

Genetic diversity in small cell lung carcinoma

Takuo Hayashi^{1,2,3}^

¹Department of Human Pathology, Juntendo University Graduate School of Medicine, Bunkyo-ku, Tokyo, Japan; ²Diagnostic Pathology Center, Juntendo Hospital, Bunkyo-ku, Tokyo, Japan; ³Bioresource Research Center, Juntendo University School of Medicine, Bunkyo-ku, Tokyo, Japan *Correspondence to:* Takuo Hayashi, MD, PhD. Department of Human Pathology, Juntendo University Graduate School of Medicine, 2-1-1, Hongo, Bunkyo-ku, Tokyo 113-8421, Japan; Diagnostic Pathology Center, Juntendo Hospital, Bunkyo-ku, Tokyo 113-8421, Japan; Bioresource Research Center, Juntendo University School of Medicine, Bunkyo-ku, Tokyo 113-8421, Japan. Email: tkhyz@juntendo.ac.jp. *Comment on:* Sivakumar S, Moore JA, Montesion M, *et al.* Integrative Analysis of a Large Real-World Cohort of Small Cell Lung Cancer Identifies Distinct Genetic Subtypes and Insights into Histologic Transformation. Cancer Discov 2023;13:1572-91.

Keywords: Human papillomavirus (HPV); PTEN; small cell carcinoma; STK11; transformation

Submitted Jan 13, 2024. Accepted for publication Apr 22, 2024. Published online May 17, 2024. doi: 10.21037/tlcr-24-40 **View this article at:** https://dx.doi.org/10.21037/tlcr-24-40

The development of next-generation sequencing (NGS) technology has led to many pivotal advancements in oncology. In particular, recurrent targetable alterations have been identified in non-small cell lung carcinoma (NSCLC), such as mutations in EGFR, BRAF, ERBB2, KRAS, rearranged ALK, ROS1, RET, NTRK1, and MET exon 14 skipping (1,2). The discovery of such actionable alterations in NSCLC has revolutionized the treatment of patients with NSCLCs. In contrast to NSCLC, small cell lung cancer (SCLC) was previously believed to be molecularly homogenous due to almost universal mutations in TP53 and RB1 (3,4). Over the last four decades, minimal changes have occurred in the therapy and survival outcomes of SCLC. In 2021, a transcriptome analysis reported that SCLC can be categorized into subgroups based on transcrriptome profiling (5). Three of the four subgroups are enriched in the predominant expression of specific transcription factors, namely ASCL1 (SCLC-A), NEUROD1 (SCLC-N), and POU2F3 (SCLC-P), whereas the fourth is an inflamed subgroup (SCLC-I) associated with lack of expression of these transcription factors and higher levels of checkpoint proteins and interferon signaling (5). YAP1 was initially proposed to define a distinct subgroup; however, it was found to be absent or expressed only at low levels in tumors (5,6). Pathogenic mutations in SMARCA4 were identified in six of eight YAP1-expressing SCLC (SCLC-Y) cell lines

and correlated with reduced SMARCA4 RNA/protein expression, indicating that the characteristics of SCLC-Y are consistent with SMARCA4-deficient undifferentiated tumors rather than SCLC (7). The clinical implications of this transcriptional profiling are significant because each subgroup exhibits a unique susceptibility toward investigational therapies: SCLC-A, SCLC-N, and SCLC-P are sensitive to BCL-2, aurora kinase, and PARP inhibitors, respectively (5). Furthermore, patients with SCLC without the expression of these three transcription factors may benefit from immune checkpoint blockade (5). This new transcriptional profiling of SCLC has been validated at the protein level by immunohistochemistry in several studies (8-10). Nevertheless, the clinical significance of the molecular classification of SCLC based on mRNA profiling is not well defined, and the mechanisms underlying the effects of specific genetic alterations on the transcriptional landscape are yet to be elucidated.

Recently, Sivakumar *et al.* reported a genetic analysis of 3,600 patients with SCLC whose tissue samples were mainly obtained from community sites throughout the United States and submitted to Foundation Medicine, Inc. (11). This report presents several key observations related to SCLC pathobiology that may open novel therapeutic avenues for patients with SCLC. Firstly, approximately 5.5% of SCLC cases were identified as *TP53/RB1* wild-type tumors in the

[^] ORCID: 0000-0002-8544-9370.

largest cohort study of patients with SCLC. While this finding is not surprising, alterations in genes that regulate RB and p53, such as CDKN2A, which codes for p16, a positive regulator of RB, CCND1, which codes for cyclin D1, a negative regulator of RB, and MDM2, a negative regulator of p53, were frequently identified in TP53/RB1 wild-type tumors. Additionally, human papillomavirus (HPV) was also identified in 12.7% of TP53/RB1 wild-type tumors. The p53 and RB proteins are well-characterized targets of the HPV E6 and E7 oncoproteins (12). HPV E7 proteins subvert G1-S arrest and induce hyperproliferation through the inhibition of RB family members and constitutive activation of E2F-responsive genes. Co-expression of HPV E6 with E7 abrogates p53-dependent apoptosis in response to the activities of E7, allowing replication in the presence of DNA damage and increased chromosomal instability (12). Thus, HPV+TP53/RB1 wild-type tumors are a distinct genetic subtype that may be uniquely responsive to strategies targeting HPV or reactivating p53 function to induce cell death in HPV-positive cancer cells. Basal cells in the bronchial epithelium can be infected with certain HPV strains (13); however, the lungs are not a common organ for HPV infection. Notably, some HPV-positive patients with NSCLCs (14) and SCLCs (15) may experience metastases due to the presence of HPV-positive tumors at other body sites. Thus, further analyses with detailed clinicopathological information are needed to elucidate the incidence of HPV infection in patients with SCLC and NSCLC. Second, driver mutations of receptor tyrosine kinase genes, including EGFR mutations (n=107) and ALK (n=5), ROS1 (n=3), RET (n=5), and NTRK1 (n=1) fusions, have been detected in SCLC (11). Subsequent genomic analysis of 41 patients with paired NSCLC and SCLC samples showed that NSCLC and SCLC tumor samples shared driver mutations in approximately 61% of cases. However, in approximately 17% of patients, NSCLC and SCLC tumor samples shared alterations, but no previously described driver mutations were detected in either biopsy. Therefore, these patients cannot be treated with tyrosine kinase inhibitors, suggesting that other treatment modalities, such as chemotherapy and immunotherapy, can also drive transformation (11). Additionally, a smokingassociated mutational signature is rare in TP53/RB1 wildtype tumors (11). Frequently altered genes in NSCLC, such as KRAS, BRAF, FGFR1, and KEAP1, were enriched in TP53/RB1 wild-type tumors in which a smokingassociated mutational signature is rare. Patients with STK11 mutations were enriched for mutations in genes associated

with NSCLC development, such as *KRAS* and *KEAP1*. These findings suggest that several of these tumors may have originated from NSCLC (11). Finally, the mutation spectrum of different sites of metastases showed that brain metastases were enriched for *PTEN* alterations compared with primary lung tumors, indicating that the PTEN pathway can play a unique role in SCLC brain metastasis (11).

Transformation of *EGFR*-mutated adenocarcinoma to SCLC in the case of acquired resistance to tyrosine kinase inhibitors has been observed in approximately 3–10% of patients (16). However, the results of a study by Sivakumar *et al.* suggested that SCLC transformation may occur across multiple distinct molecular cohorts of NSCLC (11). Populations of different ancestries are enriched for different genetic subtypes of SCLC such as *EGFR* mutations in patients with Asian ancestry and *STK11* mutations in patients with African ancestry (11). Molecular profiling, along with NGS, using formalin-fixed paraffinembedded specimens has become essential for identifying predictive biomarkers for personalized therapy in advanced or metastatic NSCLC. The clinical application of NGS in patients with SCLC should be considered in future studies.

The major limitation of the study by Sivakumar et al. is the absence of RNA/protein expression profiles analyzed for SCLC. NSCLCs have emerged as successful examples of cancers that have been molecularly redefined through the discovery of actionable mutations. In addition to such genetic changes, differences in the pemetrexed response (17,18) and the efficacy of immune checkpoint inhibitors (19) have been reported in EGFR/ALK-negative lung adenocarcinoma with differential thyroid transcription factor 1 (TTF-1) expression, suggesting that more accurate and reliable treatments can be achieved by acquiring integrated genomic and transcriptomic information in NSCLC. Similarly, combined genomic analyses with transcriptome data of key transcription factors, including ASCL1, NEUROD1, and POU2F3, may be more efficient for patient stratification in SCLC. Another limitation of the study by Sivakumar et al. is the absence of a central pathological review because tissue samples with initial diagnosis were submitted by various practicing physicians throughout the United States. Notably, 1.5% of cases harbored inactivating mutations in SMARCA4, and these tumors should have been diagnosed as SMARCA4-deficient undifferentiated tumors. Nevertheless, before recognition of SMARCA4-deficient undifferentiated tumors as a distinct entity, these tumors were commonly categorized as neuroendocrine carcinomas in patient samples (20,21). Furthermore, the incidence

Translational Lung Cancer Research, Vol 13, No 5 May 2024

of combined SCLC in 3,600 patients with SCLC was unclear. Combined SCLC is defined as SCLC combined with elements of NSCLC within the primary tumor in which spontaneous transformation from NSCLC to SCLC may occur. Pathological analysis of surgically resected, treatment-naïve SCLC revealed that combined SCLC with adenocarcinoma accounts for approximately 9% of cases of SCLC (22). Another study showed that combined SCLC with adenocarcinoma, squamous cell carcinoma, and spindle cell carcinoma account for approximately 6.5%, 12%, and 1% of cases of SCLC, respectively (9). The driver mutations in the cohort studied by Sivakumar et al. were recurrently detected because some SCLCs with driver mutations may combine SCLCs with driver mutation-positive NSCLC components. The incidence of combined SCLC in realworld data remains largely unknown because combined SCLC is usually diagnosed in surgical specimens and not biopsy specimens. To address this limitation, a radiomic or radiogenomic approach can be useful to evaluate tumor heterogeneity in SCLC and differentiate pure SCLC and combined SCLC (23).

Subtyping of SCLC has been attempted many times since the first morphological classification in 1967 (24). In the era of precision medicine, novel subclassification of SCLC based on tissue-based biomarker assessment that can improve treatment strategies is required. Sivakumar et al. revealed the genetic diversity in SCLC. Although the number of patients with TP53/RB1 wild tumors (5%) and transformed SCLC (4%) was limited, their tumors were genetically distinct from TP53/RB1 mutated SCLC (11). Genetically, SCLC is categorized into de novo SCLC and transformed or combined SCLC. De novo SCLC is further categorized into TP53/RB1 mutated type SCLC and TP53/RB1 wild type SCLC. To validate the transcriptional profiling of SCLC and its therapeutic possibilities in BCL-2, aurora kinase, PARP inhibitors, and immunotherapy, future clinical trials should be planned for every genetically diverse group. Even when the ideal classification of SCLC is established, the limitation of access to tissue as diagnostic specimens may remain. Liquid biopsy can be used to overcome these limitations. Notably, a recent study showed that DNA methylation from tissues and liquid biopsy samples allows a subgroup of SCLC with ASCL1, NEUROD1, and POU2F3 (25). In addition to genomic and transcriptomic approaches, the ability of epigenomic approaches to predict pharmacological responses to drugs has the potential to revolutionize the subclassification of SCLC.

Acknowledgments

Funding: None.

Footnote

Provenance and Peer Review: This article was commissioned by the editorial office, *Translational Lung Cancer Research*. The article has undergone external peer review.

Peer Review File: Available at https://tlcr.amegroups.com/ article/view/10.21037/tlcr-24-40/prf

Conflicts of Interest: The author has completed the ICMJE uniform disclosure form (available at https://tlcr.amegroups. com/article/view/10.21037/tlcr-24-40/coif). The author has no conflicts of interest to declare.

Ethical Statement: The author is accountable for all aspects of the work in ensuring that questions related to the accuracy or integrity of any part of the work are appropriately investigated and resolved.

Open Access Statement: This is an Open Access article distributed in accordance with the Creative Commons Attribution-NonCommercial-NoDerivs 4.0 International License (CC BY-NC-ND 4.0), which permits the non-commercial replication and distribution of the article with the strict proviso that no changes or edits are made and the original work is properly cited (including links to both the formal publication through the relevant DOI and the license). See: https://creativecommons.org/licenses/by-nc-nd/4.0/.

References

- Hirsch FR, Suda K, Wiens J, et al. New and emerging targeted treatments in advanced non-small-cell lung cancer. Lancet 2016;388:1012-24.
- Jordan EJ, Kim HR, Arcila ME, et al. Prospective Comprehensive Molecular Characterization of Lung Adenocarcinomas for Efficient Patient Matching to Approved and Emerging Therapies. Cancer Discov 2017;7:596-609.
- 3. George J, Lim JS, Jang SJ, et al. Comprehensive genomic profiles of small cell lung cancer. Nature 2015;524:47-53.
- Mahadevan NR, Knelson EH, Wolff JO, et al. Intrinsic Immunogenicity of Small Cell Lung Carcinoma Revealed by Its Cellular Plasticity. Cancer Discov 2021;11:1952-69.

- Gay CM, Stewart CA, Park EM, et al. Patterns of transcription factor programs and immune pathway activation define four major subtypes of SCLC with distinct therapeutic vulnerabilities. Cancer Cell 2021;39:346-360.e7.
- Baine MK, Hsieh MS, Lai WV, et al. SCLC Subtypes Defined by ASCL1, NEUROD1, POU2F3, and YAP1: A Comprehensive Immunohistochemical and Histopathologic Characterization. J Thorac Oncol 2020;15:1823-35.
- Ng J, Cai L, Girard L, et al. Molecular and Pathologic Characterization of YAP1-Expressing Small Cell Lung Cancer Cell Lines Leads to Reclassification as SMARCA4-Deficient Malignancies. Clin Cancer Res 2024;30:1846-58.
- Qu S, Fetsch P, Thomas A, et al. Molecular Subtypes of Primary SCLC Tumors and Their Associations With Neuroendocrine and Therapeutic Markers. J Thorac Oncol 2022;17:141-53.
- Handa T, Hayashi T, Ura A, et al. Comparison of ASCL1, NEUROD1, and POU2F3 expression in surgically resected specimens, paired tissue microarrays, and lymph node metastases in small cell lung carcinoma. Histopathology 2023;82:860-9.
- Megyesfalvi Z, Barany N, Lantos A, et al. Expression patterns and prognostic relevance of subtype-specific transcription factors in surgically resected small-cell lung cancer: an international multicenter study. J Pathol 2022;257:674-86.
- Sivakumar S, Moore JA, Montesion M, et al. Integrative Analysis of a Large Real-World Cohort of Small Cell Lung Cancer Identifies Distinct Genetic Subtypes and Insights into Histologic Transformation. Cancer Discov 2023;13:1572-91.
- Moody CA, Laimins LA. Human papillomavirus oncoproteins: pathways to transformation. Nat Rev Cancer 2010;10:550-60.
- Fortes HR, von Ranke FM, Escuissato DL, et al. Recurrent respiratory papillomatosis: A state-of-the-art review. Respir Med 2017;126:116-21.
- Yanagawa N, Wang A, Kohler D, et al. Human papilloma virus genome is rare in North American non-small cell lung carcinoma patients. Lung Cancer 2013;79:215-20.
- 15. Bishop JA, Ogawa T, Chang X, et al. HPV analysis in distinguishing second primary tumors from lung metastases in patients with head and neck squamous cell carcinoma. Am J Surg Pathol 2012;36:142-8.
- Marcoux N, Gettinger SN, O'Kane G, et al. EGFR-Mutant Adenocarcinomas That Transform to Small-Cell

Lung Cancer and Other Neuroendocrine Carcinomas: Clinical Outcomes. J Clin Oncol 2019;37:278-85.

- 17. Sun JM, Han J, Ahn JS, et al. Significance of thymidylate synthase and thyroid transcription factor 1 expression in patients with nonsquamous non-small cell lung cancer treated with pemetrexed-based chemotherapy. J Thorac Oncol 2011;6:1392-9.
- Frost N, Zhamurashvili T, von Laffert M, et al. Pemetrexed-Based Chemotherapy Is Inferior to Pemetrexed-Free Regimens in Thyroid Transcription Factor 1 (TTF-1)-Negative, EGFR/ALK-Negative Lung Adenocarcinoma: A Propensity Score Matched Pairs Analysis. Clin Lung Cancer 2020;21:e607-21.
- Iso H, Hisakane K, Mikami E, et al. Thyroid transcription factor-1 (TTF-1) expression and the efficacy of combination therapy with immune checkpoint inhibitors and cytotoxic chemotherapy in non-squamous non-small cell lung cancer. Transl Lung Cancer Res 2023;12:1850-61.
- Rekhtman N, Montecalvo J, Chang JC, et al. SMARCA4-Deficient Thoracic Sarcomatoid Tumors Represent Primarily Smoking-Related Undifferentiated Carcinomas Rather Than Primary Thoracic Sarcomas. J Thorac Oncol 2020;15:231-47.
- Rekhtman N. Lung neuroendocrine neoplasms: recent progress and persistent challenges. Mod Pathol 2022;35:36-50.
- 22. Nicholson SA, Beasley MB, Brambilla E, et al. Small cell lung carcinoma (SCLC): a clinicopathologic study of 100 cases with surgical specimens. Am J Surg Pathol 2002;26:1184-97.
- Chen BT, Chen Z, Ye N, et al. Differentiating Peripherally-Located Small Cell Lung Cancer From Nonsmall Cell Lung Cancer Using a CT Radiomic Approach. Front Oncol 2020;10:593.
- Kreyberg L. Histological lung cancer types. A morphological and biological correlation. Acta Pathol Microbiol Scand Suppl 1962;Suppl 157:1-92.
- Heeke S, Gay CM, Estecio MR, et al. Tumor- and circulating-free DNA methylation identifies clinically relevant small cell lung cancer subtypes. Cancer Cell 2024;42:225-237.e5.

Cite this article as: Hayashi T. Genetic diversity in small cell lung carcinoma. Transl Lung Cancer Res 2024;13(5):1169-1172. doi: 10.21037/tlcr-24-40