DOI: 10.2478/bjmg-2023-0003

\$ sciendo

THE PREDISPOSITION FOR TYPE 2 DIABETES MELLITUS AND METABOLIC SYNDROME

Zenoaga-Barbăroșie C¹, Berca L², Vassu-Dimov T¹, Toma M³, Nica MI⁴, Alexiu-Toma OA¹, Ciornei C^{5,6}, Albu A⁶, Nica S^{6,7*}, Nistor C^{5,8}, Nica R^{9,10}

*Corresponding Author: Ass. Prof. Silvia Nica MD, PhD, Bucharest Emergency University Hospital, Splaiul Independentei Street, no 169, 5 District, Bucharest, Romania, 050098 Tel.: +40-21-31880500, email: silvia.nica@umfcd.ro

ABSTRACT

Type 2 diabetes mellitus (T2DM) and metabolic syndrome (MetS) are diseases caused by the interaction of genetic and non-genetic factors. Therefore, the aim of our study was to investigate the association between six common genetic polymorphisms and T2DM and MetS in males. A total of 120 T2DM, 75 MetS, and 120 healthy controls (HC) were included in the study. *ACE* ID, *eNOS* 4a/b, *ATR1* A1166C, *OXTR* (A>G), *SOD1* +35A/C, *CAT*-21A/T gene polymorphisms were genotyped by PCR or PCR-RFLP techniques. T2DM was diagnosed at an earlier age compared to MetS (54 vs 55 years old, p=0.0003) and the difference was greater in carriers of the *OXTR* G allele (54 vs 56 years old, p=0.0002) or both *OXTR* G and *eNOS* b alleles

¹ Department of Genetics, University of Bucharest, Bucharest, Romania

- ² Molecular Biology Department, National Research and Development Institute for Food Bioresources – IBA Bucharest, Bucharest, Romania
- ³ Emergency Department, Central Military Emergency Hospital Dr. Carol Davila, Bucharest, Romania
- ⁴ University of Medicine and Pharmacy Carol Davila, Bucharest, Romania
- ⁵ Preclinical Department, University of Medicine and Pharmacy Carol Davila, Bucharest, Romania
- ⁶ Emergency Department, Bucharest Emergency University Hospital, Bucharest, Romania.
- ⁷ Clinic Department 4, University of Medicine and Pharmacy Carol Davila, Bucharest, Romania
- ⁸ Thoracic Surgery, Central Military Emergency Hospital Dr. Carol Davila, Bucharest, Romania
- ⁹ Surgery 2, Central Military Emergency Hospital Dr. Carol Davila, Bucharest, Romania
- ¹⁰ Special Disciplines, University of Medicine and Pharmacy Carol Davila, Bucharest, Romania

(54 vs 56, p=0.00016). The *SOD1* AA genotype (O.R.=0.11, p=0.0006) and the presence of both *ACE* I and *OXTR1* A (O.R.=0.39, p=0.0005) alleles revealed to be protective for T2DM. *SOD1* AA and AC genotypes were protective factors for triglyceride (p=0.0002 and p=0.0005, respectively) and HDL cholesterol (p=0.0002 and p=0.0004, respectively) levels in T2DM patients. *ACE* DD was identified more frequently in hypertensive T2DM patients (O.R.=3.77, p=0.0005) and in those who reported drinking alcohol (p=0.0001) comparing to HC and T2DM patients who did not drink alcohol, respectively. We observed that T2DM patients who reported drinking alcohol had an increased frequency of *ACE* DD and *eNOS* bb (p<0.0001), or *ACE* DD and *OXTR* G (p<0.0001) compared to non-drinkers. No gene polymorphisms were associated with MetS.

Keywords: *ACE* ID, *eNOS* VNTR 4a/b, metabolic syndrome, *OXTR* (A>G), *SOD1* +35A/C, type 2 diabetes mellitus

INTRODUCTION

It has been estimated that the worldwide prevalence of diabetes mellitus and metabolic syndrome (MetS) are 10.5% and 12.5 - 31.4%, respectively, and the values are predicted to increase during the following years (1,2).

Type 2 diabetes mellitus (T2DM) is a complex metabolic disorder mainly characterized by insulin resistance and β -cell dysfunction (3). Besides insulin resistance, MetS is described by a cluster of conditions, namely high blood pressure, abdominal obesity, high triglyceride levels, low HDL cholesterol level, and impaired fasting glucose. Several risk factors have been identified for T2DM and MetS, such as high adiposity, abnormal blood biomarkers levels, medical history, regional and psychosocial factors. In addition, lifestyle factors such as daily caloric intake, smoking and alcohol consumption are considered to be related to the prevalence of T2DM and MetS (4,5).

Dysfunction of many biological pathways may be involved in the pathophysiology of the diseases. Mutations in protein-coding genes involved in oxidative stress (OS) reduction (*SOD1* and *CAT*) (6,7), endothelial functions (*eNOS*) (8) and hemodynamics (*ACE*, *ATR1*) (9–12) or in the carbohydrate metabolism (*OXTR*) (13,14) may to be involved in predisposition for T2DM or MetS. There are gender-related differences regarding the relative contribution of risk factors for these diseases (15,16). Therefore, we evaluated the susceptibility in men of the association of common polymorphisms in six genes with T2DM and MetS.

MATERIAL AND METHODS

Clinical data

The case-control study included Caucasian men considered healthy (n=120) or diagnosed with T2DM (n=120)or MetS (n=75). The American Diabetes Association 2016 and NCEP ATP III criteria were used for the diagnosis of these diseases (17). Healthy individuals were selected based on a standard clinical evaluation and on paraclinical data. Patients with a diagnosis of chronic kidney diseases, retinopathy, diabetic peripheral neuropathy or with an addiction to drugs were not included in the study. We collected clinical, paraclinical, and lifestyle data from all patients. Subjects were considered smokers if they smoked between 2 and 25 cigarettes per day for at least a year. Alcohol consumers were considered those who drunk at most 50 g alcohol per day for at least a year, but were not heavy drinkers (56g alcohol/day). The research was approved by the ethics committee of the National Research and Development Institute for Food Bioresources (966/27.08.2019).

Research methods

Genomic DNA was extracted from peripheral venous blood using the Promega Wizard Genomic DNA purification kit (Promega Corporation, Madison, WI), followed by a Polymerase Chain Reaction (PCR) to genotype the rs4646994 (*ACE I/D*) (18), rs1799983 (*eNOS* VNTR 4a/b) (18), and rs53576 (*OXTR* A>G) (19) polymorphisms. A PCR-restriction fragment length polymorphism was used to genotype the *ATR1* rs5186 (A1166C) (18), *CAT* rs7943316 (-21A/T) (20), and *SOD1* rs2234694 (+35A/C) polymorphismes (20).

Statistical analysis

Statistical analysis was performed with the MedCalc software (version 20.111, Ostend, Belgium). After Bonfer-

age compared to MetS (p=0.0003). The statistical signifitribucance of difference was greater when comparing patients

RESULTS

cance of difference was greater when comparing patients who are carriers of *OXTR* G (54.04 vs 56.05 years old, p=0.0002) or both *OXTR* G and *eNOS* b alleles (54.00 vs 56.05, p=0.00016). Our results showed that the *SOD1* AA genotype

roni correction for multiple hypotheses (n=33) a p value at

The clinical data of subjects enrolled in the study are

We found that T2DM was diagnosed at an earlier

shown in Table 1. MetS patients have 3 (64%), 4 (32%) or

p<0.0015 was considered statistically significant.

5 (4%) NCEP ATP III criteria for diagnosis.

(p=0.0006) and the presence of both ACE I and OXTR A alleles (p=0.0005) are protective factors for T2DM. T2DM patients with lower triglyceride levels (<150 mg/dl) were more frequent carriers of SOD1 AA and AC genotypes when compared to HC subjects (p=0.0002 and p=0.0005, respectively). Similarly, SOD1 AA and AC genotypes were more frequent in T2DM patients with HDL levels over 40 mg/dl when comparing to HC (p=0.0002 and p=0.0004, respectively). Hypertensive T2DM patients were more frequent carriers of ACE DD genotype than HC (56.76%) vs 25.83%, p=0.0005) (Table 3). In addition, this genotype, in association with eNOS bb or OXTR G, was found more frequently in alcohol consumers compared to those without this habit (p<0.0001) (Table 3). No other statistically significant associations were found between the investigated groups or subgroups of subjects.

DISCUSSION

In this study, we evaluated the association between six common polymorphisms in the ACE, eNOS, OXTR, ATR1, CAT, and SOD1 genes with characteristics of T2DM and MetS in Romanian Caucasian men. Thus, we tried to avoid the impact of gender on these associations. Patients with acute or chronic hyperglycemia have increased OS levels can be predisposed to long term complications of diabetes (21). SOD1 gene codes for an antioxidant enzyme and therefore, it is a functional candidate for obesity (22), T2DM, and its long-term complications (23). In our study, SOD1 AA was a protective factor for T2DM (p<0.0006). Concordant results were reported in the study by Flekac et al., where both Czech males and females were included. The authors reported that SOD1 +35A/C had potential effect on enzyme activity and genotype AA was protective for T2DM (p<0.05) (24). Additionally, in our study SOD1 AA and AC genotypes were associated with triglycerides (p=0.0002 and p=0.0005, respectively) and HDL choZenoaga-Barbăroșie C, Berca L, Vassu-Dimov T, Toma M, Nica MI, Alexiu-Toma OA, Ciornei C, Albu A, Nica S, Nistor C, Nica R

Parameters	T2DM	MetS	НС
Number of subjects	120	75	120
Age ^d	54.00 (51–56) ^a	55.00 (54–58)	NA
Age ⁱ	58 (54.5–59) ^{a, b}	59 (57–61) ^b	55 (53–59)
BMI ^d	32.99 (29.41–35.22)	33.66 (29.74–35.29)	NA
BMI ⁱ	30.95 (27.97-33.07) ^{a,b}	32.83 (28.8–33.74) ^b	24.55 (23.71–27.74)
Glycemia ⁱ (mg/dl)	113.5 (102.5–123) ^{a,b}	109 (98–116)	98 (93–104.5)
Obesity ^d	68	54	0
Hypertension	37	50	0
Hyperglycemia ^d (≥110 mg/dl)	120	33	0
HDL cholesterol ^d (<40 mg/dl)	39	55	12
Triglycerides ^d (≥150 mg/dl)	48	63	8
Stroke ⁱ	10	9	0
Coronary heart diseases ⁱ	9	6	0
Sexual dysfunctions ⁱ (yes/ no/ refused to respond)	17/80/23	10/47/18	12/91/17
Subjects with offspring	45	25	100
Smokers ⁱ	19	42	57
Alcohol consumers ⁱ	43	28	27

Values are presented as number of subjects (n), median (range), or ratio as specified; ⁱ at study inclusion; ^d at disease diagnosis; a p<0.0015 compared to MetS group, b p<0.0015 compared to HC group.

Sample genotyping was performed for all patients included in the study. The distribution of the studied genotypes was in accordance with Hardy-Weinberg Equilibrium (Table 2).

Polymorphisms Genotypes T2DM MetS HC ACE I/D (rs4646994) DD 44 25 31 ID 49 35 61 II 27 15 28 ATRI A1166C (rs5186) AA 70 43 65 ACC 38 31 44 CC 12 1 11 e/NOS VNTR 4a/b (rs1799983) bb 90 52 82 ba 26 23 36 aa 4 0 2 OXTR A>G (rs53576) GG 59 31 42 GA 47 34 49 AA 14 10 29 AA 27 13 33 SOD1 +35A/C (rs2234694) AA 27 13 36 CC 31 18 36 AA 104 71 118 CC 31 18 2 CC </th <th></th> <th>C 11</th> <th></th> <th>e</th> <th></th>		C 11		e	
ACE I/D (rs4646994) ID 49 35 61 II 27 15 28 ATRI A1166C (rs5186) AA 70 43 65 AC 38 31 44 CC 12 1 11 e/NOS VNTR 4a/b (rs1799983) bb 90 52 82 ba 26 23 36 aa 4 0 2 OXTR A>G (rs53576) GG 59 31 42 GA 47 34 49 AA 14 10 29 AA 14 10 29 AA 14 10 29 SOD1 +35A/C (rs2234694) AA 27 13 33 CC 31 18 36 AA 104 71 118 CAT-21A/T (rs7943316) AA 104 71 118	Polymorphisms	Genotypes	T2DM	MetS	НС
ID493561II271528ATRI A1166C (rs5186)AA704365AC383144CC12111 $eNOS VNTR 4a/b$ (rs1799983)bb905282bb905282ba262336aa402OXTR A>G (rs53576)GA473449AA141029AA271333SOD1 +35A/C (rs2234694)AA271333CAT-21A/T (rs7943316)AA10471118AT15422		DD	44	25	31
II 27 15 28 ATRI A1166C AA 70 43 65 AC 38 31 44 CC 12 1 11 eNOS VNTR 4a/b bb 90 52 82 ba 26 23 36 aa 4 0 2 OXTR A>G GG 59 31 42 OXTR A>G GA 47 34 49 AA 14 10 29 SOD1 +35A/C AC 62 44 51 CC 31 18 36 AA 104 71 118 AA 104 71 118 AA 104 71 118 AA 104 71 118		ID	49	35	61
ATR1 A1166C (rs5186) AC 38 31 44 CC 12 1 11 eNOS VNTR 4a/b (rs1799983) bb 90 52 82 ba 26 23 36 aa 4 0 2 OXTR A>G (rs53576) GG 59 31 42 GA 47 34 49 AA 14 10 29 SOD1 +35A/C (rs2234694) AA 27 13 33 CC 31 18 36 AA 104 71 118 CAT-21A/T (rs7943316) AT 15 4 2		II	27	15	28
AC383144CC12111 $eNOS VNTR 4a/b$ (rs1799983)bb905282ba262336aa402OXTR A>G (rs53576)GG593142GA473449AA141029AA141029SOD1 +35A/C (rs2234694)AA271333CAT-21A/T (rs7943316)AA10471118AT15422		AA	70	43	65
CC12111 $eNOS VNTR 4a/b$ (rs1799983)bb905282ba262336aa402OXTR A>G (rs53576)GG593142OXTR A>G (rs2234694)GA473449AA141029AA271333AC624451CC311836AA10471118CAT-21A/T 		AC	38	31	44
$\begin{array}{c c c c c c c c c c c c c c c c c c c $	(196100)	CC	12	1	11
ba262336aa402 $OXTR A>G$ (rs53576)GG593142GA473449AA141029AA271333AC624451CC311836AA10471118AA10471118AA10471154AA1542		bb	90	52	82
aa 4 0 2 OXTR A>G GG 59 31 42 GA 47 34 49 AA 14 10 29 AA 14 10 29 SOD1 +35A/C (rs2234694) AA 27 13 33 CC 62 44 51 51 CC 31 18 36 AA 104 71 118 CAT-21A/T (rs7943316) AT 15 4 2		ba	26	23	36
OXTR A>G (rs53576) GA 47 34 49 AA 14 10 29 AA 14 10 29 SOD1 +35A/C (rs2234694) AA 27 13 33 AC 62 44 51 CC 31 18 36 AA 104 71 118 CAT-21A/T (rs7943316) AT 15 4 2	(1317)))00)	aa	4	0	2
(rs53576) GA 47 34 49 AA 14 10 29 AA 14 10 29 SOD1 +35A/C (rs2234694) AA 27 13 33 AC 62 44 51 CC 31 18 36 AA 104 71 118 CAT-21A/T (rs7943316) AT 15 4 2		GG	59	31	42
AA 14 10 29 SOD1 +35A/C (rs2234694) AA 27 13 33 AC 62 44 51 CC 31 18 36 AA 104 71 118 CAT-21A/T (rs7943316) AT 15 4 2	••••••	GA	47	34	49
SOD1 +35A/C (rs2234694) AC 62 44 51 CC 31 18 36 CAT-21A/T (rs7943316) AA 104 71 118	(1500010)	AA	14	10	29
AC 62 44 51 CC 31 18 36 AA 104 71 118 CAT-21A/T (rs7943316) AT 15 4 2		AA	27	13	33
CC 31 18 36 CAT-21A/T (rs7943316) AA 104 71 118		AC	62	44	51
CAT-21A/T (rs7943316) AT 15 4 2		CC	31	18	36
(rs7943316) AI 15 4 2		AA	104	71	118
		AT	15	4	2
		TT	1	0	0

lesterol (p=0.0002 and p=0.0004, respectively) levels in T2DM patients. Our results are supported by previous publications in which *SOD1* concentration was negatively correlated with HDL cholesterol concentration. However, unlike previously published data, we found no statistically significant association between *SOD1* polymorphism and obesity (p<0.05) (25).

Ethnicity and gender may explain, at last partially, previous conflicting results regarding the impact of ACE I/D in predisposition for T2DM (26). The ACE DD genotype was found to increased risk of hypertension and/ or diabetes in Egyptian (27), Malaysian (28), Chinese (29) populations, but not in Turkish (30) or Emirati (31). Meta-analyses have also described a positive association with subjects from Middle East, North Africa (26) or Asia (32), whereas, among Europeans, the results are more heterogenous (33,34). The number of ACE D variants was correlated with an increase in ACE activity (35) and angiotensin II signal transduction influences secretion of oxytocin (36,37). Long-term ACE hyperactivity may predispose to insulin hypersecretion and impairment of vessel walls compliance which increase the risk for T2DM and hypertension development (38). Although the distribution

Groups compared	Genetic variants	Distribution	O.R., 95% CI, p value
T2DM vs HC	SOD1 AA	104/16 vs 118/2	0.11, 0.03 - 0.49, 0.0006
T2DM vs HC	ACE I and OXTR A	32/88 vs 58/62	0.39, 0.23 – 0.67, 0.0005
T2DM with triglycerides level <150 mg/dl vs HC	SOD1 AA	60/12 vs 110/2	0.09, 0.02 - 0.42, 0.0002
	SOD1 AC	11/61 vs 2/110	9.92, 2.13 – 46.21, 0.0005
T2DM with HDL level > 40 mg/dl vs HC	SOD1 AA	67/14 vs 106/2	0.09, 0.02 - 0.41, 0.0002
	SOD1 AC	13/68 vs 2/106	10.13, 2.22 – 46.31, 0.0004
T2DM with hypertension vs HC	ACE DD	21/16 vs 31/89	3.77, 1.75 – 8.12, 0.0005
T2DM alcohol drinkers vs T2DM non-drinkers	ACE DD	30/13 vs 14/63	10.38, 4.35 – 24.82, <0.0001
	ACE DD and eNOS bb	25/18 vs 10/67	9.31, 3.79 – 22.87, <0.0001
	ACE DD and OXTR1 G	26/17 vs 12/65	8.28, 3.48 – 19.73, <0.0001

Table 3. Statistically significant results in the studied groups

of polymorphisms in *ACE* or *OXTR* did not differ between our groups, the presence of both *ACE* I and *OXTR* A alleles could be a protective factor for T2DM (OR=0.39, p<0.0005).

OXTR mediates the impact of stressful experience and influences social support seeking during distress (39). *OXTR* polymorphisms may influence the response to stress, via hypothalamic–pituitary–adrenal axis, and the risk for stress-related disorders, including T2DM (13,39,40). Carriers of the rs53576 G allele were more sensitive to both favorable or negative surroundings and individuals with GG genotype had altered cortisol levels and blood pressure after rejection (39).

Predisposition to T2DM involves the interaction between different genetic and non-genetic factors. It was considered that moderate alcohol consumption (24 g/day) is protective for T2DM development, while higher quantities (60 g/day alcohol) represents a risk factor (41). In our T2DM group, patients carrying the *ACE* DD genotype were more often alcohol consumers. The association of *ACE* DD with *eNOS* bb or with *OXTR* G were found more frequently in drinking compared to non-drinking T2DM patients. We found no association regarding the *OXTR* gene polymorphisms and smoking habits in T2DM patients.

No association was identified concerning gene polymorphisms in the MetS patients who reported being smokers or drinking alcohol.

Our study indicated no significant association between MetS and tested polymorphisms or between T2DM and *ATR1* A1166C or *CAT*-21A/T.

ACKNOWLEDGEMENTS

Declaration of Interest: The authors report no conflicts of interest. The authors alone are responsible for the content and writing of this article. Ethics approval: All procedures performed in this study involving human participants were in accordance with the ethical standards of the institutional research committee and with the 1964 Helsinki Declaration and its later amendments of comparable ethical standards.

Funding: This study was supported by the Ministry of Research and Education, grant number PN 23 01 03 03.

REFERENCES

- Sun H, Saeedi P, Karuranga S, Pinkepank M, Ogurtsova K, Duncan BB, et al. IDF Diabetes Atlas: Global, regional and country-level diabetes prevalence estimates for 2021 and projections for 2045. Diabetes Res Clin Pract. 2022;183:109119.
- Noubiap JJ, Nansseu JR, Lontchi-Yimagou E, Nkeck JR, Nyaga UF, Ngouo AT, et al. Geographic distribution of metabolic syndrome and its components in the general adult population: A meta-analysis of global data from 28 million individuals. Diabetes Res Clin Pract. 2022;188:109924.
- Kahn SE, Cooper ME, Del Prato S. Pathophysiology and treatment of type 2 diabetes: perspectives on the past, present, and future. Lancet. 2014;383(9922):1068–83.
- Bellou V, Belbasis L, Tzoulaki I, Evangelou E. Risk factors for type 2 diabetes mellitus: An exposurewide umbrella review of meta-analyses. PLoS One. 2018;13(3):e0194127.
- Damiri B, Badran L, Safadi D, Sawalha A, Yasin Y, Sawalha M, et al. Metabolic syndrome and related risk factors among adults in the northern West Bank, a cross-sectional study. Int Health. 2022;14(4):339–45.
- 6. Tavares AM, Silva JH, Bensusan C de O, Ferreira ACF, Matos LP de L, e Souza KL de A, et al.

Zenoaga-Barbăroșie C, Berca L, Vassu-Dimov T, Toma M, Nica MI, Alexiu-Toma OA, Ciornei C, Albu A, Nica S, Nistor C, Nica R

Altered superoxide dismutase-1 activity and intercellular adhesion molecule 1 (ICAM-1) levels in patients with type 2 diabetes mellitus. PLoS One. 2019;14(5):e0216256.

- Hebert-Schuster M, Fabre EE, Nivet-Antoine V. Catalase polymorphisms and metabolic diseases. Curr Opin Clin Nutr Metab Care. 2012;15(4):397–402.
- Raina P, Sikka R, Gupta H, Matharoo K, Bali SK, Singh V, et al. Association of eNOS and MCP-1 Genetic Variants with Type 2 Diabetes and Diabetic Nephropathy Susceptibility: A Case-Control and Meta-Analysis Study. Biochem Genet. 2021;59(4):966–96.
- Stephens JW, Dhamrait SS, Cooper JA, Acharya J, Miller GJ, Hurel SJ, et al. The D allele of the ACE I/D common gene variant is associated with Type 2 diabetes mellitus in Caucasian subjects. Mol Genet Metab. 2005;84(1):83–9.
- Xi B, Ruiter R, Chen J, Pan H, Wang Y, Mi J. The ACE insertion/deletion polymorphism and its association with metabolic syndrome. Metabolism. 2012;61(6):891–7.
- Alavi-Shahri J, Behravan J, Hassany M, Tatari F, Kasaian J, Ganjali R, et al. Association between angiotensin II type 1 receptor gene polymorphism and metabolic syndrome in a young female Iranian population. Arch Med Res. 2010;41(5):343–9.
- Hou L, Quan X, Li X, Su X. Correlation between gene polymorphism in angiotensin II type 1 receptor and type 2 diabetes mellitus complicated by hypertension in a population of Inner Mongolia. BMC Med Genet. 2020;21(1):83.
- Saravani R, Esmaeeli E, Kordi Tamendani M, Nejad MN. Oxytocin Receptor Gene Polymorphisms in Patients With Diabetes. Gene, Cell Tissue. 2015;2(2):e60171.
- Winterton A, Bettella F, de Lange A-MG, Haram M, Steen NE, Westlye LT, et al. Oxytocin-pathway polygenic scores for severe mental disorders and metabolic phenotypes in the UK Biobank. Transl Psychiatry. 2021;11(1):599.
- Kautzky-Willer A, Harreiter J, Pacini G. Sex and Gender Differences in Risk, Pathophysiology and Complications of Type 2 Diabetes Mellitus. Endocr Rev. 2016;37(3):278–316.
- Regitz-Zagrosek V, Lehmkuhl E, Weickert MO. Gender differences in the metabolic syndrome and their role for cardiovascular disease. Clin Res Cardiol. 2006;95(3):136–47.
- 17. Chamberlain JJ, Rhinehart AS, Shaefer CFJ, Neuman A. Diagnosis and Management of Diabetes: Synopsis

of the 2016 American Diabetes Association Standards of Medical Care in Diabetes. Ann Intern Med. 2016;164(8):542–52.

- Akcay A, Sezer S, Ozdemir FN, Arat Z, Atac FB, Verdi H, et al. Association of the genetic polymorphisms of the renin-angiotensin system and endothelial nitric oxide synthase with chronic renal transplant dysfunction. Transplantation. 2004;78(6):892–8.
- Truzzi A, Poquérusse J, Setoh P, Shinohara K, Bornstein MH, Esposito G. Oxytocin receptor gene polymorphisms (rs53576) and early paternal care sensitize males to distressing female vocalizations. Dev Psychobiol. 2018;60(3):333–9.
- Saygi S, Erol İ, Alehan F, Yalçın YY, Kubat G, Ataç FB. Superoxide Dismutase and Catalase Genotypes in Pediatric Migraine Patients. J Child Neurol. 2015;30(12):1586–90.
- Volpe CMO, Villar-Delfino PH, Dos Anjos PMF, Nogueira-Machado JA. Cellular death, reactive oxygen species (ROS) and diabetic complications. Cell Death Dis. 2018;9(2):119.
- Lewandowski Ł, Kepinska M, Milnerowicz H. Alterations in Concentration/Activity of Superoxide Dismutases in Context of Obesity and Selected Single Nucleotide Polymorphisms in Genes: SOD1, SOD2, SOD3. Int J Mol Sci. 2020;21(14):5069
- Panduru NM, Cimponeriu D, Cruce M, Ion DA, Moţa E, Moţa M, et al. Association of +35A/C (intron3/ exon3) polymorphism in SOD1-gene with diabetic nephropathy in type 1 diabetes. Rom J Morphol Embryol. 2010;51(1):37–41.
- 24. Flekac M, Skrha J, Hilgertova J, Lacinova Z, Jarolimkova M. Gene polymorphisms of superoxide dismutases and catalase in diabetes mellitus. BMC Med Genet. 2008;9:30.
- Lewandowski Ł, Urbanowicz I, Kepinska M, Milnerowicz H. Concentration/activity of superoxide dismutase isozymes and the pro-/antioxidative status, in context of type 2 diabetes and selected single nucleotide polymorphisms (genes: INS, SOD1, SOD2, SOD3) - Preliminary findings. Biomed Pharmacother. 2021;137:111396.
- 26. El Alami H, Ghazal H, Abidi O, Al Idrissi N, Wakrim L, Naamane A, et al. Relationship between insertion/ deletion (I/D) polymorphism of angiotensin converting enzyme (ACE) gene and susceptibility to type 2 diabetes mellitus in the Middle East and North Africa Region: A meta-analysis. Diabetes Metab Syndr. 2022;16(1):102386.
- 27. Zarouk WA, Hussein IR, Esmaeil NN, Raslan HM, Reheim HAA, Moguib O, et al. Association of angio-

tensin converting enzyme gene (I/D) polymorphism with hypertension and type 2 diabetes. Bratisl Lek Listy. 2012;113(1):14–8.

- Ramachandran V, Ismail P, Stanslas J, Shamsudin N, Moin S, Mohd Jas R. Association of insertion/deletion polymorphism of angiotensin-converting enzyme gene with essential hypertension and type 2 diabetes mellitus in Malaysian subjects. J Renin Angiotensin Aldosterone Syst. 2008;9(4):208–14.
- Zhou L, Xue Y, Luo R, Gao F. [Association of insertion/deletion polymorphism in angiotensinconverting enzyme gene with hypertensive type 2 diabetes mellitus]. Di Yi Jun Yi Da Xue Xue Bao. 2002;22(9):808–10.
- Araz M, Aynacioglu S, Aktaran S, Alasehirli B, Okan V. Association between polymorphism of the angiotensin I converting enzyme gene and hypertension in Turkish type II diabetic patients. Acta medica(Hradec Kralove). 2001;44(1):29-32.
- 31. Alsafar H, Hassoun A, Almazrouei S, Kamal W, Almaini M, Odama U, et al. Association of Angiotensin Converting Enzyme Insertion-Deletion Polymorphism with Hypertension in Emiratis with Type 2 Diabetes Mellitus and Its Interaction with Obesity Status. Dis Markers. 2015;2015:536041.
- 32. Ahmad N, Jamal R, Shah SA, Gafor AHA, Murad NAA. Renin-Angiotensin-Aldosterone System Gene Polymorphisms and Type 2 Diabetic Nephropathy in Asian Populations: An Updated Meta-analysis. Curr Diabetes Rev. 2019;15(4):263–76.
- 33. Al-Rubeaan K, Siddiqui K, Saeb ATM, Nazir N, Al-Naqeb D, Al-Qasim S. ACE I/D and MTHFR C677T polymorphisms are significantly associated with type 2 diabetes in Arab ethnicity: A meta-analysis. Gene. 2013;520(2):166–77.

- 34. Niu W, Qi Y, Gao P, Zhu D. Angiotensin converting enzyme D allele is associated with an increased risk of type 2 diabetes: evidence from a meta-analysis. Endocr J. 2010;57(5):431–8.
- 35. Jalil JE, Córdova S, Ocaranza M a, Schumacher E, Braun S, Chamorro G, et al. Angiotensin I-converting enzyme insertion/deletion polymorphism and adrenergic response to exercise in hypertensive patients. Med Sci Monit. 2002;8(8):CR566-71.
- Srinivasa S, Aulinas A, O'Malley T, Maehler P, Adler GK, Grinspoon SK, et al. Oxytocin response to controlled dietary sodium and angiotensin II among healthy individuals. Am J Physiol Endocrinol Metab. 2018;315(4):E671–5.
- 37. Bealer SL, Crowley WR. Angiotensin II-induced release of oxytocin: interaction with norepinephrine and role in lactation. Regul Pept. 2003;111(1):41–6.
- Dietze GJ, Henriksen EJ. Angiotensin-converting enzyme in skeletal muscle: sentinel of blood pressure control and glucose homeostasis. J Renin Angiotensin Aldosterone Syst. 2008;9(2):75–88.
- McQuaid RJ, McInnis OA, Matheson K, Anisman H. Distress of ostracism: oxytocin receptor gene polymorphism confers sensitivity to social exclusion. Soc Cogn Affect Neurosci. 2015;10(8):1153–9.
- 40. Chang HH, Chang WH, Chi MH, Peng YC, Huang C-C, Yang YK, et al. The OXTR Polymorphism Stratified the Correlation of Oxytocin and Glucose Homeostasis in Non-Diabetic Subjects. Diabetes Metab Syndr Obes. 2019;12:2707–13.
- Baliunas DO, Taylor BJ, Irving H, Roerecke M, Patra J, Mohapatra S, et al. Alcohol as a risk factor for type 2 diabetes: A systematic review and meta-analysis. Diabetes Care. 2009;32(11):2123–32.