CLINICAL AND TRANSLATIONAL MEDICINE



REVIEW

Emerging roles of the solute carrier family in pancreatic cancer

Zijian Wu ^{1,2,3,4}		Jin Xu ^{1,2,3,4}	Cł	nen Liang ^{1,2,3,4}	Qingcai Meng ^{1,2,3,4}	Jie Hua ^{1,2,3,4}
Wei Wang ^{1,2,3,4}	I	Bo Zhang ^{1,2,3,4}		Jiang Liu ^{1,2,3,4}	Xianjun Yu ^{1,2,3,4} 🝺	Si Shi ^{1,2,3,4} 🝺

¹ Department of Pancreatic Surgery, Fudan University Shanghai Cancer Center, Shanghai, China

² Department of Oncology, Shanghai Medical College, Fudan University, Shanghai, China

³ Shanghai Pancreatic Cancer Institute, Shanghai, China

⁴ Pancreatic Cancer Institute, Fudan University, Shanghai, China

Correspondence

Si Shi, Department of Pancreatic Surgery, Fudan University Shanghai Cancer Center, No. 270 Dong'An Road, Shanghai 200032, China.

Email: shisi@fudanpci.org

Xianjun Yu, Shanghai Pancreatic Cancer Institute, No. 270 Dong'An Road, Shanghai 200032, China.

Email: yuxianjun@fudanpci.org



Pancreatic cancer is a rare but highly fatal disease with poor prognosis. Some solute carriers (SLCs) have been identified to participate in several steps of pancreatic tumorigenesis, which would be a hot spot of future research and serve as new therapeutic strategies.

REVIEW



Emerging roles of the solute carrier family in pancreatic cancer

Zijian Wu^{1,2,3,4}Jin Xu^{1,2,3,4}Chen Liang^{1,2,3,4}Qingcai Meng^{1,2,3,4}Jie Hua^{1,2,3,4}Wei Wang^{1,2,3,4}Bo Zhang^{1,2,3,4}Jiang Liu^{1,2,3,4}Xianjun Yu^{1,2,3,4}Si Shi^{1,2,3,4}

¹ Department of Pancreatic Surgery, Fudan University Shanghai Cancer Center, Shanghai, China

² Department of Oncology, Shanghai Medical College, Fudan University, Shanghai, China

³ Shanghai Pancreatic Cancer Institute, Shanghai, China

⁴ Pancreatic Cancer Institute, Fudan University, Shanghai, China

Correspondence

Si Shi, Department of Pancreatic Surgery, Fudan University Shanghai Cancer Center, No. 270 Dong'An Road, Shanghai 200032, China.

Email: shisi@fudanpci.org Xianjun Yu, Shanghai Pancreatic Cancer Institute, No. 270 Dong'An Road, Shanghai 200032, China. Email: yuxianjun@fudanpci.org

Zijian Wu, Jin Xu, and Chen Liang contributed equally to this work.

Abstract

Pancreatic cancer is a gastrointestinal tumor with a high mortality rate, and advances in surgical procedures have only resulted in limited improvements in the prognosis of patients. Solute carriers (SLCs), which rank second among membrane transport proteins in terms of abundance, regulate cellular functions, including tumor biology. An increasing number of studies focusing on the role of SLCs in tumor biology have indicated their relationship with pancreatic cancer. The mechanism of SLC transporters in tumorigenesis has been explored to identify more effective therapies and improve survival outcomes. These transporters are significant biomarkers for pancreatic cancer, the functions of which include mainly proliferative signaling, cell death, angiogenesis, tumor invasion and metastasis, energy metabolism, chemotherapy sensitivity and other functions in tumor biology. In this review, we summarize the different roles of SLCs and explain their potential applications in pancreatic cancer treatment.

KEYWORDS

pancreatic cancer, solute carrier, tumor biology

Abbreviations: 5-FU, 5-fluorouracil; AIF, apoptosis-inducing factor; ASCT2, alanine–serine–cysteine transporter 2; CREB, cAMP response element-binding protein; EMT, epithelial–mesenchymal transition; GLUT1, glucose transporter 1; HGNC, HUGO Gene Nomenclature Committee; LAT1, L-type amino acid transporter 1; MgtE, magnesium (Mg^{2+}) transporter E; MMPs, matrix metalloproteases; mTOR, mammalian target of rapamycin; NBC3, bicarbonate (Na^+/HCO_3^-) cotransporter 3; NRP-1, neuropilin-1; PCD, programmed cell death; PDTC, pyrrolidine dithiocarbamate; Pfam, protein family; PTGR2, prostaglandin reductase 2; ROS, reactive oxygen species; RREB1, Ras responsive element-binding protein 1; SGLT1, sodium-dependent glucose transporter 1; SLC, solute carrier; STAT3, signal transducer and activator of transcription 3; TPEN, *N*,*N*,*N*',*N*'-tetrakis (2-pyridylmethyl) ethylenediamine; VEGF, vascular endothelial growth factor; VNUT, vesicular nucleotide transporter; ZIP, Zrt-IRF-like proteins; ZnT, zinc transporter protein

This is an open access article under the terms of the Creative Commons Attribution License, which permits use, distribution and reproduction in any medium, provided the original work is properly cited.

© 2021 The Authors. Clinical and Translational Medicine published by John Wiley & Sons Australia, Ltd on behalf of Shanghai Institute of Clinical Bioinformatics

1 | BACKGROUND

Although pancreatic cancer has a low incidence of $\sim 2.5\%$, it is a highly fatal malignancy with a 5-year overall survival rate of ~8%, making it the seventh leading cause of cancerrelated death worldwide; it accounts for 458,918 (2.5%) new cases and 432,242 (4.5%) new deaths annually according to GLOBOCAN 2018.¹⁻³ However, because of the location of the pancreas, deep in the abdomen, and the limited number of typical symptoms and lack of early and accurate detection methods, many patients are diagnosed with advanced or metastatic stages; consequently, only 15-20% of patients with pancreatic cancer are eligible for surgical resection at diagnosis.⁴ In recent decades, some progress has been achieved in the diagnosis, perioperative management, and radiotherapy and systemic therapy approaches for advanced diseases, but these advances have had limited effects on overall survival outcomes.⁵ Therefore, the molecular mechanism of pancreatic cancer tumorigenesis is a key point in pancreatic oncology research. Studies on solute carriers (SLCs) in cancer pharmacology and drug discovery have become a hotspot in recent years, particularly due to their roles as chemotherapeutic drug targets or mediators of drug disposition. Since studies by Li et al. revealed the significant role of SLC39A4 in pancreatic cancer tumor progression, our interest in research on SLCs in pancreatic cancer has grown.6

2 | INTRODUCTION TO SLC AND THEIR RELATIONSHIP WITH CANCER

The SLC superfamily encodes the second largest group of membrane transport proteins after G protein-coupled receptors, comprising 65 families with over 400 SLC transporters reported in humans to date.^{7–9} Most of these proteins are located in the membranes of cells and organelles, and their main functions are to maintain cellular homeostasis by facilitating the exchange of a wide variety of soluble molecular substrates across lipid membranes. SLC transporters rely on electrochemical gradients or ion gradients to transport a diverse array of substrates, including glucose, amino acids, vitamins, nucleotides, inorganic ions, organic ions, neurotransmitters, and drugs.^{8,10,11} The nomenclature for SLCs is based on the combination and overlap of two databases: the HUGO Gene Nomenclature Committee (HGNC) (http://www.genenames.org/) and the SLC tables (http://www.bioparadigms.org/slc/intro. htm). The HGNC provides each human gene with a certain approved symbol that also curates it into large families based on its substrates, structure, mechanism, and other characteristics.^{7,12} SLCs usually have the gene symbol name that represents the superfamily and the original pro-

tein name, for example, SLC39 and Zrt-IRF-like proteins (ZIPs). SLCs with over 20% protein sequence identity are classified into the same family.¹³ Concerning the species, two multiorganism classifications have been developed: the transporter classification (TC) system (http://www. tcdb.org/) based on functional and phylogenetic information of transporter proteins and the protein family (Pfam) system (http://pfam.xfam.org/) based on protein sequence alignment using Markov models. SLCs are divided by phylogenetic analyses into four groups denoted as α , β , δ , γ and simultaneously clustered into five Pfam clans by sequence similarity, namely, major facilitator superfamily (MFS), amino acid/polyamine/organocation (APS), cation proton antiporter/anion transporter (CPA/AT), drug/metabolite transporter (DMT), and others without clans.¹⁴ Additionally, the transporters have also been divided into two types according to the intracellular and extracellular concentrations of their substrates: passive (facilitated) transporters mediate the passage of substrates down their electrochemical gradients without using energy and active transporters (symporters and antiporters) create gradients by utilizing energy. Members of the SLC superfamily play crucial roles in human physiology through process such as the intake of amino acids in the intestine for cell growth, absorption of glucose for cellular energy metabolism, transport of metal ions to serve as essential cofactors for enzymes, and uptake of water-soluble vitamins for vital processes or reabsorption of neurotransmitters into presynaptic neurons.⁸ Given the significant roles of SLCs in regulating normal cellular functions, dysregulation of certain SLC proteins is associated with various diseases, including diabetes mellitus, hypertension, chronic kidney disease, dermatosis, cancer, and many others.13

According to the hallmarks of cancer proposed by Hanahan and Weinberg, tumorigenesis is a multistep process that gradually transforms normal cells into highly malignant derivatives.^{15,16} Previous studies have reported that the SLC superfamily participates in various steps of tumorigenesis, including proliferation, apoptosis, invasion and metastasis, chemotherapy resistance, and other processes related to cancer.¹⁰ The overexpression or suppression of SLCs may provide new strategies for determining the diagnosis, treatment, or prognosis. Some tumors must transport nutrients into cells to fuel their growth and survival, and thus the inhibition or activation of SLC transporter function may be a possible therapeutic strategy. In addition, some SLC transporters have the ability to deliver certain drugs to cancer cells, suggesting that they can serve as new targets for improving chemotherapy sensitivity and overcoming drug resistance. The next few sections aim to summarize examples of SLCs that have been reported to play a role in one or more hallmarks of pancreatic tumor biology and describe their functions (Figure 1 and Table 1).



FIGURE 1 Functions of solute carriers (SLCs) in pancreatic tumor biology

TABLE 1 List of solute carriers (SLCs) playing different roles in pancreatic cancer

Gene		Protein		-	Promote/		Other	D.C
name	Aliases	name(s)	Substrates	Transport type	suppress	Mechanism	phenotypes	Reference
	tive signaling			·				17.10
SLC1A5	System ASC	ASCT2	L-Ala, L-Ser, L-Cys, L-Thr, L-Gln, L-Asn	Na ⁺ /amino acid cotrans- porter/exchan		HIF-2α, mTOR	5	17,19
SLC4A7	NBC2, NBC3, SLC4A6	NBCn1	Na ⁺ , HCO ₃ ⁻	Na ⁺ /HCO ₃ ⁻ cotransporter	Promote	Ras, TGFβ/EMT	NA	20
SLC7A5	System L	LAT1	Large neutral L-amino acids, T3, T4, L-DOPA, BCH	Amino acid exchanger	Promote	mTOR, c-Myc	5	21 23-25
SLC7A11	System X _c ⁻	хСТ	Cys (anionic form), L-glutamate	Amino acid exchanger	Promote	PI3K/Akt, STAT	2, 4, 6	27
SLC39A4	NA	ZIP4	Zn^{2+}	NA	Promote	IL-6/STAT3	2, 3, 4	6,31–34
SLC39A6	NA	ZIP6/LIV-1	Zn^{2+}	NA	Promote	STAT3/EMT	NA	35
SLC5A8	AIT	SMCT1	Short chain fatty acids	Na ⁺ /Cl ⁻ cotransporter	Suppress	NA	NA	41,42
SLC39A3	NA	ZIP3	Zn^{2+}	NA	Suppress	REEB1	NA	43-46
SLC41A1	NA	MgtE	Mg^{2+}	Channel-like	Suppress	Akt/mTOR	NA	50
2. Cell dea	th							
SLC6A14	ATB ^{0,+}	NA	Neutral, cationic amino acids	Na ⁺ /Cl⁻- dependent	Suppress	mTOR	1	61
SLC7A11	System X _c ⁻	xCT	Cystine, L-glutamate	Amino acid exchanger	Suppress	ROS	1, 4, 6	53,54
SLC31A1	CTR1	hCtr1, Ctr1, COPT1	Cu ²⁺ , cisplatin	K ⁺ -dependent	Suppress	ROS	NA	60
SLC39A4	NA	ZIP4	Zn ²⁺	NA	Controversy	Caspase-3, 7, 8, 9	1, 3,4	56-58
3. Sustaine	ed angiogenesi	s						
SL4A4	NBC, NBC1	NBCe1	Na ⁺ , HCO ₃ ⁻	Na ⁺ /HCO ₃ ⁻ cotransporter	Suppress	NA	4	63
SLC39A4	NA	ZIP4	Zn^{2+}	NA	Promote	VEGF	1, 2, 4	32,64
4. Invasior	n and metastas	is						
SLC2A1	NA	GLUT1	Glucose, galactose, mannose, glucosamine	Facilitated transporter	Promote	AMPK/FOXO3A/ PUMA, MMP-2	5	69–71
SLC4A4	NBC, NBC1	NBCe1	Na ⁺ , HCO ₃ ⁻	Na ⁺ /HCO ₃ ⁻ cotransporter	Promote	NA	3	63
SLC7A11	System X _c ⁻	хСТ	Cys (anionic form), L-glutamate	Amino acid exchanger	Promote	PI3K/Akt	1, 2, 6	27
SLC9A1	APNH	NHE1	Na ⁺ , Li ⁺ , H ⁺ , NH ₄ ⁺	Na ⁺ /H ⁺ exchanger	Promote	EGFR	7	73–75
SLC39A4	NA	ZIP4	Zn^{2+}	NA	Promote	ZO-1/claudin-1	1, 2, 3	76
SLC39A6	NA	ZIP6/LIV-1	Zn^{2+}	NA	Promote	STAT3/EMT	1	35
5. Cellular metabolism								
SLC1A5	System ASC	ASCT2	L-Ala, L-Ser, L-Cys, L-Thr, L-Gln, L-Asn	Na ⁺ /amino acid cotrans- porter/exchan		HIF-2α	1, 6	17
			. ,	1, 5.1011411	0			(Continues)

(Continues)

TABLE 1 (Continued)

IADLE I	(Continued)								
Gene		Protein			Promote/		Other		
name	Aliases	name(s)	Substrates	Transport type	suppress	Mechanism	phenotypes	References	
SLC2A1	NA	GLUT1	Glucose, galactose, mannose, glucosamine	Facilitated transporter	Promote	hnRNPA2/B1/ PKM2	4	68,69,71,80-82	
SLC5A1	NA	SGLT1	Glucose, galactose	Na ⁺ /glucose cotransporter	Promote	AMPK/mTOR	NA	83,84	
SLC7A5	System L	LAT1	Large neutral L-amino acids, T3, T4, L-DOPA, BCH	Amino acid exchanger	Promote	c-Myc	1	25,85,86	
SLC17A9	NA	VNUT	Purine nucleotides	Cl⁻-dependent	Promote	NA	NA	92	
6. Chemotherapy sensitivity									
SLC1A5	System ASC	ASCT2	L-Ala, L-Ser, L-Cys, L-Thr, L-Gln, L-Asn	Na ⁺ /amino acid cotrans- porter/exchan		HIF-2α	1, 5	17	
SLC7A11	System X _c ⁻	хСТ	Cys (anionic form), L-glutamate	Amino acid exchanger	Promote	ROS	1, 2, 4	98,99	
SLC22A3	EMT (uptake-2 system)	OCT2	Organic cations	Facilitated transporter	Promote	NA	NA	101	
SLC22A7								128	
SLC28A1	NA	CNT1	Nucleosides	Na ⁺ /nucleoside cotransporter	Suppress	NA	NA	105	
SLC28A2	NA	CNT2	Nucleosides	Na ⁺ /nucleoside cotransporter	Suppress	NA	NA	105	
SLC29A1	NA	ENT1	Nucleosides	Facilitated transporter	Controversy	NA	NA	102,114	
SLC29A2	NA	ENT2	Nucleosides	Facilitated transporter	Suppress	NA	NA	115	
SLC29A3	NA	ENT3	Nucleosides	NA	Suppress	NA	NA	116	
SLC39A4	NA	ZIP4	Zn ²⁺	NA	Promote	STAT3/ZEB1/ JNK/ENT1	1, 2, 3, 4	117	

Abbreviations: Ala, alanine; Asn, asparagine; Cys, cystine; ENT1, equilibrative nucleoside transporter 1; Gln, glutamine; MgtE, magnesium (Mg2+) transporter E; NA, not available; Ser, serine; SGLT1, sodium-dependent glucose transporter 1; TGF β , transforming growth factor β ; Thr, threonine; VNUT, vesicular nucleotide transporter.

2.1 | Proliferation

The most fundamental characteristic of cancer cells is their unlimited proliferation ability. Normal cells carefully regulate the generation and release of growth-promoting signals, ensuring cellular homeostasis and maintaining the normal structure and function of the tissue. However, cancer cells control their own fate by deregulating these signals and disrupting normal checkpoints, thereby allowing the cells to proliferate continuously.¹⁶ Alternatively, cancer cells themselves produce growth factor ligands and bind to cognate receptors to promote proliferation in an autocrine manner. In addition, these cells also upregulate receptor expression, resulting in a hyperresponse to growth factor ligands.¹⁶

SLC1A5 encodes the alanine–serine–cysteine transporter 2 (ASCT2) protein and is a Na⁺-dependent neutral amino acid cotransporter that mainly functions to transport glutamine and other neutral amino acids into mitochondria.^{17,18} Kaira et al. evaluated 97 surgically resected samples from patients with pancreatic cancer and concluded that the expression of ASCT2 in the pancreatic cancer group was significantly higher than that in the benign pancreatic lesion group.¹⁹ A recent study also showed that oncogenic growth was dramatically suppressed by the knockdown of SLC1A5 and that overexpression of the strange of

sion of this gene restored cell growth, which indicated a critical role for SLC1A5 in pancreatic tumor growth.¹⁷

SLC4A7, also known as electroneutral Na^+/HCO_3^- cotransporter 3 (NBC3), regulates the transport of sodium and bicarbonate ions at a 1:1 ratio and plays a critical role in regulating the intracellular pH. In a recent study published in *Nature*, researchers found that knockout of SLC4A7 in Ras mutant pancreatic ductal adenocarcinoma (PDAC) cells induces micropinocytosis to support tumor growth.²⁰

SLC7A5, which encodes the light chain of the L-type amino acid transporter 1 (LAT1) protein, is a Na⁺independent amino acid transporter that regulates the influx of L-leucine and the efflux of L-glutamine when it interacts with SLC3A2.21,22 Another clinicopathological study by Kaira et al. showed that high expression of CD147 was associated with the expression of SLC7A5 and SLC1A5, which correlated with proliferation, angiogenesis, and mammalian target of rapamycin (mTOR) signaling in pancreatic cancer cells.²³ Fuchs and Bode hypothesized that LAT1 provides essential amino acids for tumor growth and proliferation and is overexpressed in cancer tissues via the mTOR signaling pathway, whereas SLC1A5 drives the function of SLC7A5 through glutamine delivery.²⁴ As shown in a study by Nicklin et al., L-glutamine activates mTOR signaling to mediate cell growth and proliferation, a process that is regulated by the transporters SLC1A5 and SLC7A5/SLC3A2.²¹ The results of a study by Hayashi et al. also revealed the biological significance of c-Myc in pancreatic cancer growth, indicating that overexpression of c-Myc increases SLC7A5 activity by binding to a canonical sequence in the promoter (Figure 2).²⁵

SLC7A11 encodes the light chain subunit xCT, which forms a complex with the cystine/glutamate antiporter system X_c^- and the heavy chain subunit SLC3A2. The main function of the X_c^- system is to promote glutathione biosynthesis through the uptake of cystine and the concomitant release of glutamate.²⁶ Zhu et al. indicated that the proliferation of pancreatic cancer cells is promoted by SLC7A11 overexpression and inhibited by miR-139-5p expression via the PI3K/AKT signaling pathway.²⁷

SLC30, which encodes zinc transporter proteins (ZnTs) and SLC39, which encodes ZIPs, are members of two zinc transporter families in human cells with opposite functions.²⁸ ZnTs mediate zinc flow from the cytoplasm to the extracellular environment or absorb zinc from the cytoplasm in the intracellular compartment to reduce the cytosolic zinc concentration, while ZIPs mediate the absorption of zinc from the extracellular environment and transport into the cytoplasm or induce the release of zinc from intracellular organelles (including the endoplasmic reticulum, mitochondria, and Golgi) into the cytoplasm to increase the cytosolic zinc concentration.^{29,30}

Regarding SLC39A4 (ZIP4), Li et al. first reported in 2007 that the ZIP4 mRNA is overexpressed 5.5-fold in pancreatic cancer tissue specimens compared with normal tissues, and ZIP4 overexpression was also confirmed to increase cell proliferation by 101% in pancreatic cancer tissue.⁶ Then, in 2009, the authors further deciphered the function of SLC39A4 by silencing it in a mouse pancreatic cancer model and found that cell proliferation, migration, and invasion were all suppressed and that survival outcomes were significantly improved.³¹ However, the detailed mechanism by which SLC39A4 regulates pancreatic cancer growth is not completely clear. Zhang et al. explained a potential mechanism in their study: SLC39A4 mediates pancreatic cancer cell growth by upregulating the expression of neuropilin-1 (NRP-1), vascular endothelial growth factor (VEGF), and matrix metalloproteases (MMPs) in cell lines and xenografts.³² In addition, Zhang et al. revealed another mechanism: overexpression of SLC39A4 increases the transcription of interleukin 6 (IL-6) through cAMP response element-binding protein (CREB), which activates signal transducer and activator of transcription 3 (STAT3) and increases the expression of cyclin D1 to increase cell proliferation and tumor progression in pancreatic cancer.³³ Furthermore, they identified a novel mechanism in 2013 in which ZIP4 transcriptionally induced miR-373 expression in pancreatic cancer to promote tumor growth by activating CREB, which formed a novel signaling pathway.³⁴

Another zinc transporter, SLC39A6, which encodes the ZIP6 protein (also known as LIV-1), is a downstream target of STAT3 that is crucial for the regulation of the epithelial to mesenchymal transition (EMT). Unno et al. reported higher expression of the LIV-1 mRNA in both pancreatic cancer cell lines and cancer tissues than in normal cell lines and tissues.³⁵ Moreover, downregulation of SLC39A6 inhibits pancreatic cancer cell proliferation in vitro and reduces tumor growth and metastasis in vivo, which indicates the aggressive role of SLC39A6 in pancreatic cancer.

In addition to sustaining proliferative signaling, cancer cells also have the capability to circumvent tumor suppressor genes that limit tumor cell growth and proliferation. SLC5A8, also known as sodium-coupled monocarboxylate transporter 1 (SMCT1), is a plasma membrane cotransporter for monocarboxylates such as lactate, butyrate, pyruvate, acetate, propionate, nicotinate, and β -hydroxybutyrate.^{36–39} Among its substrates, butyrate, propionate and pyruvate are histone deacetylase (HDAC) inhibitors that function as tumor suppressors by inducing apoptosis.⁴⁰ Park et al. reported that aberrant promoter DNA methylation induces the silencing of SLC5A8 in pancreatic cancer, which may contribute to the carcinogenesis and progression of pancreatic tumors.⁴¹ Similarly, Helm et al. assessed the effect of SLC5A8 expression on



FIGURE 2 Solute carriers (SLCs) associated with PDAC cell proliferation, angiogenesis, invasion, and metastasis. (A) Glutamine is transported into tumor cells by SLC1A5 and can be used as an efflux substrate by the SLC7A5/SLC3A2 complex to mediate the uptake of essential amino acids (EAAs), finally activating the mTOR pathway to promote cell proliferation and angiogenesis.^{21,23,24} Overexpression of c-Myc induces the expression of SLC7A5, which further activates mTOR signaling.²⁵ (B) The SLC7A11/SLC3A2 complex (system X_c^{-}) regulates the uptake of cystine against the efflux of L-glutamate, which further activates the PI3K/Akt signaling pathway to promote cell proliferation.²⁶ The proliferation of tumor cells is also inhibited by miR-139-5p expression via the PI3K/AKT signaling pathway.²⁷ (C) Overexpression of SLC39A4 upregulates the transcription of IL-6 through CREB, which activates STAT3 to increase cell proliferation and tumor progression.³⁴ SLC39A4 induces the transcription of miR-373, which inhibits the transcription of TP53INP1, LATS2, and CD44 to promote tumor growth.³⁴ (D) Inhibition of SLC6A14 decreases tumor cell proliferation by suppressing the mTOR pathway⁶¹

the survival outcomes of patients with pancreatic cancer and found that low expression and nuclear translocation of SLC5A8 were associated with a poor prognosis.⁴²

Regarding SLC39A3, Costello et al. reported that the physiological zinc level exerts cytotoxic effects on pancreatic cancer cells and that the downregulation of Ras responsive element-binding protein 1 (RREB1) represses the expression of SLC39A3, resulting in a decrease in zinc levels, which alleviates these cytotoxic effects and promotes the proliferation of pancreatic cancer cells.^{43,44} An in vivo study by Takagishi et al. indicated that ZIP3 expression is regulated by RREB1 in normal ductal/acinar epithelial cells.⁴⁵ Similarly, Franklin et al. found that concurrent downregulation of SLC39A3 and its positive regulator RREB1 decreases zinc uptake and increases cell proliferation in pancreatic cancer cells.⁴⁶

SLC41A1, the homolog of the prokaryotic magnesium (Mg^{2+}) transporter E (Mgt E) family, was the first discovered as a sodium-dependent eukaryotic protein that mediates the efflux of Mg^{2+} in human cells.^{47–49} Xie et al. reported that overexpression of SLC41A1 suppressed the growth of human PDAC cell lines by inducing the expression of Bax while suppressing the expression of Bcl-2.⁵⁰ In addition, overexpression of SLC41A1 increased the efflux of

cellular Mg²⁺ and suppressed the activation of Akt/mTOR signaling, which regulates Bcl-2 upstream.⁵⁰

2.2 | Cell death

Programmed cell death (PCD) describes the death of a cell via apoptosis, which serves as a natural barrier for cancer cell development.^{51,52} PCD is a regulated process that usually confers advantages during an organism's life cycle. The second important characteristic of cancer cells is their ability to evolve to limit or circumvent PCD.¹⁶ Most cells evade apoptosis through a loss of function mutation of the tumor suppressor gene TP53, upregulation of the expression of the antiapoptotic genes Bcl-2 or Bcl- x_L and survival signals (IGF1/2), downregulation of proapoptotic genes (Bax, Bim, and PUMA), or the inhibition of ligand-induced cell death.¹⁶

In addition to cell proliferation, SLC7A11 (xCT) also participates in PCD in pancreatic cancer. Chang et al. reported that the inhibition of prostaglandin reductase 2 (PTGR2) induces reactive oxygen species (ROS)-dependent apoptosis by suppressing the expression of SLC7A11 (xCT).⁵³ Similarly, in 2020, a study by Badgley et al. revealed the mechanism of ferroptosis (a form of cell death caused by lipid ROS) in pancreatic cancer in mice.⁵⁴ They found that knockdown of SLC7A11 (xCT) in engineered mice induced tumor cell ferroptosis and repressed pancreatic cancer tumor growth, indicating that pancreatic cancer cells rely on cysteine import to evade ferroptosis, which may be a promising anticancer therapeutic approach for pancreatic cancer.⁵⁴

According to previous studies, zinc is a crucial trace element that regulates cell proliferation, apoptosis, cellular metabolism, and gene expression in cancer.55 In a report by Cui et al., SLC39A4 conferred apoptosis resistance in pancreatic cancer cells through the caspase-9/caspase-7/PARP cascade.⁵⁶ Donadelli et al. also found that use of the zinc chelator N,N,N',N'-tetrakis (2-pyridylmethyl) ethylenediamine (TPEN) to deplete cellular zinc levels induced pancreatic cancer cell apoptosis by regulating the expression of mitochondria-related genes in the Bcl family (Bcl-X, Bcl-2, Bcl-W, and Bcl-XL), and this effect was reversed by zinc addition.⁵⁷ The authors confirmed that TPEN induced cell death by activating the caspase-3 and caspase-8 pathways.⁵⁷ However, they also reported that the application of the ionophore compound pyrrolidine dithiocarbamate (PDTC) increased zinc levels and inhibited the growth of pancreatic cancer cells by inducing ROS-dependent cell apoptosis.⁵⁸ Their experiments also showed that zinc-dependent ROS production induced mitochondrial damage through the nuclear

translocation of mitochondrial apoptosis-inducing factor (AIF), which results in cell apoptosis instead of caspase pathway activation.⁵⁸ Therefore, the role of zinc in preventing pancreatic cancer cell apoptosis requires further study.

Another potent mechanism by which SLC mediates resistance to cell death/apoptosis is via cancer cell autophagy. SLC31A1, also known as CTR1, is a cell membrane transporter that regulates the intracellular copper concentration.⁵⁹ Yu et al. found that copper intake is important for the progression of pancreatic cancer and that the knockdown of SLC31A1 blocked copper absorption to inhibit pancreatic cancer progression by increasing ROS-induced autophagy, which also contributed to resistance to cell apoptosis.⁶⁰

SLC6A14, also known as amino acid transporter $B^{0,+}$ (ATB^{0,+}), mediates the uptake of a broad range of neutral and cationic amino acids in a Na⁺/Cl⁻-dependent manner.⁶¹ As shown in a study by Coothankandaswamy et al., pharmacological and genetic inhibition of SLC6A14 decreases the proliferation of pancreatic cancer cells both in vitro and in vivo, and blockade of this gene induces tumor cell autophagy and suppresses the mTOR signaling pathway (Figures 2 and 3).⁶¹

2.3 | Angiogenesis

In addition to the traits mentioned above, tumor cells must also absorb oxygen and nutrients and release carbon dioxide and waste, which could occur via tumor angiogenesis.¹⁶ The process of angiogenesis in normal cells is transiently activated in certain situations, such as wound healing or endometrial repair; however, in tumor cells, angiogenesis is always activated. The essential cytokine mediating this process is VEGF, a signaling protein secreted by tumor cells to stimulate the creation of new blood vessels, which is associated with tumor metastasis and a poor prognosis.³²

SLC4A4, which encodes the protein NBCe1, is a sodium bicarbonate transporter that was identified as a target of microRNA-223.⁶² Zhang et al. reported that overexpression of human circular RNA 001587 upregulates the expression of SLC4A4 to inhibit angiogenesis in pancreatic cancer by binding to microRNA-223.⁶³

In addition to its function in cell proliferation, SLC39A4 is reported to upregulate the expression of VEGFs in pancreatic cancer by upregulating its receptor NRP-1, revealing an autocrine signaling pathway involving the ZIP4– VEGF–NRP-1 axis.^{32,64}

However, limited studies have been conducted on SLCs and their involvement in the angiogenesis of pancreatic cancer, necessitating further studies.



FIGURE 3 Solute carrier (SLC) expression is associated with cell death. (A) Inhibition of PTGR2 suppresses the expression of the SLC7A11/SLC3A2 complex (system X_c^-), resulting in the activation of ROS-dependent apoptosis.⁵³ The suppression of SLC7A11 (xCT) also induces ferroptosis via lipid ROS production.⁵⁴ (B) Cyt-C, which is released as a result of mitochondrial damage caused by Bax and Bak, binds to Apaf1 to activate the caspase-9/caspase-7/PARP pathway and induce cell apoptosis.⁵⁶ Overexpression of SLC39A4 increases zinc levels and represses the expression of Bax and Bak, subsequently conferring cell apoptosis resistance.⁵⁶ Conversely, the use of TPEN to reduce zinc levels induces apoptosis by inhibiting the antiapoptotic genes Bcl-2, Bcl-W, and Bcl-XL.⁵⁷ Paradoxically, the use of pyrrolidine dithiocarbamate (PDTC) to increase zinc levels also induces apoptosis through ROS-induced mitochondrial damage mediated apoptosis-inducing factor (AIF), resulting in cell apoptosis instead of caspase pathway activation.⁵⁸ (C) The inhibition of SLC31A1 to block copper absorption leads to the suppression of pancreatic cancer progression by increasing ROS-induced autophagy, which also promotes the resistance to cell apoptosis.⁶⁰ (E) Blockade of SLC6A14 induces tumor cell autophagy⁶¹

2.4 | Invasion and metastasis

After tumors are formed at the original site, malignant cells can leave the primary site via different mechanisms and travel (metastasize) to distant tissues or organs with environments suitable for their growth. This process contributes to the death of most patients with cancer.^{65–67} Several SLCs have been reported to regulate the invasion and metastasis of pancreatic cancer.

SLC2A1-encoding glucose transporter 1 (GLUT1) mediates glucose uptake into cells.⁶⁸ In a study by Nagarajan et al., paraoxonase 2 (PON2) promoted

tumor cell growth and metastasis by activating SLC2A1/GLUT1-mediated glucose transport and inhibiting the AMPK/FOXO3A/PUMA pathway.⁶⁹ According to another study, overexpression of GLUT-1 upregulated the expression of MMP-2 and promoted pancreatic cancer cell invasiveness.⁷⁰ The prognostic value of GLUT1 has also been explored by Sharen et al., who found that a high level of GLUT1 expression was associated with a tumor size greater than 2 cm and with lymph node metastasis in patients with pancreatic cancer, and these features resulted in shorter overall survival.⁷¹

In addition to its involvement in pancreatic cancer angiogenesis, the sodium bicarbonate transporter SLC4A4 is also related to invasiveness.⁶³ Zhang et al. reported that overexpression of human circular RNA 001587 inhibits the migration and invasion of pancreatic cancer cells, changes that are counteracted by upregulating the expression of SLC4A4.⁶³

As mentioned above, SLC7A11 encodes a subunit of the amino acid transport system X_c that mediates cystine uptake and glutamate biosynthesis.²⁶ In addition to its function in proliferation, Zhu et al. also found that overexpression of SLC7A11 promotes tumor cell invasion and metastasis, changes that are suppressed by miR-139-5p via the PI3K/Akt signaling pathway (Figure 2).²⁷ Furthermore, the authors verified this result with clinical data showing that low expression of miR-139-5p and high expression of SLC7A11 are associated with a poor prognosis, indicating that miR-139-5p may represent a new biomarker of pancreatic cancer.²⁷

SLC9A1, also known as Na⁺/H⁺ exchanger isoform 1 (NHE1), is a membrane transporter regulating pH in tumor cells, and its main function is to maintain the proton gradient.⁷² A study by Cardone et al. showed that NHE1 interacts with epidermal growth factor receptor (EGFR) to promote the growth and invasion of pancreatic cancer tumor cells via the digestion of the extracellular matrix.⁷³

A study by Olszewski et al. also showed that neurotensin receptor signaling promotes pancreatic cancer cell metastasis by activating SLC9A1.⁷⁴ Malinda et al. observed upregulated expression of SLC9A1 and SLC4A7 during the aberrant transforming growth factor β (TGF β)-induced EMT in pancreatic cancer cell lines.⁷⁵

A recent study by Liu et al. substantiated the role of SLC39A4 in pancreatic cancer progression.⁷⁶ Their results showed that overexpression of SLC39A4 promoted migration and invasion in pancreatic cancer by downregulating the expression of ZO-1 and Claudin-1 through upregulation of the expression of transcriptional repressor ZEB1.⁷⁶ Moreover, SLC39A6 encodes the protein LIV-1. A study by Unno et al. indicated that LIV-1 might be involved in tumor invasion and metastasis through the EMT.³⁵

2.5 | Cellular metabolism

The uncontrolled cell proliferation in neoplastic diseases increases their demand for nutrients to fuel cell growth and division, and they meet these needs by reprogramming metabolism.¹⁶ Tumor cells must upregulate the expression of the transporters responsible for the uptake or secretion of nutrients such as glucose, lactate, and amino acids to alter metabolism.⁷⁷ Otto Warburg first observed the abnormal energy metabolism or "aerobic glycolysis" of cancer cells; namely, tumor cells reprogram their metabolism by oxidizing glucose and producing lactate as waste mainly through glycolysis under aerobic conditions.^{16,77} Tumor cells must upregulate the expression of the glucose transporters SLC2A1 (GLUT1) and SLC5A1 (SGTL1) to increase glucose uptake.

Many researchers have reported the overexpression of the SLC2A1 gene in patients with pancreatic cancer compared to normal controls.^{78,79} Brandi et al. reported that the antioxidant mitochondrial uncoupling protein 2 (UCP2) promotes the proliferation of pancreatic cancer cells by upregulating the expression of SLC2A1 to limit metabolism to glycolysis.⁸⁰ Nagarajan et al. also found that human paraoxonase 2 (PON2) promotes pancreatic cancer cell growth and metastasis through SLC2A1-mediated glucose transport.⁶⁹ The prognostic value of this gene has been studied, and the results indicate that the expression of SLC2A1 is negatively correlated with the prognosis of patients with pancreatic cancer.^{68,71,81,82}

Another glucose transporter, SLC5A1, encodes sodiumdependent glucose transporter 1 (SGLT1). Gao et al. observed SLC5A1 overexpression in pancreatic cancer cells, and knockdown of this gene suppresses the growth and progression of pancreatic cancer by reducing glucose uptake through the AMPK/mTOR signaling pathway.⁸³ The prognostic value of SLC5A1 has also been illustrated by the significant association between the high expression of SLC5A1 in pancreatic cancer and the disease-free survival (DFS) of patients.⁸⁴

Tumor cells also need to increase the uptake of amino acids to fuel cell growth. The induction of SLC1A5 expression by hypoxia promotes the transport of mitochondrial glutamine to facilitate metabolism reprogramming in pancreatic cancer.¹⁷ A study by Hayashi et al. showed an important role for SLC7A5 in tumor cell growth in pancreatic cancer.²⁵ The prognostic significance of this gene has been illustrated by the association of high SLC7A5 expression with shorter overall survival in patients with pancreatic cancer.^{85,86}

In addition to glucose and amino acids, nucleotides are also quite important for tumor cells. Vesicular ATP release through exocytosis is the main process inducing purinergic chemical transmission, which has also been shown to play a crucial role in regulating the function of pancreatic exocrine cells, from which PDACs most likely originate.^{87,88} The purinergic signaling that transmits signals from nucleotides, particularly ATP, ADP, and adenosine, affects several steps of tumor biology ranging from the tumor microenvironment to proliferation, migration, EMT and metastasis, energy metabolism, and immune evasion.⁸⁹

The vesicular nucleotide transporter (VNUT) encoded by the SLC17A9 gene mediates vesicular ATP accumulation and release in secretory granules in different tissues with abundant vesicular ATP transporters, such as the brain, adrenal glands, pancreas, and other tissues.^{87,90,91} Novak et al. summarized the purinergic signaling in pancreatic cancer and indicated that PDAC cell lines release ATP through VNUT in response to several stimuli.⁹² Furthermore, VNUT regulates the viability of pancreatic cancer cells by functioning as lysosomal ATP transporters, and extracellular nucleosides also promote anticancer immunity by inducing immunogenic cell death.^{93,94}

2.6 | Chemosensitivity

The current cornerstones of chemotherapy in patients with pancreatic cancer are the uracil analog 5-fluorouracil (5-FU) and the cytidine analog gemcitabine (GEM). In the past few decades, with the emergence of new chemotherapy drugs, such as tegafur/gimeracil/oteracil potassium capsule (S-1), albumin-bound paclitaxel (Abraxane), oxaliplatin and irinotecan, more options have been developed for combined chemotherapy for the treatment of pancreatic cancer. However, the prognosis remains poor because of multidrug resistance (MDR), with a 5-year relative survival rate of less than 8%.95 MDR is the major obstacle that affects the efficacy of systematic treatment in patients with pancreatic cancer. Because the SLC family transports a wide variety of substrates, including anticancer drugs, the mechanisms by which SLC proteins modulate MDR are mainly due to the decrease in drug uptake into tumor cells mediated by the low expression of certain SLCs.^{96,97}

In addition to its function in sustained proliferative signaling, SLC1A5 also plays a role in chemotherapy sensitivity. A study by Yoo et al. showed that overexpression of an SLC1A5 variant conferred resistance to gemcitabine in pancreatic cancer cells.¹⁷

SLC7A11 functions in proliferation and invasion, but Yang et al found that overexpression of the antisense transcript of SLC7A11 (SLC7A11-AS1) in pancreatic cancer cells promotes gemcitabine resistance by reducing ROS levels through the inhibition of the ubiquitination and degradation of nuclear factor erythroid-2-related factor 2 (NRF2).⁹⁸ d that unregulation of SL C7A1

11 of 19

Similarly, Lo et al. also found that upregulation of SLC7A11 (X_c^-) is associated with gemcitabine resistance.⁹⁹

SLC22 subfamily members such as organic cation transporter 1 (OCT1, SLC22A1), OCT2 (SLC22A2), and OCT3 (SLC22A3) have been reported to transport platinum agents in vitro, but the results remain controversial.¹⁰⁰ Mohelnikova-Duchonova et al. reported the upregulated expression of SLC22A3 in pancreatic cancer tissues compared with nonneoplastic tissues, and this change predicted a favorable overall survival in patients treated with 5-FU or gemcitabine-based chemotherapy (p = .004).¹⁰¹

SLC28, a concentrative nucleoside transporter (CNT) subfamily member, was also reported to mediate the transport of gemcitabine and 5-FU in pancreatic cancer.^{102–104} Mackey et al. identified important roles for SLC28A1 and SLC28A2 in gemcitabine transfer in vitro and showed that a deficiency in SLCs might contribute to gemcitabine resistance.¹⁰⁵ García-Manteiga et al. concluded that most cell lines take up gemcitabine via SLC29A1 instead of SLC28A1, which is expressed in human pancreatic cancer at high levels, and that the expression of SLC28A1 and SLC29A1 may increase the sensitivity of pancreatic cancer cells to gemcitabine.¹⁰²

SLC29A1, also known as equilibrative nucleoside transporter 1 (ENT1), mediates both the influx and efflux of nucleosides across the membrane. According to previous studies, pancreatic cancer cell lines take up gemcitabine via SLC29A1.¹⁰² A large number of studies have documented that high SLC29A1 expression is associated with a favorable prognosis for patients with pancreatic cancer undergoing gemcitabine chemotherapy.¹⁰⁶⁻¹⁰⁸ Furthermore, the prospective randomized studies ESPA-3 and RTOG9704 are assessing this gene, and results have shown a significantly longer median survival for patients undergoing gemcitabine chemotherapy with high SLC29A1 protein expression compared with patients with low or negative SLC29A1 protein expression.^{109,110} However, the number of studies is small, and inconsistent results have been obtained.¹¹¹ Kawada et al. did not observe an association between the expression level of SLC29A1 and the prognosis of patients with pancreatic cancer treated with neoadjuvant gemcitabine chemotherapy, potentially due to the limitation that the SLC29A1 evaluation occurred after neoadjuvant chemoradiation.¹¹¹ A randomized multicenter phase II study also showed that SLC29A1 did not predict gemcitabine sensitivity in patients with metastatic pancreatic cancer.¹¹² A multicenter phase III study of AIO-PK0104 from Germany comparing two adjuvant chemotherapy regimens in patients with advanced pancreatic cancer reported similar results.¹¹³ In general, the role of SLC29A1 in gemcitabine chemotherapy resistance remains controversial and requires further study.



FIGURE 4 Solute carrier (SLC) expression is associated with chemotherapy resistance. (A) The expression of an SLC1A5 variant that encodes the mitochondrial glutamine transporter is controlled by hypoxia through HIF2α. Overexpression of the SLC1A5 variant confers resistance to gencitabine in pancreatic cancer cells.¹⁷ (B) The expression of the antisense transcript of SLC7A11 (SLC7A11-AS1) in pancreatic cancer cells promotes gencitabine resistance by reducing ROS levels through the inhibition of the ubiquitination and degradation of NRF2.⁹⁸ (C) Deficiency in SLC28A1 and SLC28A2, which mediate the transport of gencitabine and 5-FU in pancreatic cancer, contributes to chemotherapy resistance.¹⁰⁵ (D) Pancreatic cancer cells take up gencitabine and 5-FU via SLC29A1.^{102,114} ZIP4 upregulates the expression of ZEB1 to suppress the expression of SLC29A1, which reduces gencitabine uptake in pancreatic cancer cells¹¹⁷

Additionally, 5-FU sensitivity has also been reported to be associated with SLC29A1 expression.¹¹⁴ This sensitivity is significantly increased when SLC29A1 is inhibited by nitrobenzylmercaptoprine ribonucleoside (NBMPR).¹¹⁴ Nevertheless, these results are not consistent with those of the prospective studies RTOG9704 and ESPAC-3.^{109,110} Limited data are available on other members of the SLC29 subfamily. Nishio et al. found that a change in localization of SLC29A2 disrupted gemcitabine uptake, resulting in chemotherapy resistance in pancreatic cancer.¹¹⁵ High expression of SLC29A3 is significantly associated with favorable overall survival in patients with pancreatic cancer, but the underlying mechanism remains unknown.¹⁰¹ Unlike SLC29A1 and SLC29A2, Endo et al. reported that SLC29A3 and SLC29A4 are not prominent nucleoside transporters and that a significant improvement in chemotherapy sensitivity is not observed in cell lines expressing these two genes.¹¹⁶ ZIP4 upregulates the expression of ZEB1 to suppress the expression of SLC29A1, a gemcitabine transporter, which reduces gemcitabine uptake in pancreatic cancer cells (Figure 4).¹¹⁷

3 | FUTURE PERSPECTIVES

With the gradual discovery of the functions of some SLCs over the past few decades, SLCs have increasingly become targets of anticancer drugs and a future hotspot of this field.^{118,119} The advantages of SLCs as anticancer drug targets might be their easily accessibility without crossing any cellular barriers and the ability to reduce the cytotoxicity caused by nonselective drugs.¹¹ SLCs encode over 40% of transporter proteins, and more than 80 SLCs have been reported to be associated with genetic disorders, but only a small number of these proteins have been approved or are currently undergoing testing.¹²⁰ According to review by Zhang et al., drugs targeting the SLC5, 6, 12, 18, 22, 25, and 29 subfamilies have been approved as treatments for hema-

tological, nervous system, motor, and other diseases.¹²¹ An additional subfamily of nine SLCs, including SLC2, 5, 7, 9, 10, 22, and 40, is being tested in clinical trials.¹²¹ However, the potential of SLCs in precision treatment for pancreatic cancer still remains uncertain but has great therapeutic potential, and prospective clinical trials are needed to support the results of basic research studies.

SLCs regulate metabolic homeostasis, but researchers have not yet determined whether they alter the immune microenvironment in pancreatic cancer. According to the immunosurveillance theory, tumor cells have the ability to avoid detection and evade eradication by the immune system. The depletion of L-Arg in T cells inhibits mTOR through SLC38A9, which in turn suppresses the growth and proliferation of T cells.¹²²⁻¹²⁴ Kynurenine, which is transported by SLC7A5, SLC7A8, and SLC36A4, can lead to Trp degradation in effector T cells, subsequently impairing the immune function and inducing PD-1 expression.^{125,126} Furthermore, tumor cells have also been reported to outcompete T cells for methionine by expressing a high level of the transporter SLC34A2.¹²⁷ However, no studies have examined the role of SLCs in the immune evasion of pancreatic cancer, necessitating in-depth studies to decipher this problem. We believe this topic will be a hot area in the future.

4 | CONCLUSIONS

In summary, SLCs play various roles in pancreatic cancer, including proliferative signaling, cell death, angiogenesis, tumor invasion and metastasis, cellular metabolism, and chemotherapy sensitivity, providing possible therapeutic strategies for cancer treatment. Although an increasing number of studies have revealed the important functions of SLC family members in tumor biology, they have received relatively little attention to date. The functions and substrates of many SLCs remain unknown. In addition, the data are inconsistent and the conclusions are ambiguous. As a result, research on SLCs in pancreatic cancer is still in its infancy. Furthermore, membrane proteins are difficult to crystallize, which hinders the elucidation of the physiological function, and three-dimensional structures need to identify substrate binding sites and important domains. Despite these obstacles, the therapeutic potential of SLCs in pancreatic cancer is substantial. We propose that with the development of molecular biology, new SLCs and new functions will be discovered and translation from basic advances into clinical treatment will become a reality.

ACKNOWLEDGMENTS

This study was jointly funded by the National Natural Science Foundation of China (Nos. 81802352, 81772555,

and 81902428), the National Science Foundation for Distinguished Young Scholars of China (No. 81625016), the Shanghai Sailing Program (Nos. 19YF1409400 and 20YF1409000), the Shanghai Rising Star Program (No. 20QA1402100), the Shanghai Anticancer Association Young Eagle Program (No. SACA-CY19A06), the Clinical and Scientific Innovation Project of Shanghai Hospital Development Center (Nos. SHDC12018109 and SHDC12019109), and the Scientific Innovation Project of Shanghai Education Committee (No. 2019-01-07-00-07-E00057). We thank the editors at AJE (American Journal Experts) for their assistance with editing the manuscript, and all figures were created with BioRender.com.

CONFLICT OF INTEREST

The authors declare that there is no conflict of interest.

AUTHOR CONTRIBUTIONS

Zijian Wu, Jin Xu, and Chen Liang wrote the manuscript and created the illustrations. Qingcai Meng, Jie Hua, Wei Wang, Bo Zhang, and Jiang Liu contributed to the acquisition of literature. Si Shi and Xianjun Yu contributed to the conception and design, revised the article, provided final approval of the version to be published, and agreed to be accountable for all aspects of the work.

ORCID

Xianjun Yu https://orcid.org/0000-0002-6697-7143 *Si Shi* https://orcid.org/0000-0002-6652-0629

REFERENCES

- Bray F, Ferlay J, Soerjomataram I, Siegel RL, Torre LA, Jemal A. Global cancer statistics 2018: GLOBOCAN estimates of incidence and mortality worldwide for 36 cancers in 185 countries. *CA Cancer J Clin.* 2018;68(6):394-424.
- Siegel RL, Miller KD, Jemal A. Cancer statistics, 2018. CA Cancer J Clin. 2018;68(1):7-30.
- Rahib L, Smith BD, Aizenberg R, Rosenzweig AB, Fleshman JM, Matrisian LM. Projecting cancer incidence and deaths to 2030: the unexpected burden of thyroid, liver, and pancreas cancers in the United States. *Cancer Res.* 2014;74(11):2913-2921.
- 4. Labori KJ, Katz MH, Tzeng CW, et al. Impact of early disease progression and surgical complications on adjuvant chemotherapy completion rates and survival in patients undergoing the surgery first approach for resectable pancreatic ductal adenocarcinoma - a population-based cohort study. *Acta Oncol.* 2016;55(3):265-277.
- 5. Mizrahi JD, Surana R, Valle JW, Shroff RT. Pancreatic cancer. *Lancet*. 2020;395(10242):2008-2020.
- Li M, Zhang Y, Liu Z, et al. Aberrant expression of zinc transporter ZIP4 (SLC39A4) significantly contributes to human pancreatic cancer pathogenesis and progression. *Proc Natl Acad Sci* USA. 2007;104(47):18636-18641.
- 7. Schlessinger A, Matsson P, Shima JE, et al. Comparison of human solute carriers. *Protein Sci.* 2010;19(3):412-428.

- Schumann T, König J, Henke C, et al. Solute carrier transporters as potential targets for the treatment of metabolic disease. *Pharmacol Rev.* 2020;72(1):343-379.
- Höglund PJ, Nordström KJ, Schiöth HB, Fredriksson R. The solute carrier families have a remarkably long evolutionary history with the majority of the human families present before divergence of Bilaterian species. *Mol Biol Evol.* 2011;28(4):1531-1541.
- El-Gebali S, Bentz S, Hediger MA, Anderle P. Solute carriers (SLCs) in cancer. *Mol Aspects Med.* 2013;34(2-3):719-734.
- Panda S, Banerjee N, Chatterjee S. Solute carrier proteins and c-Myc: a strong connection in cancer progression. *Drug Discov Today*. 2020;25(5):891-900.
- Gray KA, Seal RL, Tweedie S, Wright MW, Bruford EA. A review of the new HGNC gene family resource. *Hum Genomics*. 2016;10:6.
- Hediger MA, Romero MF, Peng JB, Rolfs A, Takanaga H, Bruford EA. The ABCs of solute carriers: physiological, pathological and therapeutic implications of human membrane transport proteins. Introduction. *Pflugers Arch.* 2004;447(5):465-468.
- Perland E, Fredriksson R. Classification systems of secondary active transporters. *Trends Pharmacol Sci.* 2017;38(3):305-315.
- Hanahan D, Weinberg RA. The hallmarks of cancer. *Cell*. 2000;100(1):57-70.
- 16. Hanahan D, Weinberg RA. Hallmarks of cancer: the next generation. *Cell*. 2011;144(5):646-674.
- 17. Yoo HC, Park SJ, Nam M, et al. A variant of SLC1A5 is a mitochondrial glutamine transporter for metabolic reprogramming in cancer cells. *Cell Metab.* 2020;31(2):267-283.e12.
- 18. Liu Y, Zhao T, Li Z, Wang L, Yuan S, Sun L. The role of ASCT2 in cancer: a review. *Eur J Pharmacol*. 2018;837:81-87.
- Kaira K, Sunose Y, Arakawa K, et al. Clinicopathological significance of ASC amino acid transporter-2 expression in pancreatic ductal carcinoma. *Histopathology*. 2015;66(2):234-243.
- Ramirez C, Hauser AD, Vucic EA, Bar-Sagi D. Plasma membrane V-ATPase controls oncogenic RAS-induced macropinocytosis. *Nature*. 2019;576(7787):477-481.
- Nicklin P, Bergman P, Zhang B, et al. Bidirectional transport of amino acids regulates mTOR and autophagy. *Cell*. 2009;136(3):521-534.
- 22. Kanai Y, Segawa H, Miyamoto K, Uchino H, Takeda E, Endou H. Expression cloning and characterization of a transporter for large neutral amino acids activated by the heavy chain of 4F2 antigen (CD98). *J Biol Chem.* 1998;273(37):23629-23632.
- Kaira K, Arakawa K, Shimizu K, et al. Relationship between CD147 and expression of amino acid transporters (LAT1 and ASCT2) in patients with pancreatic cancer. *Am J Transl Res.* 2015;7(2):356-363.
- Fuchs BC, Bode BP. Amino acid transporters ASCT2 and LAT1 in cancer: partners in crime? *Semin Cancer Biol*. 2005;15(4):254-266.
- Hayashi K, Jutabha P, Endou H, Anzai N. c-Myc is crucial for the expression of LAT1 in MIA Paca-2 human pancreatic cancer cells. *Oncol Rep.* 2012;28(3):862-866.
- Koppula P, Zhang Y, Zhuang L, Gan B. Amino acid transporter SLC7A11/xCT at the crossroads of regulating redox homeostasis and nutrient dependency of cancer. *Cancer Commun (Lond)*. 2018;38(1):12.

- Zhu JH, De Mello RA, Yan QL, et al. MiR-139-5p/SLC7A11 inhibits the proliferation, invasion and metastasis of pancreatic carcinoma via PI3K/Akt signaling pathway. *Biochim Biophys Acta Mol Basis Dis.* 2020;1866(6):165747.
- Liuzzi JP, Cousins RJ. Mammalian zinc transporters. Annu Rev Nutr. 2004;24:151-172.
- 29. Song ZH, Ke YL, Xiao K, Jiao LF, Hong QH, Hu CH. Diosmectite-zinc oxide composite improves intestinal barrier restoration and modulates TGF-β1, ERK1/2, and Akt in piglets after acetic acid challenge. *J Anim Sci.* 2015;93(4):1599-1607.
- Myers S, Shastri MD, Adulcikas J, Sohal SS, Norouzi S. Zinc and gastrointestinal disorders: a role for the zinc transporters Zips and ZnTs. *Curr Pharm Des.* 2017;23(16):2328-2332.
- 31. Li M, Zhang Y, Bharadwaj U, et al. Down-regulation of ZIP4 by RNA interference inhibits pancreatic cancer growth and increases the survival of nude mice with pancreatic cancer xenografts. *Clin Cancer Res.* 2009;15(19):5993-6001.
- Zhang Y, Chen C, Yao Q, Li M. ZIP4 upregulates the expression of neuropilin-1, vascular endothelial growth factor, and matrix metalloproteases in pancreatic cancer cell lines and xenografts. *Cancer Biol Ther*. 2010;9(3):236-242.
- 33. Zhang Y, Bharadwaj U, Logsdon CD, Chen C, Yao Q, Li M. ZIP4 regulates pancreatic cancer cell growth by activating IL-6/STAT3 pathway through zinc finger transcription factor CREB. *Clin Cancer Res.* 2010;16(5):1423-1430.
- Zhang Y, Yang J, Cui X, et al. A novel epigenetic CREB-miR-373 axis mediates ZIP4-induced pancreatic cancer growth. *EMBO Mol Med.* 2013;5(9):1322-1334.
- 35. Unno J, Satoh K, Hirota M, et al. LIV-1 enhances the aggressive phenotype through the induction of epithelial to mesenchymal transition in human pancreatic carcinoma cells. *Int J Oncol.* 2009;35(4):813-821.
- Gopal E, Miyauchi S, Martin PM, et al. Transport of nicotinate and structurally related compounds by human SMCT1 (SLC5A8) and its relevance to drug transport in the mammalian intestinal tract. *Pharm Res.* 2007;24(3):575-584.
- Thangaraju M, Gopal E, Martin PM, et al. SLC5A8 triggers tumor cell apoptosis through pyruvate-dependent inhibition of histone deacetylases. *Cancer Res.* 2006;66(24):11560-11564.
- Martin PM, Gopal E, Ananth S, et al. Identity of SMCT1 (SLC5A8) as a neuron-specific Na+-coupled transporter for active uptake of L-lactate and ketone bodies in the brain. *J Neurochem*. 2006;98(1):279-288.
- Gupta N, Martin PM, Prasad PD, Ganapathy V. SLC5A8 (SMCT1)-mediated transport of butyrate forms the basis for the tumor suppressive function of the transporter. *Life Sci.* 2006;78(21):2419-2425.
- 40. Coothankandaswamy V, Elangovan S, Singh N, Prasad PD, Thangaraju M, Ganapathy V. The plasma membrane transporter SLC5A8 suppresses tumour progression through depletion of surviving without involving its transport function. *Biochem J.* 2013;450(1):169-178.
- Park JY, Helm JF, Zheng W, et al. Silencing of the candidate tumor suppressor gene solute carrier family 5 member 8 (SLC5A8) in human pancreatic cancer. *Pancreas*. 2008;36(4):e32-e39.
- 42. Helm J, Coppola D, Ganapathy V, et al. SLC5A8 nuclear translocation and loss of expression are associated with

CLINICAL AND TRANSLATIONAL MEDICINE

poor outcome in pancreatic ductal adenocarcinoma. *Pancreas*. 2012;41(6):904-909.

- Costello LC, Levy BA, Desouki MM, et al. Decreased zinc and downregulation of ZIP3 zinc uptake transporter in the development of pancreatic adenocarcinoma. *Cancer Biol Ther.* 2011;12(4):297-303.
- Costello LC, Zou J, Desouki MM, Franklin RB. Evidence for changes in RREB-1, ZIP3, and zinc in the early development of pancreatic adenocarcinoma. *J Gastrointest Cancer*. 2012;43(4):570-578.
- Takagishi T, Hara T, Fukada T. Recent advances in the role of SLC39A/ZIP zinc transporters in vivo. *Int J Mol Sci.* 2017;18(12):2708.
- Franklin RB, Zou J, Costello LC. The cytotoxic role of RREB1, ZIP3 zinc transporter, and zinc in human pancreatic adenocarcinoma. *Cancer Biol Ther*. 2014;15(10):1431-1437.
- Goytain A, Quamme GA. Functional characterization of human SLC41A1, a Mg2+ transporter with similarity to prokaryotic MgtE Mg2+ transporters. *Physiol Genomics*. 2005;21(3):337-342.
- Wabakken T, Rian E, Kveine M, Aasheim HC. The human solute carrier SLC41A1 belongs to a novel eukaryotic subfamily with homology to prokaryotic MgtE Mg2+ transporters. *Biochem Biophys Res Commun.* 2003;306(3):718-724.
- Kolisek M, Nestler A, Vormann J, Schweigel-Röntgen M. Human gene SLC41A1 encodes for the Na+/Mgš+ exchanger. *Am J Physiol Cell Physiol*. 2012;302(1):C318-C326.
- 50. Xie J, Cheng CS, Zhu XY, et al. Magnesium transporter protein solute carrier family 41 member 1 suppresses human pancreatic ductal adenocarcinoma through magnesium-dependent Akt/mTOR inhibition and bax-associated mitochondrial apoptosis. *Aging (Albany NY)*. 2019;11(9):2681-2698.
- Adams JM, Cory S. The Bcl-2 apoptotic switch in cancer development and therapy. *Oncogene*. 2007;26(9):1324-1337.
- 52. Lowe SW, Cepero E, Evan G. Intrinsic tumour suppression. *Nature*. 2004;432(7015):307-315.
- 53. Chang EY, Chang YC, Shun CT, et al. Inhibition of prostaglandin reductase 2, a putative oncogene overexpressed in human pancreatic adenocarcinoma, induces oxidative stress-mediated cell death involving xCT and CTH gene expressions through 15-Keto-PGE2. *PLoS One*. 2016;11(1):e0147390.
- Badgley MA, Kremer DM, Maurer HC, et al. Cysteine depletion induces pancreatic tumor ferroptosis in mice. *Science*. 2020;368(6486):85-89.
- Franklin RB, Costello LC. The important role of the apoptotic effects of zinc in the development of cancers. *J Cell Biochem*. 2009;106(5):750-757.
- Cui X, Zhang Y, Yang J, et al. ZIP4 confers resistance to zinc deficiency-induced apoptosis in pancreatic cancer. *Cell Cycle*. 2014;13(7):1180-1186.
- Donadelli M, Dalla Pozza E, Costanzo C, Scupoli MT, Scarpa A, Palmieri M. Zinc depletion efficiently inhibits pancreatic cancer cell growth by increasing the ratio of antiproliferative/proliferative genes. *J Cell Biochem*. 2008;104(1):202-212.
- Donadelli M, Dalla Pozza E, Scupoli MT, Costanzo C, Scarpa A, Palmieri M. Intracellular zinc increase inhibits p53(-/-) pancreatic adenocarcinoma cell growth by ROS/AIF-mediated apoptosis. *Biochim Biophys Acta*. 2009;1793(2):273-280.

- Schweigel-Röntgen M. The families of zinc (SLC30 and SLC39) and copper (SLC31) transporters. *Curr Top Membr*. 2014;73:321-355.
- Yu Z, Zhou R, Zhao Y, et al. Blockage of SLC31A1-dependent copper absorption increases pancreatic cancer cell autophagy to resist cell death. *Cell Prolif.* 2019;52(2):e12568.
- Coothankandaswamy V, Cao S, Xu Y, et al. Amino acid transporter SLC6A14 is a novel and effective drug target for pancreatic cancer. *Br J Pharmacol.* 2016;173(23):3292-3306.
- Romero MF, Chen AP, Parker MD, Boron WF. The SLC4 family of bicarbonate (HCO₃⁻) transporters. *Mol Aspects Med*. 2013;34(2-3):159-182.
- Zhang X, Tan P, Zhuang Y, Du L. Hsa_circRNA_001587 upregulates SLC4A4 expression to inhibit migration, invasion and angiogenesis of pancreatic cancer cells via binding to microRNA-223. *Am J Physiol Gastrointest Liver Physiol*. 2020;319(6):G703-G717.
- Soker S, Takashima S, Miao HQ, Neufeld G, Klagsbrun M. Neuropilin-1 is expressed by endothelial and tumor cells as an isoform-specific receptor for vascular endothelial growth factor. *Cell*. 1998;92(6):735-745.
- 65. Meyer T, Hart IR. Mechanisms of tumour metastasis. *Eur J Cancer*. 1998;34(2):214-221.
- Kaplan RN, Rafii S, Lyden D. Preparing the "soil": the premetastatic niche. *Cancer Res.* 2006;66(23):11089-11093.
- Geiger TR, Peeper DS. Metastasis mechanisms. *Biochim Bio-phys Acta*. 2009;1796(2):293-308.
- Basturk O, Singh R, Kaygusuz E, et al. GLUT-1 expression in pancreatic neoplasia: implications in pathogenesis, diagnosis, and prognosis. *Pancreas*. 2011;40(2):187-192.
- Nagarajan A, Dogra SK, Sun L, et al. Paraoxonase 2 facilitates pancreatic cancer growth and metastasis by stimulating GLUT1-mediated glucose transport. *Mol Cell*. 2017;67(4):685-701.e6.
- Ito H, Duxbury M, Zinner MJ, Ashley SW, Whang EE. Glucose transporter-1 gene expression is associated with pancreatic cancer invasiveness and MMP-2 activity. *Surgery*. 2004;136(3):548-556.
- Sharen G, Peng Y, Cheng H, Liu Y, Shi Y, Zhao J. Prognostic value of GLUT-1 expression in pancreatic cancer: results from 538 patients. *Oncotarget*. 2017;8(12):19760-19767.
- Reshkin SJ, Cardone RA, Harguindey S. Na+-H+ exchanger, pH regulation and cancer. *Recent Pat Anticancer Drug Discov*. 2013;8(1):85-99.
- Cardone RA, Greco MR, Zeeberg K, et al. A novel NHE1centered signaling cassette drives epidermal growth factor receptor-dependent pancreatic tumor metastasis and is a target for combination therapy. *Neoplasia*. 2015;17(2):155-166.
- Olszewski U, Hamilton G. Neurotensin signaling induces intracellular alkalinization and interleukin-8 expression in human pancreatic cancer cells. *Mol Oncol.* 2009;3(3):204-213.
- 75. Malinda RR, Zeeberg K, Sharku PC, et al. TGFβ signaling increases net acid extrusion, proliferation and invasion in Panc-1 pancreatic cancer cells: SMAD4 dependence and link to Merlin/NF2 signaling. *Front Oncol.* 2020;10:687.
- Liu M, Yang J, Zhang Y, et al. ZIP4 promotes pancreatic cancer progression by repressing ZO-1 and Claudin-1 through a ZEB1-dependent transcriptional mechanism. *Clin Cancer Res.* 2018;24(13):3186-3196.

- Koppenol WH, Bounds PL, Dang CV. Otto Warburg's contributions to current concepts of cancer metabolism. *Nat Rev Cancer*. 2011;11(5):325-337.
- 78. Reske SN, Grillenberger KG, Glatting G, et al. Overexpression of glucose transporter 1 and increased FDG uptake in pancreatic carcinoma. *J Nucl Med.* 1997;38(9):1344-1348.
- Higashi T, Tamaki N, Torizuka T, et al. FDG uptake, GLUT-1 glucose transporter and cellularity in human pancreatic tumors. *J Nucl Med.* 1998;39(10):1727-1735.
- Brandi J, Cecconi D, Cordani M, et al. The antioxidant uncoupling protein 2 stimulates hnRNPA2/B1, GLUT1 and PKM2 expression and sensitizes pancreas cancer cells to glycolysis inhibition. *Free Radic Biol Med*. 2016;101:305-316.
- Pizzi S, Porzionato A, Pasquali C, et al. Glucose transporter-1 expression and prognostic significance in pancreatic carcinogenesis. *Histol Histopathol*. 2009;24(2):175-185.
- Kurahara H, Maemura K, Mataki Y, et al. Significance of glucose transporter type 1 (GLUT-1) expression in the therapeutic strategy for pancreatic ductal adenocarcinoma. *Ann Surg Oncol.* 2018;25(5):1432-1439.
- Gao HF, Chen LY, Cheng CS, Chen H, Meng ZQ, Chen Z. SLC5A1 promotes growth and proliferation of pancreatic carcinoma via glucose-dependent AMPK/mTOR signaling. *Cancer Manag Res.* 2019;11:3171-3185.
- Casneuf VF, Fonteyne P, Van Damme N, et al. Expression of SGLT1, Bcl-2 and p53 in primary pancreatic cancer related to survival. *Cancer Invest*. 2008;26(8):852-859.
- Kaira K, Sunose Y, Arakawa K, et al. Prognostic significance of L-type amino-acid transporter 1 expression in surgically resected pancreatic cancer. *Br J Cancer*. 2012;107(4): 632-638.
- Altan B, Kaira K, Watanabe A, et al. Relationship between LAT1 expression and resistance to chemotherapy in pancreatic ductal adenocarcinoma. *Cancer Chemother Pharmacol.* 2018;81(1):141-153.
- Geisler JC, Corbin KL, Li Q, Feranchak AP, Nunemaker CS, Li C. Vesicular nucleotide transporter-mediated ATP release regulates insulin secretion. *Endocrinology*. 2013;154(2): 675-684.
- Haanes KA, Novak I. ATP storage and uptake by isolated pancreatic zymogen granules. *Biochem J.* 2010;429(2):303-311.
- Campos-Contreras ADR, Díaz-Muñoz M, Vázquez-Cuevas FG. Purinergic signaling in the hallmarks of cancer. *Cells*. 2020;9(7):1612.
- Sawada K, Echigo N, Juge N, et al. Identification of a vesicular nucleotide transporter. *Proc Natl Acad Sci U S A*. 2008;105(15):5683-5686.
- Moriyama Y, Hiasa M, Sakamoto S, Omote H, Nomura M. Vesicular nucleotide transporter (VNUT): appearance of an actress on the stage of purinergic signaling. *Purinergic Signal*. 2017;13(3):387-404.
- 92. Novak I, Yu H, Magni L, Deshar G. Purinergic signaling in pancreas-from physiology to therapeutic strategies in pancreatic cancer. *Int J Mol Sci.* 2020;21(22):8781.
- Cao Q, Zhao K, Zhong XZ, et al. SLC17A9 protein functions as a lysosomal ATP transporter and regulates cell viability. *J Biol Chem.* 2014;289(33):23189-23199.
- Kepp O, Loos F, Liu P, Kroemer G. Extracellular nucleosides and nucleotides as immunomodulators. *Immunol Rev.* 2017;280(1):83-92.

- Zeng H, Chen W, Zheng R, et al. Changing cancer survival in China during 2003–15: a pooled analysis of 17 population-based cancer registries. *Lancet Glob Health*. 2018;6(5):e555-e567.
- Huang Y. Pharmacogenetics/genomics of membrane transporters in cancer chemotherapy. *Cancer Metastasis Rev.* 2007;26(1):183-201.
- 97. Mohelnikova-Duchonova B, Melichar B. Human equilibrative nucleoside transporter 1 (hENT1): do we really have a new predictive biomarker of chemotherapy outcome in pancreatic cancer patients? *Pancreatology*. 2013;13(6):558-563.
- Yang Q, Li K, Huang X, et al. lncRNA SLC7Al1-AS1 promotes chemoresistance by blocking SCF(β-TRCP)-mediated degradation of NRF2 in Pancreatic Cancer. *Mol Ther Nucleic Acids*. 2020;19:974-985.
- Lo M, Ling V, Wang YZ, Gout PW. The xc-cystine/glutamate antiporter: a mediator of pancreatic cancer growth with a role in drug resistance. *Br J Cancer*. 2008;99(3):464-472.
- Sprowl JA, Ness RA, Sparreboom A. Polymorphic transporters and platinum pharmacodynamics. *Drug Metab Pharmacokinet*. 2013;28(1):19-27.
- 101. Mohelnikova-Duchonova B, Brynychova V, Hlavac V, et al. The association between the expression of solute carrier transporters and the prognosis of pancreatic cancer. *Cancer Chemother Pharmacol.* 2013;72(3):669-682.
- 102. García-Manteiga J, Molina-Arcas M, Casado FJ, Mazo A. Pastor-Anglada M. Nucleoside transporter profiles in human pancreatic cancer cells: role of hCNT1 in 2',2'difluorodeoxycytidine-induced cytotoxicity. *Clin Cancer Res.* 2003;9(13):5000-5008.
- Damaraju VL, Damaraju S, Young JD, et al. Nucleoside anticancer drugs: the role of nucleoside transporters in resistance to cancer chemotherapy. *Oncogene*. 2003;22(47):7524-7536.
- Mohelníková-Duchonová B, Soucek P. The role of membrane transporters in cellular resistance of pancreatic carcinoma to gemcitabine. *Klin Onkol.* 2010;23(5):306-310.
- 105. Mackey JR, Mani RS, Selner M, et al. Functional nucleoside transporters are required for gemcitabine influx and manifestation of toxicity in cancer cell lines. *Cancer Res.* 1998;58(19):4349-4357.
- 106. Maréchal R, Mackey JR, Lai R, et al. Human equilibrative nucleoside transporter 1 and human concentrative nucleoside transporter 3 predict survival after adjuvant gemcitabine therapy in resected pancreatic adenocarcinoma. *Clin Cancer Res.* 2009;15(8):2913-2919.
- 107. Maréchal R, Bachet JB, Mackey JR, et al. Levels of gemcitabine transport and metabolism proteins predict survival times of patients treated with gemcitabine for pancreatic adenocarcinoma. *Gastroenterology*. 2012;143(3):664-674. e6.
- 108. Spratlin J, Sangha R, Glubrecht D, et al. The absence of human equilibrative nucleoside transporter 1 is associated with reduced survival in patients with gemcitabine-treated pancreas adenocarcinoma. *Clin Cancer Res.* 2004;10(20):6956-6961.
- 109. Greenhalf W, Ghaneh P, Neoptolemos JP, et al. Pancreatic cancer hENT1 expression and survival from gemcitabine in patients from the ESPAC-3 trial. J Natl Cancer Inst. 2014;106(1):djt347.
- 110. Farrell JJ, Elsaleh H, Garcia M, et al. Human equilibrative nucleoside transporter 1 levels predict response to gemcitabine in patients with pancreatic cancer. *Gastroenterology*. 2009;136(1):187-195.

CLINICAL AND TRANSLATIONAL MEDICINE

- 111. Kawada N, Uehara H, Katayama K, et al. Human equilibrative nucleoside transporter 1 level does not predict prognosis in pancreatic cancer patients treated with neoadjuvant chemoradiation including gemcitabine. *J Hepatobiliary Pancreat Sci.* 2012;19(6):717-722.
- 112. Poplin E, Wasan H, Rolfe L, et al. Randomized, multicenter, phase II study of CO-101 versus gemcitabine in patients with metastatic pancreatic ductal adenocarcinoma: including a prospective evaluation of the role of hENT1 in gemcitabine or CO-101 sensitivity. *J Clin Oncol.* 2013;31(35):4453-4461.
- 113. Ormanns S, Heinemann V, Raponi M, et al. Human equilibrative nucleoside transporter 1 is not predictive for gemcitabine efficacy in advanced pancreatic cancer: translational results from the AIO-PK0104 phase III study with the clone SP120 rabbit antibody. *Eur J Cancer*. 2014;50(11):1891-1899.
- 114. Tsujie M, Nakamori S, Nakahira S, et al. Human equilibrative nucleoside transporter 1, as a predictor of 5-fluorouracil resistance in human pancreatic cancer. *Anticancer Res.* 2007;27(4b):2241-2249.
- 115. Nishio R, Tsuchiya H, Yasui T, et al. Disrupted plasma membrane localization of equilibrative nucleoside transporter 2 in the chemoresistance of human pancreatic cells to gemcitabine (dFdCyd). *Cancer Sci.* 2011;102(3):622-629.
- 116. Endo Y, Obata T, Murata D, et al. Cellular localization and functional characterization of the equilibrative nucleoside transporters of antitumor nucleosides. *Cancer Sci.* 2007;98(10):1633-1637.
- 117. Liu M, Zhang Y, Yang J, et al. ZIP4 increases expression of transcription factor ZEB1 to promote integrin $\alpha 3\beta 1$ signaling and inhibit expression of the gemcitabine transporter ENT1 in pancreatic cancer cells. *Gastroenterology*. 2020;158(3):679-692. e1.
- 118. Al-Abdulla R, Perez-Silva L, Abete L, Romero MR, Briz O, Marin JJG. Unraveling 'The Cancer Genome Atlas' information on the role of SLC transporters in anticancer drug uptake. *Expert Rev Clin Pharmacol.* 2019;12(4):329-341.
- Nyquist MD, Prasad B, Mostaghel EA. Harnessing solute carrier transporters for precision oncology. *Molecules*. 2017;22(4):539.

- Lin L, Yee SW, Kim RB, Giacomini KM. SLC transporters as therapeutic targets: emerging opportunities. *Nat Rev Drug Discov*. 2015;14(8):543-560.
- 121. Zhang Y, Zhang Y, Sun K, Meng Z, Chen L. The SLC transporter in nutrient and metabolic sensing, regulation, and drug development. *J Mol Cell Biol.* 2019;11(1):1-13.
- 122. Wang S, Tsun ZY, Wolfson RL, et al. Lysosomal amino acid transporter SLC38A9 signals arginine sufficiency to mTORC1. *Science*. 2015;347(6218):188-194.
- Rebsamen M, Pochini L, Stasyk T, et al. SLC38A9 is a component of the lysosomal amino acid sensing machinery that controls mTORC1. *Nature*. 2015;519(7544):477-481.
- 124. Geiger R, Rieckmann JC, Wolf T, et al. L-arginine modulates T cell metabolism and enhances survival and anti-tumor activity. *Cell.* 2016;167(3):829-842.e13.
- 125. Liu Y, Liang X, Dong W, et al. Tumor-repopulating cells induce PD-1 expression in CD8(+) T cells by transferring kynurenine and AhR activation. *Cancer Cell*. 2018;33(3):480-494.e7.
- 126. Sinclair LV, Neyens D, Ramsay G, Taylor PM, Cantrell DA. Single cell analysis of kynurenine and System L amino acid transport in T cells. *Nat Commun.* 2018;9(1):1981.
- 127. Bian Y, Li W, Kremer DM, et al. Cancer SLC43A2 alters T cell methionine metabolism and histone methylation. *Nature*. 2020;585(7824):277-282.
- 128. Kobayashi Y, Ohshiro N, Sakai R, Ohbayashi M, Kohyama N, Yamamoto T. Transport mechanism and substrate specificity of human organic anion transporter 2 (hOat2 [SLC22A7]). J Pharm Pharmacol. 2005;57(5):573-578.

How to cite this article: Wu Z, Xu J, Liang C, et al. Emerging roles of the solute carrier family in pancreatic cancer. *Clin Transl Med.* 2021;11:e356. https://doi.org/10.1002/ctm2.356