al., 2006; Peng et al., 2009; Liang et al., 2012). The experiments in vivo with C57BL/6 mice found that icariin could prevent decreased BMD and bone strength in femur by oestrogen deficiency after ovariectomy surgery (Mok et al., 2010). The ratio of OPG/RANKL expression in tibia has been improved (Mok et al., 2010). In the OVX rat experiment, orally treated rats with icariin at the concentration of 125 mg/kg body weight enhanced the activity of bone mineralization and formation, obtaining higher BMD, biomechanical, and histopathological parameters. And the decreased concentrations of Ca^{2+} , P, and E_2 in the serum were prevented (Nian et al., 2009). In the glucocorticoidinduced osteoporosis (GIOP) model study, icariin significantly attenuated the bone deteriorations, less BMD, hypocalcemia, and hypercalciuria of glucocorticoid positive group. The bone formation level of ALP, calcium, OCN, and fibroblast growth factor-23 in serum were increased. The bone resorption markers of carboxyterminal collagen cross-links, C-terminal telopeptide of type I collagen, and TRAP were reduced (Feng et al., 2013; Zhang et al., 2015). The antiosteoporotic effects by icariin maybe act via involvement of the ERK, PI3K/Akt/GSK3b/β-catenin integrated signalling pathways (Feng et al., 2013; Zhang et al., 2015). Liu et al. found that icariin had beneficial effects for osteoporotic rats via the inhibition of peroxisome proliferator-activated receptor y (PPARy) and Notch2 mRNA expression (Liu et al., 2017a). And Ma et al. found that icariin appears to be a therapeutic drug to manage glucocorticoid-induced bone loss via the activation of microRNA-186-mediated suppression on cathepsin K (Ma et al., 2018). Additionally, icariin could significantly reduce particleinduced bone resorption by suppressing osteoclast formation (Shao et al., 2015). Oral administration of icariin improved the abilities of bone formation with higher BMD in the regenerated bone area during the distraction osteogenesis of mandibular, indicating the icariin might be a potential medicine could shorten the course and improve the activity of distraction osteogenesis (Wei R. et al., 2011).

In clinical, a double-blind placebo-controlled clinical trial showed that the flavonoids treatment (containing the compounds of icariin, daidzein, and genistein in Epimedium) possessed the beneficial ability to inhibit the serious bone loss in postmenopausal women. The BMD could be maintained at 12 and 24 months with treatment. However, no significant changes in serum estradiol or uterus tissue were found, indicating the safety to endometrium during the application (Zhang et al., 2007a).

Therefore, being the main ingredient of *E. brevicornum*, icariin could act as a potential useful medicine to affect the imbalance of bone metabolism by increasing osteogenesis and inhibiting bone resorption. More importantly, despite the low number of clinical trials with Chinese medicine compounds, and three kinds of flavonoids in the Epimedium treated group, it has effectively indicated the antiosteoporotic effects by *Epimedium Brevicornum Maxim* clinically. Numerous studies in this review based on osteoporotic animal models, osteoblasts and osteoclasts cells have deeply and consistently confirmed the potential effects and mechanisms by which icariin regulates bone metabolism to treat osteoporosis. Furthermore, high-quality clinical research is needed to test the antiosteoporotic effects by the single compound and to compare their representive effects.

Pueraria montana (Lour.) Merr

The Chinese herb of *Pueraria montana* (Lour.) Merr. (PM, "Ge Gen") has been famously used for the daily diet and medicine in China and other Asia countries from ancient years. Being a classical and antioxidant agent, it had more recently exhibited benefits for the treatment of angina pectoris and hypertension (Yang et al., 2010b; Tan et al., 2017), neurological health (Gao et al., 2009), blood glucose homeostasis (Prasain et al., 2012), and bone metabolism (Manonai et al., 2008).

Puerarin is an active and famous isoflavone compound extracted from the classical Chinese medicine P. montana. Puerarin treatment with intragastric administration protected against the decreased levels of BMD and BMC, and the poor structure of femur trabecular bone in ovariectomized rats was improved (Wang et al., 2012a). In the in vivo study with orchidectomized (ORX) osteoporotic model, the BMD of the femur was significantly decreased. PM treatment of diet intake effectively decreased the impaired BMD, and the analysis of the femoral metaphysis indicated that PM significantly decreased the levels of BV/TV and trabecular number. And the enhancement of trabecular separation in ORX mice was restored (Wang et al., 2005; Yuan et al., 2016). In the experiment with natural menopausal monkeys, the treatment of 1000 mg/kg body weight of Puerarin powder for 16 months could significantly alleviate the loss of cortical bone. And the bone turnover levels of serum ALP and osteocalcin were decreased (Kittivanichkul et al., 2016). Puerarin 6"-O-xyloside (PXY), one of the major isoflavones of the P. montana had the beneficial effects to improve the levels of calcium, phosphorus, ALP activity, and OPG which had been decreased after OVX surgery in ICR mice serum. The destructive femur osseous tissues of enlarged bone marrow cavity and sparse trabecular bone were alleviated with PXY treatment. Correspondingly, PXY effectively improved the proliferation of osteoblasts via the improvement in the expression of OPG/ RANKL ratio (Li et al., 2016b).

In the *vitro* study, Puerarin could stimulate and improve the proliferation and differentiation of osteoblast cells (Wang et al., 2013a; Wang et al., 2014). The stimulation of osteoprotegerin and inhibition of RANKL and interleukin-6 production may act *via* the classic estrogen response element (ERE) pathway in MG-63

PG mRNA was Salvianolate could control t Ils (Yuan et al., in glucocorticoid-treated lupus-

cells (Wang et al., 2014). And the expression of OPG mRNA was increased by Puerarin in MC3T3-E1 osteoblast cells (Yuan et al., 2016). Puerarin at the dose of 2.5-100 µM would increase the growth of human BMSCs concentration-dependently (Lv et al., 2015). The osteoblastic maturation would be stimulated with the increased ALP activity, as well as the formation of mineralized nodules by Puerarin (Wang et al., 2012a; Lv et al., 2015; Zeng et al., 2018). The signalling pathways of classical ER, MAPK, and Wnt/ β -catenin were involved in the osteogenesis and bone formation effects stimulated by Puerarin treatment (Wang et al., 2012a). Ly et al. found that the osteogenesis marker expressions of Runx2, osterix, and osteocalcin were enhanced via the increased nitric oxide production and cyclic guanosine monophosphate content in hBMSCs (Lv et al., 2015). And Zeng et al. reported that the expression of transient receptor potential Melastatin 3 (TRPM3) and miR-204 were decreased and the activation of Runx2 was promoted following puerarin treatment in MC3T3-E1 osteoblastic cells (Zeng et al., 2018). Additionally, Puerarin opposed the apoptosis of human osteoblast cells induced by cisplatin or in the serum-free condition. The expression of Bcl-xL and Bcl-2 was up-regulated and Bax was decreased via the activation of MEK/ERK and PI3K/Akt signalling (Liu et al., 2013; Wang et al., 2013a).

PM could also inhibit the formation of osteoclasts *in vitro*. *Pueraria montana* extract (PME) could dose-dependently inhibit osteoclast differentiation and formation from the precursor cells. Consistently, the expression of osteoclast differentiation markers including *c-Fos* and *NFATc1* genes were downregulated (Park et al., 2017). MAPK activity induced by RANKL had also been effectively inhibited by PME treatment (Park et al., 2017). In the *vitro* experiment with RAW 264.7 cells, PM reduced the formation of TRAP-positive cells induced by the stimulation of RANKL. Correspondingly, the mRNA expression of RANKL was inhibited (Yuan et al., 2016).

These results strongly suggest that *P. montana* could act as both effective promotors of osteogenesis and inhibitor of RANKL-induced osteoclastogenesis, and it appears the isoflavon compounds of Puerarin and PXY have the great promotion on osteogenesis ability in the *in vivo* and *in vitro* studies. Even Pueraria Montana may be a potential therapeutic agent for the treatment of bone loss diseases, while the definite extracts of PM to inhibit osteoclastogenesis were still not well known and studied. Further research is necessary to characterize the bioactive compounds of CM which contains anticatabolic or anabolic benefits for the treatment of osteoporosis, and their molecular mechanisms providing the antiosteoporotic effects.

Salvia miltiorrhiza Bunge

Salvia miltiorrhiza Bunge (SMB, "Dan Shen") has been widely and classically used in clinical practice and trial for the treatment and prevention of vascular diseases in liver and heart, as well as commonly used for treating trauma wounds and fractures and correcting blood stasis in TCM for its antioxidant properties (Chen et al., 2017b; Zhang et al., 2017a; Chen et al., 2019b). The application of Salvianolate, Salvianolic acid B on the treatment of osteoporosis has been deeply studied (Guo et al., 2014).

Salvianolate could control the metabolism of bones in glucocorticoid-treated lupus-prone mice. Lupus mice usually have a marked bone loss and deterioration due to an imbalance of bone formation and resorption. Glucocorticoid treatment would deeply restrain their bone formation. After the treatment, Salvianolate increased the trabecular qualities of BV/TV, Conn.D, and Tb.Th, and decreased the SMI number in both the untreated and GC-treated lupus mice. The mechanical parameters of bone ultimate load, yield load, and stiffness in treated lupus mice were significantly improved (Liu et al., 2016). Correspondingtly, the bone resorption maker of serum TRAcP was down-regulated and OPG level was increased. The expression of RANKL, IL-6, ROS, and PPARy was inhibited, while the Runx2 expression was increased in the mice. These results indicated that Salvianolate treatment significantly affected bone metabolism to inhibit bone loss in lupus mice (Liu et al., 2016). The compound of Salvianolic acid B could prevent glucocorticoid-induced decreased BMD, bone strength, and serious architecture, and could effectively enhance the bone formation rate and the local microcirculation with more capillary dilation (Cui et al., 2012).

There are many compounds in S. miltiorrhiza having the proosteogenesis abilities including water solution, Salvianic acid A, Salvianolic acid B, Tanshinol, and Tanshinone IIA. The water solution of Salvia miltiorrhiza improved bone remodelling by enhancing the gene expression of ALP, OCN, and OPG (Chin et al., 2011). Salvianic acid A protected bone metabolism from serious impairment by the stimulation on osteogenesis and the depression of adipogenesis induced by prednisone (Cui et al., 2009). It was reported that Salvianolic acid B had the potential to stimulate the ALP activity of osteoblastic cells (Liu et al., 2007). It could also protect BMSCs differentiation and increase osteoblast activities via the increase of Runx2 mRNA expression even with the exposure of glucocorticoid. The glucocorticoid associated adipogenic differentiation was decreased by the regulation of PPARy mRNA expression (Cui et al., 2012). In the vivo study with rat tibia fracture model, Salvianolic acid B could accelerate the early-stage fracture healing for that the callus growth in the fractured bone was significantly greater in the Salvianolic acid B treated group. And the serum ALP level of the fracture rats was enhanced at weeks 1 and 3 postfracture. These findings indicate that Salvianolic acid B is a potential candidate to treat bone fracture and osteoporosis by the promoting effects on bone formation (He and Shen, 2014). In another experiment with zebrafish in vivo, dexamethasone exposure had a series of serious impairment to the bone formation, bone mass, and osteoblastspecific genes. While Tanshinol protectively promoted bone formation and bone mass via the inhibition of oxidative stress, and the osteoblast-specific genes expression of Runx2, osteocalcin, ALP, and osterix were stimulated (Luo et al., 2016). Additionally, Tanshinone IIA blocked the apoptosis of osteoblasts induced by glucocorticoids via the inhibition on the Nox4-derived overexpressed reactive oxygen species activities (Li et al., 2015a). And Tanshinone IIA enhanced the differentiation of C2C12 cells to osteoblasts via activating the signalling pathways of p38, BMP2/Smad, and Runx2 (Kim and Kim, 2010). It could also enhance the osteogenic differentiation of human periodontal

тсм	Compound	Animal model	Beneficial effects
Gynochthodes officinalis (F.C.How)	MO extract	OVX rats Disuse OP rats	BMC, BMD, serum P, Ca2+ and OPG† AKP and TRAP ↓(Li et al., 2009; Li et al., 2014b) tibia BMD, histomorphometrical parameters† osteoblasts† osteoclast ↓ (Seo et al., 2005)
Razafim. & B.Bremer (syn. <i>Morinda</i> <i>officinalis</i> F.C.How)	polysaccharides Monotropein	OVX rats	BMC, BMD, mineral element levels† (Zhu et al., 2008; Zhang et al., 2016c)
Curculigo orchioides	CO extract	OVX rats rabbits	BMC, BMD, and serum OPG↑ serum DPD/Cr, TRAcP ↓(Cao et al., 2008) bone defects↓ (Wong et al., 2007a)
Psoralea corylifolia	PC extract	OVX rats rachitic rats	BMD, ash weight and calcium content↑ (Tsai et al., 2007) serum phosphorus, bone calcification and hyperosteoidosis ↑ (Miura et al., 1996)
	Bavachin, bakuchiol Psoralen	OVX rats	BMD, trabecular parameters† (Lim et al., 2009; Weng et al., 2015) BV. Tb. Th. osteocalcin † (Yang et al., 2012)
Eucommia ulmoides	EU cortex extract	OVX rats	BMD, biomechanical parameters and microarchitecture↑ (Zhang et al., 2009; Pan et al., 2014; Zhang et al., 2014)
	т	disused OP rats	
	FU leaf extract	OVX rats	RMD biomechanical parameters serum OCt (7bang et al. 2012)
	EU seed extract	healthy rats	BMD, microarchitecturet (Li et al., 2011)
Dipsacus inermis	RD extract	healthy rats	bone density, bone histomorphologyt (Wong et al., 2007b)
	RDE RTS	OVX rats	BMD, BMC, microstructure, Young's modulus, serum OC and ALP↑ (Liu et al., 2009; Niu et al., 2012; Liu et al., 2012b; Niu et al., 2015a)
		Disuse OP rats	
Cibotium barometz	CB extract	OVX rats	BMD, bone strength, bone metaphysis↑ (Zhao et al., 2011)
Velvet Antler	blood	OVX rats	BMD, IGF-1, testosterone↑ (Yang et al., 2010a)
	VA and blood combination	OVX rats	Microarchitecture, strength, serum ALP↑ (Tseng et al., 2012)
	TVAPL	OVX rats	BWC, BMC, BMD, microarchitecture↑ (Zhang et al., 2013)
Cistanche deserticola	CD extract	OVX rats	BMD, BMC, biomechanical parameters \uparrow (Liang et al., 2011) E ₂ , Smad1, Smad5, TGF- β 1 and TIEG1 \uparrow (Liang et al., 2013)
	Echinacoside	OVX rats	BMD, microarchitecture and biomechanical parameters† (Li et al., 2013a)
Cuscuta chinensis	Kaempferol	OVX rats	BMD, Young's modulus↑(Nowak et al., 2017)
Cnidium monnieri	osthole	bone fracture	bone growth, maximum load (Zhang et al., 2017b)
		OVX rats	maximal load↑ (Li et al., 2002)
		heathy rats	peak bone mass, serum OC, micro-architecture, biomechanical parameters† (Cheng et al., 2014)
Epimedium brevicornum	Icariin flavonoids	OVX rats C57BL/6 mice and GIOP	BMD, serum ALP, OC, micro-architecture, biomechanical parameters†(Zhang et al., 2006; Peng et al., 2009; Liang et al., 2012) Serum TRACP 5b, CTX↓(Feng et al., 2013; Zhang et al., 2015)
Pueraria montana	Puerarin Puerarin 6"-O-xyloside	ORX and OVX rats, monkeys	BMD and BMC, serum ALP, OCN↑ (Wang et al., 2005; Kittivanichkul et al., 2016; Yuan et al., 2016)
Salvia miltiorrhiza	Salvianolate Salvianolic acid B	Lupus mice GIOP	bone mechanical parameters, RUNX2 expression↑ Serum TRACP, RANKL, IL-6, ROS↓(Cui et al., 2012; Liu et al., 2016)

TABLE 1 | Summary of in vivo studies for the antiosteoporotic effects of natural Chinese medicine.

They could enhance the BMC, BMD, and biomechanical parameters of the bones in osteoporosis model animals. And some of the osteogenesis makers of phosphorus, Ca²⁺, osteocalcin, osteoprotegerin, and ALP in serum would be alleviated during the dynamic metabolism progress of bone formation and resorption.

ligament stem cells *via* enhancing the activation of both ERK and Runx2 (Liu et al., 2019).

In the *in vivo* study, after SMB treatment at the concentration of 5 g/kg for 14 weeks, the unbalanced levels of serum ALP, OPG, TRACP, and RANKL of OVX rats were attenuated. The decreased BMD and bone strength was inhibited, and the impaired bone microstructures were improved. Moreover, the decreased expression of p-LRP6, IGF-1, ALP, and OPG were enhanced. While the increased expression of RANKL and CTSK in the tibias and femurs of OVX rats were effectively inhibited by SMB treatment (Liu et al., 2018). Tanshinone VI, extracted from the root of *S. miltiorrhiza*, which could greatly inhibit osteoclast differentiation and bone resorption by disrupting the formation of actin ring. Tanshinone VI appears to prevent osteoclast differentiation by the downregulation of RANKL expression (Nicolin et al., 2010). Kwak et al. reported that Tanshinone IIA inhibited the osteoclast differentiation from the precursors *via* the down-regulation of RANKL-induced high levels of c-Fos and NFATc1 (Kwak et al., 2006). Additionally, in the natural drug screening experiment, maybe tanshinone 1, cryptotanshinone, and 15,16-dihydrotanshinone I diterpenoids and other unknown compounds had a synergistic effect with tanshinone, possessing the antiosteoclastogenesis effects by reducing the formation and function of TRAP-positive multinuclear osteoclasts (Lee et al., 2005; Kim et al., 2008).

These studies highlight the antiosteoporotic effects of *S. miltiorrhiza in vivo* and *in vitro*. Most of the compounds of *S. miltiorrhiza* including Salvianolate, Salvianic acid A, Salvianolic acid B, Tanshinol, and Tanshinone IIA, and so

тсм	Compound	Cells	Beneficial effects
Gynochthodes	Bajijiasu	OCs	osteoclast formation↓ (Hong et al., 2017a)
officinalis (F.C.How)	anthraquinone	OBs	proliferation↑ (Wu et al., 2009)
Razafim. & B.Bremer		OCs	differentiation, TRAcP activity↓, apoptosis↑ (Wu et al., 2009; Bao et al., 2011)
(syn. <i>Morinda</i> officinalis F.C.How)	Monotropein	OBs	Proliferation, mineralization† (Zhang et al., 2016c)
Curculigo orchioides	Curculigoside (CCG)	hAFSCs	osteogenesis, ALP activity, calcium deposition↑ osteoclastogenesis↓ (Liu et al., 2014a)
		BMSCs	Proliferation, differentiation↑ (Shen et al., 2013)
		OBs	oxidative damage↓, proliferation, differentiation↑ (Wang et al., 2012b)
	M2, CCG-A, CCG-B etc.	OBs	proliferation, differentiation (Wang et al., 2017a)
		OCs	TRAcP activity↓ (Jiao et al., 2009; Wang et al., 2013b; Wang et al., 2017c)
Psoralea corylifolia	bavachin and bakuchiol	OBs	Proliferation, differentiation, ALP activity† (Lim et al., 2009)
	PSO	MSCs	differentiation, ALP activity† (Yang et al., 2012)
	Neobavaisoflavone	OBs	differentiation, ALP activity (Don et al., 2012)
	bavachalcone	OCs	osteoclastogenesis, resorption pits↓ (Chai et al., 2018)
Eucommia ulmoides	5-HMF	MSCs	osteogenesis Mineralization† (Tan et al., 2014)
			adipogenesis↓ (Tan et al., 2014)
	AU, GP, GA, TL	OBs	proliferation↑ (Ha et al., 2003; Zhang et al., 2014)
		OCs	proliferation, differentiation.)(Ha et al., 2003; Zhang et al., 2014)
Dipsacus inermis	RDE, RTS	OBs, MSCs	proliferation, differentiation (Liu et al., 2012b; Niu et al., 2015b)
		OCs	osteoclastogenesis 1 (Niu et al., 2012)
	Asperosaponin VI	OBs	proliferation, differentiation, mineralization (Niu et al., 2011)
	RD extract	MSCs	differentiation↑ (Kim et al., 2011)
Cibotium barometz	RW-Cb	OBs	proliferation, differentiation (Xu et al., 2014)
	CB extract	OCs	Differentiation, TRAcP activity (Nguyen et al., 2009)
Velvet Antler	TVAPL	OBs	Proliferation, ↑ (Zhang et al., 2013)
	FA-, NFA-		Proliferation, mineralization (Lee et al., 2011)
	FA	OCs	Differentiation, TRAcP activity (Choi et al., 2013)
	CE-C		Differentiation, actin ring↓, apoptosis↑ (Li et al., 2007)
Cistanche deserticola	CD extract	OBs	mineralization† (Li et al., 2012b)
	Echinacoside	OBs	proliferation, mineralization, ALP activity† (Li et al., 2012a)
	CD extract	OCs	Differentiation, actin ring↓, ROS↓ (Song et al., 2018)
Cuscuta chinensis	CC extract	OBs	Proliferation, differentiation mineralization (Yao et al., 2005; Yang et al., 2009)
			ALP activity, collagen synthesis, BMP-2↑ (Yang et al., 2009; Yang et al., 2011)
			oxidative damage↑, apoptosis↓ (Gao et al., 2013)
	Kaempferol, hyperoside	OBs	ALP activity† (Yang et al., 2011)
	Campesterol	OCs	osteoclasts activities↓ (Yao et al., 2005)
Cnidium monnieri	Osthole	OBs	Proliferation, ALP, mineralization†(Jia et al., 2016; Zhang et al., 2017b)
	Bergapten	OCs	Osteoclastogenesis and bone resorption 1 (Chen et al., 2019a)
Epimedium	lcariin, etc.	OBs and	Proliferation, ALP, mineralization, COL1a2, OSX, RUNX-2, BMP-2/Smad4 Notch21(Chen et al.,
brevicornum		MSCs	2007b; Mok et al., 2010; Fan et al., 2011; Cao et al., 2012; Liang et al., 2012)
		OCs	Osteoclastogenesis, bone resorption, RANKL, CTSK, TRAcP and MMP91(Chen et al., 2007a)
Pueraria montana	Puerarin	OBs, MSCs	proliferation and differentiation, ALP activity↑ (Wang et al., 2013a; Wang et al., 2014)
		OCs	osteoclast differentiation and formation (Park et al., 2017)
Salvia miltiorrhiza	Salvianic acid A	OBs,	ALP, OCN, OPG, Runx2↑(Chin et al., 2011; Cui et al., 2009)
	Salvianolic acid		
	B Tanshinol and		
	Tanshinone IIA		
		OCs	Osteoclast formation and function, c-Fos and NFATc11(Nicolin et al., 2010; Liu et al., 2018)

They have beneficial osteogenetic effects by enhancing the proliferation and differentiation of osteoblasts and bone mesenchymal stem cells, improving the activity of ALP and the formation of mineralized nodules. While the osteoclastogenesis and function of osteoclasts are inhibited.

on, have potential antiosteoporosis effects by promoting bone formation *via* increased expression of osteogenesisrelated genes and proteins, and by decreasing bone resorptive osteoclastogenesis through the inhibition of reactive oxygen species activity. Compounds in the research of Kim et al. also have the antiosteoclastogenic effects which are not further studied. More research is needed to provide the evidence of the herb and its potent compounds to target osteoporosis in clinical trials, including their mode of application and mechanisms of action.

DISCUSSION

In summary, with the increasingly ageing population worldwide, the osteoporotic fracture has become a major health and social issue. The side effects caused by hormone therapy and alendronate antiosteoporotic agents have prompted researchers to study natural therapeutic compounds, which may be effective and safe for the treatment of osteoporosis, and with less adverse side-effects.

The pathophysiology of osteoporosis is complicated in terms of occurrence, development, and progression,



including much more numerous mechanisms of mechanistic/ mammalian target of rapamycin (mTOR), autophagy, and notch involved (Shen et al., 2016; Zanotti et al., 2018; Hiraiwa et al., 2019), except for RANKL, MAPK, Wnt, and Smad signalling pathways discussed above. Natural Chinese medicine may contain compounds that are effective for the treatment of osteoporosis and this review documents current evidence as to their potential bio-pharmacological effects and possible mechanisms of actions. A summary of the in vivo and in vitro antiosteoporosis effects of the natural herbs reviewed by this article is presented in Table 1 and Table 2, respectively. Natural Chinese medicine appears to promote bone formation activity, including the osteogenesis of MSCs and osteoblasts. Some medicines could protect them from oxidative damage due to ROS activity. Additionally, the bone resorption activity of osteoclasts may be significantly inhibited by certain herbal compounds, thus potentially alleviating the imbalance between bone formation by osteoblasts and bone resorption by osteoclasts. Figure 3 summarizes the signalling pathways that appear to mediate the antiosteoporotic effects of the natural medicine reviewed by this article.

The natural Chinese medicines in this review are classic and bone-specific medicines. As we know, clinical experiences are very important to Chinese medicine. Chinese medicines were classified into different categories with special functions according to the rich practices and experiences in the clinic and the Chinese medicine theories. Some of them were the classic and bone-specific drugs to treat skeleton fractures and bone loss diseases for their beneficial improvement on bone formation. Most of them have the effects and functions to tonify the "Yang" in traditional Chinese medicine, which has an improvement on bone development and metabolism. "Yang-tonifying" medicines are the most popular and classic kind of natural drugs to treat osteoporosis in Chinese medicine (Ju et al., 2014; Li et al., 2015b). Furthermore, all of them are deeply studied possessing both anabolic and anticatabolic effects. They have potential bone-formation effects by enhancing the proliferation and differentiation of osteoblasts and BMSCs, improving the activity of ALP and mineralization formation. Some of them could protect osteoblasts and BMSCs from apoptosis induced by oxidative stress (Liu et al., 2013). While the osteoclastogenesis and bone-resorption function of osteoclasts were inhibited by the treatment of these medicines (Wang et al., 2014). Interestingly, they have the phytoestrogenic or phytoandrogenic effects which might act as the natural and potential alternatives for hormone replacement treatment or alendronate therapies to significantly inhibit the bone loss and improve skeleton development of osteporotic patients. It has been reported that testosterone played a vital basic and clinical role in the homeostasis of skeletal tissue (Ebeling, 2010). In vivo study indicated that the androgen

deficiency would significantly lead to an increase of osteopenia in the aged male rats (Erben et al., 2000). Clinically, testicular malfunction induced by androgen deficiency may cause the osteoporosis in old men with increasing bone resorption (Foresta et al., 1984). Numerous studies indicated that these bone-specific drugs contain phytoandrogens (Edouard et al., 2014), which could act as a natural and potential alternative for testosterone replacement therapy (TRT). They could effectively restore the level of serum testosterone and thus significantly improve the bone health and physical condition of patients (George and Henkel, 2014). Some studies found that the compounds from these classical drugs may also possess phytoestrogenic effects (Jiao et al., 2009; Ma et al., 2011; Zhang et al., 2016b), having a similar structure of estrogen conformation and capabilities to bind with estrogen receptors. Therefore, they may regulate bone remodeling via estrogen receptor pathway (Wiseman, 2000). More importantly, the application of these drugs exhibiting phytoandrogen and phytoestrogen effects do not appear to cause obvious or harmful side-effects including cardiovascular disease, prostate cancer, and breast cancer, which might be induced by the long-term and large dosage use of testosterone or estrogen replacement therapy (Wiseman, 2000).

However, the development of osteoporosis is very complex in postmenopausal women, elderly men, glucocorticoidoveruse patients, and other patients with metabolic diseases. The mechanisms of action of natural Chinese medicines effective for the treatment of osteoporosis have not yet been well investigated, thus indicating the need for further studies (Ju et al., 2014). Besides, large dosages or long-term usage warrants caution and certain methodologies should be observed. Further research to isolate and characterize the bioactive antiosteoporotic compounds from the classical and bone-specific drugs is necessary to extensively profile compounds for pharmacological usage, especially their safety, efficacy, and potential chemical interactions with other drugs. Studies to determine the special and targeted cellular and molecular mechanisms of natural Chinese medicine compounds are required to develop their potential application for the treatment of osteoporosis, as an effective, safe alternative to primary therapeutic strategies, or in combination with current primary pharmacological treatments. Additionally, few high-quality clinical studies have documented the antiosteoporosis effects of structure well-known

REFERENCES

- Albright, F. (1947). Osteoporosis. Ann. Intern. Med. 27, 861–882. doi: 10.7326/0003-4819-27-6-861
- Bao, L. L., Qin, L. P., Liu, L., Wu, Y. B., Han, T., Xue, L. M., et al. (2011). Anthraquinone compounds from Morinda officinalis inhibit osteoclastic bone resorption *in vitro*. *Chem-Biol. Interact.* 194, 97–105. doi: 10.1016/j.cbi.2011.08.013
- Bian, Q., Huang, J. H., Liu, S. F., Ning, Y., Yang, Z., Zhao, Y. J., et al. (2012). Different molecular targets of Icariin on bMSCs in CORT and OVX -rats. *Front. Biosci.* (*Elite Ed.*) 4, 1224–1236. doi: 10.2741/e453
- Cao, D. P., Zheng, Y. N., Qin, L. P., Han, T., Zhang, H., Rahman, K., et al. (2008). Curculigo orchioides, a traditional Chinese medicinal plant, prevents bone loss in ovariectomized rats. *Maturitas* 59, 373–380. doi: 10.1016/j.maturitas.2008.03.010
- Cao, H., Ke, Y., Zhang, Y., Zhang, C. J., Qian, W., and Zhang, G. L. (2012). Icariin stimulates MC3T3-E1 cell proliferation and differentiation through

compounds, for example, Epimedium-derived phytoestrogen flavonoids were used to treat and inhibit osteoporosis and bone loss of the postmenopausal women in a clinical trial (Zhang et al., 2007a). There are still some limitations and deficiencies of these clinical drug findings, which are studied together with combined medicines in traditional formulas, due to the potential for unknown interactions between the various drugs and nonspecific compounds in this medicine (Wei H. et al., 2011; Wei R. et al., 2011; Shi et al., 2012). Therefore, more high-quality clinical studies with natural Chinese medicines possessing the anabolic and anticatabolic effects are needed in the future.

CONCLUSION

Recent *in vivo* and *in vitro* findings suggest that natural Chinese medicine may provide potential therapeutic benefits for the treatment of osteoporosis. Further research is necessary to ensure the safety, efficacy, and specificity of the compounds in Chinese medicines to develop their therapeutic potential. More high-quality clinical researches with these natural medicines are needed to provide greater evidence for the candidate to beneficial and safer antiosteoporotic application.

AUTHOR CONTRIBUTIONS

JH, XL, and ZW contributed equally to this work. JH and XL conceived the idea and wrote the manuscript. ZW, SB, and KC helped modify the language and the revision. ZX collected the literature. JX, DL, and SW helped supervise the research and contribute to the final draft of the paper. We thanked JZ, SC, YH, and JC for the help of this review. All authors reviewed and approved the final manuscript.

FUNDING

This work was supported by the National Natural Science Foundation of China (No. 81673992), National Natural Science Foundation of China Youth Fund (No. 81904091) and Fundamental Research Funds for the Central Universities (No. 21619307).

up-regulation of bone morphogenetic protein-2. *Int. J. Mol. Med.* 29, 435–439. doi: 10.3892/ijmm.2011.845

- Chai, L., Zhou, K., Wang, S., Zhang, H., Fan, N., Li, J., et al. (2018). Psoralen and Bakuchiol ameliorate M-CSF Plus RANKL-induced osteoclast differentiation and bone resorption via inhibition of AKT and AP-1 pathways in vitro. *Cell Physiol. Biochem.* 48, 2123–2133. doi: 10.1159/000492554
- Chen, D. L., Li, N., Lin, L., Long, H. M., Lin, H., Chen, J., et al. (2014). Confocal mirco-Raman spectroscopic analysis of the antioxidant protection mechanism of the oligosaccharides extracted from Morinda officinalis on human sperm DNA. J. Ethnopharmacol. 153, 119–124. doi: 10.1016/j. jep.2014.01.021
- Chen, D. L., Yang, X., Yang, J., Lai, G. X., Yong, T. Q., Tang, X. C., et al. (2017a). Prebiotic effect of fructooligosaccharides from Morinda officinalis on Alzheimer's disease in rodent models by targeting the microbiota-gut-brain axis. *Front. Aging Neurosci.* 9, 1–28. doi: 10.3389/fnagi.2017.00403

- Chen, F., Li, L., and Tian, D. D. (2017b). Salvia miltiorrhiza roots against cardiovascular disease: consideration of herb-drug interactions. *BioMed. Res. Int.* 2017, 9868694. doi: 10.1155/2017/9868694
- Chen, G., Xu, Q., Dai, M., and Liu, X. (2019a). Bergapten suppresses RANKLinduced osteoclastogenesis and ovariectomy-induced osteoporosis via suppression of NF-kappaB and JNK signaling pathways. *Biochem. Biophys. Res. Commun.* 509, 329–334. doi: 10.1016/j.bbrc.2018.12.112
- Chen, K. M., Ge, B. F., Liu, X. Y., Ma, P. H., Lu, M. B., Bai, M. H., et al. (2007a). Icariin inhibits the osteoclast formation induced by RANKL and macrophagecolony stimulating factor in mouse bone marrow culture. *Pharmazie* 62, 388– 391. doi: 10.1691/ph.2007.5.6682
- Chen, K. M., Ma, H. P., Ge, B. F., Liu, X. Y., Ma, L. P., Bai, M. H., et al. (2007b). Icariin enhances the osteogenic differentiation of bone marrow stromal cells but has no effects on the differentiation of newborn calvarial osteoblasts of rats. *Pharmazie* 62, 785–789. doi: 10.1691/ph.2007.10.7504
- Chen, Z., Gao, X., Jiao, Y., Qiu, Y., Wang, A., Yu, M., et al. (2019b). Tanshinone IIA exerts anti-inflammatory and immune-regulating effects on vulnerable atherosclerotic plaque partially via the TLR4/MyD88/NF-kappaB signal pathway. Front. Pharmacol. 10, 850. doi: 10.3389/fphar.2019.00850
- Cheng, K., Ge, B. F., Zhen, P., Chen, K. M., Ma, X. N., Zhou, J., et al. (2014). [Comparative study on effect of osthole and genistein on peak bone mass in rats]. *Zhongguo Gu Shang* 27, 587–591. doi: 10.3969/j.issn.1003-0034.2014.07.013
- Chin, A., Yang, Y., Chai, L., Wong, R. W., and Rabie, A. B. (2011). Effects of medicinal herb salvia militorrhiza on osteoblastic cells *in vitro*. J. Orthop. Res. 29, 1059–1063. doi: 10.1002/jor.21376
- Choi, S. W., Moon, S. H., Yang, H. J., Kwon, D. Y., Son, Y. J., Yu, R., et al. (2013). Antiresorptive Activity of Bacillus-Fermented Antler Extracts: Inhibition of Osteoclast Differentiation. *Evid-Based Complement. Alternat.* 2013, 1–9 doi: 10.1155/2013/748687
- Cui, J., Zhu, M., Zhu, S., Wang, G., Xu, Y., and Geng, D. (2014). Inhibitory effect of icariin on Ti-induced inflammatory osteoclastogenesis. J. Surg. Res. 192, 447–453. doi: 10.1016/j.jss.2014.05.038
- Cui, L., Li, T., Liu, Y., Zhou, L., Li, P., Xu, B., et al. (2012). Salvianolic acid B prevents bone loss in prednisone-treated rats through stimulation of osteogenesis and bone marrow angiogenesis. *PloS One* 7, e34647. doi: 10.1371/journal. pone.0034647
- Cui, L., Liu, Y. Y., Wu, T., Ai, C. M., and Chen, H. Q. (2009). Osteogenic effects of D+beta-3,4-dihydroxyphenyl lactic acid (salvianic acid A, SAA) on osteoblasts and bone marrow stromal cells of intact and prednisone-treated rats. *Acta Pharmacol. Sin.* 30, 321–332. doi: 10.1038/aps.2009.9
- Curtis, E. M., van der Velde, R., Moon, R. J., van den Bergh, J. P., Geusens, P., de Vries, F., et al. (2016). Epidemiology of fractures in the United Kingdom 1988-2012: variation with age, sex, geography, ethnicity and socioeconomic status. *Bone* 87, 19–26. doi: 10.1016/j.bone.2016.03.006
- Dinger, J., Bardenheuer, K., and Heinemann, K. (2016a). Drospirenone plus estradiol and the risk of serious cardiovascular events in postmenopausal women. *Climacteric* 19, 349–356. doi: 10.1080/13697137.2016.1183624
- Dinger, J., Minh, T. D., and Heinemann, K. (2016b). Impact of estrogen type on cardiovascular safety of combined oral contraceptives. *Contraception* 94, 328– 339. doi: 10.1016/j.contraception.2016.06.010
- Do, M. H., Hur, J., Choi, J., Kim, M., Kim, M. J., Kim, Y., et al. (2018). Eucommia ulmoides ameliorates glucotoxicity by suppressing advanced glycation endproducts in diabetic mice kidney. *Nutrients* 10, 1–13. doi: 10.3390/nu10030265
- Don, M. J., Lin, L. C., and Chiou, W. F. (2012). Neobavaisoflavone stimulates osteogenesis via p38-mediated up-regulation of transcription factors and osteoid genes expression in MC3T3-E1 cells. *Phytomedicine* 19, 551–561. doi: 10.1016/j.phymed.2012.01.006
- Donnapee, S., Li, J., Yang, X., Ge, A. H., Donkor, P. O., Gao, X. M., et al. (2014). Cuscuta chinensis Lam.: a systematic review on ethnopharmacology, phytochemistry and pharmacology of an important traditional herbal medicine. J. Ethnopharmacol. 157, 292–308. doi: 10.1016/j.jep.2014.09.032
- Ebeling, P. R. (2010). Androgens and osteoporosis. Curr. Opin. Endocrinol. Diabetes Obes. 17, 284–292. doi: 10.1097/MED.0b013e328339658c
- Edouard, M. J., Miao, L., Fan, G. W., Ojong, B. B., Zhen, H., Zhang, J., et al. (2014). Yang-tonifying traditional Chinese medicinal plants and their potential phytoandrogenic activity. *Chin. J. Nat. Med.* 12, 321–334. doi: 10.1016/ S1875-5364(14)60040-6

- Erben, R. G., Eberle, J., Stahr, K., and Goldberg, M. (2000). Androgen deficiency induces high turnover osteopenia in aged male rats: a sequential histomorphometric study. *J. Bone Miner. Res.* 15, 1085–1098. doi: 10.1359/ jbmr.2000.15.6.1085
- Fan, J. J., Cao, L. G., Wu, T., Wang, D. X., Jin, D., Jiang, S., et al. (2011). The doseeffect of icariin on the proliferation and osteogenic differentiation of human bone mesenchymal stem cells. *Molecules* 16, 10123–10133. doi: 10.3390/ molecules161210123
- Fan, X. L., Chen, Y., Li, L., Wang, Y., Zhang, Y., Lu, S., et al. (2019). Efficacy of Chinese herb Cistanche Yishen granules in treatment of tinnitus for patients with chronic nephritis. J. Cell Biochem. 120, 5505–5509. doi: 10.1002/jcb.27833
- Feng, R., Feng, L., Yuan, Z., Wang, D., Wang, F., Tan, B., et al. (2013). Icariin protects against glucocorticoid-induced osteoporosis *in vitro* and prevents glucocorticoid-induced osteocyte apoptosis *in vivo*. *Cell Biochem. Biophys.* 67, 189–197. doi: 10.1007/s12013-013-9533-8
- Foresta, C., Ruzza, G., Mioni, R., Guarneri, G., Gribaldo, R., Meneghello, A., et al. (1984). Osteoporosis and decline of gonadal function in the elderly male. *Horm. Res.* 19, 18–22. doi: 10.1159/000179855
- Fu, Z. F., Fan, X., Wang, X. Y., and Gao, X. M. (2018). Cistanches Herba: an overview of its chemistry, pharmacology, and pharmacokinetics property. *J. Ethnopharmacol.* 219, 233–247. doi: 10.1016/j.jep.2017.10.015
- Gao, J. M., Li, R., Zhang, L., Jia, L. L., Ying, X. X., Dou, D. Q., et al. (2013). Cuscuta chinensis seeds water extraction protecting murine osteoblastic MC3T3-E1 cells against tertiary butyl hydroperoxide induced injury. *J. Ethnopharmacol.* 148, 587–595. doi: 10.1016/j.jep.2013.05.005
- Gao, L., Ji, X., Song, J., Liu, P., Yan, F., Gong, W., et al. (2009). Puerarin protects against ischemic brain injury in a rat model of transient focal ischemia. *Neurol. Res.* 31, 402–406. doi: 10.1179/174313209X444017
- George, A., and Henkel, R. (2014). Phytoandrogenic properties of Eurycoma longifolia as natural alternative to testosterone replacement therapy. *Andrologia* 46, 708–721. doi: 10.1111/and.12214
- Gilbey, A., and Perezgonzalez, J. D. (2012). Health benefits of deer and elk velvet antler supplements: a systematic review of randomised controlled studies. N. Z. Med. J. 125, 80–86.
- Guo, Y., Li, Y., Xue, L., Severino, R. P., Gao, S., Niu, J., et al. (2014). Salvia miltiorrhiza: an ancient Chinese herbal medicine as a source for anti-osteoporotic drugs. *J. Ethnopharmacol.* 155, 1401–1416. doi: 10.1016/j.jep.2014.07.058
- Ha, H., Ho, J. Y., Shin, S., Kim, H., Koo, S., Kim, I. H., et al. (2003). Effects of eucommiae cortex on osteoblast-like cell proliferation and osteoclast inhibition. Arch. Pharm. Res. 26, 929–936. doi: 10.1007/BF02980202
- He, J. B., Chen, M. H., and Lin, D. K. (2017). New insights into the tonifying kidney-yin herbs and formulas for the treatment of osteoporosis. Arch. Osteoporos. 12, 1–14. doi: 10.1007/s11657-016-0301-4
- He, X., and Shen, Q. (2014). Salvianolic acid B promotes bone formation by increasing activity of alkaline phosphatase in a rat tibia fracture model: a pilot study. BMC Complement. Altern. Med. 14, 493. doi: 10.1186/1472-6882-14-493
- He, Y. Q., Zhang, Q., Shen, Y., Han, T., Zhang, Q. L., Zhang, J. H., et al. (2018). Rubiadin-1-methyl ether from Morinda officinalis How. Inhibits osteoclastogenesis through blocking RANKL-induced NF-kappa B pathway. *Biochem. Biophys. Res. Commun.* 506, 927–931. doi: 10.1016/j.bbrc.2018.10.100
- Hejazi, I. I., Khanam, R., Mehdi, S. H., Bhat, A. R., Rizvi, M. M. A., Thakur, S. C., et al. (2018). Antioxidative and anti-proliferative potential of Curculigo orchioides Gaertn in oxidative stress induced cytotoxicity: in vitro, ex vivo and in silico studies. *Food Chem. Toxicol.* 115, 244–259. doi: 10.1016/j.fct.2018.03.013
- Hiraiwa, M., Ozaki, K., Yamada, T., Iezaki, T., Park, G., Fukasawa, K., et al. (2019). mTORC1 activation in osteoclasts prevents bone loss in a mouse model of osteoporosis. *Front. Pharmacol.* 10, 684. doi: 10.3389/fphar.2019.00684
- Hong, G. J., Zhou, L., Shi, X. G., He, W., Wang, H. B., Wei, Q. S., et al. (2017a). Bajijiasu Abrogates osteoclast differentiation *via the* suppression of RANKL signaling pathways through NF-kappa B and NFAT. *Int. J. Mol. Sci.* 18, 1–12. doi: 10.3390/ijms18010203
- Hong, H., An, J. C., De La Cruz, J. F., and Hwang, S. G. (2017b). Cnidium officinale Makino extract induces apoptosis through activation of caspase-3 and p53 in human liver cancer HepG2 cells. *Exp. Ther. Med.* 14, 3191–3197. doi: 10.3892/ etm.2017.4916
- Hsieh, T. P., Sheu, S. Y., Sun, J. S., and Chen, M. H. (2011). Icariin inhibits osteoclast differentiation and bone resorption by suppression of MAPKs/NF-kappaB

regulated HIF-1alpha and PGE(2) synthesis. *Phytomedicine* 18, 176–185. doi: 10.1016/j.phymed.2010.04.003

- Hsieh, T. P., Sheu, S. Y., Sun, J. S., Chen, M. H., and Liu, M. H. (2010). Icariin isolated from Epimedium pubescens regulates osteoblasts anabolism through BMP-2, SMAD4, and Cbfa1 expression. *Phytomedicine* 17, 414–423. doi: 10.1016/j.phymed.2009.08.007
- Huang, J., Yuan, L., Wang, X., Zhang, T. L., and Wang, K. (2007). Icaritin and its glycosides enhance osteoblastic, but suppress osteoclastic, differentiation and activity *in vitro*. *Life Sci.* 81, 832–840. doi: 10.1016/j.lfs.2007.07.015
- Hussain, T., Tan, B., Liu, G., Oladele, O. A., Rahu, N., Tossou, M. C., et al. (2016). Health-promoting properties of Eucommia ulmoides: a review. *Evid-Based Complement. Alternat.* 2016, 1–9. doi: 10.1155/2016/5202908
- Jia, M., Li, Y., Xin, H. L., Hou, T. T., Zhang, N. D., Xu, H. T., et al. (2016). Estrogenic activity of osthole and imperatorin in MCF-7 cells and their osteoblastic effects in Saos-2 cells. *Chin. J. Nat. Medicines* 14, 413–420. doi: 10.1016/ S1875-5364(16)30037-1
- Jiang, K. M., Huang, D., Zhang, D. W., Wang, X. L., Cao, H. J., Zhang, Q., et al. (2018). Investigation of inulins from the roots of Morinda officinalis for potential therapeutic application as anti-osteoporosis agent. *Int. J. Biol. Macromol.* 120, 170–179. doi: 10.1016/j.ijbiomac.2018.08.082
- Jiang, Z. H., Wang, J., Li, X. P., and Zhang, X. Y. (2016). Echinacoside and Cistanche tubulosa (Schenk) R. wight ameliorate bisphenol A-induced testicular and sperm damage in rats through gonad axis regulated steroidogenic enzymes. *J. Ethnopharmacol.* 193, 321–328. doi: 10.1016/j.jep.2016.07.033
- Jiao, L., Cao, D. P., Qin, L. P., Han, T., Zhang, Q. Y., Zhu, Z., et al. (2009). Antiosteoporotic activity of phenolic compounds from Curculigo orchioides. *Phytomedicine* 16, 874–881. doi: 10.1016/j.phymed.2009.01.005
- Jing, H. R., Wang, S. X., Wang, M., Fu, W. L., Zhang, C., and Xu, D. G. (2017). Isobavachalcone attenuates MPTP-induced Parkinson's disease in mice by inhibition of microglial activation through NF-kappa B pathway. *PloS One* 12, 1–15. doi: 10.1371/journal.pone.0169560
- Ju, D., Liu, M., Zhao, H., and Wang, J. (2014). Mechanisms of "kidney governing bones" theory in traditional Chinese medicine. *Front. Med.* 8, 389–393. doi: 10.1007/s11684-014-0362-y
- Kim, B. S., Kim, Y. C., Zadeh, H., Park, Y. J., Pi, S. H., Shin, H. S., et al. (2011). Effects of the dichloromethane fraction of Dipsaci Radix on the osteoblastic differentiation of human alveolar bone marrow-derived mesenchymal stem cells. *Biosci. Biotech. Biochem.* 75, 13–19. doi: 10.1271/bbb.100379
- Kim, H. J., and Kim, S. H. (2010). Tanshinone IIA enhances BMP-2-stimulated commitment of C2C12 cells into osteoblasts *via* p38 activation. *Amino Acids* 39, 1217–1226. doi: 10.1007/s00726-010-0557-8
- Kim, H. K., Kim, M. G., and Leem, K. H. (2016). Comparison of the effect of velvet antler from different sections on longitudinal bone growth of adolescent rats. *Evid-Based Complement. Alternat.* 2016, 1–9. doi: 10.1155/2016/1927534
- Kim, H. K., Woo, E. R., Lee, H. W., Park, H. R., Kim, H. N., Jung, Y. K., et al. (2008). The correlation of Salvia miltiorrhiza extract-induced regulation of osteoclastogenesis with the amount of components tanshinone I, tanshinone IIA, cryptotanshinone, and dihydrotanshinone. *Immunopharmacol. Immunotoxicol.* 30, 347–364. doi: 10.1080/08923970801949133
- Kim, J. Y., Lee, J. I., Song, M., Lee, D., Song, J., Kim, S. Y., et al. (2015). Effects of Eucommia ulmoides extract on longitudinal bone growth rate in adolescent female rats. *Phytother. Res.* 29, 148–153. doi: 10.1002/ptr.5195
- Kittivanichkul, D., Charoenphandhu, N., Khemawoot, P., and Malaivijitnond, S. (2016). Pueraria mirifica alleviates cortical bone loss in naturally menopausal monkeys. J. Endocrinol. 231, 121–133. doi: 10.1530/JOE-16-0277
- Kwak, H. B., Yang, D., Ha, H., Lee, J. H., Kim, H. N., Woo, E. R., et al. (2006). Tanshinone IIA inhibits osteoclast differentiation through down-regulation of c-Fos and NFATc1. *Exp. Mol. Med.* 38, 256–264. doi: 10.1038/emm.2006.31
- Lee, H. S., Kim, M. K., Kim, Y. K., Jung, E. Y., Park, C. S., Woo, M. J., et al. (2011). Stimulation of osteoblastic differentiation and mineralization in MC3T3-E1 cells by antler and fermented antler using Cordyceps militaris. *J. Ethnopharmacol.* 133, 710–717. doi: 10.1016/j.jep.2010.10.047
- Lee, S. Y., Choi, D. Y., and Woo, E. R. (2005). Inhibition of osteoclast differentiation by tanshinones from the root of Salvia miltiorrhiza bunge. *Arch. Pharm. Res.* 28, 909–913. doi: 10.1007/BF02973876
- Li, C., Dong, J., Tian, J., Deng, Z., and Song, X. (2016a). LC/MS/MS determination and pharmacokinetic study of iridoid glycosides monotropein and deacetylasperulosidic acid isomers in rat plasma after oral administration of

Morinda officinalis extract. *BioMed. Chromatogr.* 30, 163–168. doi: 10.1002/bmc.3532

- Li, F., Yang, X., Yang, Y., Guo, C., Zhang, C., Yang, Z., et al. (2013a). Antiosteoporotic activity of echinacoside in ovariectomized rats. *Phytomedicine* 20, 549–557. doi: 10.1016/j.phymed.2013.01.001
- Li, F., Yang, Y. N., Zhu, P. P., Chen, W. N., Qi, D. L., Shi, X. P., et al. (2012a). Echinacoside promotes bone regeneration by increasing OPG/RANKL ratio in MC3T3-E1 cells. *Fitoterapia* 83, 1443–1450. doi: 10.1016/j.fitote.2012.08.008
- Li, H., Chen, B., Pang, G., Chen, J., Xie, J., and Huang, H. (2016b). Anti-osteoporotic activity of puerarin 6"-O-xyloside on ovariectomized mice and its potential mechanism. *Pharm. Biol.* 54, 111–117. doi: 10.3109/13880209.2015.1017885
- Li, J., He, C., Tong, W., Zou, Y., Li, D., Zhang, C., et al. (2015a). Tanshinone IIA blocks dexamethasone-induced apoptosis in osteoblasts through inhibiting Nox4-derived ROS production. *Int. J. Clin. Exp. Pathol.* 8, 13695–13706.
- Li, N., Qin, L. P., Han, T., Wu, Y. B., Zhang, Q. Y., and Zhang, H. (2009). Inhibitory effects of Morinda officinalis extract on bone loss in ovariectomized rats. *Molecules* 14, 2049–2061. doi: 10.3390/molecules14062049
- Li, T. M., Huang, H. C., Su, C. M., Ho, T. Y., Wu, C. M., Chen, W. C., et al. (2012b). Cistanche deserticola extract increases bone formation in osteoblasts. J. Pharm. Pharmacol. 64, 897–907. doi: 10.1111/j.2042-7158.2012.01483.x
- Li, W. D., Yan, C. P., Wu, Y., Weng, Z. B., Yin, F. Z., Yang, G. M., et al. (2014a). Osteoblasts proliferation and differentiation stimulating activities of the main components of Fructus Psoraleae corylifoliae. *Phytomedicine* 21, 400–405. doi: 10.1016/j.phymed.2013.09.015
- Li, X. F., Xu, H., Zhao, Y. J., Tang, D. Z., Xu, G. H., Holz, J., et al. (2013b). Icariin augments bone formation and reverses the phenotypes of osteoprotegerindeficient mice through the activation of Wnt/beta - Catenin-BMP signaling. *Evid Based Complement. Alternat. Med.* 2013, 652317. doi: 10.1155/2013/652317
- Li, X. X., Hara, I., and Matsumiya, T. (2002). Effects of osthole on postmenopausal osteoporosis using ovariectomized rats; Comparison to the effects of estradiol. *Biol. Pharm. Bull.* 25, 738–742. doi: 10.1248/bpb.25.738
- Li, Y., Liang, W., Li, X., Gao, B., Gan, H., Yin, L., et al. (2015b). Effect of serum from postmenopausal women with osteoporosis exhibiting the Kidney-Yang deficiency pattern on bone formation in an hFOB 1.19 human osteoblastic cell line. *Exp. Ther. Med.* 10, 1089–1095. doi: 10.3892/etm.2015.2616
- Li, Y., Lu, S. S., Tang, G. Y., Hou, M., Tang, Q., Zhang, X. N., et al. (2014b). Effect of Morinda officinalis capsule on osteoporosis in ovariectomized rats. *Chin. J. Nat. Medicines* 12, 204–212. doi: 10.1016/S1875-5364(14)60034-0
- Li, Y., Wang, M. J., Li, S., Zhang, Y. M., Zhao, Y., Xie, R. M., et al. (2011). Effect of total glycosides from Eucommia ulmoides seed on bone microarchitecture in rats. *Phytother. Res.* 25, 1895–1897. doi: 10.1002/ptr.3543
- Li, Y. J., Kim, T. H., Kwak, H. B., Lee, Z. H., Lee, S. Y., and Jhon, G. J. (2007). Chloroform extract of deer antler inhibits osteoclast differentiation and bone resorption. *J. Ethnopharmacol.* 113, 191–198. doi: 10.1016/j.jep.2007.04.020
- Li, Y. M., Jia, M., Li, H. Q., Zhang, N. D., Wen, X., Rahman, K., et al. (2015c). Cnidium monnieri: a review of traditional uses, phytochemical and ethnopharmacological properties. *Am. J. Chin. Med.* 43, 835–877. doi: 10.1142/ S0192415X15500500
- Liang, H. D., Yu, F., Tong, Z. H., and Huang, Z. G. (2011). Effect of Cistanches Herba aqueous extract on bone loss in ovariectomized rat. *Int. J. Mol. Sci.* 12, 5060–5069. doi: 10.3390/ijms12085060
- Liang, H. D., Yu, F., Tong, Z. H., Zhang, H. Q., and Liang, W. (2013). Cistanches Herba aqueous extract affecting serum BGP and TRAP and bone marrow Smad1 mRNA, Smad5 mRNA, TGF-beta 1 mRNA and TIEG1 mRNA expression levels in osteoporosis disease. *Mol. Biol. Rep.* 40, 757–763. doi: 10.1007/s11033-012-2065-2
- Liang, W., Lin, M., Li, X., Li, C., Gao, B., Gan, H., et al. (2012). Icariin promotes bone formation via the BMP-2/Smad4 signal transduction pathway in the hFOB 1.19 human osteoblastic cell line. *Int. J. Mol. Med.* 30, 889–895. doi: 10.3892/ijmm.2012.1079
- Lim, S. H., Ha, T. Y., Kim, S. R., Ahn, J., Park, H. J., and Kim, S. (2009). Ethanol extract of Psoralea corylifolia L. and its main constituent, bakuchiol, reduce bone loss in ovariectomised Sprague-Dawley rats. *Br. J. Nutr.* 101, 1031–1039. doi: 10.1017/S0007114508066750
- Lin, C. H., Funayama, S., Peng, S. F., Kuo, C. L., and Chung, J. G. (2018). The ethanol extraction of prepared Psoralea corylifolia induces apoptosis and autophagy and alteres genes expression assayed by cDNA microarray in human prostate cancer PC-3 cells. *Environ. Toxicol.* 33, 770–788. doi: 10.1002/tox.22564

- Liu, H., Xiong, Y., Zhu, X., Gao, H., Yin, S., Wang, J., et al. (2017a). Icariin improves osteoporosis, inhibits the expression of PPARgamma, C/EBPalpha, FABP4 mRNA, N1ICD and jagged1 proteins, and increases Notch2 mRNA in ovariectomized rats. *Exp. Ther. Med.* 13, 1360–1368. doi: 10.3892/ etm.2017.4128
- Liu, H., Zhu, R., Wang, L., Liu, C., Ma, R., Qi, B., et al. (2018). Radix Salviae miltiorrhizae improves bone microstructure and strength through Wnt/betacatenin and osteoprotegerin/receptor activator for nuclear factor-kappaB ligand/cathepsin K signaling in ovariectomized rats. *Phytother. Res.* 32, 2487– 2500. doi: 10.1002/ptr.6188
- Liu, L. J., Liu, L. Q., Bo, T., Li, S. J., Zhu, Z., Cui, R. R., et al. (2013). Puerarin suppress apoptosis of human osteoblasts via ERK signaling pathway. Int. J. Endocrinol. 2013, 786574. doi: 10.1155/2013/786574
- Liu, M., Li, Y., and Yang, S. T. (2014a). Curculigoside improves osteogenesis of human amniotic fluid-derived stem cells. *Stem Cells Dev.* 23, 146–154. doi: 10.1089/scd.2013.0261
- Liu, M., Zhong, C., He, R. X., and Chen, L. F. (2012a). Icariin associated with exercise therapy is an effective treatment for postmenopausal osteoporosis. *Chin. Med.* J. (Engl.) 125, 1784–1789. doi: 10.3760/cma.j.issn.0366-6999.2012.10.018
- Liu, M. J., Xiao, G. G., Rong, P. J., Zhang, Z. G., Dong, J. Z., Zhao, H. Y., et al. (2012b). Therapeutic effects of radix dipsaci, pyrola herb, and cynomorium songaricum on bone metabolism of ovariectomized rats. *BMC Complement Altern. Med.* 12, 1–10. doi: 10.1186/1472-6882-12-67
- Liu, W., Mao, L., Ji, F., Chen, F., Wang, S., and Xie, Y. (2017b). Icariside II activates EGFR-Akt-Nrf2 signaling and protects osteoblasts from dexamethasone. *Oncotarget* 8, 2594–2603. doi: 10.18632/oncotarget.13732
- Liu, X., Niu, Y., Xie, W., Wei, D., and Du, Q. (2019). Tanshinone IIA promotes osteogenic differentiation of human periodontal ligament stem cells via ERK1/2-dependent Runx2 induction. Am. J. Transl. Res. 11, 340–350.
- Liu, Y., Cui, Y., Zhang, X., Gao, X., Su, Y., Xu, B., et al. (2016). Effects of salvianolate on bone metabolism in glucocorticoid-treated lupus-prone B6.MRL-Fas (lpr)/J mice. Drug Des. Devel. Ther. 10, 2535–2546. doi: 10.2147/DDDT.S110125
- Liu, Y. Q., Yang, Q. X., Cheng, M. C., and Xiao, H. B. (2014b). Synergistic inhibitory effect of Icariside II with Icaritin from Herba Epimedii on preosteoclastic RAW264.7 cell growth. *Phytomedicine* 21, 1633–1637. doi: 10.1016/j.phymed.2014.07.016
- Liu, Y. R., Qu, S. X., Maitz, M. F., Tan, R., and Weng, J. (2007). The effect of the major components of Salvia Miltiorrhiza Bunge on bone marrow cells. *J. Ethnopharmacol.* 111, 573–583. doi: 10.1016/j.jep.2007.01.005
- Liu, Z. G., Zhang, R., Li, C., Ma, X., Liu, L., Wang, J. P., et al. (2009). The osteoprotective effect of Radix Dipsaci extract in ovariectomized rats. *J. Ethnopharmacol.* 123, 74–81. doi: 10.1016/j.jep.2009.02.025
- Lorentzon, M., and Cummings, S. R. (2015). Osteoporosis: the evolution of a diagnosis. J. Intern. Med. 277, 650–661. doi: 10.1111/joim.12369
- Lungu, A. E., Lazar, M. A., Tonea, A., Rotaru, H., Roman, R. C., and Badea, M. E. (2018). Observational study of the bisphosphonate-related osteonecrosis of jaws. *Clujul Med.* 91, 209–215. doi: 10.15386/cjmed-838
- Luo, S., Yang, Y., Chen, J., Zhong, Z., Huang, H., Zhang, J., et al. (2016). Tanshinol stimulates bone formation and attenuates dexamethasone-induced inhibition of osteogenesis in larval zebrafish. J. Orthop. Translat. 4, 35–45. doi: 10.1016/j. jot.2015.07.002
- Luo, Z., Liu, M., Sun, L., and Rui, F. (2015). Icariin recovers the osteogenic differentiation and bone formation of bone marrow stromal cells from a rat model of estrogen deficiency-induced osteoporosis. *Mol. Med. Rep.* 12, 382– 388. doi: 10.3892/mmr.2015.3369
- Lv, H., Che, T., Tang, X., Liu, L., and Cheng, J. (2015). Puerarin enhances proliferation and osteoblastic differentiation of human bone marrow stromal cells via a nitric oxide/cyclic guanosine monophosphate signaling pathway. *Mol. Med. Rep.* 12, 2283–2290. doi: 10.3892/mmr.2015.3647
- Ma, H. P., Ming, L. G., Ge, B. F., Zhai, Y. K., Song, P., Xian, C. J., et al. (2011). Icariin is more potent than genistein in promoting osteoblast differentiation and mineralization *in vitro*. J. Cell Biochem. 112, 916–923. doi: 10.1002/jcb.23007
- Ma, X. N., Zhou, J., Ge, B. F., Zhen, P., Ma, H. P., Shi, W. G., et al. (2013). Icariin induces osteoblast differentiation and mineralization without dexamethasone *in vitro*. *Planta Med*. 79, 1501–1508. doi: 10.1055/s-0033-1350802
- Ma, Y., Yang, H., and Huang, J. (2018). Icariin ameliorates dexamethasoneinduced bone deterioration in an experimental mouse model *via* activation of

microRNA186 inhibition of cathepsin K. Mol. Med. Rep. 17, 1633–1641. doi: 10.3892/mmr.2017.8065

- Manonai, J., Chittacharoen, A., Udomsubpayakul, U., Theppisai, H., and Theppisai, U. (2008). Effects and safety of Pueraria mirifica on lipid profiles and biochemical markers of bone turnover rates in healthy postmenopausal women. *Menopause* 15, 530–535. doi: 10.1097/gme.0b013e31815c5fd8
- Meng, F., Xiong, Z., Sun, Y., and Li, F. (2004). Coumarins from Cnidium monnieri (L.) and their proliferation stimulating activity on osteoblast-like UMR106 cells. *Pharmazie* 59, 643–645.
- Meng, F. H., Li, Y. B., Xiong, Z. L., Jiang, Z. M., and Li, F. M. (2005). Osteoblastic proliferative activity of Epimedium brevicornum Maxim. *Phytomedicine* 12, 189–193. doi: 10.1016/j.phymed.2004.03.007
- Ming, L. G., Zhou, J., Cheng, G. Z., Ma, H. P., and Chen, K. M. (2013). Osthol, a coumarin isolated from common cnidium fruit, enhances the differentiation and maturation of osteoblasts *in vitro* (vol 88, pg 33, 2011). *Pharmacol.* 92, 117–119. doi: 10.1159/000328776
- Miura, H., Nishida, H., and Linuma, M. (1996). Effect of crude fractions of psoralea corylifolia seed extract on bone calcification. *Planta Med.* 62, 150–153. doi: 10.1055/s-2006-957839
- Mok, S. K., Chen, W. F., Lai, W. P., Leung, P. C., Wang, X. L., Yao, X. S., et al. (2010). Icariin protects against bone loss induced by oestrogen deficiency and activates oestrogen receptor-dependent osteoblastic functions in UMR 106 cells. Br. J. Pharmacol. 159, 939–949. doi: 10.1111/j.1476-5381.2009.00593.x
- Mukwaya, E., Xu, F., Wong, M. S., and Zhang, Y. (2014). Chinese herbal medicine for bone health. *Pharm. Biol.* 52, 1223–1228. doi: 10.3109/13880209.2014.884606
- Nguyen, X. C., Chau, V. M., Phan, V. K., Hoang, T. H., Ninh, K. B., Nguyen, X. N., et al. (2009). Inhibitors of osteoclast formation from Rhizomes of Cibotium barometz. *J. Nat. Prod.* 72, 1673–1677. doi: 10.1021/np9004097
- Nian, H., Ma, M. H., Nian, S. S., and Xu, L. L. (2009). Antiosteoporotic activity of icariin in ovariectomized rats. *Phytomedicine* 16, 320–326. doi: 10.1016/j. phymed.2008.12.006
- Nicolin, V., Dal Piaz, F., Nori, S. L., Narducci, P., and De Tommasi, N. (2010). Inhibition of bone resorption by Tanshinone VI isolated from Salvia miltiorrhiza Bunge. *Eur. J. Histochem.* 54, e21. doi: 10.4081/ejh.2010.e21
- Nie, Y., Dong, X., He, Y., Yuan, T., Han, T., Rahman, K., et al. (2013). Medicinal plants of genus Curculigo: traditional uses and a phytochemical and ethnopharmacological review. J. Ethnopharmacol. 147, 547–563. doi: 10.1016/j. jep.2013.03.066
- Niu, Y., Li, C., Pan, Y., Li, Y., Kong, X., Wang, S., et al. (2015a). Treatment of Radix Dipsaci extract prevents long bone loss induced by modeled microgravity in hindlimb unloading rats. *Pharm. Biol.* 53, 110–116. doi: 10.3109/13880209.2014.911920
- Niu, Y. B., Kong, X. H., Li, Y. H., Fan, L., Pan, Y. L., Li, C. R., et al. (2015b). Radix Dipsaci total saponins stimulate MC3T3-E1 cell differentiation via the bone morphogenetic protein-2/MAPK/Smad-dependent Runx2 pathway. Mol. Med. Rep. 11, 4468–4472. doi: 10.3892/mmr.2015.3249
- Niu, Y. B., Li, Y. H., Huang, H. T., Kong, X. H., Zhang, R., Liu, L., et al. (2011). Asperosaponin VI, a saponin component from Dipsacus asper wall, induces osteoblast differentiation through bone morphogenetic protein-2/p38 and extracellular signal-regulated kinase 1/2 pathway. *Phytother. Res.* 25, 1700– 1706. doi: 10.1002/ptr.3414
- Niu, Y. B., Li, Y. H., Kong, X. H., Zhang, R., Sun, Y., Li, Q., et al. (2012). The beneficial effect of Radix Dipsaci total saponins on bone metabolism *in vitro* and *in vivo* and the possible mechanisms of action. *Osteoporos. Int.* 23, 2649– 2660. doi: 10.1007/s00198-012-1932-y
- Nowak, B., Matuszewska, A., Nikodem, A., Filipiak, J., Landwojtowicz, M., Sadanowicz, E., et al. (2017). Oral administration of kaempferol inhibits bone loss in rat model of ovariectomy-induced osteopenia. *Pharmacol. Rep.* 69, 1113–1119. doi: 10.1016/j.pharep.2017.05.002
- Pan, Y. L., Niu, Y. B., Li, C. R., Zhai, Y. K., Zhang, R., Guo, X., et al. (2014). Du-Zhong (Eucommia ulmoides) prevents disuse-induced osteoporosis in hind limb suspension rats. *Am. J. Chin. Med.* 42, 143–155. doi: 10.1142/ S0192415X14500104
- Park, C. K., Lee, Y., Chang, E. J., Lee, M. H., Yoon, J. H., Ryu, J. H., et al. (2008). Bavachalcone inhibits osteoclast differentiation through suppression of NFATc1 induction by RANKL. *Biochem. Pharmacol.* 75, 2175–2182. doi: 10.1016/j.bcp.2008.03.007

- Park, K. H., Gu, D. R., Jin, S. H., Yoon, C. S., Ko, W., Kim, Y. C., et al. (2017). Pueraria lobate inhibits RANKL-mediated osteoclastogenesis via downregulation of CREB/PGC1beta/c-Fos/NFATc1 signaling. Am. J. Chin. Med. 45, 1725–1744. doi: 10.1142/S0192415X17500938
- Pavone, V., Testa, G., Giardina, S. M. C., Vescio, A., Restivo, D. A., and Sessa, G. (2017). Pharmacological therapy of osteoporosis: a systematic current review of literature. *Front. Pharmacol.* 8, 803. doi: 10.3389/fphar.2017.00803
- Peng, S., Zhang, G., He, Y., Wang, X., Leung, P., Leung, K., et al. (2009). Epimediumderived flavonoids promote osteoblastogenesis and suppress adipogenesis in bone marrow stromal cells while exerting an anabolic effect on osteoporotic bone. *Bone* 45, 534–544. doi: 10.1016/j.bone.2009.05.022
- Prasain, J. K., Peng, N., Rajbhandari, R., and Wyss, J. M. (2012). The Chinese Pueraria root extract (Pueraria lobata) ameliorates impaired glucose and lipid metabolism in obese mice. *Phytomedicine* 20, 17–23. doi: 10.1016/j. phymed.2012.09.017
- Qi, S. S., Zheng, H. X., Chen, C., and Jiang, H. (2019). Du-Zhong (Eucommia ulmoides Oliv.) cortex extract alleviates lead acetate-induced bone loss in rats. *Biol. Trace Elem. Res.* 187, 172–180. doi: 10.1007/s12011-018-1362-6
- Rachner, T. D., Khosla, S., and Hofbauer, L. C. (2011). Osteoporosis: now and the future. *Lancet* 377, 1276–1287. doi: 10.1016/S0140-6736(10)62349-5
- Ramchandani, D., Ganeshpurkar, A., Bansal, D., Karchuli, M. S., and Dubey, N. (2014). Protective effect of curculigo orchioides extract on cyclophosphamideinduced neurotoxicity in murine model. *Toxicol. Int.* 21, 232–235. doi: 10.4103/0971-6580.155323
- Seo, B. I., Ku, S. K., Cha, E. M., Park, J. H., Kim, J. D., Choi, H. Y., et al. (2005). Effect of Mornidae Radix extracts on experimental osteoporosis in sciatic neurectomized mice. *Phytother. Res.* 19, 231–238. doi: 10.1002/ptr.1683
- Shao, H., Shen, J., Wang, M., Cui, J., Wang, Y., Zhu, S., et al. (2015). Icariin protects against titanium particle-induced osteolysis and inflammatory response in a mouse calvarial model. *Biomaterials* 60, 92–99. doi: 10.1016/j. biomaterials.2015.04.048
- Shen, G., Ren, H., Qiu, T., Liang, D., Xie, B., Zhang, Z., et al. (2016). Implications of the Interaction Between miRNAs and Autophagy in Osteoporosis. *Calcif. Tissue Int.* 99, 1–12. doi: 10.1007/s00223-016-0122-x
- Shen, Q., Zeng, D., Zhou, Y., Xia, L., Zhao, Y., Qiao, G., et al. (2013). Curculigoside promotes osteogenic differentiation of bone marrow stromal cells from ovariectomized rats. *J. Pharm. Pharmacol.* 65, 1005–1013. doi: 10.1111/ jphp.12054
- Shi, C., Yuan, P., and Yang, X. (2012). Observation of granule of Kang-shu Jian-gu on postmenopausal osteoporosis: a report of 40 cases. *Shanxi J. Tradit. Chin. Med.* 8, 16–17.
- Shu, B., Shi, Q., and Wang, Y. J. (2015). Shen (Kidney)-tonifying principle for primary osteoporosis: to treat both the disease and the Chinese medicine syndrome. *Chin. J. Integr. Med.* 21, 656–661. doi: 10.1007/s11655-015-2306-z
- Song, D. Z., Cao, Z., Liu, Z. B., Tickner, J., Qiu, H., Wang, C., et al. (2018). Cistanche deserticola polysaccharide attenuates osteoclastogenesis and bone resorption via inhibiting RANKL signaling and reactive oxygen species production. J. Cell Physiol. 233, 9674–9684. doi: 10.1002/jcp.26882
- Song, L., Zhao, J., Zhang, X., Li, H., and Zhou, Y. (2013). Icariin induces osteoblast proliferation, differentiation and mineralization through estrogen receptormediated ERK and JNK signal activation. *Eur. J. Pharmacol.* 714, 15–22. doi: 10.1016/j.ejphar.2013.05.039
- Sozen, T., Ozisik, L., and Basaran, N. C. (2017). An overview and management of osteoporosis. Eur. J. Rheumatol. 4, 46–56. doi: 10.5152/eurjrheum.2016.048
- Spivakovsky, S. (2017). Treatment for bisphosphonate-related osteonecrosis of the jaw. *Evid Based Dent.* 18, 56. doi: 10.1038/sj.ebd.6401243
- Sui, Z. G., Zhang, L. H., Huo, Y. S., and Zhang, Y. K. (2014). Bioactive components of velvet antlers and their pharmacological properties. *J. Pharm. BioMed.* 87, 229–240. doi: 10.1016/j.jpba.2013.07.044
- Suvarna, V., Sarkar, M., Chaubey, P., Khan, T., Sherje, A., Patel, K., et al. (2018). Bone health and natural products- an insight. *Front. Pharmacol.* 9, 981. doi: 10.3389/fphar.2018.00981
- Takamura, C., Hirata, T., Ueda, T., Ono, M., Miyashita, H., Ikeda, T., et al. (2007). Iridoids from the green leaves of Eucommia ulmoides. J. Nat. Prod. 70, 1312– 1316. doi: 10.1021/np0780046
- Tan, C., Wang, A., Liu, C., Li, Y., Shi, Y., and Zhou, M. S. (2017). Puerarin improves vascular insulin resistance and cardiovascular remodeling in

salt-sensitive hypertension. Am. J. Chin. Med. 45, 1169–1184. doi: 10.1142/ S0192415X17500641

- Tan, S., Xu, J., Lai, A., Cui, R., Bai, R., Li, S., et al. (2019). Curculigoside exerts significant antiarthritic effects *in vivo* and *in vitro via* regulation of the JAK/ STAT/NFkappaB signaling pathway. *Mol. Med. Rep.* 19, 2057–2064. doi: 10.3892/mmr.2019.9854
- Tan, X. L., Zhang, Y. H., Cai, J. P., Zhu, L. H., Ge, W. J., and Zhang, X. (2014). 5-(Hydroxymethyl)-2-furaldehyde inhibits adipogenic and enhances osteogenic differentiation of rat bone mesenchymal stem cells. *Nat. Prod. Commun.* 9, 529–532. doi: 10.1177/1934578X1400900427
- Tao, Y., Huang, S. R., Li, W. D., and Cai, B. C. (2019). Simultaneous determination of ten bioactive components in raw and processed Radix Dipsaci by UPLC-Q-TOF-MS. J. Chromatogr. Sci. 57, 122–129. doi: 10.1093/chromsci/bmy093
- Tsai, M. H., Huang, G. S., Hung, Y. C., Bin, L., Liao, L. T., and Lin, L. W. (2007). Psoralea corylifolia extract ameliorates experimental osteoporosis in ovariectomized rats. Am. J. Chin. Med. 35, 669–680. doi: 10.1142/ S0192415X07005168
- Tseng, S. H., Sung, C. H., Chen, L. G., Lai, Y. J., Chang, W. S., Sung, H. C., et al. (2014). Comparison of chemical compositions and osteoprotective effects of different sections of velvet antler. *J. Ethnopharmacol.* 151, 352–360. doi: 10.1016/j.jep.2013.10.060
- Tseng, S. H., Sung, H. C., Chen, L. G., Lai, Y. J., Wang, K. T., Sung, C. H., et al. (2012). Effects of velvet antler with blood on bone in ovariectomized rats. *Molecules* 17, 10574–10585. doi: 10.3390/molecules170910574
- Wang, G. J., Tsai, T. H., and Lin, L. C. (2007). Prenylflavonol, acylated flavonol glycosides and related compounds from Epimedium sagittatum. *Phytochem.* 68, 2455–2464. doi: 10.1016/j.phytochem.2007.05.035
- Wang, J. Y., Yuan, Y., Chen, X. J., Fu, S. G., Zhang, L., Hong, Y. L., et al. (2016a). Extract from Eucommia ulmoides Oliv. ameliorates arthritis via regulation of inflammation, synoviocyte proliferation and osteoclastogenesis in vitro and in vivo. J. Ethnopharmacol. 194, 609–616. doi: 10.1016/j.jep.2016.10.038
- Wang, L., He, Y. J., Han, T., Zhao, L., Lv, L., He, Y. Q., et al. (2017a). Metabolites of curculigoside in rats and their antiosteoporotic activities in osteoblastic MC3T3-E1 cells. *Fitoterapia* 117, 109–117. doi: 10.1016/j.fitote.2017.01.009
- Wang, L., Li, Y., Guo, Y., Ma, R., Fu, M., Niu, J., et al. (2016b). Herba Epimedii: an ancient Chinese herbal medicine in the prevention and treatment of osteoporosis. *Curr. Pharm. Des.* 22, 328–349. doi: 10.2174/138161282266615 1112145907
- Wang, N., Xu, P., Wang, X., Yao, W., Yu, Z., Wu, R., et al. (2019a). Integrated pathological cell fishing and network pharmacology approach to investigate main active components of Er-Xian decotion for treating osteoporosis. *J. Ethnopharmacol.* 241, 111977. doi: 10.1016/j.jep.2019.111977
- Wang, P. P., Zhu, X. F., Yang, L., Liang, H., Feng, S. W., and Zhang, R. H. (2012a). Puerarin stimulates osteoblasts differentiation and bone formation through estrogen receptor, p38 MAPK, and Wnt/beta-catenin pathways. J. Asian Nat. Prod. Res. 14, 897–905. doi: 10.1080/10286020.2012.702757
- Wang, T., Liu, Q., Tjhioe, W., Zhao, J., Lu, A., Zhang, G., et al. (2017b). Therapeutic potential and outlook of alternative medicine for osteoporosis. *Curr. Drug Targets* 18, 1051–1068. doi: 10.2174/1389450118666170321105425
- Wang, X., Wu, J., Chiba, H., Yamada, K., and Ishimi, Y. (2005). Puerariae radix prevents bone loss in castrated male mice. *Metab.* 54, 1536–1541. doi: 10.1016/j. metabol.2005.05.022
- Wang, X. Q., Zhang, M. L., Zhang, D. W., Wang, S. M., and Yan, C. Y. (2017c). An O-acetyl-glucomannan from the rhizomes of Curculigo orchioides: Structural characterization and anti-osteoporosis activity *in vitro*. *Carbohydr. Polym.* 174, 48–56. doi: 10.1016/j.carbpol.2017.06.051
- Wang, X. Q., Zhang, M. L., Zhang, D. W., Wang, X. L., Cao, H. J., Zhang, Q., et al. (2019b). Structural elucidation and anti-osteoporosis activities of polysaccharides obtained from Curculigo orchioides. *Carbohydr. Polym.* 203, 292–301. doi: 10.1016/j.carbpol.2018.09.059
- Wang, Y., Wang, W. L., Xie, W. L., Li, L. Z., Sun, J., Sun, W. J., et al. (2013a). Puerarin stimulates proliferation and differentiation and protects against cell death in human osteoblastic MG-63 cells via ER-dependent MEK/ERK and PI3K/Akt activation. *Phytomedicine* 20, 787–796. doi: 10.1016/j.phymed.2013.03.005
- Wang, Y., Yang, C., Xie, W. L., Zhao, Y. W., Li, Z. M., Sun, W. J., et al. (2014). Puerarin concurrently stimulates osteoprotegerin and inhibits receptor activator of NF-kappaB ligand (RANKL) and interleukin-6 production in

human osteoblastic MG-63 cells. *Phytomedicine* 21, 1032–1036. doi: 10.1016/j. phymed.2014.04.012

- Wang, Y., Zhao, L., Wang, Y., Xu, J., Nie, Y., Guo, Y., et al. (2012b). Curculigoside isolated from Curculigo orchioides prevents hydrogen peroxide-induced dysfunction and oxidative damage in calvarial osteoblasts. *Acta Biochim. Biophys. Sin. (Shanghai)* 44, 431–441. doi: 10.1093/abbs/gms014
- Wang, Y., Zhou, Y., Wang, X., Zhen, F., Chen, R., Geng, D. Q., et al. (2019c). Osthole alleviates MPTP-induced Parkinson's disease mice by suppressing Notch signaling pathway. *Int. J. Neurosci.* 129, 833–841. doi: 10.1080/00207454.2019.1573171
- Wang, Z., Wang, D., Yang, D., Zhen, W., Zhang, J., and Peng, S. (2018). The effect of icariin on bone metabolism and its potential clinical application. *Osteoporos. Int.* 29, 535–544. doi: 10.1007/s00198-017-4255-1
- Wang, Z. H., Huang, J., Ma, X. C., Li, G. Y., Ma, Y. P., Li, N., et al. (2013b). Phenolic glycosides from Curculigo orchioides Gaertn. *Fitoterapia* 86, 64–69. doi: 10.1016/j.fitote.2013.01.008
- Wei, H., Zili, L., Yuanlu, C., Biao, Y., Cheng, L., Xiaoxia, W., et al. (2011). Effect of icariin on bone formation during distraction osteogenesis in the rabbit mandible. *Int. J. Maxillofac Surg.* 40, 413–418. doi: 10.1016/j.ijom.2010.10.015
- Wei, L., Zhang, H., Li, X., Yang, C., Wang, G., Zhang, L., et al. (2016). Efficacy and safety evaluation of intravenous infusion of cervus and cucumis polypeptides for treatment of avascular necrosis of the femoral head: a randomized clinical trial. J. Tradit. Chin. Med. 36, 39–44. doi: 10.1016/S0254-6272(16)30006-1
- Wei, R., Lao, R., Liu, Z., Xu, G., and Li, Z. (2011). Effect of anti-osteoporosis capsule on ostealgia and bone metabolismof postmenopausal osteoporosis patients. J. New Chin. Med. 43, 61–63.
- Weng, Z. B., Gao, Q. Q., Wang, F., Zhao, G. H., Yin, F. Z., Cai, B. C., et al. (2015). Positive skeletal effect of two ingredients of Psoralea corylifolia L. @ on estrogen deficiency-induced osteoporosis and the possible mechanisms of action. *Mol. Cell Endocrinol.* 417, 103–113. doi: 10.1016/j.mce.2015.09.025
- Wiseman, H. (2000). The therapeutic potential of phytoestrogens. Expert Opin. Invest. Drugs 9, 1829–1840. doi: 10.1517/13543784.9.8.1829
- Wong, R. W., Rabie, B., Bendeus, M., and Hagg, U. (2007a). The effects of Rhizoma Curculiginis and Rhizoma Drynariae extracts on bones. *Chin. Med.* 2, 13. doi: 10.1186/1749-8546-2-13
- Wong, R. W. K., Rabie, A. B. M., and Hagg, E. U. O. (2007b). The effect of crude extract from Radix Dipsaci on bone in mice. *Phytother. Res.* 21, 596–598. doi: 10.1002/ptr.2126
- Wu, F., Li, H., Jin, L., Li, X., Ma, Y., You, J., et al. (2013). Deer antler base as a traditional Chinese medicine: a review of its traditional uses, chemistry and pharmacology. *J. Ethnopharmacol.* 145, 403–415. doi: 10.1016/j.jep.2012.12.008
- Wu, Q., and Yang, X. W. (2009). The constituents of Cibotium barometz and their permeability in the human Caco-2 monolayer cell model. *J. Ethnopharmacol.* 125, 417–422. doi: 10.1016/j.jep.2009.07.017
- Wu, Y., Xia, L., Zhou, Y., Xu, Y., and Jiang, X. (2015a). Icariin induces osteogenic differentiation of bone mesenchymal stem cells in a MAPK-dependent manner. *Cell Prolif.* 48, 375–384. doi: 10.1111/cpr.12185
- Wu, Y. B., Zheng, C. J., Qin, L. P., Sun, L. N., Han, T., Jiao, L., et al. (2009). Antiosteoporotic activity of anthraquinones from Morinda officinalis on osteoblasts and osteoclasts. *Molecules* 14, 573–583. doi: 10.3390/ molecules14010573
- Wu, Z. Q., Chen, D. L., Lin, F. H., Lin, L., Shuai, O., Wang, J. Y., et al. (2015b). Effect of bajijiasu isolated from Morinda officinalis F. C. how on sexual function in male mice and its antioxidant protection of human sperm. *J. Ethnopharmacol.* 164, 283–292. doi: 10.1016/j.jep.2015.02.016
- Xiao, Q., Chen, A., and Guo, F. (2005). Effects of Icariin on expression of OPN mRNA and type I collagen in rat osteoblasts *in vitro*. J. Huazhong Univ. Sci. Technolog. Med. Sci. 25, 690–692. doi: 10.1007/BF02896172
- Xie, G. P., Jiang, N., Wang, S. N., Qi, R. Z., Wang, L., Zhao, P. R., et al. (2015). Eucommia ulmoides Oliv. bark aqueous extract inhibits osteoarthritis in a rat model of osteoarthritis (vol 162, pg 148, 2015). *J. Ethnopharmacol.* 172, 436– 436. doi: 10.1016/j.jep.2015.07.012
- Xu, F., Ding, Y., Guo, Y., Liu, B., Kou, Z., Xiao, W., et al. (2016). Anti-osteoporosis effect of Epimedium via an estrogen-like mechanism based on a system-level approach. J. Ethnopharmacol. 177, 148–160. doi: 10.1016/j.jep.2015.11.007
- Xu, G., Zhao, M. J., Sun, N., Ju, C. G., and Jia, T. Z. (2014). Effect of the RW-Cb and its active ingredient like P-acid and P-aldehyde on primary rat osteoblasts. *J. Ethnopharmacol.* 151, 237–241. doi: 10.1016/j.jep.2013.10.030

- Xu, X. X., Zhang, Z. Z., Wang, W. P., Yao, H. Q., and Ma, X. Q. (2017). Therapeutic effect of Cistanoside a on bone metabolism of ovariectomized mice. *Molecules* 22, 1–11. doi: 10.3390/molecules22020197
- Yang, H. M., Shin, H. K., Kang, Y. H., and Kim, J. K. (2009). Cuscuta chinensis extract promotes osteoblast differentiation and mineralization in human osteoblast-like MG-63 cells. J. Med. Food 12, 85–92. doi: 10.1089/jmf.2007.0665
- Yang, J. H., Cao, Y., Wang, R. L., Fei, Y. R., Zhang, H., Feng, P., et al. (2010a). Anti-resorptive effect of pilose antler blood (Cervus nippon Temminck) in ovariectomized rats. *Indian J. Exp. Biol.* 48, 554–558.
- Yang, L., Chen, Q., Wang, F., and Zhang, G. (2011). Antiosteoporotic compounds from seeds of Cuscuta chinensis. J. Ethnopharmacol. 135, 553–560. doi: 10.1016/j.jep.2011.03.056
- Yang, L., Lu, D., Guo, J., Meng, X., Zhang, G., and Wang, F. (2013). Icariin from Epimedium brevicornum Maxim promotes the biosynthesis of estrogen by aromatase (CYP19). *J. Ethnopharmacol.* 145, 715–721. doi: 10.1016/j.jep.2012.11.031
- Yang, X., Hu, W., Zhang, Q., Wang, Y., and Sun, L. (2010b). Puerarin inhibits C-reactive protein expression via suppression of nuclear factor kappaB activation in lipopolysaccharide-induced peripheral blood mononuclear cells of patients with stable angina pectoris. *Basic Clin. Pharmacol. Toxicol.* 107, 637–642. doi: 10.1111/j.1742-7843.2010.00548.x
- Yang, Z., Huang, J. H., Liu, S. F., Zhao, Y. J., Shen, Z. Y., Wang, Y. J., et al. (2012). The osteoprotective effect of psoralen in ovariectomy-induced osteoporotic rats via stimulating the osteoblastic differentiation from bone mesenchymal stem cells. *Menopause* 19, 1156–1164. doi: 10.1097/gme.0b013e3182507e18
- Yao, C. H., Tsai, H. M., Chen, Y. S., and Liu, B. S. (2005). Fabrication and evaluation of a new composite composed of tricalcium phosphate, gelatin, and Chinese medicine as a bone substitute. *J. BioMed. Mater. Res. B* 75b, 277–288. doi: 10.1002/jbm.b.30294
- Ye, Y., Jing, X., Li, N., Wu, Y., Li, B., and Xu, T. (2017). Icariin promotes proliferation and osteogenic differentiation of rat adipose-derived stem cells by activating the RhoA-TAZ signaling pathway. *BioMed. Pharmacother.* 88, 384–394. doi: 10.1016/j.biopha.2017.01.075
- Yuan, S. Y., Sheng, T., Liu, L. Q., Zhang, Y. L., Liu, X. M., Ma, T., et al. (2016). Puerarin prevents bone loss in ovariectomized mice and inhibits osteoclast formation *in vitro. Chin. J. Nat. Med.* 14, 265–269. doi: 10.1016/S1875-5364(16)30026-7
- Zanotti, S., Yu, J., Adhikari, S., and Canalis, E. (2018). Glucocorticoids inhibit notch target gene expression in osteoblasts. J. Cell Biochem. 119, 6016–6023. doi: 10.1002/jcb.26798
- Zeng, X., Feng, Q., Zhao, F., Sun, C., Zhou, T., Yang, J., et al. (2018). Puerarin inhibits TRPM3/miR-204 to promote MC3T3-E1 cells proliferation, differentiation and mineralization. *Phytother. Res.* 32, 996–1003. doi: 10.1002/ptr.6034
- Zhai, Y. K., Guo, X., Pan, Y. L., Niu, Y. B., Li, C. R., Wu, X. L., et al. (2013). A systematic review of the efficacy and pharmacological profile of Herba Epimedii in osteoporosis therapy. *Pharmazie* 68, 713–722. doi: 10.1691/ph.2013.2900
- Zhang, G., Qin, L., Hung, W. Y., Shi, Y. Y., Leung, P. C., Yeung, H. Y., et al. (2006). Flavonoids derived from herbal Epimedium Brevicornum Maxim prevent OVX-induced osteoporosis in rats independent of its enhancement in intestinal calcium absorption. *Bone* 38, 818–825. doi: 10.1016/j.bone.2005.11.019
- Zhang, G., Qin, L., and Shi, Y. (2007a). Epimedium-derived phytoestrogen flavonoids exert beneficial effect on preventing bone loss in late postmenopausal women: a 24-month randomized, double-blind and placebo-controlled trial. *J. Bone Miner. Res.* 22, 1072–1079. doi: 10.1359/jbmr.070405
- Zhang, J., Song, J., and Shao, J. (2015). Icariin attenuates glucocorticoid-induced bone deteriorations, hypocalcemia and hypercalciuria in mice. *Int. J. Clin. Exp. Med.* 8, 7306–7314.
- Zhang, L. Z., Xin, J. L., Zhang, X. P., Fu, Q., Zhang, Y., and Zhou, Q. L. (2013). The anti-osteoporotic effect of velvet antler polypeptides from Cervus elaphus Linnaeus in ovariectomized rats. *J. Ethnopharmacol.* 150, 181–186. doi: 10.1016/j.jep.2013.08.029
- Zhang, Q. Y., Qin, L. P., He, W. D., Van Puyvelde, L., Maes, D., Adams, A., et al. (2007b). Coumarins from Cnidium monnieri and their antiosteoporotic activity. *Planta Med.* 73, 13–19. doi: 10.1055/s-2006-951724
- Zhang, R., Liu, Z. G., Li, C., Hu, S. J., Liu, L., Wang, J. P., et al. (2009). Du-Zhong (Eucommia ulmoides Oliv.) cortex extract prevent OVX-induced osteoporosis in rats. *Bone* 45, 553–559. doi: 10.1016/j.bone.2008.08.127
- Zhang, R., Pan, Y. L., Hu, S. J., Kong, X. H., Juan, W., and Mei, Q. B. (2014). Effects of total lignans from Eucommia ulmoides barks prevent bone loss *in vivo* and *in vitro*. J. Ethnopharmacol. 155, 104–112. doi: 10.1016/j.jep.2014.04.031

- Zhang, W. P., Fujikawa, T., Mizuno, K., Ishida, T., Ooi, K., Hirata, T., et al. (2012). Eucommia Leaf Extract (ELE) prevents OVX-induced osteoporosis and obesity in rats. Am. J. Chin. Med. 40, 735–752. doi: 10.1142/S0192415X12500553
- Zhang, X. N., Zhao, W. W., Wang, Y., Lu, J. J., and Chen, X. P. (2016a). The chemical constituents and bioactivities of Psoralea corylifolia Linn.: a review. Am. J. Chin. Med. 44, 35–60. doi: 10.1142/S0192415X16500038
- Zhang, Y., Xie, Y., Liao, X., Jia, Q., and Chai, Y. (2017a). A Chinese patent medicine Salvia miltiorrhiza depside salts for infusion combined with conventional treatment for patients with angina pectoris: a systematic review and metaanalysis of randomized controlled trials. *Phytomedicine* 25, 100–117. doi: 10.1016/j.phymed.2017.01.002
- Zhang, Z., Zhang, Q., Yang, H., Liu, W., Zhang, N., Qin, L., et al. (2016b). Monotropein isolated from the roots of Morinda officinalis increases osteoblastic bone formation and prevents bone loss in ovariectomized mice. *Fitoterapia* 110, 166–172. doi: 10.1016/j.fitote.2016.03.013
- Zhang, Z. G., Zhang, Q. Y., Yang, H., Liu, W., Zhang, N. D., Qin, L. P., et al. (2016c). Monotropein isolated from the roots of Morinda officinalis increases osteoblastic bone formation and prevents bone loss in ovariectomized mice. *Fitoterapia* 110, 166–172. doi: 10.1016/j.fitote.2016.03.013
- Zhang, Z. R., Leung, W. N., Li, G., Kong, S. K., Lu, X., Wong, Y. M., et al. (2017b). Osthole enhances osteogenesis in osteoblasts by elevating transcription factor osterix via cAMP/CREB signaling in vitro and in vivo. Nutrients 9, 1–15. doi: 10.3390/nu9060588
- Zhao, J., Ohba, S., Shinkai, M., Chung, U. I., and Nagamune, T. (2008). Icariin induces osteogenic differentiation *in vitro* in a BMP- and Runx2-dependent manner. *Biochem. Biophys. Res. Commun.* 369, 444–448. doi: 10.1016/j. bbrc.2008.02.054

- Zhao, X., Wu, Z. X., Zhang, Y., Yan, Y. B., He, Q., Cao, P. C., et al. (2011). Anti-osteoporosis activity of Cibotium barometz extract on ovariectomyinduced bone loss in rats. *J. Ethnopharmacol.* 137, 1083–1088. doi: 10.1016/j. jep.2011.07.017
- Zhu, F. B., Wang, J. Y., Zhang, Y. L., Quan, R. F., Yue, Z. S., Zeng, L. R., et al. (2015). Curculigoside regulates proliferation, differentiation, and pro-inflammatory cytokines levels in dexamethasone-induced rat calvarial osteoblasts. *Int. J. Clin. Exp. Med.* 8, 12337–12346.
- Zhu, M. Y., Wang, C. J., Zhang, H. S., Pei, X. W., and Fen, J. M. (2008). Protective effect of polysaccharides from morinda officinalis on bone loss in ovariectomized rats. *Int. J. Biol. Macromol.* 43, 276–278. doi: 10.1016/j. ijbiomac.2008.06.008

Conflict of Interest: The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

The handling editor declared a shared affiliation, though no other collaboration, with one of the authors, XL.

Copyright © 2019 He, Li, Wang, Bennett, Chen, Xiao, Zhan, Chen, Hou, Chen, Wang, Xu and Lin. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.