

Targeting copper metabolism to defeat KRAS-driven colorectal cancer

Léo Aubert^a, Neethi Nandagopal^a, and Philippe P. Roux^{a,b} 

^aInstitute for Research in Immunology and Cancer (IRIC), Université De Montréal, Montreal, Quebec, Canada; ^bDepartment of Pathology and Cell Biology, Faculty of Medicine, Université De Montréal, Montreal, Quebec, Canada

ABSTRACT

KRAS-driven cancers acquire profound metabolic dependencies that are intimately linked to tumor growth. Our work revealed that colorectal cancers that harbor KRAS mutations are addicted to copper metabolism. This adaptation renders tumor cells critically dependent on the copper transporter ATP7A, which reveals copper metabolism as a promising therapeutic target for KRAS-driven colorectal cancers.

ARTICLE HISTORY

Received 31 August 2020
Revised 3 September 2020
Accepted 4 September 2020

KEYWORDS

Copper; KRAS; TTM; ATP7A; colorectal cancer; chelators; micronutrients

Activating mutations in KRAS (Kirsten rat sarcoma 2 viral oncogene homolog) contribute to nearly half of all human colorectal cancers (CRC), and are commonly associated with resistance to first-line therapies. Consequently, almost four decades of intensive research have focused on the development of monotherapeutic approaches to directly or indirectly target oncogenic KRAS.¹ Though early clinical studies have shown some promise, including the recent evaluation of KRAS^{G12C}-specific inhibitors, the emergence of resistance mechanisms has severely limited clinical outcomes. While several different combination therapies are currently being explored to counter drug resistance, the identification of novel vulnerabilities for KRAS-dependent cancers remains important for the design of tailored therapeutic approaches.²

Metabolic adaptation in cancer is widely accepted as a critical aspect of tumor growth.³ In CRC, mutant KRAS actively rewires tumor cell metabolism to fuel the inexorable biosynthetic demand associated with aberrant cell growth and proliferation.⁴ Establishment of this new metabolic program is accompanied with a unique set of cellular features, including the ability to acquire and utilize essential nutrients required to maintain cancer cell fitness. Evidence suggest that cancer cell addiction to nutrient supply can be exploited for the design of efficient therapeutic approaches. In particular, KRAS-mutant cancer cells were shown to require different modes of nutrient uptake, such as macropinocytosis, which is increasingly recognized as a potential therapeutic target.^{5,6} In recent years, tremendous efforts have helped shed light on the implication of mutant KRAS in nutrient uptake and metabolism, such as with glucose, lipid and amino acids. In contrast, much less is known regarding the role of mutant KRAS on the uptake and metabolism of micronutrients, such as trace minerals and vitamins.

Using a proteogenomic approach to study mutant KRAS-mediated transformation of intestinal epithelial cells, we have identified the copper-transporter ATP7A (Copper-transporting ATPase 1) as a potent synthetic lethal target for KRAS-mutant CRC.⁷ Combining cutting-edge cell-surface

proteomics with CRISPR/Cas9 screens, we found that ATP7A is selectively elevated at the surface of KRAS^{G12V}-transformed intestinal cells and plays essential roles in tumor growth *in vitro* and *in vivo*. Intriguingly, we found that ATP7A upregulation occurred in a transcription-independent manner that involved copper-dependent protein stabilization and intracellular transport, as described elsewhere.⁸ As these results suggested that KRAS-mutant cells have higher intracellular copper levels, we set out to measure basal copper levels in CRC cells and patient-derived colorectal tumor specimens. Using several methods, including the anti-correlated expression of the copper chaperone for superoxide dismutase (CCS), as well as inductively coupled plasma-mass spectrometry (ICP-MS), we found that KRAS mutation correlated with elevated intracellular copper levels. Using ratiometric fluorescent probes to monitor labile pools of copper,⁹ which contribute to metabolism, we found that KRAS mutation was linked to increased levels of bioavailable copper.⁷ Consistent with this, we found that ATP7A participates in the regulation of several cuproenzymes, including Cytochrome C Oxidase (CCO) and the pathway leading to ERK1/2 (extracellular signal-regulated kinases 1/2) activation, which helps explain its observed essentiality in KRAS-mutated cells.⁷

Our results have shown that KRAS-mutated cells are more sensitive to copper chelators, such as ammonium tetrathiomolybdate (TTM), than wild-type counterparts, suggesting that copper metabolism may be a potential therapeutic target for KRAS-mutated CRC. Anti-Cu agents, such as TTM, choline tetrathiomolybdate (ATN-224), trientine, and D-penicillamine were previously shown to inhibit cancer cell growth in preclinical studies.¹⁰ A limited number of clinical trials have attempted to evaluate the effect of TTM on tumor growth, and found that the drug was generally well tolerated with manageable toxicity profiles. Notably, TTM treatment sensitized melanoma cells that are resistant to BRAF (Serine/threonine-protein kinase B-raf) inhibitors, suggesting that TTM could be repurposed to counter some forms of drug resistance.

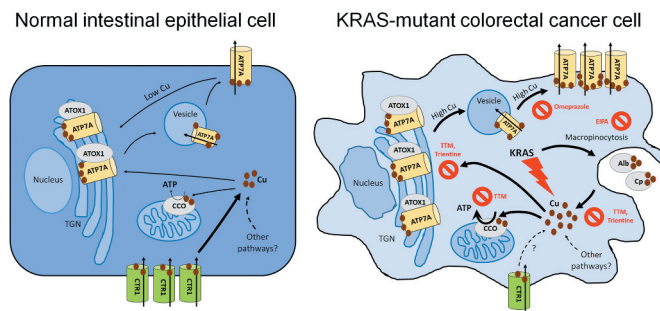


Figure 1. KRAS-driven colorectal cancer (CRC) cells are addicted to copper metabolism. In normal intestinal epithelial cells (left), copper (Cu) [depicted as brown circles] is mainly imported via the high-affinity transporter CTR1. Intracellular Cu is then transported to the mitochondria and the trans-Golgi network (TGN), where it is loaded on essential cuproenzymes. KRAS-mutant CRC cells (right) are associated with elevated intracellular Cu levels. An important Cu entry route in KRAS-mutated CRC cells is macropinocytosis, which can be inhibited using 5-(N-ethyl-N-isopropyl) amiloride (EIPA). Increased Cu levels is expected to stimulate the activity of cuproenzymes, which can be inhibited using Cu chelators, such as tetrathiomolybdate (TTM) and the related Trientine. Increased intracellular Cu levels in KRAS-mutant cells result in ATP7A upregulation and translocation to the plasma membrane, which is required for the export of excess Cu and resistance to cuproptosis. ATP7A translocation to the plasma membrane was shown to be sensitive to the ATPase inhibitor Omeprazole. Other unidentified pathways, such as autophagy or intracellular stores of Cu, could also contribute to the increased Cu pools in KRAS-mutant cells. Potential therapeutic interventions are highlighted in red. The intensity of arrows indicate the extent of contribution of this pathway to the intracellular process. Dietary Cu is incorporated into Albumin (Alb) and Ceruloplasmin (Cp), which are the major Cu-binding proteins in the plasma. CTR1, high affinity copper uptake protein 1; ATP7A, copper-transporting ATPase 1; CCO, cytochrome C oxidase; ATOX1, copper transport protein ATOX1; KRAS, Kirsten rat sarcoma 2 viral oncogene homolog.

As our data suggest that KRAS-mutant CRC are addicted to copper metabolism, an interesting possibility would be to determine the efficacy of anti-Cu agents in the context of KRAS-mutated cancers.

To determine the mechanism by which oncogenic KRAS increases intracellular copper levels, we investigated the involvement of the high-affinity copper-importer CTR1 (High affinity copper uptake protein 1). Our results have shown that KRAS-mutant cells express reduced levels of CTR1 compared to control cells, and that its expression is dispensable for KRAS-dependent tumor growth.⁷ Given that CTR1-independent routes for copper uptake are not well described in humans, we evaluated the contribution of macropinocytosis, which is upregulated in KRAS-mutated cells and previously shown to participate in nutrient uptake.⁶ Using the macropinocytosis inhibitor 5-(N-ethyl-N-isopropyl) amiloride (EIPA), we found that macropinocytosis regulates intracellular copper levels and is required for KRAS-mutated CRC growth.⁷ While macropinocytosis may not be the exclusive mode of copper uptake, our results indicate that it is required to fuel copper metabolism in KRAS-mutated CRC cells.

In summary, our findings suggest new ways to target KRAS-addicted CRC (see Figure 1). While copper metabolism could be targeted using anti-Cu drugs or ATP7A inhibitors, our results show that targeting copper entry via macropinocytosis could also be an efficient way to specifically affect KRAS-mutated cancer cells.

Disclosure of potential conflicts of interest

The authors declare no conflict of interest.

Funding

Our work is funded by grants from the Canadian Institutes for Health Research (CIHR) and the Cancer Research Society (CRS). P.P.R. is a Senior Scholar of the Fonds de la recherche du Québec-Santé (FRQS). L.A. received a Postdoctoral Fellowship from the Cole Foundation, and N.

N. was supported by Doctoral Scholarships from the FRQS and the Fonds de la recherche du Québec-Nature and Technologies (FRQNT).

ORCID

Philippe P. Roux  <http://orcid.org/0000-0002-5962-0250>

References

1. Stalneck CA, Der CJ. RAS, wanted dead or alive: advances in targeting RAS mutant cancers. *Sci Signal.* 2020;13(624):eaay6013. doi:10.1126/scisignal.aay6013.
2. Moore AR, Rosenberg SC, McCormick F, Malek S. RAS-targeted therapies: is the undruggable drugged? *Nat Rev Drug Discov.* 2020;19:533–552. doi:10.1038/s41573-020-0068-6.
3. Pavlova NN, Thompson CB. The emerging hallmarks of cancer metabolism. *Cell Metab.* 2016;23(1):27–47. doi:10.1016/j.cmet.2015.12.006.
4. Yun J, Mullarky E, Lu C, Bosch KN, Kavalier A, Rivera K, Roper J, Chio II, Giannopoulou EG, Rago C, et al. Vitamin C selectively kills KRAS and BRAF mutant colorectal cancer cells by targeting GAPDH. *Science.* 2015;350(6266):1391–1396. doi:10.1126/science.aaa5004.
5. Ramirez C, Hauser AD, Vucic EA, Bar-Sagi D. Plasma membrane V-ATPase controls oncogenic RAS-induced macropinocytosis. *Nature.* 2019;576:477–481. doi:10.1038/s41586-019-1831-x.
6. Recouvreux MV, Comisso C. Macropinocytosis: a metabolic adaptation to nutrient stress in cancer. *Front Endocrinol (Lausanne).* 2017;8:261. doi:10.3389/fendo.2017.00261.
7. Aubert L, Nandagopal N, Steinhart Z, Lavoie G, Nourredine S, Berman J, Saba-El-Leil MK, Papadopolis D, Lin S, Hart T, et al. Copper bioavailability is a KRAS-specific vulnerability in colorectal cancer. *Nat Commun.* 2020;11(1):3701. doi:10.1038/s41467-020-17549-y.
8. Chun H, Catterton T, Kim H, Lee J, Kim BE. Organ-specific regulation of ATP7A abundance is coordinated with systemic copper homeostasis. *Sci Rep.* 2017;7:12001. doi:10.1038/s41598-017-11961-z.
9. Morgan MT, Bourassa D, Harankhedkar S, McCallum AM, Zlatic SA, Calvo JS, Meloni G, Faundez V, Fahrni CJ. Ratiometric two-photon microscopy reveals attomolar copper buffering in normal and Menkes mutant cells. *Proc Natl Acad Sci U S A.* 2019;116(25):12167–12172. doi:10.1073/pnas.1900172116.
10. Denoyer D, Masaldan S, La Fontaine S, Cater MA. Targeting copper in cancer therapy: ‘copper that cancer’. *Metallomics.* 2015;7(11):1459–1476. doi:10.1039/c5mt00149h.