



## Genomic Insight into Three Marine *Micromonospora* sp. Strains from the Gulf of California

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**ABSTRACT** Three actinomycete strains, designated BL1, BL4, and CV4, were isolated from sediment samples from the Gulf of California in 2009 together with nearly 300 other actinobacteria. Genome mining and analysis of their  $\sim$ 6.4-Mb sequences confirmed the bioprospecting potential of these three bacteria belonging to the genus *Micromonospora*.

The genus *Micromonospora*, class *Actinobacteria*, contained over 80 validly described species (1) at the time of writing. Although several species have terrestrial origin, they have also been associated with marine environments (2), sea sands (3), near-shore sediments (4), deep-sea sediments (5, 6), and marine sponges (7). *Micromonospora* spp. show well-developed, branched, substrate mycelium, with nonmotile spores and usually absent aerial mycelium, although there are recent reports opposing this view (8).

Micromonosporae show 16S rRNA similarity values within the range of 96.7% to 99% (2), making the description of putative novel species difficult without full-genome sequences. Micromonosporae have also been found to be a promising source of biosynthetic metabolites (9).

Strains BL1, BL4, and CV4 were part of a previous report on the isolation of almost 300 actinobacteria (10) from the Gulf of California and Gulf of Mexico. Single colonies were selected for DNA extraction from isolates cultured on ISP2 medium (11). Genomes were sequenced by MicrobesNG using the Illumina MiSeq sequencing platform. Genomic DNA libraries were prepared using a Nextera XT library prep kit (Illumina, San Diego, CA, USA). DNA quantification and library preparation were carried out on a Hamilton Microlab STAR automated liquid-handling system. Pooled libraries were quantified using the Kapa Biosystems library quantification kit for Illumina on a Roche LightCycler 96 quantitative PCR (qPCR) machine. Libraries were sequenced using a 250-bp paired-end protocol. The reads were trimmed using Trimmomatic v.0.38, and the quality was assessed using in-house scripts combined with the following software: SAMtools, BedTools, and BWA-MEM (12–15). *De novo* assembly was performed using SPAdes 3.1.1 using default parameters (16). Contigs were annotated with the NCBI Prokaryotic Genome Annotation Pipeline (17).

Genome characteristics are summed up in Table 1. EzTaxon analysis of the 16S rRNA (18) identified that the three species belonged to the genus *Micromonospora*; BL1 is related to *Micromonospora tulbaghiae* with 100% 16S rRNA gene similarity, while BL4 and CV4 are related to *Micromonospora coriariae*, both with 99.72% 16S rRNA gene similarity. It is worth noting that recent reviews of actinobacterial phylogeny (19) have proven that 16S rRNA phylogeny is not enough for a correct identification even of the genus in these closely related genera.

An average nucleotide identity (ANI) matrix was generated using different Mi-

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lsolate	Median insert size (bp)	Mean coverage (×)	Mean coverage excluding 0s (×)	No. of reads	No. of reads with insert size >300 bp	No. of contigs	N <sub>50</sub> value	Genome size (Mb)	G+C content (%)	No. of open reading frames	No. of tRNAs/ rRNAs
BL1	496	31.9427	31.9761	592,434	416,541	422	33,132	6.46	72.9	6,153	52/6
BL4	487	41.0063	41.0232	732,201	512,449	283	53,042	6.41	72.2	6,036	49/6
CV4	532	36.5438	36.5923	631,879	502,335	454	34,445	6.37	72.2	6,122	49/7

TABLE 1 Characteristics of the Micromonospora sp. isolate genomes

*cromonospora* species (20). The ANI value between BL4 and CV4 was 98.9%, suggesting that both isolates belong to the same species and its most related species is *M. coriariae* DSM 44875<sup>T</sup>, a bacterium isolated from the plant *Coriaria myrtifolia* (21), with an ANI of 93.2%, which is below the species delimitation percentage of 95%. This suggests that BL4 and CV4 may constitute a novel species (22). As for BL1, the ANI value with *M. tulbaghiae* DSM 45142<sup>T</sup> was 96.2%. This result, along with the EZTaxon identification, shows that BL1 should be identified as *M. tulbaghiae*, a species first isolated from leaves of the South African plant *Tulbaghia violacea* (23). The fact that these micromonosporae were recovered from marine sediments and that BL1 has been reported to produce aerial hyphae, an unusual feature on micromonosporae (8), could imply different ecological niches for *Micromonospora* spp. and roles in aquatic environments.

Genome mining using antiSMASH 3.0 (24) and the Web tool NaPDoS (25) predicted the production of bleomycin, lymphostin, phosphonoglycan, actinomycin, alnumycin, epothilone, spinosad, syringomycin, and sioxanthin biosynthetic clusters among the three genomes. The xantholipin cluster was found only in BL1 and BL4, alkyl-*O*-dihydrogeranyl-methoxyhydroquinone on BL1 and CV4, and desferrioxamine B on both BL4 and CV4. Exclusive clusters were azicemicin, leinamycin, lobosamide, nocathiacin, pentalenolactone, terramycin, curacin, tyrocidin, and oxazolomycin for BL1, ochronotic pigment for BL4, and pradimicin for CV4.

**Data availability.** This whole-genome shotgun project has been deposited in GenBank under the accession no. RCVK0000000, RCVJ00000000, and RCVI00000000 and in the Sequence Read Archive under the accession no. SRR8727741, SRR8727780 and SRR8727783. The versions described in this paper are the first versions, RCVK01000000, RCVJ01000000, and RCVI01000000.

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## REFERENCES

- Kuncharoen N, Kudo T, Ohkuma M, Tanasupawat S. 2018. *Micromonospora azadirachtae* sp. nov., isolated from roots of *Azadirachta indica* A. Antonie Van Leeuwenhoek 112:1–10. https://doi.org/10.1007/s10482 -018-1152-3.
- Genilloud O. 2015. *Micromonospora. In* Whitman WB, Rainey F, Kämpfer P, Trujillo M, Chun J, DeVos P, Hedlund B, Dedysh S (ed), Bergey's manual of systematics of Archaea and Bacteria. John Wiley & Sons, Inc., Hoboken, NJ. https://doi.org/10.1002/9781118960608.gbm00148.
- Tanasupawat S, Jongrungruangchok S, Kudo T. 2010. Micromonospora marina sp. nov., isolated from sea sand. Int J Syst Evol Microbiol 60: 648–652. https://doi.org/10.1099/ijs.0.014068-0.
- Phongsopitanun P, Kudo T, Mori M, Shiomi K, Pittayakhajonwut P, Suwanborirux K, Tanasupawat S. 2015. *Micromonospora fluostatini* sp. nov., isolated from marine sediment. Int J Syst Evol Microbiol 65:4417–4423. https://doi .org/10.1099/ijsem.0.000589.

- Weyland H. 1969. Actinomycetes in North Sea and Atlantic Ocean sediments. Nature 223:858. https://doi.org/10.1038/223858a0.
- 6. Weyland H. 1981. Distribution of actinomycetes on the sea floor. Zentrabl Bakteriol Suppl 11:185–193.
- Supong K, Suriyachadkun C, Pittayakhajonwut P, Suwanborirux K, Thawai C. 2013. *Micromonospora spongicola* sp. nov., an actinomycete isolated from a marine sponge in the gulf of Thailand. J Antibiot (Tokyo) 66:505–509. https://doi.org/10.1038/ja.2013.35.
- Maldonado LA, Quintana ET. 2015. Unexpected properties of micromonoporae from marine origin. Adv Microbiol 5:452–456. https://doi.org/ 10.4236/aim.2015.56046.
- Carro L, Nouioui I, Sangal V, Meier-Kolthoff JP, Trujillo ME, Montero-Calasanz M, Sahin N, Smith DL, Kim KE, Peluso P, Deshpande S, Woyke T, Shapiro N, Kyrpides NC, Klenk HP, Göker M, Goodfellow M. 2018. Genome-based classification of micromonosporae with a focus on their

biotechnological and ecological potential. Sci Rep 8:525. https://doi.org/ 10.1038/s41598-017-17392-0.

- Maldonado LA, Fragoso-Yáñez D, Pérez-García A, Rosellón-Druker J, Quintana ET. 2009. Actinobacterial diversity from marine sediments collected in Mexico. Antonie Van Leeuwenhoek 95:111–120. https://doi .org/10.1007/s10482-008-9294-3.
- 11. Shirling EB, Gottlieb D. 1966. Methods for characterization of *Streptomyces* species. Int J Syst Bacteriol 16:317–327.
- 12. Bolger AM, Lohse M, Usadel B. 2014. Trimmomatic: a flexible trimmer for Illumina sequence data. Bioinformatics 30:2114–2120. https://doi.org/10 .1093/bioinformatics/btu170.
- Li H, Handsaker B, Wysoker A, Fennell T, Ruan J, Homer N, Marth G, Abecasis G, Durbin R, 1000 Genome Project Data Processing Subgroup. 2009. The sequence alignment/map format and SAMtools. Bioinformatics 25:2078–2079. https://doi.org/10.1093/bioinformatics/btp352.
- Quinlan AR, Hall IM. 2010. BEDTools: a flexible suite of utilities for comparing genomic features. Bioinformatics 26:841–842. https://doi .org/10.1093/bioinformatics/btq033.
- 15. Li H, Durbin R. 2009. Fast and accurate short read alignment with Burrows-Wheeler Transform. Bioinformatics 25:1754–1760. https://doi .org/10.1093/bioinformatics/btp324.
- Bankevich A, Nurk S, Antipov D, Gurevich AA, Dvorkin M, Kulikov AS, Lesin VM, Nikolenko SI, Pham S, Prjibelski AD, Pyshkin AV, Sirotkin AV, Vyahhi N, Tesler G, Alekseyev MA, Pevzner PA. 2012. SPAdes: a new genome assembly algorithm and its applications to single-cell sequencing. J Comput Biol 19:455–477. https://doi.org/10.1089/cmb.2012.0021.
- Tatusova T, DiCuccio M, Badretdin A, Chetvernin V, Ciufo S, Li W. 2013. Prokaryotic Genome Annotation Pipeline. *In* Beck J, Benson D, Coleman J, Hoeppner M, Johnson M, Maglott M, Mizrachi I, Morris R, Ostell J, Pruitt K, Rubinstein W, Sayers E, Sirotkin K, Tatusova T (ed), The NCBI handbook, 2nd ed. NCBI, Bethesda, MD.
- 18. Yoon SH, Ha SM, Kwon S, Lim J, Kim Y, Seo H, Chun J. 2017. Introducing

EzBioCloud: a taxonomically united database of 16S rRNA and whole genome assemblies. Int J Syst Evol Microbiol 67:1613–1617. https://doi .org/10.1099/ijsem.0.001755.

- Nouioui I, Carro L, García-López M, Meier-Kolthoff JP, Woyke T, Kyrpides NC, Pukall R, Klenk HP, Goodfellow M, Göker M. 2018. Genome-based taxonomic classification of the phylum *Actinobacteria*. Front Microbiol 9:2007. https://doi.org/10.3389/fmicb.2018.02007.
- Rodriguez-R LM, Konstantinidis KT. 2016. The enveomics collection: a toolbox for specialized analyses of microbial genomes and metagenomes. PeerJ Prepr 4:e1900v1. https://peerj.com/preprints/1900/.
- Trujillo ME, Kroppenstedt RM, Schumann P, Carro L, Martínez-Molina E. 2006. *Micromonospora coriariae* sp. nov., isolated from root nodules of *Coriaria myrtifolia*. Int J Syst Evol Microbiol 56:2381–2385. https://doi .org/10.1099/ijs.0.64449-0.
- Chun J, Oren A, Ventosa A, Christensen H, Arahal DR, da Costa MS, Rooney AP, Yi H, Xu XW, De Meyer S, Trujillo ME. 2018. Proposed minimal standards for the use of genome data for the taxonomy of prokaryotes. Int J Syst Evol Microbiol 68:461–466. https://doi.org/10.1099/ijsem.0 .002516.
- Kirby BM, Meyers PR. 2010. *Micromonospora tulbaghiae* sp. nov., isolated from the leaves of wild garlic, *Tulbaghia violacea*. Int J Syst Evol Microbiol 60:1328–1333. https://doi.org/10.1099/ijs.0.013243-0.
- Weber T, Blin K, Duddela S, Krug D, Kim HU, Bruccoleri R, Lee SY, Fischbach MA, Müller R, Wohlleben W, Breitling R, Takano E, Medema MH. 2015. antiSMASH 3.0—a comprehensive resource for the genome mining of biosynthetic gene clusters. Nucleic Acids Res 43:W237–243. https://doi.org/10.1093/nar/gkv437.
- Ziemert N, Podell S, Penn K, Badger JH, Allen E, Jensen PR. 2012. The natural product domain seeker NaPDoS: a phylogeny based bioinformatic tool to classify secondary metabolite gene diversity. PLoS One 7:e34064. https://doi.org/10.1371/journal.pone.0034064.