

GOPEN ACCESS

Citation: Del Carro KB, Leite GR, de Oliveira Filho AG, dos Santos CB, de Souza Pinto I, Fux B, et al. (2020) Assessing geographic and climatic variables to predict the potential distribution of the visceral leishmaniasis vector *Lutzomyia longipalpis* in the state of Espírito Santo, Brazil. PLoS ONE 15 (9): e0238198. https://doi.org/10.1371/journal. pone.0238198

Editor: Abdallah M. Samy, Faculty of Science, Ain Shams University (ASU), EGYPT

Received: February 28, 2020

Accepted: August 11, 2020

Published: September 18, 2020

Copyright: © 2020 Del Carro et al. This is an open access article distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

Data Availability Statement: Collection data is deposited in the Harvard Dataverse (DOI: <u>https://</u>doi.org/10.7910/DVN/0JYIXS).

Funding: The authors received no specific funding for this work.

Competing interests: The authors have declared that no competing interests exist.

RESEARCH ARTICLE

Assessing geographic and climatic variables to predict the potential distribution of the visceral leishmaniasis vector *Lutzomyia longipalpis* in the state of Espírito Santo, Brazil

Karina Bertazo Del Carro^{1*}, Gustavo Rocha Leite¹, Amandio Gonçalves de Oliveira Filho², Claudiney Biral dos Santos^{1,3}, Israel de Souza Pinto¹, Blima Fux¹, Aloísio Falqueto^{1¤}

1 Tropical Medicine Unit, Federal University of Espírito Santo, Vitória, Espírito Santo, Brazil, 2 Department of Civil Engineering, Federal University of Espírito Santo, Vitória, Espírito Santo, Brazil, 3 Center for Entomology and Malacology, Espírito Santo State Health Department, Serra, Espírito Santo, Brazil

¤ Current address: Departamento de Patologia, Universidade Federal do Espírito Santo, Vitória, Espírito Santo, Brazil

* bertazokarina@gmail.com

Abstract

Visceral leishmaniasis (VL) is an infectious disease caused by the protozoa Leishmania chagasi, whose main vector in South America is Lutzomyia longipalpis. The disease was diagnosed in the Brazilian state of Espírito Santo (ES) for the first time in 1968. Currently, this disease has been considered endemic in 10 municipalities. Furthermore, the presence of L. longipalpis has been detected in eight other municipalities where the transmission has not been reported thus far. In this study, we performed species distribution modeling (SDM) to identify new and most likely receptive areas for VL transmission in ES. The sandflies were both actively and passively collected in various rural area of ES between 1986 and 2017. The collection points were georeferenced using a global positioning system device. Climatic data were retrieved from the WorldClim database, whereas geographic data were obtained from the National Institute for Space Research and the Integrated System of Geospatial Bases of the State of Espírito Santo. The maximum entropy algorithm was used through the MIAmaxent R package to train and test the distribution models for L. longipalpis. The major contributor to model generation was rocky outcrops, followed by temperature seasonality. The SDM predicted the expansion of the L. longipalpis-prone area in the Doce River Valley and limited the probability of expanding outside its watershed. Once the areas predicted suitable for L. longipalpis occurrence are determined, we can avoid the inefficient use of public resources in conducting canine serological surveys where the vector insect does not occur.

Introduction

Visceral leishmaniasis (VL), also known as kala-azar, is a chronic infectious disease with high mortality if not properly treated [1]. In Latin America, the etiological agent of VL *Leishmania*

infantum is transmitted by mosquitoes of the Phlebotominae subfamily, with *Lutzomyia longipalpis* (Lutz & Neiva, 1912) as the main vector [1]. In the state of Espírito Santo (ES), located in southeastern Brazil, the disease was first observed in the Doce River Valley in 1966. The first five cases of VL were diagnosed in the municipalities of Baixo Guandu and Colatina [2]. Currently, the disease affects 10 of the 78 municipalities; all the 10 municipalities are located in the central-west and northwest regions of ES [3]. However, the presence of *L. longipalpis* has been detected in eight other municipalities where the transmission has not been reported thus far [4]. These regions are characterized by sloped landscapes, warm weather, and low precipitation [3, 5].

The environment affects the composition, distribution, and behavior of the vectors and reservoirs involved in the transmission of VL, which impacts the epidemiology of the disease [6]. Therefore, since the transmission cycle of VL depends on an insect vector, the environmental characteristics that favor its occurrence must be investigated and identified.

In Brazil, with respect to canines as a potential reservoir of the disease, the frequent movement of people implies a lack of control in the transport of infected dogs from one region to another, resulting in an improper use of resources for investigating canine disease in areas where the VL vector does not exist [7]. Hence, the presence of the vector must be investigated before performing any other actions on the canine reservoir.

Species distribution modeling (SDM) is a technique that uses machine learning algorithms to predict the distribution of a species across geographic space and time [8]. This approach identifies regions containing suitable environmental conditions based on habitat characteristics at locations of known species occurrences [9]. Hence, the use of these tools to identify suitable areas for a disease vector occurrence would be instrumental in directing efforts toward regions that truly need it. SDMs can reveal areas with underestimated vector populations where disease transmission is unknown but can potentially occur [10–12].

Considering that the transmission of VL is through an insect vector, whose frequency and distribution are affected by environmental variables, geographic information coupled with SDM tools can be used to identify factors associated with the presence of *L. longipalpis* in ES. Therefore, predictive spatial modeling can help identify new and potential receptive areas for the transmission of VL in ES.

Materials and methods

Study area

ES is located on the Atlantic coast of southeastern Brazil, between parallels $17^{\circ}53'29''-21^{\circ}$ 18'03"S and meridians $39^{\circ}41'18''-41^{\circ}52'45''W$. It has an area of approximately 46,000 km², with a maximum north-south extension of 374 km and a width of 130–150 km [13]. ES accounts for 0.5% of the Brazilian territory and comprises 78 municipalities.

The ES territory is longitudinally intersected by the Serra do Mar, a 1,500-km long system of mountain ranges and escarpments. The elevation reaches up to 2,900 m above sea level in Pico da Bandeira, Serra do Caparaó. It has a tropical climate, with an average annual mean temperature of 22 °C (9–24 °C) and average annual precipitation of 1,200 mm (900–1,600 mm) [14]. It also has extensive flat areas along its coast, with a wide variety of ecosystems determined by their natural geographical features [13].

The state encompasses the southern part of the Central Corridor of the Atlantic Forest, which comprises one of the primary areas of ombrophilous dense forest within this biome. There is a high level of endemism and species diversity in this region, which continues to be threatened by human activities [15].

Entomological data

Phlebotomine occurrence data were obtained from samples collected from several localities in rural areas of the 78 municipalities of ES between 1986 and 2017. The phlebotomines were collected in the first three hours after sunset by active searching with a manual suction tube (Castro collector) and by passive capture using light-based traps. The active search for sandflies was performed within 30 m from the human houses, collecting resting insects on the inner and outer walls of the houses and their annexes, animal shelters, tree trunks, and Shannon traps. Passive collections were performed simultaneously using the Centers for Disease Control and Prevention traps installed in wild areas. Experienced technicians performed the sampling, regardless of the occurrence of any disease outbreak. The sampled specimens were identified according to the identification keys reported by Galati (2003) and Young and Duncan (1994) [16, 17]. All sampled localities were georeferenced using a global positioning system device.

The sampled localities where *L. longipalpis* was not found were considered absence points for one of the model evaluation techniques.

To reduce the sampling bias of the presence and absence occurrence points of *L. longipalpis*, we spatially rarefied the occurrence data using SDM Toolbox v2.4 extension for ArcGis. This tool removes spatially autocorrelated occurrence points by reducing multiple occurrence records to a single record within a specified distance.

Environmental variables

The environmental variables used for the modeling included 19 bioclimatic variables and three topographic variables (elevation, terrain slope, and rocky outcrops).

The 19 bioclimatic variables, derived from temperature and precipitation, were obtained from the WorldClim database (version 2.1, released in January 2020, https://www.worldclim. org/). The climatic data for the creation of these bioclimatic variables were generated through interpolations of climate data obtained from approximately 50,000 weather stations distributed worldwide from 1970 to 2000 [14].

The topographic variables—elevation and terrain slope—were retrieved from the Brazilian Geomorphometric Database (TOPODATA, *Banco de Dados Geomorfométricos do Brasil*, http://www.dsr.inpe.br/topodata/) of the National Institute for Space Research (INPE, *Instituto Nacional de Pesquisas Espaciais*). Data from the Shuttle Radar Topography Mission from the US Geological Survey were used to generate these two topographic variables.

The variable rocky outcrops was acquired from the Integrated System of Geospatial Bases of the State of Espírito Santo (GEOBASES, *Sistema Integrado de Bases Geoespaciais do Estado do Espírito Santo*, https://geobases.es.gov.br/). Mapping of the rocky outcrops was performed on an orthomosaic using photointerpretation analysis and manual vectoring procedures. For better applicability in this study, this variable was transformed from categorical to continuous using the kernel density estimation with a bandwidth of 0.1°.

All variables were raster datasets, with a resolution of 30 arc seconds, which is approximately equivalent to 1 km.

The Geographic Information System ArcGIS version 10.8 (ESRI, Redlands, CA, USA) with the World Geodetic System (WGS84) datum were used for the spatial procedures of this study.

Species distribution modeling

SDM uses machine learning computer algorithms to predict the distribution of a species using its known occurrence data and environmental variables [18]. Maximum entropy fitting is a general-purpose machine learning approach, which uses a mathematical formulation for modeling species distributions with presence-only data [18, 19]. This algorithm is based on the

principle of maximum entropy, a method used to choose the best probability distribution that fits the study data [18, 20].

We used the MIAmaxent R package v1.1.1, running under R version 4.0.0, for the modeling procedures of this study. MIAmaxent is a presence-only modeling approach that provides a statistical method to model species distributions similar to Maxent's, but it has subset selection instead of LASSO regularization. The simpler models typically produced by subset selection are ecologically more interpretable, making distribution models more grounded in ecological theory [21]. The package also executes a variable transformation based on expected occurrence–environment relationships [22].

We split the occurrence data into training and test sets for model evaluation. The final model, however, was trained using all the occurrence records and replicated 100 times, randomizing only the background points.

Model evaluation

For model evaluation, we used two types of train-test splits—(1) random cross-validation, which splits the data randomly into 75% for training the model and 25% for testing the model, and (2) spatially stratified cross-validation, which partitions data according to the latitude and longitude lines that divide the occurrence localities into four bins of equal numbers. Both occurrence and background localities are assigned to each of the four bins based on their position concerning these lines.

Spatially stratified cross-validation is an alternative method of dealing with spatial autocorrelation, which is very likely to inflate performance in random cross-validation [23].

To account for the out-of-sample prediction error in the random cross-validation, we replicated the process 100 times. Spatially stratified cross-validation was performed using the "block" method, available in the ENMeval R package, that conducts spatially independent evaluations and estimates optimal model complexity for SDM [24].

Each of the two train-test splits described above was applied for model evaluation using random background points as species pseudo-absence and using real localities defined as species absence.

Then, we used the receiver operating characteristic (ROC) plot method. An ROC plot is created by plotting the sensitivity values (the true-positive fraction) against the 1 –specificity (the false-positive fraction) for all available probability thresholds [25]. The area under the curve (AUC) derived from the ROC plot can be interpreted as a measure of the ability of the algorithm to discriminate between a suitable environmental condition and a random analysis pixel [18]. The closer the AUC value is to 1.0, the better the performance of the model. For this study, we used the interpretation by Hosmer and Lemeshow (2000), which considers AUC values of 0.5–0.6 (no discrimination), 0.6–0.7 (discrimination), 0.7–0.8 (acceptable model), 0.8–0.9 (excellent model), and 0.9–1.0 (outstanding discrimination) [26].

To minimize problems with this measure, we constrained our analysis to a well-studied region, averaged the AUC from several replicates of the original presence data, and used the real absence data [27, 28].

The predictive contribution of each variable for the final model was obtained using the fraction of total variation explained [29]. This is represented by the increase in the fraction of deviance explained when a given variable is added to the model [12].

Results

During the three decades of sandfly collection in ES, more than 13,600 specimens of *L. longipalpis* were collected from 82 unique localities throughout the state. Although the sampling



Fig 1. Sampling localities of phlebotomine sand flies in the state of Espírito Santo, southeastern Brazil, during the **period between 1986** and 2017. Green dots represent the localities where *L. longipalpis* was collected, and red dots the localities where other phlebotomine species were collected but not *L. longipalpis*.

effort covered the entire state, the species were only recorded in 18 of the 78 municipalities. These collection data revealed 78 presence and 262 absence points for *L. longipalpis* after the application of the spatially rarefying method (Fig 1).

The model evaluation for (1) random cross-validation using background points revealed a mean AUC of 0.91 (n = 100; IC95 = 0.006); (2) random cross-validation using the species



absences revealed a mean AUC of 0.83 (n = 100; IC95 = 0.007); (3) spatially stratified cross-validation using background points revealed a median AUC of 0.88 (n = 4; range = 0.85–0.95); and (4) spatially stratified cross-validation using the species absences revealed a median AUC of 0.82 (n = 4; range = 0.68–0.83). We used median and range for the spatially stratified crossvalidation AUC representation because they are more appropriate measures to deal with only four bins of the block method area stratification (Fig 2).

The fraction of total variation explained, used for accounting the contribution of each variable in the final model, indicated the following variable contributions: (1) a rocky outcrop density mean of 66.79% (n = 100; 95% confidence interval [CI] = 0.41); a temperature seasonality (BIO4) mean of 19.97% (n = 100; 95% CI = 0.31), a temperature annual range (BIO7) mean of 8.38% (n = 99; 95% CI = 0.31); and a precipitation of the wettest quarter (BIO16) mean of 4.06% (n = 72; 95% CI = 0.18).

The frequency of the observed presence plot shows how commonly *L. longipalpis* occurs across the range of the environmental variables kept in the final model, making it possible to recognize patterns in the observed frequency of occurrence (Fig 3).

The final model projected on the map indicates areas predicted suitable for *L. longipalpis* throughout ES. The predicted areas are consistent with the current knowledge regarding species distribution. However, seven municipalities—Marilândia, São Domingos do Norte, Alto Rio Novo, Barra de São Francisco, Ecoporanga, Vila Valério, and Linhares—where the species was never found in several years of entomological studies showed areas predicted to be suitable for its occurrence (Fig 4).



Fig 3. Frequency of observed presence of *Lutzomyia longipalpis* represented by the black dots and the red line as a local regression smoother across the significant environmental variables retained in the final model: Rocky outcrops, temperature seasonality (BIO04), temperature annual range (BIO07), and precipitation of the wettest quarter (BIO16).

Discussion

This study defines the areas predicted to be suitable for the occurrence of *L. longipalpis*, the VL vector, in the state of ES, suggesting possible areas of environmental risk for the transmission of the disease to both dogs and humans. This finding can be used to help public health agencies identify areas conducive to vector development, enabling their serological and entomological research efforts specifically focused on these regions rather than throughout the state.

The number of studies assessing occurrence of leishmaniasis using SDM is increasing, both in Brazil and abroad. These studies, in addition to covering all links in the disease cycle, such as vectors, reservoirs (dogs), and human diseases, included climate change scenarios [30-35].

In Brazil, the municipality of Itapira, located in the state of São Paulo (SP), was the focus of an SDM study for predicting a risk area for the spread of cutaneous leishmaniasis [36], whereas environmental niches for VL were studied in the state of Bahia (BA) [37]. VL risk areas were also investigated in the Brazilian states of Piauí [38], Mato Grosso do Sul [39, 40], Minas Gerais (MG) [41], and Rio de Janeiro (RJ) [42]. In ES, Meneguzzi et al. (2016), using SDM, investigated the association between cutaneous leishmaniasis cases and five sandfly vectors, with *Lutzomyia intermedia* considered the main vector in the state for its strongest associations with disease cases [43].

In this study, we used a modeling protocol that is at the forefront of current methods. The MIAmaxent package modeling process deals with several known flaws of SDM such as variable



Fig 4. Final model normalized predicted distribution for *Lutzomyia longipalpis* **in the state of Espírito Santo, southeastern Brazil.** The occurrence points used in the modeling are also shown on the map.

transformation and model selection [22]. We applied the best practices in our modeling effort, including accounting for sampling bias/spatial autocorrelation rarefying the species occurrence, setting a suitable study area extent, averaging the AUC from several replicates to account for the out-of-sample performance of the model, and using spatially stratified cross-validation and species absence for model validation [27, 28, 44]. The four distinct types of AUC validations were applied to our models to compare the extremes—at one extreme was the most widely used random cross-validation with background points and to the other extreme was spatially stratified cross-validation with species absence. The former is known to inflate the AUC, and the latter is known to provide a more reliable and realistic AUC value. Even the relatively rigorous methods for model validation resulted in a median of 0.82, which according to Hosmer and Lemeshow (2000) classifies it as an excellent model.

The occurrence region of VL in ES spans across the Doce River Valley. Furthermore, SDM for *L. longipalpis* showed that the environmental suitability areas match the VL occurrence areas, revealing that most of the state does not have favorable geoclimatic conditions for vector development.

This study emphasizes the planning of VL-control actions by restricting entomological and canine serological surveys to areas predicted to be suitable for vector occurrence, thereby saving financial resources and increasing the probability of finding more receptive sites for the disease. For example, the Brazilian Ministry of Health has recently decided to conduct a sero-logical survey of 10% of the dog population of the entire state, which is an expensive affair and could be deemed unnecessary by our study.

According to our prediction, the coastal region of ES is an unlikely region for the occurrence of *L. longipalpis*, as observed in BA, where the vector was observed in Caatinga [37]. However, in RJ, where climatic and geographical conditions were similar to those of ES, this insect has been found in 17 municipalities thus far, including those near coastal areas where the precipitation is higher [45–49].

A possible explanation is that several studies identified *L. longipalpis* as a polymorphic species i.e., a complex of species [50–57]. Additionally, the genetic variant found in RJ, 9MGB ([S]-9-methyl-germacrene-B), differs from the variant that is found in ES (cembrene-1) [58]. Thus, the ecological needs of these two genetic variants may differ, helping them adapt to different environmental conditions [59, 60]. Hence, other *L. longipalpis* genetic variants may eventually occupy previously unoccupied areas of ES, like they occurred in the locality of Cemitério do Caju, RJ by the transport of soil in plant pots from regions where the species occurred [61]. The states of Ceará and MG, for example, have records of these two morphospecies occurring in sympatry. Additionally, the states of MG, SP, and Tocantins recorded the occurrence of more genetic variants in their territories, three in the first and two in the last two states [58].

In 2017 and 2018, phlebotomine collections were carried out and the SDM was validated by confirming the presence of *L. longipalpis* in new locations in three municipalities of ES—Barra de São Francisco, Nova Venécia, and Santa Teresa (personal communication).

In our study, the climatic variable that was most associated with the occurrence of *L. longipalpis* was rocky outcrops. This finding corroborates the theory formulated by us during several years of work in ES. The preliminary arbitrary *L. longipalpis* collections were carried out primarily in areas with hot and dry climates, in elevations not exceeding 500 m, and with the presence of rocky outcrops. In this context, the northwest region of ES, which is the intermediate portion of the Doce River basin, appears to be studded with rocks that dominate the local landscape.

In the southern region of ES, the locality of Estrela do Norte (20°34'48"S and 41°19'1.99"W) in the municipality of Castelo represents another area with geographic and climatic characteristics

similar to those found in the Doce River basin. This geographical area, located in the Itapemirim River watershed, exhibits an important cluster of rocky outcrops. However, although this region seems to be adequate for the environmental needs of *L. longipalpis*, no specimen has been found there thus far. The absence of the LV vector in this area could be explained by the fact that the municipality of Castelo is geographically isolated from the Doce River Valley by a mountain range, which is impossible to be transposed naturally by the insect. The mountainous region that separates the two main hydrographic basins of ES has elevations above 1,000 meters, presenting unsuitable climatic characteristics for the occurrence of the species.

We believe that the colonization of *L. longipalpis* could occur in the municipality of Castelo, ES. This process could occur by mechanical transport of soil or agricultural plant seedlings from one region to another; however, there is currently no evidence of such transit between the regions. It is also plausible to think that in the Doce River Valley, the insect was introduced in ES by the transportation of cargo from the neighboring state of MG, where LV has been detected since 1959 [62].

A theory to explain the substantial relevance of rocky outcrops in predicting the occurrence of *L. longipalpis* would be that during the day, the rocks accumulate a great amount of heat. The slow dissipation of heat during the night would allow minor oscillations in the local temperature, generating favorable conditions for the occurrence of the vector. In contrast, in more elevated regions of ES, where there are also some rocky outcrops and the species does not occur, the thermal oscillation would be greater since temperature varies inversely to the elevation.

Temperature seasonality was the second most influential variable in the model. In this study, we noticed that in ES, *L. longipalpis* were predominantly observed in regions with a hot climate and a small temperature variability throughout the year. These areas have low elevations, corroborating the influence of temperature. In ES, the LV vector is observed in regions with elevations not exceeding 500 m.

In other Brazilian regions, *L. longipalpis* occurrence is associated with elevations lower than 900 m. In Belo Horizonte, MG, Margonari et al. (2006) [63] found a positive, albeit nonsignificant, association between VL and elevations up to 880 m, whereas Saraiva et al. (2011) [64] noted that both canine and human cases of VL were concentrated at elevations from 750 to 850 m. Conversely, in the state of São Paulo, VL vectors and hosts preferentially occurred at elevations from 274 to 539 m [65].

Most of the Doce River watershed is located in MG, the ES bordering state. There are many areas in these states sharing similar geographic and climatic conditions, mainly those located in the Atlantic Forest biome. Thus, we believe that our findings can be extrapolated to these neighboring states.

In this context, SDM enhances knowledge and provides a path to researchers and the government toward allocating human and financial resources to areas where they are significantly needed. Additionally, once the potential areas for *L. longipalpis* occurrence are determined, we can avoid the inefficient use of public resources in conducting canine serological surveys where the vector insect does not occur.

Acknowledgments

We acknowledge the Laboratory of Entomology and Malacology from Espírito Santo State Health Department; Viviane Coutinho Meneguzzi from the Federal University of Espírito Santo, Vitória, Brazil for clarifying uncertainties during the modeling process; and Julien Vollering from Western Norway University of Applied Sciences, Sogndal, Norway for kindly helping and clarifying some aspects of MIAmaxent application.

Author Contributions

Conceptualization: Karina Bertazo Del Carro.

Data curation: Karina Bertazo Del Carro, Claudiney Biral dos Santos, Israel de Souza Pinto.

Formal analysis: Karina Bertazo Del Carro, Amandio Gonçalves de Oliveira Filho, Aloísio Falqueto.

Methodology: Karina Bertazo Del Carro, Gustavo Rocha Leite, Aloísio Falqueto.

Project administration: Aloísio Falqueto.

Resources: Karina Bertazo Del Carro, Aloísio Falqueto.

Supervision: Gustavo Rocha Leite, Blima Fux, Aloísio Falqueto.

Validation: Gustavo Rocha Leite.

Writing - original draft: Karina Bertazo Del Carro.

Writing – review & editing: Karina Bertazo Del Carro, Gustavo Rocha Leite, Blima Fux, Aloísio Falqueto.

References

- Brasil, Brasil. Ministério da Saúde (MS). Secretaria de Vigilância em Saúde. Manual de VigiLância e Controle da Leishmaniose Visceral [Internet]. 1 ed. Ministério da Saúde. Brasília/DF; 2014. 122 p. http://portalsaude.saude.gov.br/index.php/o-ministerio/principal/leia-mais-o-ministerio/727-secretariasvs/vigilancia-de-a-a-z/leishmaniose-visceral-lv/l1-leishmaniose-visceral-lv/11858-vacinacaoleishmaniose
- Martins J, Souza JC, Silva E. Primeiros casos autóctones de calazar no Espírito Santo. O Hosp. 1968; 73:69–97.
- Pinto I de S, Ferreira AL, Valim V, Carvalho F dos S, da Silva GM, Falcão AL, et al. Sand fly vectors (Diptera, Psychodidae) of American visceral leishmaniasis areas in the Atlantic Forest, state of Espírito Santo, southeastern Brazil. J Vector Ecol [Internet]. 2012 Jun; 37(1):90–6. Available from: http://www. scielo.br/scielo.php?script=sci_arttext&pid=S0074-02762007000200007&lng=en&tlng=en PMID: 22548541
- 4. Pinto I de S. Associação entre variáveis geográficas e climáticas e a ocorrência de Lutzomyia longipalpis (Lutz & Neiva, 1912) (Diptera, Psychodidae, Phlebotominae) no estado do Espírito Santo, Brasil. Universidade Federal do Espírito Santo; 2009.
- Sessa A, Falqueto A, Feitoza LR, Feitoza HN. Avaliação do potencial de disseminação da leishmaniose visceral no estado do Espírito Santo através de um sistema de informações geográficas. Rev Soc Bras Med Trop. 2000; 33(Suppl I):327.
- 6. WHO. Leishmaniasis [Internet]. Internet. 2019. https://www.who.int/news-room/fact-sheets/detail/ leishmaniasis
- Barcellos C, Bastos FI. Geoprocessamento, ambiente e saúde: uma união possível? Cad Saude Publica. 1996; 12(3):389–97. https://doi.org/10.1590/s0102-311x1996000300012 PMID: 10904341
- Guisan A, Tingley R, Baumgartner JB, Naujokaitis-Lewis I, Sutcliffe PR, Tulloch AIT, et al. Predicting species distributions for conservation decisions. Arita H, editor. Ecol Lett [Internet]. 2013 Dec; 16 (12):1424–35. Available from: http://doi.wiley.com/10.1111/ele.12189 PMID: 24134332
- Moreira D de O, Leite GR, de Siqueira MF, Coutinho BR, Zanon MS, Mendes SL. The Distributional Ecology of the Maned Sloth: Environmental Influences on Its Distribution and Gaps in Knowledge. Fenton B, editor. PLoS One [Internet]. 2014 Oct 22; 9(10):e110929. Available from: https://dx.plos.org/10. 1371/journal.pone.0110929 PMID: 25338139
- 10. Peterson AT. Predicting species' geographic distributions based on ecological niche modeling. Condor. 2001; 103(3):599.
- Peterson AT. Developing risk maps. In: Mapping disease transmission risk: enriching models using biogeography and ecology. Baltimore; 2014. p. 140–9.
- Guisan A, Zimmermann NE. Predictive habitat distribution models in ecology. Ecol Modell [Internet]. 2000 Dec; 135(2–3):147–86. Available from: https://linkinghub.elsevier.com/retrieve/pii/ S0304380000003549

- 13. INCAPER. Natural resources information systems for rural development: approaches for Espírito Santo state, Brazil. Feitoza LR, Stocking M, Resende M, editors. Vitória/ES; 2001. 223 p.
- Fick SE, Hijmans RJ. WorldClim 2: new 1-km spatial resolution climate surfaces for global land areas. Int J Climatol [Internet]. 2017 Oct; 37(12):4302–15. Available from: https://onlinelibrary.wiley.com/doi/ abs/10.1002/joc.5086
- Tabarelli M, Pinto LP, Silva JMC, Hirota M, Bede L. Challenges and Opportunities for Biodiversity Conservation in the Brazilian Atlantic Forest. Conserv Biol [Internet]. 2005 Jun; 19(3):695–700. Available from: http://doi.wiley.com/10.1111/j.1523-1739.2005.00694.x
- Galati EAB. Morfologia e taxonomia. In: Rangel EF, Laison R, editors. Flebotomíneos do Brasil. Rio de Janeiro; 2003. p. 23–51.
- Young DG, Duncan MA. Guide to the identification and geographic distribution of Lutzomyia sand flies in Mexico, the West Indies, Central and South America (Diptera: Psychodidae). Memoirs of the American Entomological Institute. 1994. 1–881 p.
- Phillips SJ, Anderson RP, Schapire RE. Maximum entropy modeling of species geographic distributions. Ecol Modell. 2006 Jan; 190(3–4):231–59.
- Phillips SJ D M. Modeling of species distributions with Maxent: new extensions and a comprehensive evaluation. Ecography (Cop). 2008; 31:161–175.
- **20.** Phillips SJ, Dudík M, Schapire RE. A maximum entropy approach to species distribuition modeling. In: Proceedings of the 21st International Conference on Machine Learning. 2004. p. 655–62.
- Merow C, Smith MJ, Silander JA. A practical guide to MaxEnt for modeling species' distributions: what it does, and why inputs and settings matter. Ecography (Cop) [Internet]. 2013 Oct; 36(10):1058–69. Available from: http://doi.wiley.com/10.1111/j.1600-0587.2013.07872.x
- Vollering J, Halvorsen R, Mazzoni S. The MIAmaxent R package: Variable transformation and model selection for species distribution models. Ecol Evol [Internet]. 2019 Nov 30; 9(21):12051–68. Available from: https://onlinelibrary.wiley.com/doi/abs/10.1002/ece3.5654 PMID: 31832144
- 23. Valavi R, Elith J, Lahoz-Monfort JJ, Guillera-Arroita G. blockCV: An package for generating spatially or environmentally separated folds for k-fold cross-validation of species distribution models. Warton D, editor. Methods Ecol Evol [Internet]. 2019 Feb 8; 10(2):225–32. Available from: https://onlinelibrary. wiley.com/doi/abs/10.1111/2041-210X.13107
- 24. Muscarella R, Galante PJ, Soley-Guardia M, Boria RA, Kass JM, Uriarte M, et al. ENMeval: An R package for conducting spatially independent evaluations and estimating optimal model complexity for MAX-ENT ecological niche models. McPherson J, editor. Methods Ecol Evol [Internet]. 2014 Nov; 5(11):1198– 205. Available from: http://doi.wiley.com/10.1111/2041-210X.12261
- Manel S, Williams HC O S. Evaluating presence–absence models in ecology: the need to account for prevalence. J Appl Ecol. 2001; 38:921–931.
- Hosmer DW, Lemeshow S. Applied Logistic Regression [Internet]. Second Edi. Applied Logistic Regression. Hoboken, NJ, USA: John Wiley & Sons, Inc.; 2000. Available from: <u>http://doi.wiley.com/</u> 10.1002/0471722146
- Lobo JM, Jiménez-Valverde A RR. AUC: a misleading measure of the performance of predictive distribution models. Glob Ecol Biogeogr. 2008; 17:145–151.
- Peterson AT, Papes M S J. Rethinking receiver operating characteristic analysis applications in ecological niche modeling. Ecol Model. 2008; 213:63–72.
- Halvorsen R, Mazzoni S, Bryn A, Bakkestuen V. Opportunities for improved distribution modelling practice via a strict maximum likelihood interpretation of MaxEnt. Ecography (Cop) [Internet]. 2015 Feb; 38 (2):172–83. Available from: http://doi.wiley.com/10.1111/ecog.00565
- Chamaillé L, Tran A, Meunier A, Bourdoiseau G, Ready P, Dedet J-PP. Environmental risk mapping of canine leishmaniasis in France. Parasit Vectors [Internet]. 2010; 3(1):31. Available from: http:// parasitesandvectors.biomedcentral.com/articles/10.1186/1756-3305-3-31 PMID: 20377867
- Franco AO, Davies CR, Mylne A, Dedet J-P, Gállego M, Ballart C, et al. Predicting the distribution of canine leishmaniasis in western Europe based on environmental variables. Parasitology [Internet]. 2011 Dec 14; 138:1878–91. Available from: https://www.cambridge.org/core/product/identifier/ S003118201100148X/type/journal_article PMID: 21914251
- 32. Fischer D, Thomas SM, Beierkuhnlein C. Modelling climatic suitability and dispersal for disease vectors: the example of a phlebotomine sandfly in Europe. Procedia Environ Sci [Internet]. 2011; 7:164–9. Available from: http://dx.doi.org/10.1016/j.proenv.2011.07.029
- 33. González C, Wang O, Strutz SE, González-Salazar C, Sánchez-Cordero V, Sarkar S. Climate change and risk of leishmaniasis in North America: predictions from ecological niche models of vector and reservoir species. Galvani AP, editor. PLoS Negl Trop Dis [Internet]. 2010 Jan 19; 4(1):e585. Available from: https://dx.plos.org/10.1371/journal.pntd.0000585 PMID: 20098495

- González C, Paz A, Ferro C. Predicted altitudinal shifts and reduced spatial distribution of Leishmania infantum vector species under climate change scenarios in Colombia. Acta Trop [Internet]. 2014 Jan; 129(1):83–90. Available from: http://dx.doi.org/10.1016/j.actatropica.2013.08.014 The remote server returned an error: (404) Not Found.PMID: 23988300
- 35. Peterson AT, Campbell LP, Moo-Llanes DA, Travi B, González C, Ferro MC, et al. Influences of climate change on the potential distribution of Lutzomyia longipalpis sensu lato (Psychodidae: Phlebotominae). Int J Parasitol [Internet]. 2017 Sep; 47(10–11):667–74. Available from: http://dx.doi.org/10.1016/j.ijpara. 2017.04.007 PMID: 28668326
- Aparicio C, Bitencourt MD. Modelagem espacial de zonas de risco da leishmaniose tegumentar americana. Rev Saude Publica. 2004; 38(4):511–6. https://doi.org/10.1590/s0034-89102004000400005 PMID: 15311290
- Nieto P, Malone JB, Bavia ME. Ecological niche modeling for visceral leishmaniasis in the state of Bahia, Brazil, using genetic algorithm for rule-set prediction and growing degree day-water budget analysis. Geospat Health. 2006; 1:115–26. https://doi.org/10.4081/gh.2006.286 PMID: 18686237
- de Almeida AS, Werneck GL, da Costa Resendes AP. Classificação orientada a objeto de imagens de sensoriamento remoto em estudos epidemiológicos sobre leishmaniose visceral em área urbana. Cad Saude Publica. 2014; 30(8):1639–53. https://doi.org/10.1590/0102-311x00059414 PMID: 25210905
- 39. de Almeida PS, Sciamarelli A, Batista PM, Ferreira AD, Nascimento J, Raizer J, et al. Predicting the geographic distribution of Lutzomyia longipalpis (Diptera: Psychodidae) and visceral leishmaniasis in the state of Mato Grosso do Sul, Brazil. Mem Inst Oswaldo Cruz [Internet]. 2013 Dec 6; 108(8):992–6. Available from: http://www.scielo.br/scielo.php?script=sci_arttext&pid=S0074-02762013000800992&Ing=en&tIng=en PMID: 24402151
- 40. de Oliveira EF, Galati EAB, de Oliveira AG, Rangel EF, de Carvalho BM. Ecological niche modelling and predicted geographic distribution of Lutzomyia cruzi, vector of Leishmania infantum in South America. Carvalho MS, editor. PLoS Negl Trop Dis [Internet]. 2018 Jul 30; 12(7):e0006684. Available from: https://dx.plos.org/10.1371/journal.pntd.0006684 PMID: 30059494
- 41. de Araújo VEM, Pinheiro LC, de M Almeida MC, de Menezes FC, Morais MHF, Reis IA, et al. Relative risk of visceral leishmaniasis in Brazil: a spatial analysis in urban area. Kamhawi S, editor. PLoS Negl Trop Dis [Internet]. 2013 Nov 7; 7(11):e2540. Available from: <u>https://dx.plos.org/10.1371/journal.pntd.</u> 0002540 PMID: 24244776
- 42. Abrantes TR, Werneck GL, de Almeida AS, Figueiredo FB. Fatores ambientais associados à ocorrência de leishmaniose visceral canina em uma área de recente introdução da doença no estado do Rio de Janeiro, Brasil. Cad Saude Publica [Internet]. 2018 Feb 5; 34(1):e00021117. Available from: http://www.scielo.br/scielo.php?script=sci_arttext&pid=S0102-311X2018000105013&Ing=pt&tIng=pt PMID: 29412315
- 43. Meneguzzi VC, dos Santos CB, Leite GR, Fux B, Falqueto A. Environmental niche modelling of phlebotomine sand flies and cutaneous leishmaniasis identifies Lutzomyia intermedia as the main vector species in Southeastern Brazil. Gatton ML, editor. PLoS One [Internet]. 2016 Oct 26; 11(10):e0164580. Available from: http://dx.plos.org/10.1371/journal.pone.0164580 PMID: 27783641
- 44. Guisan A, Thuiller W, Zimmermann NE. Habitat Suitability and Distribution Models: With Applications in R. In: Ecology, Biodiversity and Conservation [Internet]. 1st ed. Cambridge: Cambridge University Press; 2017. http://ebooks.cambridge.org/ref/id/CBO9781139028271
- 45. Novo SPC, de Souza MB, Villanova CB, Meródio JC, de Medeiros Meira A. Survey of sandfly vectors of leishmaniasis in Marambaia Island, municipality of Mangaratiba, state of Rio de Janeiro, Brazil. Rev Soc Bras Med Trop [Internet]. 2013 Apr; 46(2):231–3. Available from: http://www.scielo.br/scielo.php? script=sci_arttext&pid=S0037-86822013000200231&Ing=en&tIng=en PMID: 23740054
- 46. de Azevedo ACR, de Andrade-Coelho CA, da Silva VC, Sena CAP, de Souza FJMS, de Souza NA. Abundance and monthly frequency of phlebotomine sand flies (Diptera: Phlebotominae) in some municipalities in the state of Rio de Janeiro, Brazil. EntomoBrasilis [Internet]. 2015; 8(3):201–8. Available from: http://www.periodico.ebras.bio.br/ojs/index.php/ebras/article/view/498
- de Aguiar GM, de Medeiros WM, de Marco TS, dos Santos SC, Gambardella S. Ecologia dos flebotomíneos da Serra do Mar, Itaguaí, estado do Rio de Janeiro, Brasil. I–A fauna flebotomínica e prevalência pelo local e tipo de captura (Diptera, Psychodidae, Phlebotominae). Cad Saude Publica [Internet]. 1996; 12(2):195–206. Available from: http://www.ncbi.nlm.nih.gov/pubmed/10904321 PMID: 10904321
- 48. Fuzari AA, Delmondes AF dos S, Barbosa V de A, Marra F de A, Brazil RP. Presence of Lutzomyia longipalpis (Diptera: Psychodidae) in the Parque Estadual da Serra da Tiririca, state of Rio de Janeiro, Southeastern Brazil. Rev Soc Bras Med Trop [Internet]. 2016 Oct; 49(5):616–9. Available from: http://www.scielo.br/scielo.php?script=sci_arttext&pid=S0037-86822016000500616&Ing=en&tIng=en PMID: 27812657
- 49. Carvalho BM, Dias CMG, Rangel EF. Phlebotomine sand flies (Diptera, Psychodidae) from Rio de Janeiro state, Brazil: species distribution and potential vectors of leishmaniases. Rev Bras Entomol

[Internet]. 2014 Mar; 58(1):77–87. Available from: http://www.scielo.br/scielo.php?script=sci_arttext&pid=S0085-56262014000100013&Ing=en&nrm=iso&tIng=en

- Mangabeira Filho O. Sôbre a sistemática e biologia dos Phlebotomus do Ceará. Rev Bras Mariologia e Doenças Trop. 1969; 21:3–26.
- Watts PC, Hamilton JGC, Ward RD, Noyes HA, Souza NA, Kemp SJ, et al. Male sex pheromones and the phylogeographic structure of the Lutzomyia longipalpis species complex (Diptera: Psychodidae) from Brazil and Venezuela. Am J Trop Med Hyg. 2005; 73(4):734–43. PMID: 16222018
- 52. Araki AS, Ferreira GEM, Mazzoni CJ, Souza NA, Machado RC, Bruno R V., et al. Multilocus analysis of divergence and introgression in sympatric and allopatric sibling species of the Lutzomyia longipalpis complex in Brazil. Warburg A, editor. PLoS Negl Trop Dis [Internet]. 2013 Oct 17; 7(10):e2495. Available from: https://dx.plos.org/10.1371/journal.pntd.0002495 PMID: 24147172
- 53. Ward RD, Ribeiro AL, Ready PD, Murtagh A. Reproductive isolation between different forms of Lutzomyia longipalpis (Lutz & Neiva), (Diptera: Psychodidae), the vector os Leishmania donovani chagasi Cunha & Chagas and its significance to kala-zar distribution in South America. Mem Inst Oswaldo Cruz. 1983; 78(3):269–80.
- 54. Arrivillaga J, Mutebi J-P, Piñago H, Norris D, Alexander B, Feliciangeli MD, et al. The taxonomic status of genetically divergent populations of Lutzomyia longipalpis (Diptera: Psychodidae) based on the distribution of mitochondrial and isozyme variation. J Med Entomol. 2003; 40(5):615–27. https://doi.org/10. 1603/0022-2585-40.5.615 PMID: 14596274
- 55. Pech-May A, Ramsey JM, González Ittig RE, Giuliani M, Berrozpe P, Quintana MG, et al. Genetic diversity, phylogeography and molecular clock of the Lutzomyia longipalpis complex (Diptera: Psychodidae). Al-Salem WS, editor. PLoS Negl Trop Dis [Internet]. 2018 Jul 5; 12(7):e0006614. Available from: https://dx.plos.org/10.1371/journal.pntd.0006614 PMID: 29975695
- 56. Araki AS, Vigoder FM, Bauzer LGSR, Ferreira GEM, Souza NA, Araújo IB, et al. Molecular and behavioral differentiation among brazilian populations of Lutzomyia longipalpis (Diptera: Psychodidae: Phlebotominae). Caccone A, editor. PLoS Negl Trop Dis [Internet]. 2009 Jan 27; 3(1):e365. Available from: https://dx.plos.org/10.1371/journal.pntd.0000365 PMID: 19172187
- 57. Bauzer LGSRG, Souza NA, Maingon RDDC, Peixoto AA. Lutzomyia longipalpis in Brazil: a complex or a single species? A mini-review. Mem Inst Oswaldo Cruz [Internet]. 2007 Feb; 102(1):1–12. Available from: http://www.scielo.br/scielo.php?script=sci_arttext&pid=S0074-02762007000100001&Ing= en&ting=en PMID: 17293992
- Spiegel CN, Dias DB dos S, Araki AS, Hamilton JGC, Brazil RP, Jones TM. The Lutzomyia longipalpis complex: a brief natural history of aggregation-sex pheromone communication. Parasit Vectors [Internet]. 2016 Dec 14; 9(1):580. Available from: http://dx.doi.org/10.1186/s13071-016-1866-x PMID: 27842601
- Bauzer LGSR, Souza NA, Ward RD, Kyriacou CP, Peixoto AA. The period gene and genetic differentiation between three Brazilian populations of Lutzomyia longipalpis. Insect Mol Biol [Internet]. 2002 Aug; 11(4):315–23. Available from: http://doi.wiley.com/10.1046/j.1365-2583.2002.00340.x PMID: 12144696
- 60. Souza NA, Andrade-Coelho CA, Vigoder FM, Ward RD, Peixoto AA. Reproductive isolation between sympatric and allopatric Brazilian populations of Lutzomyia longipalpis s.l. (Diptera: Psychodidae). Mem Inst Oswaldo Cruz [Internet]. 2008 Mar; 103(2):216–9. Available from: http://www.scielo.br/scielo.php?script=sci_arttext&pid=S0074-02762008000200017&Ing=en&tIng=en PMID: 18425278
- Brazil RP. The dispersion of *Lutzomyia longipalpis* in urban areas. Rev Soc Bras Med Trop. 2013; 46 (3):263–4. https://doi.org/10.1590/0037-8682-0101-2013 PMID: 23856862
- 62. Rezende C, Bastos O. Calazar infantil. Relato de 1 caso ocorrido em Belo Horizonte. J Pediatr. 1959; 24:323–31.
- 63. Margonari C, Freitas CR, Ribeiro RC, Moura ACM, Timbó M, Gripp AH, et al. Epidemiology of visceral leishmaniasis through spatial analysis, in Belo Horizonte municipality, state of Minas Gerais, Brazil. Mem Inst Oswaldo Cruz [Internet]. 2006 Feb; 101(1):31–8. Available from: http://www.scielo.br/scielo.php?script=sci_arttext&pid=S0074-02762006000100007&Ing=en&tIng=en PMID: 16699707
- 64. Saraiva L, Andrade Filho JD, Falcão AL, de Carvalho DAA, de Souza CM, Freitas CR, et al. Phlebotominae fauna (Diptera: Psychodidae) in an urban district of Belo Horizonte, Brazil, endemic for visceral leishmaniasis: Characterization of favored locations as determined by spatial analysis. Acta Trop [Internet]. 2011 Feb; 117(2):137–45. Available from: http://dx.doi.org/10.1016/j.actatropica.2010.11.007 PMID: 21110938
- 65. da P Sevá A, Mao L, Galvis-Ovallos F, Tucker Lima JM, Valle D. Risk analysis and prediction of visceral leishmaniasis dispersion in São Paulo state, Brazil. Carvalho EM, editor. PLoS Negl Trop Dis. 2017 Feb; 11(2):e0005353. https://doi.org/10.1371/journal.pntd.0005353 PMID: 28166251