


Advances in the Biological Functions and Mechanisms of miRNAs in the Development of Osteosarcoma

Technology in Cancer Research & Treatment
 Volume 21: 1-16
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 DOI: 10.1177/15330338221117386
journals.sagepub.com/home/tct


Zihe Dong, BS¹ , Zhipeng Liao, BS², Yonglin He, BS¹, Chengye Wu, BS¹, Zixiang Meng, BS¹, Baolong Qin, BS¹, Ge Xu, BS¹, Zeyang Li, BS¹, Tianxin Sun, BS¹, Yuyan Wen, BS¹, and Guangjie Li, PhD³

Abstract

Osteosarcoma is one of the most common primary malignant bone tumors, mainly occurring in children and adolescents, and is characterized by high morbidity and poor prognosis. MicroRNAs, a class of noncoding RNAs consisting of 19 to 25 nucleotides, are involved in cell proliferation, invasion, metastasis, and apoptosis to regulate the development and progression of osteosarcoma. Studies have found that microRNAs are closely related to the diagnosis, treatment, and prognosis of osteosarcoma patients and have an important role in improving drug resistance in osteosarcoma. This paper reviews the role of microRNAs in the pathogenesis of osteosarcoma and their clinical value, aiming to provide a new research direction for diagnosing and treating osteosarcoma and achieving a better prognosis.

Keywords

microRNAs, osteosarcoma, biomarkers, targeted therapy, molecular mechanisms

Abbreviations

CDK2, cyclin-dependent kinase 2; CDK4, cyclin-dependent kinase 4; CDKN1A, cyclin-dependent kinase inhibitor 1A; CDKN1B, cyclin-dependent kinase inhibitor 1B; DKK1, Dickkopf WNT Signaling Pathway Inhibitor 1; DKK3, Dickkopf WNT Signaling Pathway Inhibitor 3; DNMT3B, DNA Methyltransferase 3 Beta; E2F5, E2F Transcription Factor 5; EBF2, Early B-Cell Factor-2; FBXW11, F-box and WD repeat domain containing 11; FBXW7, F-Box And WD Repeat Domain Containing 7; GPX4, glutathione peroxidase 4; HES1, Hes Family BHLH Transcription Factor 1; HIF-1 α , Hypoxia inducible factor-1 α ; IFI44L, interferon-induced protein 44-like; IRF1, interferon regulatory factor 1; IRF2, interferon regulatory factor 2; ITGAV, integrin subunit alpha V; JMJD2C, Jumonji C domain-containing oxygenase D2C; KLF12, Kruppel Like Factor 12; MAFG, MAF BZIP Transcription Factor G; MAKPL, NF- κ B-activating protein-like; PTEN, phosphatase and tensin homolog; PTPN14, Non-receptor tyrosine phosphatase 14; ROCK1, Rho-associated protein kinase 1; Runx2, Runt-related transcription factor 2; SIX1, sine oculis homeobox 1; SOX4, SRY-Box Transcription Factor 4; STAT3, Signal transducer and activator of transcription 3; TAZ, Transcriptional co-activator with PDZ-binding motif; TGF β 1, transforming growth factor- β 1; TRAF3, TNF receptor-associated factor 3; VEPHI, Ventricular Zone Expressed PH Domain Containing 1; ZIF, zeolitic imidazolate framework

Received: March 22, 2022; Revised: June 6, 2022; Accepted: July 13, 2022.

Introduction

Osteosarcoma (OS) originates from primitive mesenchymal cells, and it is the most common malignant bone tumor in children and adolescents.¹ The most common sites of osteosarcoma are the metaphysis of long bones, especially the distal femur, proximal tibia, and proximal humerus.^{2,3} Most patients with OS have pain and swelling in the affected area.⁴ Due to surgical resection, combined chemotherapy and targeted radiotherapy in

¹ The First School of Clinical Medicine, Lanzhou University, Lanzhou, Gansu, China

² The Second School of Clinical Medicine, Lanzhou University, Lanzhou, Gansu, China

³ The First Hospital of Lanzhou University, Lanzhou, Gansu, China

Corresponding Author:

Guangjie Li, PhD, The First Hospital of Lanzhou University, Lanzhou, Gansu, China.

Email: ligj@lzu.edu.cn



treating osteosarcoma, the 5-year survival rate for patients without metastases can be around 60% to 70%.^{5,6} However, metastases and recurrences often lead to a poor prognosis, so the 5-year survival rate for OS patients with metastases or recurrences is only 10% to 20%.^{7,8} The malignant grade of osteosarcoma is high, the micro-lesion metastasis may be possible in the diagnosis, and lung tissue is a common metastatic site.⁹ At the same time, osteosarcoma cells can become resistant to various chemotherapeutic agents during treatment, which poses a huge dilemma for the clinical management of osteosarcoma.¹⁰ Although overall survival is universally recognized as the gold standard when assessing prognostic information or measuring treatment effects in clinical research, the complexity of cancer death, including invasion, recurrence, and metastasis, still limits the practicality and reliability of OS in the estimation of cancer progress and prognosis.¹¹ In addition, a novel technique, positron emission tomography/computed tomography (PET/CT), which is widely used in clinical practice, shows more accuracy, sensitivity, and specificity in the diagnosis of osteosarcoma. However, finance and cost limits have prevented its widespread use.^{12,13} Therefore, it is necessary to conduct in-depth research on osteosarcoma diagnosis and treatment options and actively pursue clinical translation to improve the current diagnosis and treatment methods and obtain a better prognosis.

MicroRNAs (miRNAs) are a class of highly conserved endogenous non-coding RNAs, approximately 19 to 25 nucleotides in length.¹⁴ They perform their biological functions mainly through the regulation of gene expression.¹⁵ Abnormal expression of miRNAs is associated with various types of cancer, and these abnormal miRNAs often function as oncogenes or tumor suppressors during cancer development and progression.^{16–18} Therefore, attempts can be made to improve the expression of aberrant miRNAs by inhibiting the function of highly expressed miRNAs in cancer or by supplementing the amount of downstream specific products of lowly expressed miRNAs through various pathways, thereby inhibiting the development and progression of cancer.^{19,20} Recently, many studies have shown that numerous miRNAs are either overexpressed or underexpressed in osteosarcoma and are often associated with the entire process of tumor development.²¹ Therefore, this paper reviews the research progress of miRNAs in osteosarcoma to explore the potential application of miRNAs in the diagnosis and treatment of osteosarcoma and prognosis and lay the theoretical foundation for the update of clinical diagnosis and treatment methods.

The Production and Biological Functions of miRNAs

The Generation of miRNAs

Most miRNAs are produced through the typical miRNAs biogenesis pathway.²² MicroRNA genes are usually expressed as single transcription units, and mature miRNAs are produced through 2 steps, nuclear and cytoplasmic synthesis, and this

process requires the involvement of multiple enzymes.²³ The formation of miRNAs begins in the nucleus, where most miRNAs are transcribed by RNA polymerase II (Pol II), producing primary miRNAs (pri-miRNAs) that encode miRNAs sequences in a “hairpin” structure.²⁴ Pri-miRNAs are excised in the nucleus by the Drosha-DGCR8 complex to produce precursor miRNAs (pre-miRNAs) that retain their “hairpin” structure.^{23,25} The pre-miRNAs are then translocated to the top of the hairpin to produce precursor miRNAs (pre-miRNAs) that still retain their “hairpin” structure. The pre-miRNAs are then transported to the cytoplasm by the transporter protein Exportin-5. In the cytoplasm, pre-miRNAs bind to the double-stranded RNA (dsRNA) endoribonuclease/transactivation response (TAR) RNA binding protein (DICER/TRBP) complex, which cleaves the hairpin and produces a complementary RNA duplex. One of the nucleotide chains preferentially binds to the RNA-induced silencing complex (RISC) containing the Argonaut (Ago) protein to form the miRNA-RISC complex, while the other chain is degraded. The choice of the strand is largely dependent on the local thermodynamic stability of the miRNAs duplex—RISC tends to load onto the less stably paired 5' end. This thermodynamic difference occurs partly because miRNAs tend to start with uracil and partly because miRNAs duplexes contain mismatches and bumps that favor the loading of miRNAs strands into RISC. Finally, Argonaut proteins guide the miRNAs strand to the 3'-untranslated region (3'-UTR) of the target sequence on the mRNAs and allow it to bind to the RISC.^{26–29} In the middle, the structural characteristics of the 5'-UTR in mRNA, including the upstream UTR, secondary structure, start codon, open-reading frame (ORF), and ribosome binding sites, regulate the efficiency of translation.³⁰ Although most transcription regulation of mRNA takes place in 5'-UTRs, the 3'-UTRs of mRNAs play an important role in the posttranscriptional regulation of gene expression through the presence of cis-acting elements and through interaction with miRNAs.^{30,31} The commonly recognized function of miRNAs is through interacting with the 3'-UTR regions of targeted mRNAs and hinders gene expression, which subsequently leads to the alteration of multiple cellular functions, including growth, migration, and angiogenesis.³² Moreover, the 3'-UTR of mRNA contains terminal processing signals, and its miRNA targeted binding region and AU-rich region regulate gene expression at the translational level.³³ In addition, it has been widely reported that miRNA expression is remarkably deregulated in cancer.³⁴ So any mutations in miRNAs has the power to attack specific domains such as the coding region of mRNA domains that serve as binding targets and result in oncogenic or tumor suppressor activity of the miRNA.³⁴ miRNAs can also be secreted outside the cell and may be involved in intercellular communication.³⁵ Extracellular miRNAs are stably present within vesicles, including apoptotic vesicles and exosomes, or bound to RNA-binding proteins such as AGO and HDL.³⁶ The biogenesis pathway of miRNAs is depicted in Figure 1.

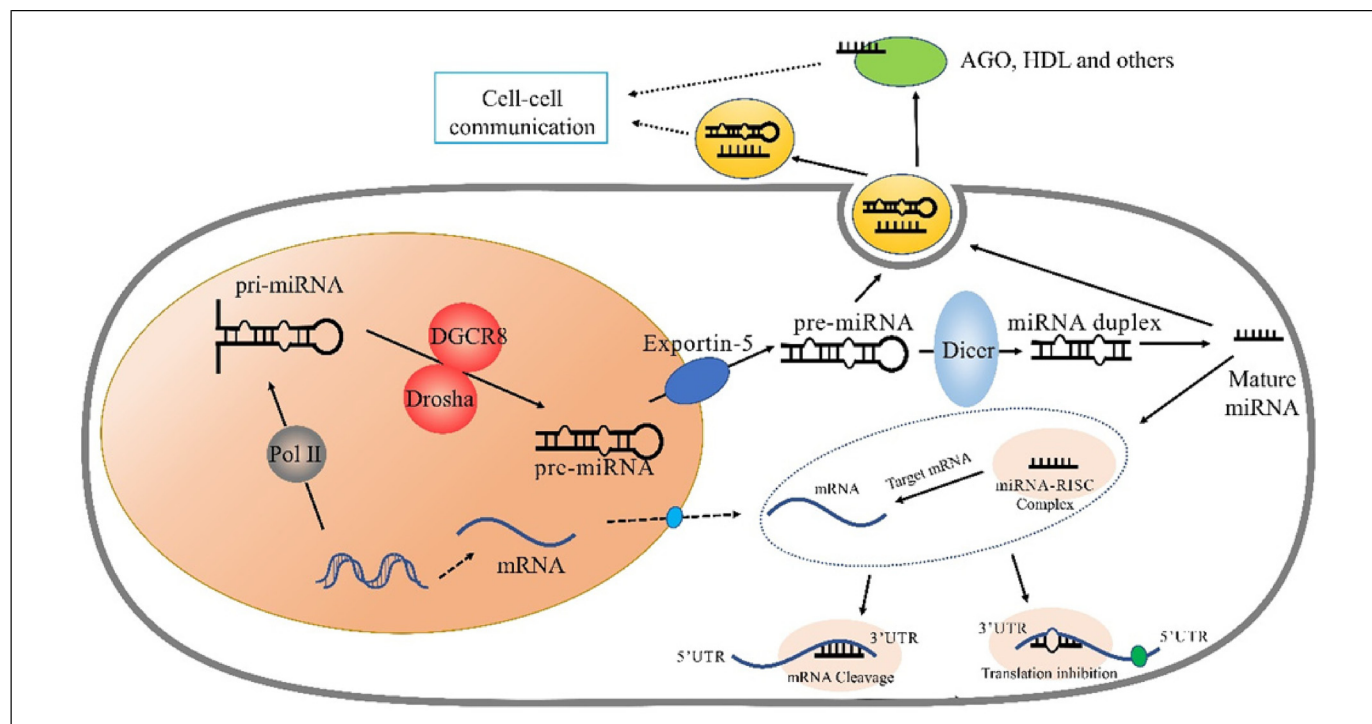


Figure 1. The biogenesis of miRNA begins in the nucleus. First of all, the miRNA gene is transcribed by pol II to produce pri-miRNA. Subsequently, a complex of Drosha and DGCR8 proteins cleave pri-miRNA to produce pre-miRNA. Then, pre-miRNA is transported to the cytoplasm by Exportin-5, and Dicer guides TRBP and PACT to further process pre-miRNA to produce miRNA duplex. Finally, the guide chain of miRNA duplex and AGO protein is loaded into RISC to recognize the target mRNA through sequence complementarity, and gene silencing is caused by mRNA degradation or translation inhibition. In addition, miRNA is also secreted out of the cell and may be involved in intercellular communication.

The Biological Functions of miRNAs

Among the various types of small molecule RNAs, miRNAs have a wide range of gene regulatory functions.³⁷ Although miRNA genes only account for 1% to 3% of the human genome.³⁸ However, it is estimated that miRNAs regulate about 30% to 60% of protein-coding genes in the human body.^{39,40} miRNAs can be detected in cells and various biological fluids, such as plasma, serum, and follicular fluid.⁴¹⁻⁴³ Mature miRNAs can form complexes that induce RNA to silence through 2 processes: (1) when miRNAs are not perfectly complementarily paired with sequences in the 3'-UTR of target mRNAs, miRNAs inhibit protein synthesis by repressing translation or promoting the deadenylation and decay of mRNAs; (2) when miRNAs are almost perfectly complementarily paired with target mRNAs, these mRNAs inhibit protein synthesis by repressing translation or promoting the decay of mRNAs. When paired, these mRNAs are cleaved and degraded by nucleic acid endonucleases.⁴⁴ MiRNAs can regulate normal cellular genetic activities such as differentiation, proliferation, apoptosis, intracellular signaling, gene expression, tissue development, material metabolism, and other aspects through both these 2 ways.^{45,46} For example, miRNA-143 can promote angiogenesis and osteoblast differentiation *in vivo*⁴²; miRNA-375 can regulate mammalian islet cell development

and insulin secretion⁴³; miRNA-181 can control mammalian hematopoietic stem cell differentiation,⁴⁷ and so on.

Numerous studies have revealed that a single mRNA may contain multiple binding sites complementary to multiple miRNAs, thus forming a complex regulatory network that participates in various biological mechanisms to regulate biological processes.⁴⁸ For example, circ-XPO1 regulates XPO1 expression by using miR-23a-3p, miR-23b-3p, miR-23c, and miR-130a-5p as competing endogenous RNAs (ceRNAs).⁴⁹ In addition, individual miRNAs can also target various mRNAs to exert regulatory effects on biological gene activity.⁵⁰ As a result of this complex regulatory network of miRNAs, one or more specific miRNAs can affect the expression of many downstream target genes through direct or indirect regulation and thus perform a wide range of biological functions.

In addition, miRNAs have been closely associated with human cancer development. Since the first evidence of the involvement of miRNAs in tumorigenesis was discovered in 2002,⁵¹ for example, miRNA-372 and miRNA-373 in testicular germ cell tumors can neutralize p53-mediated CDK inhibition by directly inhibiting the expression of the tumor suppressor LATS2, thereby promoting tumorigenesis.⁵² Furthermore, high expression of miRNA-421 was found in various tumors, including neuroblastoma, pancreatic cancer, and prostate cancer.⁵³⁻⁵⁵ This suggests that miRNA-421 can act as a pro-

carcinogenic gene in human cancers.⁵⁶ However, miRNA-125a-5p can act as a tumor suppressor miRNA, and its regulatory mechanism is to regulate cell proliferation by regulating the cell cycle during the G1/S transition, thereby inhibiting the development of head and neck cancer.⁵⁷ The overexpression of miRNA-217 in hepatocellular carcinoma cells inhibits proliferation and migration and induces apoptosis.⁵⁸ These findings on the pro- or anticancer effects of miRNAs in tumor tissues may provide new ideas for clinical diagnosis, drug delivery, and targeted therapy.

The Role and Mechanism of miRNAs in the Development of Osteosarcoma Cells

The Role of miRNAs in the Proliferation of Osteosarcoma Cells

An in-depth study of the molecular mechanisms involved in the development of osteosarcoma is essential to improving the survival rate of patients with osteosarcoma.⁵⁹ The regulatory role of miRNAs in the development of osteosarcoma is diverse. Some miRNAs have been shown to act as oncogenes, others can act as a tumor suppressor by specifically regulating the expression of their target genes.⁵⁹ The regulatory role of miRNAs in the development of osteosarcoma is related to the activation status of various signaling pathways and the expression levels of some protein molecules.⁶⁰ Therefore, studying the signaling pathways including PI3K/AKT, PTEN/AKT, Wnt/ β -catenin, Wnt, and NF- κ B, and proteins including P21, IRF2, VEPH1, and BCL9L, may contribute to the development of osteosarcoma studies. We summarized the progress of miRNAs regulating osteosarcoma through these pathways (Figure 2).

miRNAs promote the proliferation of osteosarcoma cells. Various studies have shown that miRNAs play a role in developing osteosarcoma.⁶¹ In this process, the role of miRNAs in promoting osteosarcoma proliferation is exerted through specific molecular mechanisms. Therefore, it is of great clinical value and significance to elucidate the regulatory mechanisms of miRNAs in this aspect of osteosarcoma. The regulatory mechanisms, upstream regulatory factors, and other malignant tumors with abnormal expression of some miRNAs that promote the progression of osteosarcoma are shown in Table 1.

P21 is involved in various cellular metabolic pathways, plays an essential role in promoting tumor proliferation, and is considered a promising target molecule in anticancer research.⁸¹ The expression of miRNA-95-3p in osteosarcoma is significantly correlated with the clinical stage. The overexpression of miRNA-95-3p increases cell growth and inhibits apoptosis of osteosarcoma cells through TGF- β /CDKN1A/p21/cyclin D1.⁸² P21 has an extensive inhibiting effect on CDKs (cyclin-dependent kinases), especially on CDK2 and CDK4. The Cyclin-CDK complex can help cells quickly pass checkpoints of the cell cycles, and its overexpression can reduce cell mass and promote proliferation and transformation. P21waf1/cip1

plays a role in checkpoint control in inhibiting Cyclin-CDK.⁸³ The expression of CDKN1A/p21 is tightly controlled at the transcriptional level primarily by the tumor suppressor protein p53 in response to DNA damage.^{83,84} In addition, the overexpression of miRNA-95-3p promotes tumor cell proliferation and migration in cultured cells and tumor growth in xenograft mouse models through negative posttranscriptional regulation of p21 by directly targeting the 3'-UTR.⁸⁵

At the same time, miRNAs can also inhibit the development of osteosarcoma by targeting many proteins and molecules. For example, it has been found that miRNA-92a can promote the occurrence and development of hepatocellular carcinoma and cervical cancer expression.^{86,87} In osteosarcoma, miRNA-92a can promote osteosarcoma cell growth and cell cycle progression by directly targeting and inhibiting FBXW7 while inhibiting apoptosis.⁸⁸ In addition, miRNA-92a can negatively regulate DKK3 expression and thus promote osteosarcoma cell progression.⁸⁹

PI3K/AKT is another critical intracellular signaling pathway with essential regulatory functions in the cell cycle. This signaling pathway is activated in various tumors, including osteosarcoma.⁹⁰⁻⁹² The signaling pathway promotes proliferation, survival, and epithelial-mesenchymal transition (EMT) while suppressing apoptosis.⁹³ The carcinogenesis of miRNA-23b-3p in colon adenocarcinoma,⁹⁴ hepatocellular carcinoma,⁹⁵ and esophageal squamous cell carcinomas⁹⁶ has been confirmed. In osteosarcoma, miRNA-23b-3p can promote osteosarcoma by targeting VEPH1/PI3K/AKT signaling pathway.⁹⁷ MiRNA-23b-3p can also promote the apoptosis and inhibit the proliferation and invasion of osteosarcoma cells by targeting SIX1.⁹⁸ In another study, miRNA-18a-5p can promote the invasion and migration of OS cells by inhibiting IRF2 expression.⁹⁹ However, LncRNA FER1L4 can downregulate the expression of miRNA-18a-5p and block PI3K/AKT signaling pathway, effectively inhibiting the progress of OS.¹⁰⁰

PTEN is a well-established tumor suppressor gene and is one of the most commonly mutated genes in various human cancers.⁸⁹ PTEN acts as a lipid phosphatase to dephosphorylate the 3' position of phosphatidylinositol-3,4,5-trisphosphate (PIP3), the product of a potent proto-oncogenic phosphatidylinositide 3-kinase (PI3K), and triggers activation of the PI3K pathway. AKT kinase plays a key role in cell survival, cell proliferation, angiogenesis, and anabolism and is a major target for cancer therapy. One of the main targets for cancer therapy.¹⁰¹ miRNA-21, miRNA-216, miRNA-155, and miRNA-524 can enhance the proliferation of osteosarcoma cells by targeting PTEN to activate the PI3K/Akt signaling pathway.¹⁰²⁻¹⁰⁵ The PI3K/AKT signaling pathway is unregulated in most localized diseases and in 100% of advanced diseases, implying that alterations in this pathway may be a prerequisite for inhibiting osteosarcoma progression.¹⁰⁶

Finally, the activation of the WNT/ β -catenin signaling pathway has been highly conserved throughout evolution and plays a key role in regulating tissue development and maintaining homeostasis *in vivo*.¹⁰⁷ In human cancers, Wnt/ β -catenin signaling is highly activated, such as cervical

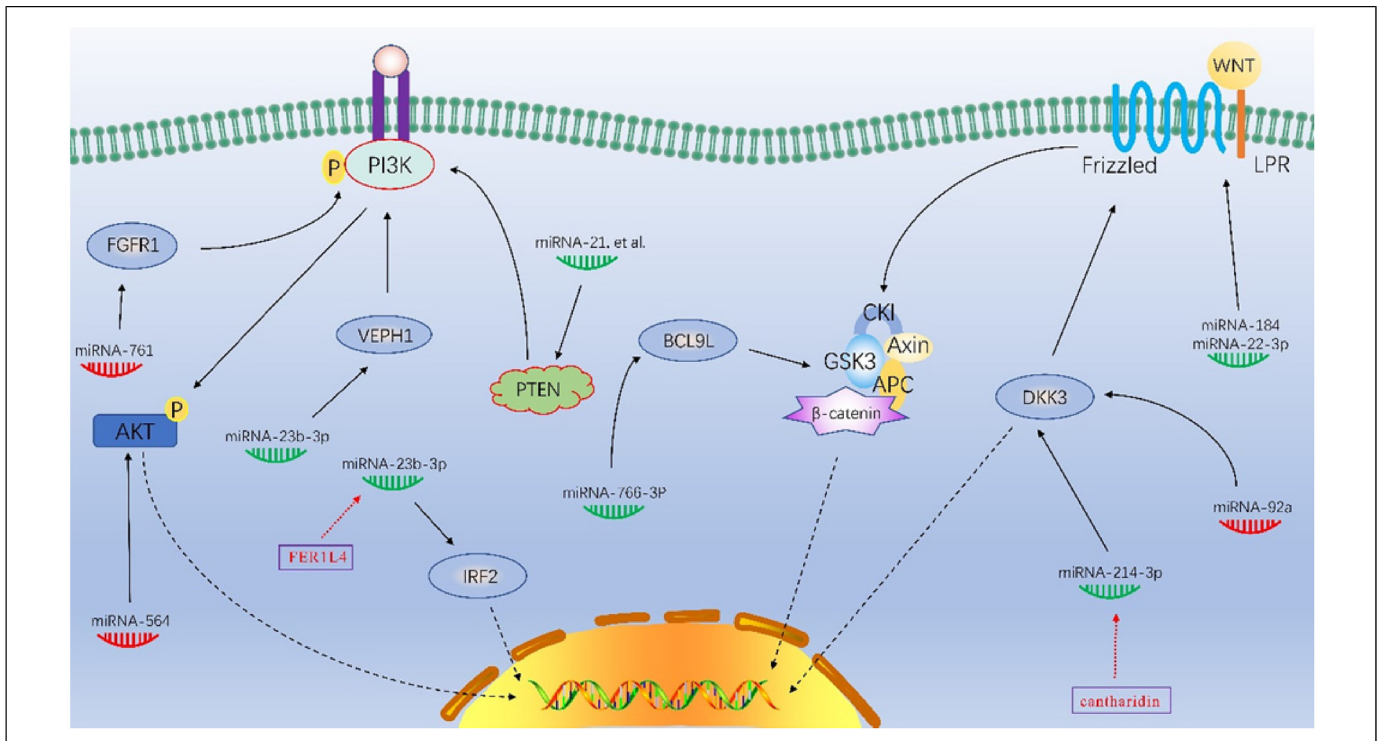


Figure 2. miRNAs can regulate PI3K/Akt and Wnt/ β -catenin signaling pathways in many ways, thus affecting the progression of osteosarcoma (Green represents the promotion of osteosarcoma; Red represents the inhibition of osteosarcoma).

cancer,¹⁰⁸ nasopharyngeal carcinoma,¹⁰⁹ and gastric cancer.¹¹⁰ And genetic and epigenetic deregulation of Wnt/ β -catenin signaling contributes to human cancer, which has led to the development of extensive approaches targeting Wnt/ β -catenin signaling as cancer therapies.¹¹¹ The recent research found that the inhibitory effect of baicalein,¹¹² alantolactone,¹¹³ and schisandrin B¹¹⁴ on osteosarcoma through Wnt/ β -catenin signaling pathway. Some studies have found several miRNAs whose expression is specific to osteosarcoma, which can promote its occurrence and development by targeting Wnt/ β -catenin signaling pathway. For example, miRNA-377-3p can inhibit osteosarcoma progression by targeting Cullin-1 (CUL1) and regulating Wnt/ β -catenin signaling pathway.¹¹⁵ In addition, miRNA-184 and miRNA-22-3p can also target this signaling pathway to promote the progress of osteosarcoma.^{115,116} It has been studied that miRNA-214-3p can mediate Wnt/ β -catenin/LEF1 signaling activation by targeting DKK3 to promote the oncogenesis of osteosarcoma, but this role of miRNA-214-3p is inhibited by cantharidin.¹¹⁷ This indicates that cantharidin may be a prospective candidate for osteosarcoma treatment by targeting miRNA-214-3p/DKK3/ β -catenin signaling pathways.

miRNAs inhibit the proliferation of osteosarcoma cells. In addition, a large number of studies have found many miRNAs are under-expressed in osteosarcoma.⁶¹ These miRNAs can be amplified in the treatment of osteosarcoma by means of reducing the proliferation of osteosarcoma and thus inhibiting the tumor inhibition. By studying the targets and signaling pathways of these

miRNAs, it may be possible to provide therapeutic targets for this purpose. The regulatory mechanisms, upstream regulatory factors, and other malignant tumors with abnormal expression of some miRNAs that inhibit the progression of osteosarcoma are shown in Table 2.

Astrocyte elevated gene-1 (AEG-1) is associated with various oncogenic signaling pathways such as Wnt and NF- κ B, regulates tumor growth and metastasis, and plays an important role in osteosarcoma.¹³³ For example, miRNA-342-3p and miRNA-448 inhibit the progression of osteosarcoma by targeting AEG-1 to inhibit the Wnt and NF- κ B signaling pathways.^{134, 135} The NF- κ B signaling pathway is responsible for cell proliferation, apoptosis prevention, and transcriptional regulation of various genes in response to damaging agents and cytokines and plays an important role in the pathogenesis of osteosarcoma.¹³⁶ For example, miRNA-29a inhibits the proliferation of osteosarcoma cells by negatively regulating its target DNMT3B, which inhibits the SOCS1/NF- κ B signaling pathway.¹³⁷ Meanwhile, downregulation of miRNA-155 expression also inhibited osteosarcoma cell proliferation and induced apoptosis through the NF- κ B signaling pathway.¹³⁸

It has also been found that inhibition of PI3K/Akt and Wnt/ β -catenin signaling pathways can also exert anti-osteosarcoma effects. For example, miRNA-761 inhibits the proliferation, migration and invasion of osteosarcoma cells and promotes apoptosis by directly targeting FGFR1 and inactivating the PI3K/Akt pathway. miRNA-564 downregulates osteosarcoma and inhibits the proliferation of osteosarcoma cells by targeting Akt.¹³⁹ In addition, miRNA-152 inhibits osteosarcoma cell proliferation by directly targeting the Wnt/ β -catenin signaling

Table 1. miRNAs Which Are Downregulated in Osteosarcoma.

miRNAs	Gene targets	Signaling pathways	Effects	miRNA regulators	Other tumors
miRNA-4295 ⁶²	PTPN14	YAP1	Restrict the growth and invasion of osteosarcoma cells	-	Gastric cancer ⁶³ Head and neck squamous cell carcinoma ⁶⁴
miRNA-218 ⁶⁵	Runx2	-	Inhibit proliferation, migration, and invasion of osteosarcoma cells	circEIF4G2 ⁶⁶ lncRNA SNHG10 ⁶⁷	Glioma ⁶⁸ Non-small cell lung cancer ⁶⁹
miRNA-744-5p ⁷¹	TGFB1	p38 MAPK	Suppress tumorigenesis and metastasis of osteosarcoma	-	Multiple myeloma ⁷⁰ Hepatocellular carcinoma ⁷² Glioblastoma ⁷³ Multiple myeloma ⁷⁴ Colorectal cancer ⁷⁵
miRNA-139 ⁷⁶	ITGAV	-	Inhibit proliferation, migration, and invasion of osteosarcoma cell line MG63	LncRNA SNHG20 ⁷⁷ LncRNA LINC00858 ⁷⁸	Pancreatic cancer ⁷⁹ Prostate cancer ⁸⁰

Table 2. miRNAs Which Are Upregulated in Osteosarcoma.

miRNAs	Gene targets	Signaling pathways	Effects	miRNA regulators	Other tumors
miRNA-182-5p ¹¹⁸	NKAPL	Notch	Regulate the cell cycle and promote proliferation of osteosarcoma	lncRNA SNHG10 ¹¹⁹	Bladder cancer ¹²⁰ Hepatocellular carcinoma ¹²¹ Clear cell renal cell carcinoma ¹²²
miRNA-628-5p ¹²³	IFI44L	-	Promote the growth and movement of osteosarcoma	-	Cervical carcinoma ¹²⁴ Colorectal cancer ¹²⁵ Pancreatic ductal adenocarcinoma ¹²⁶
miRNA-4295 ¹²⁷	IRF1	-	Promote proliferation, migration, and invasion of osteosarcoma	-	Gastric cancer ⁶³ Head and neck squamous cell carcinoma ⁶⁴
miRNA-221 ¹²⁸	FBXW11	Wnt	Promote cell proliferation and inhibit apoptosis in osteosarcoma	Lnc RNA GAS5 ¹²⁹	Breast cancer ¹³⁰ Thyroid cancer ¹³¹
miRNA-221 ¹³²	CDKN1B/ p27	-	Regulate proliferation, apoptosis, migration, and invasion of osteosarcoma		

pathway in a DKK1-dependent manner.¹⁴⁰ LncRNA PVT1 can be used as its upstream regulatory factor to enhance the chemical resistance of osteosarcoma.¹⁴¹ However, LncRNA HAGLROS could promote osteosarcoma progression by sponging miRNA-152 to promote ROCK1 expression.¹⁴² There are also some miRNAs that inhibit the progression of osteosarcoma, as shown in Table 1. Therefore, the discovery of these signaling pathways will help to enhance the role of these cancer-inhibiting miRNAs in osteosarcoma by using drugs or other auxiliary means in the future and provide a basis for the treatment of osteosarcoma.

The Role of miRNAs in Invasion and Metastasis of Osteosarcoma Cell

Metastasis is a complex process that describes the spread of cancer cells from the primary tumor site to other organs.¹⁴³ Metastasis is a complex multistep event and invasion of a

tumor through the basement membrane into surrounding tissue is the first crucial step of the metastasis process. In other words, migration away from the primary tumor to adjacent tissues or organs, followed by migration of cancer cells through the stroma to blood vessels or lymphatic vessels to be transported to other organs where metastasis eventually occurs.¹⁴⁴ Metastasis is the hallmark of cancer and the leading cause of death in cancer patients.¹⁴⁵ In recent years, miRNAs have been reported to be involved in the invasion and metastasis of various tumors and have promised applications in many different cancers, including osteosarcoma.⁵⁹ Therefore, inhibiting osteosarcoma invasion and metastasis by selectively promoting and blocking fundamental molecular mechanisms associated with miRNAs may be a potential strategy for osteosarcoma treatment.

Similar to other cancer types, osteosarcoma cells undergo EMT in the early metastatic stages to facilitate their movement

and migration, and once osteosarcoma cells arrive and settle in a new metastatic site, such as the lung region, bone cancer cells, they can undergo EMT to allow them to adapt to and grow in their new environment.¹⁴⁶ Therefore, strategies that can prevent the development of early EMT osteosarcoma may help inhibit the spread of bone cancer. miRNA-486 is downregulated in many tumor cells. It acts as a tumor suppressor, regulating osteosarcoma cell invasion and EMT by targeting PIM1.¹⁴⁷ In addition, miRNA-766-3p can target BCL9L to inhibit EMT and metastasis of osteosarcoma through the β -catenin signaling pathway. It has also been shown that the miRNA-135b-TAZ positive feedback loop promotes EMT and migration and invasion of osteosarcoma cells and can also block metastasis of osteosarcoma cells by inhibiting TAZ protein expression and activity.¹⁴⁸

In addition, several functional targets have been identified in osteosarcoma cells that bind to miRNAs, and miRNAs can inhibit osteosarcoma invasion and metastasis by binding to these targets. For example, E2F5 is a direct-binding target of miRNA-154-5p, and the overexpression of miRNA-154-5p can exert an inhibitory effect on osteosarcoma cell invasion and metastasis by inhibiting E2F5.¹⁴⁹ The expression of miRNA-25-3p is downregulated in osteosarcoma tissue samples and cell lines, and the overexpression of miRNA-25-3p could directly target SOX4 to inhibit osteosarcoma cell invasion and metastasis.¹⁵⁰ Besides, miRNA-92a inhibitors can inhibit osteosarcoma invasion and metastasis by upregulating DKK3 expression.⁸⁹ Downregulation of miRNA-4660 in osteosarcoma cells and reduction of the inhibition of MAFG expression by miRNA-4660 can also inhibit the invasion and metastasis of osteosarcoma cells.¹⁵¹ By investigating and exploiting these potent targets of miRNAs, it may provide a strategy to inhibit osteosarcoma invasion and metastasis.

The Role of miRNAs in the Regulated Cell Death of Osteosarcoma

Selective elimination of cancer cells without damaging nonmalignant cells is essential in cancer treatment.¹⁵² Unlike accidental cell death or an uncontrolled and passive process, regulated cell death can occur through a range of molecular mechanisms and signaling.¹⁵³ Therefore, rationally promoting regulated cell death in osteosarcoma cells may offer hope for osteosarcoma treatment. Regulated cell death can be further classified into subtypes based on the signaling and degradation pathways and the morphological characteristics of the biochemical endpoints of the dead cells.¹⁵³ Apoptosis was once thought to be the only form of regulated cell death. Still, recent studies have demonstrated that iron death and cell scorching are 2 other forms of regulated cell death.¹⁵⁴

Apoptosis. Apoptosis is a regulated form of cell death that is triggered primarily by the activation of proteases of the cysteine aspartase family.¹⁵⁵ Apoptosis is a process of regulated cell

death during normal development and aging. It is a mechanism for maintaining stable cell numbers.¹⁵⁶ It was found that miRNAs play an essential role in promoting apoptosis of osteosarcoma cells. MiRNA-204-5p, an oncogenic miRNA in osteosarcoma, promotes apoptosis and inhibits the migration of osteosarcoma via targeting EBF2.¹⁵⁷ Moreover, miRNA-497, for instance, can activate P21 expression by inhibiting the expression of MAPK/Erk signaling pathway and promoting the apoptosis of osteosarcoma cells.¹⁵⁸ MiRNAs have also been found to inhibit apoptosis in osteosarcoma cells. For example, miRNA-216 is an oncogene in osteosarcoma. miRNA-216 knockdown promotes apoptosis through PTEN/PI3K/AKT and related downstream genes P53 and MMP-2/9, thus exerting antitumor effects.¹⁰⁴ High expression of miRNA-1226-3p was associated with lower overall survival. It can inhibit the proliferation, migration, and invasion of osteosarcoma cells by targeting TRIAP3 and promote the apoptosis of osteosarcoma cells.¹⁵⁹ Successful treatment of osteosarcoma requires selective destruction of osteosarcoma cancer cells and attempting to selectively induce apoptosis in these osteosarcoma cells. It may offer hope for osteosarcoma treatment.

Ferroptosis. Despite the insight into the expression and function of some miRNAs in osteosarcoma, there are still many difficulties to be overcome by miRNAs inducing apoptosis in osteosarcoma cells and thus improving the treatment of osteosarcoma. Studies on ferroptosis in osteosarcoma may provide some effective ways to address these problems.^{104,159,160} It plays a vital role in cancer and many degenerative diseases.^{72,161} Ferroptosis was initially described as a non-apoptotic form of cell death characterized by impaired cellular uptake of cysteine, glutathione (GSH) depletion, and iron-dependent lipid peroxidation.¹⁶² Ferroptosis differs morphologically and biochemically from apoptosis. For example, ferroptosis is accompanied by cell swelling and plasma membrane rupture. In contrast, apoptotic cells usually exhibit cell shrinkage and blistering of the plasma membrane.¹⁶³ With the discovery of various regulators and pathways, fundamental research into ferroptosis is rapidly expanding. Targeting ferroptosis may help develop novel therapeutic agents for treating malignancies offering new opportunities to treat various pathological conditions and diseases.^{155,164} The mechanism of action related to ferroptosis in osteosarcoma is shown in.¹⁶⁵ Recently, it was also found that miRNA-1287-5p could inhibit the progression of osteosarcoma cells by inhibiting GPX4 to promote ferroptosis in osteosarcoma cells.¹⁶⁶ However, there are few literatures about the mechanism of miRNAs regulating ferroptosis in osteosarcoma, and a lot of research is needed to make up for this deficiency in the future.

Pyroptosis. More and more studies have recently found that pyroptosis can also be involved in the progression of osteosarcoma and may be a viable target for future osteosarcoma therapy.¹⁶⁷ The study also found that pyroptosis is an inflammatory form of cell death that releases specific cytokines and other immunostimulatory factors that protect mammals from

pathogens.¹⁶⁸ As a type of regulated cell death, pyroptosis is characterized by cell swelling, lysis, and the release of many pro-inflammatory factors.¹⁶⁹ Pyroptosis is also a highly immunogenic form of cell death, which can cause local inflammation and attract the infiltration of inflammatory cells, providing an excellent opportunity to alleviate the immunosuppression of the tumor microenvironment and induce the systemic immune response to treat solid tumors.¹⁷⁰ So pyroptosis can effectively eliminate malignant cells by specifically activating regulated cell death at their site of action, offering a new strategy for cancer treatment.¹⁵⁴ In osteosarcoma, it has been found that miRNA-181a can be aberrantly elevated in osteosarcoma tissues and cells. That knockdown of miRNA-181a can activate NLRP3-dependent pyroptosis to inhibit cell proliferation and invasion.¹⁷¹ However, some studies have shown that pyroptosis is related to the negative effects of chemotherapy and radiotherapy. For example, chemotherapy and radiotherapy in tumor treatment may promote cell death of immune cells or hematopoietic cells, resulting in the damage of antitumor immune function.¹⁷⁰ Therefore, it is crucial to prevent normal cells from scorching induced by anticancer therapy to reduce the side effects of traditional therapy.

Overall, miRNAs regulate the occurrence and progression of osteosarcoma through their involvement in cell proliferation, invasion, metastasis, and apoptosis. These findings provide potential therapeutic targets and prognostic biomarkers for the involvement of miRNAs in the effective treatment of osteosarcoma. However, the specific mechanisms of action of many of these miRNAs are still unclear and need to be further investigated. Studies on the mechanisms of competitive endogenous RNAs (ceRNAs) have been continuously reported in recent years.¹⁷² And more recently, the lncRNA-miRNA-mRNA axis,¹⁷³ circRNA-miRNA-mRNA regulatory networks,¹⁷⁴ etc, lay a theoretical foundation for in-depth study of the specific action mechanism of miRNAs, which can also provide better strategies for the diagnosis and treatment of osteosarcoma. Through continuous research on the role and mechanism of miRNAs in the development of osteosarcoma, we will guide the continuous improvement of the theoretical system of miRNAs, so that miRNAs can play a more significant role in clinical benefits.

The Potential Utility of miRNAs in Osteosarcoma

The Role of miRNAs in the Diagnosis and Prognosis of Osteosarcoma

Extracellular miRNAs (circulating miRNAs) can be stabilized by binding to proteins or encapsulated in vesicles, to be examined in biological fluids such as whole blood, plasma, and serum.¹⁷⁵ The high stability and ease of detection of miRNAs make them a good source of biomarkers for various diseases.³⁶ They are a good source of biomarkers for various diseases. Altered levels of circulating miRNAs are associated with tumor growth, progression, metastasis, and drug resistance,

suggesting that they could potentially be used to optimize therapeutic approaches for tumor patients.¹⁷⁶ Analysis of circulating miRNA levels in patients' blood could therefore provide a new approach to the diagnosis or prognosis of cancer.

Existing studies have found that miRNA-139-5p expression in the serum of patients with osteosarcoma is significantly lower than that of healthy individuals and that miRNA-139-5p is more pronounced in patients with distant metastases or higher clinical stage.¹⁷⁷ In addition, increased serum levels of miRNA-542-3p are significantly associated with progressive tumor staging and shorter survival.¹⁷⁸ In addition, low plasma miRNA-375 levels are correlated with large tumor size, advanced clinical stages, positive distant metastasis, and poor tumor response to preoperative chemotherapy.¹⁷⁹ Other studies have found a significant reduction in serum miRNA-375 expression in osteosarcoma patients. Other studies have found that low serum miR-194 expression is strongly associated with positive metastasis, clinical stage, and poor survival and that serum miRNA-194 levels are significantly higher in osteosarcoma patients after surgery.¹⁸⁰ Therefore, detection of serum miRNA-139-5p, miRNA-542-3p, miRNA-375, and miRNA-194 expression can be used as reliable biomarkers to determine the diagnosis and prognosis of osteosarcoma.

As a promising biomarker, circulating miRNAs may open the door for diagnosing and prognosis of osteosarcoma and guide designing treatment plans and personalized surgical plans for osteosarcoma. However, the relatively difficult kinetic detection, low expression level, and short nucleotide sequence of circulating miRNAs challenge circulating miRNAs as a diagnostic and prognostic marker of osteosarcoma. Firstly, it must be clear that this difference in circulating miRNAs is not disease-specific. For example, it has been found that the expression of circulating miRNA-139-5p in osteosarcoma and lung cancer patients is downregulated.^{177,181} Moreover, some drugs can target their targets and affect the expression of downstream miRNAs. For example, demethylating drugs such as 5-azacytidine (5-aza) can inhibit the proliferation and migration of hepatocellular carcinoma cells by upregulating the expression of miRNA-139-5p.¹⁸² Therefore, the accuracy of circulating miRNAs will be interfered by diseases complicated with osteosarcoma or the use of some drugs. In addition, the low expression level of miRNAs in cells makes them exist less in biological fluids.¹⁸³ And miRNAs are sequenced with short base lengths, which lead to a high possibility of sequence similarity between homologous miRNAs.¹⁸³ So the potential application value of circulating miRNAs in osteosarcoma still needs further exploration.

The Role of miRNAs in Drug Resistance in Osteosarcoma

The standard treatment for patients with osteosarcoma includes chemotherapy for 10 weeks before surgery, surgical resection, and 20 weeks after surgery. Although there is some variation in chemotherapy regimens worldwide, the most commonly used regimens include cisplatin, doxorubicin, and high-dose methotrexate.¹⁸⁴ Long-term use of these chemotherapeutic

agents in patients with osteosarcoma tends to induce genetic mutations and resistance to the drugs, thus making osteosarcoma more difficult to treat.¹⁰ In recent decades, treatment outcomes for osteosarcoma patients have not improved with newer chemotherapy drugs.⁶¹ Many studies have found that miRNAs play an important role in improving drug resistance of osteosarcoma. Research in this area could provide a reliable and effective solution for future exploration of various related drugs and clinical treatment of osteosarcoma.¹⁸⁵

miRNAs can be used as adjuvants in chemotherapy. Drug resistance remains a key challenge in current cancer chemotherapy, and miRNAs are strongly associated with changes in cancer chemosensitivity.¹⁸⁶ The use of miRNAs as chemotherapeutic adjuvants in the treatment of cancer would ameliorate this problem. For example, overexpression of miRNA-329-3p can also increase the sensitivity of osteosarcoma cells to cisplatin by targeting LDHA to inhibit glucose metabolism.¹⁸⁷ MiRNA-216b is significantly lower in osteosarcoma tissues than in paraneoplastic tissues, and increased miRNA-216b expression is associated with higher overall survival in osteosarcoma patients. Further studies revealed that JMJD2C is a target of miRNA-216b and that miRNA-216b could enhance cisplatin-induced apoptosis by regulating the JMJD2C/HIF1 α /HES1 signaling axis in osteosarcoma cells.¹⁸⁸ Therefore, miRNA-216b can be used as a chemotherapeutic adjuvant than cisplatin to effectively treat osteosarcoma. In addition, low expression of miRNA-382 is closely associated with low survival rates in osteosarcoma patients. A related study found that overexpression of miRNA-382 could target and negatively regulate the expression of KLF12 and HIPK3, thereby enhancing the sensitivity of osteosarcoma cells to methotrexate and inhibiting the growth of osteosarcoma cells.¹⁸⁹ This resulted in enhanced sensitivity to methotrexate and inhibition of osteosarcoma cell growth. Increased miRNA-221 expression in osteosarcoma correlates with tumor staging, metastasis, and response to chemotherapy. miRNA-221 has been shown to induce resistance to adriamycin-induced apoptosis in osteosarcoma cells by activating the STAT3 signaling pathway and upregulating the expression of P-GP and BCL-2 proteins. STAT3-IN-3, a chemical inhibitor of the STAT3 signaling pathway, could effectively interfere with this effect and thus reverse the miRNA-221-induced resistance of osteosarcoma cells to adriamycin.¹⁹⁰ As the clinical treatment of osteosarcoma often uses a combination of chemotherapeutic agents, the development of multidrug resistance in osteosarcoma will seriously affect the efficacy of chemotherapy in osteosarcoma.¹⁹¹ For example, SDC2 is a positive regulator of multidrug resistance in osteosarcoma. miRNA-20a-5p can regulate multidrug resistance to adriamycin and etoposide in osteosarcoma by directly acting on and inhibiting the target gene Syndecan-2(SDC2).¹⁹² Overexpression of miRNA-20a-5p in osteosarcoma may help to overcome osteosarcoma drug resistance and be a candidate for preventing osteosarcoma chemoresistance.

miRNAs are involved in the regulation of autophagy in osteosarcoma. Autophagy is a cytoprotective mechanism that promotes cell survival by maintaining energy production

under stressful conditions such as chemotherapy, hypoxia, and metabolic stress, maintaining intracellular homeostasis, development, differentiation, and processes essential for cell growth and survival.¹⁹³ However, in the presence of anticancer drugs, osteosarcoma cells can activate autophagy as a protective mechanism, making chemotherapy treatment of osteosarcoma difficult.¹⁹⁴ This would make chemotherapy for osteosarcoma difficult. Existing studies have shown that miRNAs can enhance chemoresistance or chemosensitivity through autophagy regulation in various tumors such as osteosarcoma, and inhibition of autophagy may be a new approach to improve the efficiency of chemotherapy in cancer treatment.¹⁹⁵ miRNA-22 can increase the sensitivity of osteosarcoma cells to cisplatin treatment by regulating autophagy-related genes. MTDH has been identified as a target of miRNA-22 in osteosarcoma cells and inhibition of MTDH-triggered autophagy by miRNA-22 plays a key role in the sensitivity to chemotherapy.¹⁹⁶ In addition, miRNA-22 is also found to play a key role in the sensitivity to chemotherapy. In addition, miRNA-410 is also found to enhance chemosensitivity by inhibiting autophagy in osteosarcoma cells. Apoptosis was significantly enhanced in cells treated with miRNA-410 combined with chemotherapeutic agents such as rapamycin, adriamycin, and cisplatin, compared to cells treated with chemotherapeutic agents alone.¹⁹⁷ Based on the above findings, miRNAs have an important role in enhancing the chemosensitivity of cells. However, finding effective inhibitors of autophagy remains a challenge. Further studies are needed to identify better and more appropriate inhibitors of autophagy to improve drug resistance in osteosarcoma.

The Application of miRNAs in Novel Nanomaterials

Despite tremendous advances in the treatment of osteosarcoma, significant challenges remain in terms of poor metabolism kinetics, high cytotoxicity, and drug resistance of drugs used in the treatment of osteosarcoma.¹⁹⁸ Developing new delivery mechanisms is another way to overcome resistance to conventional chemotherapeutic agents.¹⁸⁴ The use of drug-loaded nanomaterials with the ability to kill cancer cells and superior bioactivity is promising in the treatment of osteosarcoma. Nanotechnology may offer opportunities to overcome these difficulties by providing carriers for transporting molecules or other factors that affect gene expression in tumor cells.¹⁹⁸ For example, in one experiment, ZIF was used as a carrier to deliver miRNAs to achieve efficient cellular uptake and payload release at specific sites within the target cells. The results showed that the ZIF-8 vector exhibited high loading efficiency, promoted cellular uptake, and enhanced the endosomal escape of miRNAs. Both *in vitro* and *in vivo* evaluations also demonstrated that the miRNAs@ZIF-8 nanocomposite is a potential carrier for the effective delivery of therapeutic nucleic acid drugs.¹⁹⁹ In addition, miRNA-214 has been shown to playing an oncogenic role in osteosarcoma, promoting osteosarcoma invasion and migration, and mediating osteosarcoma drug resistance. miRNA-214 knockdown and

miRNA-214 inhibitors increase the radiosensitivity or chemosensitivity of osteosarcoma in osteosarcoma cells and xenograft mouse models.^{200,201} Available studies have found that a highly positively charged nonviral vector (GO-PEI) complex can bind to negatively charged miRNA-214 and encapsulate miRNA-214 to synthesize miRNA-214 inhibitors, which can then be introduced into cells from *in vitro* or *in vivo* to exert tumor-suppressive effects. This inhibitor effectively inhibited the migration and invasion of MG63, and U2OS cells showed superior antitumor activity in an MG63 xenograft mouse model and enhanced cisplatin's killing effect on osteosarcoma cells.²⁰² Novel nanomaterials for drug delivery have the advantages of targeting specific cells for drug delivery, overcoming barriers to drug penetration, improving the bioavailability and therapeutic performance of antitumor drugs, and exploiting these advantages will offer hope for improving treatment strategies for osteosarcoma.

Summary and Outlook

In summary, this study reviewed the mechanism of miRNAs in the development of osteosarcoma and the application value of miRNAs in the diagnosis and drug resistance of osteosarcoma in recent years. The relationship between different miRNAs and patient prognosis, cancer diagnosis, cancer progression, chemotherapy resistance, and their application as possible drug targets has been demonstrated, and research on miRNAs in osteosarcoma development has achieved outstanding success. However, most miRNAs are still being studied *in vitro* studies of osteosarcoma cells. The specific mechanisms of many miRNAs in osteosarcoma, and genes and enzymes involved in the process of miRNAs acting on target genes, are still unknown. The accuracy of diagnostic and therapeutic target prediction needs to be improved. In addition, the literature on ferroptosis, pyroptosis, and other regulatory death pathways in osteosarcoma research is very limited and still needs much exploration by researchers. Although the differential expression of extracellular miRNAs has been repeatedly reported in osteosarcoma, due to the limitations of current detection methods and costs, no reliable miRNAs have been identified in the sera of osteosarcoma patients that can be used as diagnostic criteria for osteosarcoma, and the diagnosis and poor prognosis of osteosarcoma remains a serious problem for osteosarcoma patients. In addition, although miRNAs are becoming increasingly mature as early diagnostic indicators and combination chemotherapeutic agents in osteosarcoma, and relevant antagonistic drugs or gene mimetic drugs have been designed for cancer treatment in the course of therapy, the drug resistance mechanism of most osteosarcomas is still not clearly studied, and the research progress of miRNAs in drug resistance of osteosarcoma is still at a preliminary stage. In addition, the continuous development of nanomaterials will bring broader therapeutic prospects for osteosarcoma and may bring new therapeutic strategies to improve the problems of poor drug metabolism kinetics, high cytotoxicity, and drug

resistance in osteosarcoma treatment. However, this aspect is limited by many factors, and the development is very slow and still needs much exploration by researchers.

In the research process, we found that miRNAs do not regulate the development of osteosarcoma through a single signaling pathway but through a complex network of signaling pathways, such as lncRNAs and miRNAs, circRNAs, and miRNAs, which together regulate the development of osteosarcoma. Therefore, future research on the role of miRNAs in the development of osteosarcoma should not be limited to the study of single miRNAs but should comprehensively evaluate and deeply explore these regulatory networks of miRNAs and identify common therapeutic targets from them. In addition, the combined diagnosis and treatment of osteosarcoma using one or more drugs that simultaneously target signaling pathways associated with different miRNAs may also amplify the effect of drugs in the treatment of osteosarcoma. This approach to identifying new targets for developing new therapeutic strategies and drugs may provide new ideas to improve these problems. Besides, the development and delivery of targeted therapeutic strategies using novel nanomaterials could be an effective treatment for diseases such as osteosarcoma, taking advantage of their therapeutic benefits. Osteosarcoma could be better treated by taking advantage of current developments and advances in technology.

In conclusion, there are still many challenges in the study of miRNAs in osteosarcoma. We believe that miRNAs will continue to be studied in depth in osteosarcoma in the future and will play an important role in the diagnosis, treatment, and prognosis of osteosarcoma. miRNAs are expected to be applied in the clinical treatment and diagnosis of osteosarcoma.

Acknowledgments

This work was generously supported by grants from the Gansu Provincial Party Committee Organization Department 2021 Longyuan Youth Innovation and Entrepreneurship Talent Project (21LQTD26); the Youth Project of Natural Science Foundation of Gansu Province (21JR1RA098); the Health Industry Scientific Research And Management Project of Health Commission of Gansu Province (GSWSKY2019-73); and the Hospital Fund project of Lanzhou University First Hospital (ldyyyn2018-01).

Authors' Note

This article is a review article, which does not include human or animal experiments and does not involve ethical issues.

Declaration of Conflicting Interests


The author(s) declared no potential conflicts of interest with respect to the research, authorship, and/or publication of this article.

Funding

The author(s) disclosed receipt of the following financial support for the research, authorship, and/or publication of this article: This work was supported by the Hospital Fund project of Lanzhou University First Hospital, the Health Industry Scientific Research And Management Project of Health Commission of Gansu Province,

Gansu Provincial Party Committee Organization Department, 2021 Longyuan Youth Innovation and Entrepreneurship Talent Project, Natural Science Foundation of Gansu Province (grant number ldyyyyn2018-01, GSWSKY2019-73, 21LQTD26, 21JR1RA098).

ORCID iD

Zihe Dong  <https://orcid.org/0000-0003-2898-1325>

References

- Ottaviani G, Jaffe N. The epidemiology of osteosarcoma. *Cancer Treat Res*. 2009;152:3-13.
- Clark JC, Dass CR, Choong PF. A review of clinical and molecular prognostic factors in osteosarcoma. *J Cancer Res Clin Oncol*. 2008;134(3):281-297.
- Arndt CAS, Crist WM. Medical progress—common musculoskeletal tumors of childhood and adolescence. *N Engl J Med*. 1999;341(5):342-352.
- Marina N, Gebhardt M, Teot L, Gorlick R. Biology and therapeutic advances for pediatric osteosarcoma. *Oncologist*. 2004;9(4):422-441.
- Zhi CS, Wu B. Serum miRNA-34a serves as a diagnostic and prognostic bio-marker in osteosarcoma. *Int J Clin Exp Pathol*. 2016;9(3):3459-3464.
- Wang Y, Wang Y, Jia LS, et al. Low miR-34a and miR-192 are associated with unfavorable prognosis in patients suffering from osteosarcoma. *Am J Transl Res*. 2015;7(1):111-119.
- Wu PK, Chen WM, Chen CF, Lee OK, Huang CK, Chen TH. Primary osteogenic sarcoma with pulmonary metastasis: clinical results and prognostic factors in 91 patients. *Jpn J Clin Oncol*. 2009;39(8):514-522.
- Bielack SS, Kempf-Bielack B, Delling G, et al. Prognostic factors in high-grade osteosarcoma of the extremities or trunk: an analysis of 1,702 patients treated on neoadjuvant cooperative osteosarcoma study group protocols. *J Clin Oncol*. 2002;20(3):776-790.
- Liu Q, Xu B, Zhou W. Correlation between chemotherapy resistance in osteosarcoma patients and PAK5 and ezrin gene expression. *Oncol Lett*. 2018;15(1):879-884.
- Gorelik L, Flavell RA. Transforming growth factor-beta in T-cell biology. *Nat Rev Immunol*. 2002;2(1):46-53.
- Bi G, Liang JQ, Zheng YS, et al. Multi-omics characterization and validation of invasiveness-related molecular features across multiple cancer types. *J Transl Med*. 2021;19(1):124.
- Costelloe CM, Chuang HH, Daw NC. PET/CT of osteosarcoma and Ewing sarcoma. *Semin Roentgenol*. 2017;52(4):255-268.
- Zhang X, Guan Z. PET/CT in the diagnosis and prognosis of osteosarcoma. *Front Biosci (Landmark Ed)*. 2018;23:2157-2165. doi: 10.2741/4696
- Wang JC, Liu SZ, Shi JY, et al. The role of miRNA in the diagnosis, prognosis, and treatment of osteosarcoma. *Cancer Biother Radiopharm*. 2019;34(10):605-613.
- Macfarlane LA, Murphy PR. MicroRNA: biogenesis, function and role in cancer. *Curr Genomics*. 2010;11(7):537-561.
- Vanacore D, Boccellino M, Rossetti S, et al. MicroRNAs in prostate cancer: an overview. *Oncotarget*. 2017;8(30):50240-50251.
- Iqbal MA, Arora S, Prakasam G, Calin GA, Syed MA. MicroRNA in lung cancer: role, mechanisms, pathways and therapeutic relevance. *Mol Aspects Med*. 2019;70:3-20.
- Shin VY, Chu K-M. MiRNA as potential biomarkers and therapeutic targets for gastric cancer. *World J Gastroenterol*. 2014;20(30):10432-10439.
- Zahedipour F, Bolourinezhad M, Teng Y, Sahebkar A. The multifaceted therapeutic mechanisms of curcumin in osteosarcoma: state-of-the-art. *J Oncol*. 2021;2021:3006853.
- Wang T, Zhang C, Wang S. Ginsenoside Rg3 inhibits osteosarcoma progression by reducing circ_0003074 expression in a miR-516b-5p/KPNA4-dependent manner. *J Orthop Surg Res*. 2021;16(1).
- Otoukesh B, Abbasi M, Gorgani HOL, et al. MicroRNAs signatures, bioinformatics analysis of miRNAs, miRNA mimics and antagonists, and miRNA therapeutics in osteosarcoma. *Cancer Cell Int*. 2020;20(1).
- Yang JS, Lai EC. Alternative miRNA biogenesis pathways and the interpretation of core miRNA pathway mutants. *Mol Cell*. 2011;43(6):892-903.
- Lee Y, Jeon K, Lee JT, Kim S, Kim VN. MicroRNA maturation: stepwise processing and subcellular localization. *EMBO J*. 2002;21(17):4663-4670.
- Davis BN, Hata A. Regulation of MicroRNA biogenesis: a miRiad of mechanisms. *Cell Commun Signal*. 2009;7:18.
- Zeng Y, Cullen BR. Sequence requirements for micro RNA processing and function in human cells. *RNA*. 2003;9(1):112-123.
- Han J, Lee Y, Yeom KH, Kim YK, Jin H, Kim VN. The drosha-DGCR8 complex in primary microRNA processing. *Genes Dev*. 2004;18(24):3016-3027.
- Bernstein E, Caudy AA, Hammond SM, Hannon GJ. Role for a bidentate ribonuclease in the initiation step of RNA interference. *Nature*. 2001;409(6818):363-366.
- Du T, Zamore PD. Microprimer: the biogenesis and function of microRNA. *Development*. 2005;132(21):4645-4652.
- Bravo-Vazquez LA, Medina-Rios I, Marquez-Gallardo LD, et al. Functional implications and clinical potential of MicroRNAs in irritable bowel syndrome: a concise review. *Dig Dis Sci*. 2022.
- Wilkie GS, Dickson KS, Gray NK. Regulation of mRNA translation by 5'- and 3'-UTR-binding factors. *Trends Biochem Sci*. 2003;28(4):182-188.
- Mayr C, Bartel DP. Widespread shortening of 3' UTRs by alternative cleavage and polyadenylation activates oncogenes in cancer cells. *Cell*. 2009;138(4):673-684.
- Zhao ZH, Li G, Han YG, et al. Circular RNA ZNF609 enhances proliferation and glycolysis during glioma progression by miR-378b/SLC2A1 axis. *Aging (Albany NY)*. 2021;13(17):21122-21133.
- Vasudevan S, Tong Y, Steitz JA. Switching from repression to activation: microRNAs can up-regulate translation. *Science*. 2007;318(5858):1931-1934.
- Bagheri M, Sarabi PZ, Mondanizadeh M. The role of miRNAs as a big master regulator of signaling pathways involved in lymphoblastic leukemia. *J Cell Physiol*. 2022;237(4):2128-2139.
- Ha TY. The role of MicroRNAs in regulatory T cells and in the immune response. *Immune Netw*. 2011;11(1):11-41.

36. Cortez MA, Bueso-Ramos C, Ferdin J, Lopez-Berestein G, Sood AK, Calin GA. MicroRNAs in body fluids-the mix of hormones and biomarkers. *Nat Rev Clin Oncol*. 2011;8(8):467-477.
37. Chen CZ. MicroRNAs as oncogenes and tumor suppressors. *N Engl J Med*. 2005;353(17):1768-1771.
38. Zhao Y, Srivastava D. A developmental view of microRNA function. *Trends Biochem Sci*. 2007;32(4):189-197.
39. Bartel DP. MicroRNAs: genomics, biogenesis, mechanism, and function. *Cell*. 2004;116(2):281-297.
40. Friedman RC, Farh KKH, Burge CB, Bartel DP. Most mammalian mRNAs are conserved targets of microRNAs. *Genome Res*. 2009;19(1):92-105.
41. O'Brien J, Hayder H, Zayed Y, Peng C. Overview of MicroRNA biogenesis, mechanisms of actions, and circulation. *Front Endocrinol (Lausanne)*. 2018;9:402.
42. Van den Brande S, Gijbels M, Wynant N, et al. The presence of extracellular microRNAs in the media of cultured drosophila cells. *Sci Rep*. 2018;8(1):17312.
43. Max KEA, Bertram K, Akat KM, et al. Human plasma and serum extracellular small RNA reference profiles and their clinical utility. *Proc Natl Acad Sci U S A*. 2018;115(23):E5334-E5343.
44. Krol J, Loedige I, Filipowicz W. The widespread regulation of microRNA biogenesis, function and decay. *Nat Rev Genet*. 2010;11(9):597-610.
45. Ebert MS, Sharp PA. Roles for microRNAs in conferring robustness to biological processes. *Cell*. 2012;149(3):515-524.
46. Kosik KS. The neuronal microRNA system. *Nat Rev Neurosci*. 2006;7(12):911-920.
47. Xu Z, Jiang JF, Xu C, et al. MicroRNA-181 regulates CARM1 and histone arginine methylation to promote differentiation of human embryonic stem cells. *PLoS One*. 2013;8(1).
48. Wang B, Ricardo S. Role of microRNA machinery in kidney fibrosis. *Clin Exp Pharmacol Physiol*. 2014;41(8):543-550.
49. Jiang Y, Hou JY, Zhang XD, et al. Circ-XPO1 upregulates XPO1 expression by sponging multiple miRNAs to facilitate osteosarcoma cell progression. *Exp Mol Pathol*. 2020;117.
50. Chen C, Liu YM, Fu BL, Xu LL, Wang B. MicroRNA-21: an emerging player in bone diseases. *Front Pharmacol*. 2021;12:722804.
51. Calin GA, Dumitru CD, Shimizu M, et al. Frequent deletions and down-regulation of micro-RNA genes miR15 and miR16 at 13q14 in chronic lymphocytic leukemia. *Proc Natl Acad Sci U S A*. 2002;99(24):15524-15529.
52. Voorhoeve PM, le Sage C, Schrier M, et al. A genetic screen implicates miRNA-372 and miRNA-373 as oncogenes in testicular germ cell tumors. *Cell*. 2006;124(6):1169-1181.
53. Hu HL, Du LT, Nagabayashi G, Seeger RC, Gatti RA. ATM is down-regulated by N-myc-regulated microRNA-421. *Proc Natl Acad Sci U S A*. 2010;107(4):1506-1511.
54. Hao J, Zhang SY, Zhou YQ, Liu C, Hu XG, Shao CH. MicroRNA 421 suppresses DPC4/Smad4 in pancreatic cancer. *Biochem Biophys Res Commun*. 2011;406(4):552-557.
55. Ostling P, Leivonen SK, Aakula A, et al. Systematic analysis of microRNAs targeting the androgen receptor in prostate cancer cells. *Cancer Res*. 2011;71.
56. Jiang SA, Zhang HW, Lu MH, et al. MicroRNA-155 functions as an OncomiR in breast cancer by targeting the suppressor of cytokine signaling 1 gene. *Cancer Res*. 2010;70(8):3119-3127.
57. Vo DT, Karanam NK, Ding LH, et al. miR-125a-5p functions as tumor suppressor microRNA and is a marker of locoregional recurrence and poor prognosis in head and neck cancer. *Neoplasia*. 2019;21(9):849-862.
58. Zhang M, Li M, Li N, et al. miR-217 suppresses proliferation, migration, and invasion promoting apoptosis via targeting MTDH in hepatocellular carcinoma. *Oncol Rep*. 2017;37(3):1772-1778.
59. Reddy KB. MicroRNA (miRNA) in cancer. *Cancer Cell Int*. 2015;15:38.
60. Kushlinskii NE, Fridman MV, Braga EA. Molecular mechanisms and microRNAs in osteosarcoma pathogenesis. *Biochemistry (Mosc)*. 2016;81(4):315-328.
61. Chen RL, Wang GY, Zheng Y, Hua YQ, Cai ZD. Long non-coding RNAs in osteosarcoma. *Oncotarget*. 2017;8(12):20462-20475.
62. Liang G, Duan CP, He J, Ma W, Dai X. PTPN14, A target gene of miR-4295, restricts the growth and invasion of osteosarcoma cells through inactivation of YAP1 signalling. *Clin Exp Pharmacol Physiol*. 2020;47(7):1301-1310.
63. Yang X, Yang J, Tang YL, et al. MiR-4295 promotes the malignant progression of gastric cancer via targeting PTEN. *Comb Chem High Throughput Screen*. 2021.
64. Lu S, Zhou C, Zou BW, Zhang HY, Feng M. MiR-4295 facilitates cell proliferation and metastasis in head and neck squamous cell carcinoma by targeting NPTX1. *Genes Immun*. 2020;21(1):4-12.
65. Guo Q, Ma J, Wu J. MiRNA-218 inhibits cell proliferation, migration and invasion by targeting runt-related transcription factor 2 (Runx2) in human osteosarcoma cells. *Regen Ther*. 2021;18:508-515.
66. Lin E, Liu S, Xiang W, Zhang HB, Xie CF. CircEIF4G2 promotes tumorigenesis and progression of osteosarcoma by sponging miR-218. *Biomed Res Int*. 2020;2020:8386936.
67. He P, Xu Y, Wang Z. LncRNA SNHG10 increases the methylation of miR-218 gene to promote glucose uptake and cell proliferation in osteosarcoma. *J Orthop Surg Res*. 2020;15(1).
68. Dang SW, Zhang R, Tian SJ, Hou P, Li G, Ji MJ. MicroRNA-218 inhibits the malignant phenotypes of glioma by modulating the TNC/AKT/AP-1/TGF beta 1 feedback signaling loop. *Int J Mol Med*. 2021;48(5).
69. Tian WX, Yuan XF, Song YN, et al. miR-218 inhibits glucose metabolism in non-small cell lung cancer via the NF-kappa B signaling pathway. *Exp Ther Med*. 2021;21(2).
70. Chen HF, Cao WL, Chen J, et al. miR-218 contributes to drug resistance in multiple myeloma via targeting LRRC28. *J Cell Biochem*. 2021;122(3-4):305-314.
71. Liang H, Li L, Zhu S, et al. MicroRNA-744-5p suppresses tumorigenesis and metastasis of osteosarcoma through the p38 mitogen-activated protein kinases pathway by targeting transforming growth factor-beta 1. *Bioengineered*. 2022;13(5):12309-12325.
72. Huang W, Chen QS, Dai JW, et al. miR-744-5p suppresses tumor proliferation and metastasis by targeting transforming growth

- factor-beta 1 (TGF-beta 1) in hepatocellular carcinoma (HCC). *J Gastrointest Oncol*. 2021;12(4):1811-1822.
73. Fan F, Yao DX, Yan PF, Jiang XB, Hu J. MicroRNA-744-5p inhibits glioblastoma malignancy by suppressing replication factor C subunit 2. *Oncol Lett*. 2021;22(2).
74. Guo B, Xiao CY, Liu YM, et al. miR-744-5p inhibits multiple myeloma proliferation, epithelial mesenchymal transformation and glycolysis by targeting SOX12/Wnt/beta-catenin signaling. *Onco Targets Ther*. 2021;14:1161-1172.
75. Zhang W, Liao K, Liu D. MicroRNA-744-5p is downregulated in colorectal cancer and targets SEPT2 to suppress the malignant phenotype. *Mol Med Rep*. 2021;23(1).
76. Li Z, Deng LR, Li YG, Wang YJ. MiR-139 inhibits proliferation, migration and invasion of osteosarcoma cell line MG63 via down-regulating integrin subunit alpha V(ITGAV). *Tissue Cell*. 2022;75:101720.
77. Wang W, Luo P, Guo WJ, et al. LncRNA SNHG20 knockdown suppresses the osteosarcoma tumorigenesis through the mitochondrial apoptosis pathway by miR-139/RUNX2 axis. *Biochem Biophys Res Commun*. 2018;503(3):1927-1933.
78. Gu Z, H, Hou ZH, Zheng LB, Wang XQ, Wu LB, Zhang C. Long noncoding RNA LINC00858 promotes osteosarcoma through regulating miR-139-CDK14 axis. *Biochem Biophys Res Commun*. 2018;503(2):1134-1140.
79. Wang Y, Zheng Y, Chen Q, Dai YM, Li T. MicroRNA-139 inhibits pancreatic-cancer carcinogenesis by suppressing RalB via the Ral/RAC/PI3K pathway. *Arch Biochem Biophys*. 2021;704.
80. Nam RK, Benatar T, Amemiya Y, Sherman C, Seth A. Mir-139 regulates autophagy in prostate cancer cells through beclin-1 and mTOR signaling proteins. *Anticancer Res*. 2020;40(12):6649-6663.
81. Dutto I, Tillhon M, Cazzalini O, Stivala LA, Prosperi E. Biology of the cell cycle inhibitor p21(CDKN1A): molecular mechanisms and relevance in chemical toxicology. *Arch Toxicol*. 2015;89(2):155-178.
82. Zhao X, Yang YK, Xu JT, Luo Y, Xin Y, Wan YB. Downregulation of microRNA-95-3p suppresses cell growth of osteosarcoma via CDKN1A/p21 expression. *Oncol Rep*. 2018;39(1):289-297.
83. Anttila MA, Kosma VM, Hongxiu J, et al. P21/WAF1 expression as related to p53, cell proliferation and prognosis in epithelial ovarian cancer. *Br J Cancer*. 1999;79(11-12):1870-1878.
84. Benassi MS, Molendini L, Gamberi G, et al. Alteration of pRb/p16/cdk4 regulation in human osteosarcoma. *Int J Cancer*. 1999;84(5):489-493.
85. Ye J, Yao YF, Song QX, et al. Up-regulation of miR-95-3p in hepatocellular carcinoma promotes tumorigenesis by targeting p21 expression. *Sci Rep*. 2016;6.
86. Yang W, Dou CW, Wang YF, et al. MicroRNA-92a contributes to tumor growth of human hepatocellular carcinoma by targeting FBXW7. *Oncol Rep*. 2015;34(5):2576-2584.
87. Zhou C, Shen F, Mao L, Wang B, Li Y, Yu HZ. miR-92a is up-regulated in cervical cancer and promotes cell proliferation and invasion by targeting FBXW7. *Biochem Biophys Res Commun*. 2015;458(1):63-69.
88. Jiang XS, Li XF, Wu FF, et al. Overexpression of miR-92a promotes the tumor growth of osteosarcoma by suppressing F-box and WD repeat-containing protein 7. *Gene*. 2017;606:10-16.
89. Yu HY, Song H, Liu L, et al. MiR-92a modulates proliferation, apoptosis, migration, and invasion of osteosarcoma cell lines by targeting Dickkopf-related protein 3. *Biosci Rep*. 2019;39.
90. Xu H, Ding Y, Yang XY. Overexpression of long noncoding RNA H19 downregulates miR-140-5p and activates PI3K/AKT signaling pathway to promote invasion, migration and epithelial-mesenchymal transition of ovarian cancer cells. *BioMed Res Int*. 2021;2021.
91. Li D, Lin FF, Li GP, Zeng FC. Exosomal microRNA-15a from ACHN cells aggravates clear cell renal cell carcinoma via the BTG2/PI3K/AKT axis. *Kaohsiung Journal of Medical Sciences*. 2021;37(11):p. 973-982.
92. Lv ZL, Li YJ, Dan ZD, Shen K. MIR-451 INHIBITS PROLIFERATION AND PROMOTES APOPTOSIS OF LUNG CANCER CELLS BY REGULATING TARGET GENE PSMB8-NOS2 AND ACTIVATING PI3K/AKT/MTOR SIGNALING PATHWAY. *Acta Medica Mediterranea*. 2021;37(5):2791-2796.
93. Zhang J, Yu XH, Yan YG, Wang C, Wang WJ. PI3K/Akt signaling in osteosarcoma. *Clin Chim Acta*. 2015;444:182-192.
94. Huang G, Yang YM, Lv MX, et al. miR-23b-3p inhibits the oncogenicity of colon adenocarcinoma by directly targeting NFE2L3. *J Oncol*. 2021;2021.
95. Hayashi M, Yamada S, Kurimoto K, et al. miR-23b-3p plays an oncogenic role in hepatocellular carcinoma. *Ann Surg Oncol*. 2021;28(6):3416-3426.
96. Zhang J, Zhang Y, Tan XP, Zhang Q, Liu CY, Zhang YL. MiR-23b-3p induces the proliferation and metastasis of esophageal squamous cell carcinomas cells through the inhibition of EBF3. *Acta Biochim Biophys Sin*. 2018;50(6):605-614.
97. Fan L, Cao X, Lei Y. MicroRNA miR-23b-3p promotes osteosarcoma by targeting ventricular zone expressed PH domain-containing 1 (VEPH1)/phosphatidylinositol 3-kinase/protein kinase B (PI3K/AKT) pathway. *Bioengineered*. 2021;12(2):12568-12582.
98. Liu H, Wei W, Wang XJ, et al. miR-23b-3p promotes the apoptosis and inhibits the proliferation and invasion of osteosarcoma cells by targeting SIX1. *Mol Med Rep*. 2018;18(6):5683-5692.
99. Lu C, Peng K, Guo H, et al. miR-18a-5p promotes cell invasion and migration of osteosarcoma by directly targeting IRF2. *Oncol Lett*. 2018;16(3):3150-3156.
100. Fei D, Zhang XN, Liu JX, et al. Long noncoding RNA FER1L4 suppresses tumorigenesis by regulating the expression of PTEN targeting miR-18a-5p in osteosarcoma. *Cell Physiol Biochem*. 2018;51(3):1364-1375.
101. Carracedo A, Alimonti A, Pandolfi PP. PTEN level in tumor suppression: how much is too little? *Cancer Res*. 2011;71(3):629-633.
102. Yang J, Zou Y, Jiang D. Honokiol suppresses proliferation and induces apoptosis via regulation of the miR21/PTEN/PI3K/AKT signaling pathway in human osteosarcoma cells. *Int J Mol Med*. 2018;41(4):1845-1854.

103. Zhuang M, Qiu XB, Cheng D, Zhu CL, Chen L. MicroRNA-524 promotes cell proliferation by down-regulating PTEN expression in osteosarcoma. *Cancer Cell Int.* 2018;18.
104. Jiang P, Yang X, Li YL, Chen J. miRNA-216 knockdown has effects to suppress osteosarcoma via stimulating PTEN. *Food Sci Nutr.* 2020;8(9):4708-4716.
105. Wang L, Tang B, Han H, et al. miR-155 affects osteosarcoma MG-63 cell autophagy induced by Adriamycin through regulating PTEN-PI3K/AKT/mTOR signaling pathway. *Cancer Biother Radiopharm.* 2018;33(1):32-38.
106. Zhou WY, Hao MZ, Du XL, Chen KX, Wang GW, Yang JL. Advances in targeted therapy for osteosarcoma. *Discov Med.* 2014;17(96):301-307.
107. Liu D, Chen L, Zhao H, Vaziri ND, Ma SC, Zhao YY. Small molecules from natural products targeting the Wnt/beta-catenin pathway as a therapeutic strategy. *Biomed Pharmacother.* 2019;117:108990.
108. Hsu W, Liu LF, Chen X, Zhang Y, Zhu WP, LncRNA CASC11 promotes the cervical cancer progression by activating Wnt/beta-catenin signaling pathway. *Biol Res.* 2019;52.
109. Ji YQ, Wang M, Li XS, Cui Fs. The long noncoding RNA NEAT1 targets miR-34a-5p and drives nasopharyngeal carcinoma progression via Wnt/beta-catenin signaling. *Yonsei Med J.* 2019;60(4):336-345.
110. Wang H, Duan XL, Qi XL, et al. Concurrent hypermethylation of SFRP2 and DKK2 activates the Wnt/beta-catenin pathway and is associated with poor prognosis in patients with gastric cancer. *Mol Cells.* 2017;40(1):45-53.
111. Jung YS, Park JI. Wnt signaling in cancer: therapeutic targeting of Wnt signaling beyond beta-catenin and the destruction complex. *Exp Mol Med.* 2020;52(2):183-191.
112. Zhang F-W, Peng LY, Shi CJ, et al. Baicalein mediates the anti-tumor activity in osteosarcoma through lncRNA-NEF driven Wnt/beta-catenin signaling regulatory axis. *J Orthop Translat.* 2022;33:132-141.
113. Yang C, Zhang LL, Huang HK, et al. Alantolactone inhibits proliferation, metastasis and promotes apoptosis of human osteosarcoma cells by suppressing Wnt/ β -catenin and MAPKs signaling pathways. *Genes Dis.* 2022;9(2):466-478.
114. Wang Y, Chen J, Haung YR, et al. Schisandrin B suppresses osteosarcoma lung metastasis in vivo by inhibiting the activation of the Wnt/beta-catenin and PI3K/Akt signaling pathways. *Oncol Rep.* 2022;47(3).
115. Liang K, Liao L, Liu Q, Ouyang Q, Jia L, Wu G. microRNA-377-3p inhibits osteosarcoma progression by targeting CUL1 and regulating Wnt/beta-catenin signaling pathway. *Clin Transl Oncol.* 2021;23(11):2350-2357.
116. Xue Y, Guo Y, Liu N, Meng XQ. MicroRNA-22-3p targeted regulating transcription factor 7-like 2 (TCF7L2) constrains the Wnt/ β -catenin pathway and malignant behavior in osteosarcoma. *Bioengineered.* 2022;13(4):9135-9147.
117. Hu SP, Chang JL, Ruan HF, et al. Cantharidin inhibits osteosarcoma proliferation and metastasis by directly targeting miR-214-3p/DKK3 axis to inactivate beta-catenin nuclear translocation and LEF1 translation. *Int J Biol Sci.* 2021;17(10):2504-2522.
118. Yang S, Chen KX, Cao K, et al. miR-182-5p inhibits NKAPL expression and promotes the proliferation of osteosarcoma. *Biotechnol Bioprocess Eng.* 2021;26(5):758-766.
119. Zhu S, Liu Y, Wang X, Wang JY, Xi GH. lncRNA SNHG10 promotes the proliferation and invasion of osteosarcoma via Wnt/beta-catenin signaling. *Mol Ther Nucleic Acids.* 2020;22:957-970.
120. Zhang Z, Wang C, Liu TT, et al. miRNA-182-5p promotes human bladder cancer proliferation and migration through the FOXF2/SHH axis. *Neoplasma.* 2022;69(2):321-330.
121. Kong DM, Wang X, Wang XD, Wang ZX, Wang F. Downregulated miRNA-22-3p promotes the progression and leads to poor prognosis of hepatocellular carcinoma through targeting CDKN2C. *J BUON.* 2021;26(2):409-417.
122. Wu Y, Zhang CJ, Peng D, et al. MiR-182-5p inhibits the tumorigenesis of clear cell renal cell carcinoma by repressing UBE2T. *Hum Cell.* 2022;35(2):542-556.
123. Wang J-Y, Wang J-Q, Lu S-B. miR-628-5p promotes growth and migration of osteosarcoma by targeting IFI44L. *Biochem Cell Biol.* 2020;98(2):99-105.
124. Wu X, Lei JZ, Zhou B, et al. MiR-628-5p inhibits cervical carcinoma proliferation and promotes apoptosis by targeting VEGF. *Am J Med Sci.* 2021;361(4):499-508.
125. Guo F, Xue J. MicroRNA-628-5p inhibits cell proliferation and induces apoptosis in colorectal cancer through downregulating CCND1 expression levels. *Mol Med Rep.* 2020;21(3):1481-1490.
126. Zhou L, Jiao XX, Peng XQ, Yao XM, Liu L, Zhang LF. MicroRNA-628-5p inhibits invasion and migration of human pancreatic ductal adenocarcinoma via suppression of the AKT/NF-kappa B pathway. *J Cell Physiol.* 2020;235(11):8141-8154.
127. Cheng JP, Huang B, Duan JH, Yi KJ, Zhuang ZL. miR-4295 promotes cell proliferation, migration and invasion of osteosarcoma through targeting interferon regulatory factor 1. *Oncol Lett.* 2020;20(5).
128. Zhang Q, Yin X, Zhang Y. MicroRNA-221 promotes cell proliferation and inhibits apoptosis in osteosarcoma cells by directly targeting FBXW11 and regulating Wnt signaling. *Arch Med Res.* 2021;52(2):191-199.
129. Ye KS, Wang SK, Zhang HH, Han H, Ma B, Nan W. Long noncoding RNA GAS5 suppresses cell growth and epithelial-mesenchymal transition in osteosarcoma by regulating the miR-221/ARHI pathway. *J Cell Biochem.* 2017;118(12):4772-4781.
130. Huynh TK, Huang CH, Chen JY, et al. miR-221 confers lapatinib resistance by negatively regulating p27(kip1) in HER2-positive breast cancer. *Cancer Sci.* 2021;112(10):4234-4245.
131. Jiang N, Zhu S, Zhu J. MiR-221 regulates suppressors of cytokine signaling 3-Janus kinase 2/signal transducer and activator of transcription 3 (SOCS3-JAK2/STAT3) pathway and affects thyroid cancer cell proliferation and apoptosis. *J Biomater Tissue Eng.* 2022;12(5):996-1001.
132. Hu X-H, Zhao ZX, Dai J, Geng DC, Xu YZ. MicroRNA-221 regulates osteosarcoma cell proliferation, apoptosis, migration, and invasion by targeting CDKN1B/p27. *J Cell Biochem.* 2019;120(3):4665-4674.

133. Huang Y, Li LP. Progress of cancer research on astrocyte elevated gene-1/Metadherin (review). *Oncol Lett.* 2014;8(2):493-501.
134. Zhang SK, Liu LD, Lv ZS, Li Q, Gong WQ, Wu H. MicroRNA-342-3p inhibits the proliferation, migration, and invasion of osteosarcoma cells by targeting astrocyte-elevated gene-1 (AEG-1). *Oncol Res.* 2017;25(9):1505-1515.
135. Jiang WB, Wang S, Sun Y, Jiang Y, Yu T, Wang JC. Overexpression of microRNA-448 inhibits osteosarcoma cell proliferation and invasion through targeting of astrocyte elevated gene-1. *Mol Med Rep.* 2017;16(4):5713-5721.
136. Mongre RK, Sodhi SS, Ghosh M, et al. A new paradigm to mitigate osteosarcoma by regulation of MicroRNAs and suppression of the NF- κ B signaling cascade. *Dev Reprod.* 2014;18(4):197-212.
137. Gong HL, Tao Y, Mao XZ, Song DY, You D, Ni JD. MicroRNA-29a suppresses the invasion and migration of osteosarcoma cells by regulating the SOCS1/NF- κ B signalling pathway through negatively targeting DNMT3B. *Int J Mol Med.* 2019;44(4):1219-1232.
138. Lu S, Liao QS, Tang L. MiR-155 affects osteosarcoma cell proliferation and invasion through regulating NF- κ B signaling pathway. *Eur Rev Med Pharmacol Sci.* 2018;22(22):7633-7639.
139. Ru N, Zhang F, Liang J, et al. MiR-564 is down-regulated in osteosarcoma and inhibits the proliferation of osteosarcoma cells via targeting Akt. *Gene.* 2018;645:163-169.
140. Zhao X, Sun SC, Xu JT, Luo Y, Xin Y, Wang YB. MicroRNA-152 inhibits cell proliferation of osteosarcoma by directly targeting Wnt/beta-catenin signaling pathway in a DKK1-dependent manner. *Oncol Rep.* 2018;40(2):767-774.
141. Sun ZY, Jian YK, Zhu HY, Li B. lncRNAPVT1 targets miR-152 to enhance chemoresistance of osteosarcoma to gemcitabine through activating c-MET/PI3K/AKT pathway. *Pathol Res Pract.* 2019;215(3):555-563.
142. Zhou KF, Xu J, Yin XF, Xia JN. Long noncoding RNA HAGLROS promotes cell invasion and metastasis by sponging miR-152 and upregulating ROCK1 expression in osteosarcoma. *Comput Math Methods Med.* 2020;2020:7236245.
143. Man TK, Chintagumpala M, Visvanathan J, et al. Expression profiles of osteosarcoma that can predict response to chemotherapy. *Cancer Res.* 2005;65(18):8142-8150.
144. Clark AG, Vignjevic DM. Modes of cancer cell invasion and the role of the microenvironment. *Curr Opin Cell Biol.* 2015;36:13-22.
145. Hanahan D, Weinberg RA. Hallmarks of cancer: the next generation. *Cell.* 2011;144(5):646-674.
146. Berlanga P, Munoz L, Piqueras M, et al. miR-200c and phospho-AKT as prognostic factors and mediators of osteosarcoma progression and lung metastasis. *Mol Oncol.* 2016;10(7):1043-1053.
147. Liu Y, Zhang J, Xing CH, Wei SX, Guo N, Wang YL. miR-486 inhibited osteosarcoma cells invasion and epithelial-mesenchymal transition by targeting PIM1. *Cancer Biomark.* 2018;23(2):269-277.
148. Shen SY, Huang KM, Wu YZ, et al. A miR-135b-TAZ positive feedback loop promotes epithelial-mesenchymal transition (EMT) and tumorigenesis in osteosarcoma. *Cancer Lett.* 2017;407:32-44.
149. Tian Q, Gu YF, Wang FF, et al. Upregulation of miRNA-154-5p prevents the tumorigenesis of osteosarcoma. *Biomed Pharmacother.* 2020;124:109884.
150. Wu XL, Zhou HZ, Yue B, et al. Upregulation of microRNA-25-3p inhibits proliferation, migration and invasion of osteosarcoma cells in vitro by directly targeting SOX4. *Mol Med Rep.* 2017;16(4):4293-4300.
151. Shan HJ, Zhu LQ, Yao C, et al. MAFG-driven osteosarcoma cell progression is inhibited by a novel miRNA miR-4660. *Mol Ther Nucleic Acids.* 2021;24:385-402.
152. Chen X, Kang R, Kroemer G, Tang DL. Broadening horizons: the role of ferroptosis in cancer. *Nat Rev Clin Oncol.* 2021;18(5):280-296.
153. Tang D, Kang R, Vanden Berghe T, Vandenabeele P, Kroemer G. The molecular machinery of regulated cell death. *Cell Res.* 2019;29(5):347-364.
154. Wu D, Wang S, Yu GC, Chen XY. Cell death mediated by the pyroptosis pathway with the aid of nanotechnology: prospects for cancer therapy. *Angew Chem Int Ed Engl.* 2021;60(15):8018-8034.
155. Tang DL, Chen X, Kang R, Kroemer G. Ferroptosis: molecular mechanisms and health implications. *Cell Res.* 2021;31(2):107-125.
156. Jin Z, El-Deiry WS. Overview of cell death signaling pathways. *Cancer Biol Ther.* 2005;4(2):139-163.
157. Li M, Shen YJ, Wang Q, Zhou XF. MiR-204-5p promotes apoptosis and inhibits migration of osteosarcoma via targeting EBF2. *Biochimie.* 2019;158:224-232.
158. Gui ZL, Wu TL, Zhao GC, Lin ZX, Xu HG. MicroRNA-497 suppress osteosarcoma by targeting MAPK/Erk pathway. *Bratisl Lek Listy.* 2017;118(8):449-452.
159. Li Y, Song D, An T, Liu J, Yang Q, Nan SK. MicroRNA-1226-3p has a tumor-promoting role in osteosarcoma. *Oncol Lett.* 2021;21(6).
160. Liu Q, Wang K. The induction of ferroptosis by impairing STAT3/Nrf2/GPx4 signaling enhances the sensitivity of osteosarcoma cells to cisplatin. *Cell Biol Int.* 2019;43(11):1245-1256.
161. Conrad M, Lorenz SM, Proneth B. Targeting ferroptosis: new hope for as-yet-incurable diseases. *Trends Mol Med.* 2021;27(2):113-122.
162. Dixon SJ, Lemberg KM, Lamprecht MR, et al. Ferroptosis: an iron-dependent form of nonapoptotic cell death. *Cell.* 2012;149(5):1060-1072.
163. Chen X, Kang R, Kroemer G, Tang DL. Organelle-specific regulation of ferroptosis. *Cell Death Differ.* 2021;28(10):2843-2856.
164. Stockwell BR, Angeli JPF, Bayir H, et al. Ferroptosis: a regulated cell death nexus linking metabolism, redox biology, and disease. *Cell.* 2017;171(2):273-285.
165. Qiu C, Liu TY, Luo D, Luan DY, Cheng L, Wang SG. Novel therapeutic savior for osteosarcoma: the endorsement of ferroptosis. *Front Oncol.* 2022;12:746030.
166. Xu ZQ, Chen LH, Wang CS, Zhang LQ, Xu WH. MicroRNA-1287-5p promotes ferroptosis of osteosarcoma cells through inhibiting GPX4. *Free Radic Res.* 2022.
167. Bu XX, Liu JX, Ding R, Li Z. Prognostic value of a pyroptosis-related long noncoding RNA signature associated with osteosarcoma microenvironment. *J Oncol.* 2021;2021:2182761-2182761.
168. Dhital S, Deo P, Stuart I, Naderer T. Bacterial outer membrane vesicles and host cell death signaling. *Trends Microbiol.* 2021;29(12):1106-1116.

169. Tang R, Xu J, Zhang B, et al. Ferroptosis, necroptosis, and pyroptosis in anticancer immunity. *J Hematol Oncol.* 2020;13(1):110.
170. Tan Y, Chen QZ, Li XL, et al. Pyroptosis: a new paradigm of cell death for fighting against cancer. *J Exp Clin Cancer Res.* 2021;40(1).
171. Tian BG, Hua Z, Wang ZJ, Li J. Knockdown of microRNA-181a inhibits osteosarcoma cells growth and invasion through triggering NLRP3-dependent pyroptosis. *Eur Rev Med Pharmacol Sci.* 2020;24(15):7922.
172. Salmena L, Poliseno L, Tay Y, Kats L, Pandolfi PP. A ceRNA hypothesis: the Rosetta stone of a hidden RNA language? *Cell.* 2011;146(3):353-358.
173. Wang JY, Yang Y, Ma YJ, et al. Potential regulatory role of lncRNA-miRNA-mRNA axis in osteosarcoma. *Biomed Pharmacother.* 2020;121:109627.
174. He Y, Zhou HT, Wang W, Xu HR, Cheng H. Construction of a circRNA-miRNA-mRNA regulatory network reveals potential mechanism and treatment options for osteosarcoma. *Front Genet.* 2021;12:632359.
175. Weber JA, Baxter DH, Zhang SL, et al. The microRNA spectrum in 12 body fluids. *Clin Chem.* 2010;56(11):1733-1741.
176. Bottani M, Banfi G, Lombardi G. Circulating miRNAs as diagnostic and prognostic biomarkers in common solid tumors: focus on lung, breast, prostate cancers, and osteosarcoma. *J Clin Med.* 2019;8(10).
177. Zhou L, Ma X, Yue J, et al. The diagnostic effect of serum miR-139-5p as an indicator in osteosarcoma. *Cancer Biomark.* 2018;23(4):561-567.
178. Li Q, Song SR, Ni GZ, Li Y, Wang XH. Serum miR-542-3p as a prognostic biomarker in osteosarcoma. *Cancer Biomark.* 2018;21(3):521-526.
179. Liu W, Zhao XT, Zhang YJ, Fang GW, Xue Y. MicroRNA-375 as a potential serum biomarker for the diagnosis, prognosis, and chemosensitivity prediction of osteosarcoma. *J Int Med Res.* 2018;46(3):975-983.
180. Shi L, Xie CJ, Zhu JF, Chen XM. Downregulation of serum miR-194 predicts poor prognosis in osteosarcoma patients. *Ann Diagn Pathol.* 2020;46.
181. Xu S, Yang F, Liu RW, et al. Serum microRNA-139-5p is down-regulated in lung cancer patients with lytic bone metastasis. *Oncol Rep.* 2018;39(5):2376-2384.
182. Tonon F, Cemazar M, Kamensek U, et al. 5-Azacytidine down-regulates the proliferation and migration of hepatocellular carcinoma cells in vitro and in vivo by targeting miR-139-5p/ROCK2 pathway. *Cancers (Basel).* 2022;14(7).
183. Zhao Y, Chen F, Li Q, Wang LH, Fan CH. Isothermal amplification of nucleic acids. *Chem Rev.* 2015;115(22):12491-12545.
184. O'Day K, Gorlick R. Novel therapeutic agents for osteosarcoma. *Expert Rev Anticancer Ther.* 2009;9(4):511-523.
185. Chen RL, Wang GY, Zheng Y, Hua YQ, Cai ZD. Drug resistance-related microRNAs in osteosarcoma: translating basic evidence into therapeutic strategies. *J Cell Mol Med.* 2019;23(4):2280-2292.
186. Yu X, Li Z, Yu J, Chan MTV, Wu WKK. MicroRNAs predict and modulate responses to chemotherapy in colorectal cancer. *Cell Prolif.* 2015;48(5):503-510.
187. Li G, Li Y, Wang DY. Overexpression of miR-329-3p sensitizes osteosarcoma cells to cisplatin through suppression of glucose metabolism by targeting LDHA. *Cell Biol Int.* 2021;45(4):766-774.
188. Yang D, Xu TY, Fan L, Liu KY, Li GD. microRNA-216b enhances cisplatin-induced apoptosis in osteosarcoma MG63 and SaOS-2 cells by binding to JMJD2C and regulating the HIF1 alpha/HES1 signaling axis. *J Exp Clin Cancer Res.* 2020;39(1).
189. Xu M, Jin H, Xu CX, et al. miR-382 inhibits tumor growth and enhance chemosensitivity in osteosarcoma. *Oncotarget.* 2014;5(19):9472-9483.
190. Liu Y, Liu X, Yang S. MicroRNA-221 upregulates the expression of P-gp and Bcl-2 by activating the Stat3 pathway to promote doxorubicin resistance in osteosarcoma cells. *Biol Pharm Bull.* 2021;44(6):861-868.
191. Tang Z, Lu YB, Chen YT, Zhang JR, Chen ZJ, Wang QF. Research progress of microRNA in chemotherapy resistance of osteosarcoma. *Technol Cancer Res Treat.* 2021;20:15330338211034262.
192. Zhao F, Pu YG, Cui MD, Wang HY, Cai SB. MiR-20a-5p represses the multi-drug resistance of osteosarcoma by targeting the SDC2 gene. *Cancer Cell Int.* 2017;17.
193. Mizushima N, Levine B, Cuervo AM, Klionsky DJ. Autophagy fights disease through cellular self-digestion. *Nature.* 2008;451(7182):1069-1075.
194. Jamali Z, Taheri-Anganeh M, Shabaninejad Z, et al. Autophagy regulation by microRNAs: novel insights into osteosarcoma therapy. *IUBMB Life.* 2020;72(7):1306-1321.
195. Xu R, D, Liu SZ, Chen HH, Lao LF. MicroRNA-30a downregulation contributes to chemoresistance of osteosarcoma cells through activating beclin-1-mediated autophagy. *Oncol Rep.* 2016;35(3):1757-1763.
196. Wang P, Zhao ZQ, Guo SB, et al. Roles of microRNA-22 in suppressing proliferation and promoting sensitivity of osteosarcoma cells via metadherin-mediated autophagy. *Orthop Surg.* 2019;11(2):285-293.
197. Chen R, Li X, He B, Hu W. MicroRNA-410 regulates autophagy-related gene ATG16L1 expression and enhances chemosensitivity via autophagy inhibition in osteosarcoma. *Mol Med Rep.* 2017;15(3):1326-1334.
198. Savvidou OD, Bolia IK, Chloros GD, et al. Applied nanotechnology and nanoscience in orthopedic oncology. *Orthopedics.* 2016;39(5):280-286.
199. Feng H, Li ZY, Xie WJ, et al. Delivery of therapeutic miRNAs using nanoscale zeolitic imidazolate framework for accelerating vascularized bone regeneration. *Chem Eng J.* 2022;430.
200. Li Y, Song XM, Liu ZG, et al. Upregulation of miR-214 induced radioresistance of osteosarcoma by targeting PHLDA2 via PI3K/Akt signaling. *Front Oncol.* 2019;9:298.
201. Song YD, Li DD, Guan Y, Wang YL, Zheng J. miR-214 modulates cisplatin sensitivity of osteosarcoma cells through regulation of anaerobic glycolysis. *Cell Mol Biol.* 2017;63(9):75-79.
202. Ou L, Lin HYJ, Song YW, et al. Efficient miRNA inhibitor with GO-PEI nanosheets for osteosarcoma suppression by targeting PTEN. *Int J Nanomedicine.* 2020;15:5131-5146.