

Epigenetic regulation of HOTAIR in advanced chronic myeloid leukemia

Ziye Li
Jianmin Luo

Department of Hematology, The Second Hospital of Hebei Medical University, Shijiazhuang, Hebei 050000, People's Republic of China

Purpose: Chronic myeloid leukemia (CML) accounts for ~10% of leukemia cases, and its progression involves epigenetic gene regulation. This study investigated epigenetic regulation of HOTAIR and its target microRNA, miR-143, in advanced CML.

Patients and methods: We first isolated bone marrow mononuclear cells from 70 patients with different phases of CML and from healthy donors as normal control; we also cultured K562 and KCL22 cells, treated with demethylation drug; MTT assay, flow cytometry, quantitative real-time polymerase chain reaction (qPCR), methylation-specific polymerase chain reaction (MSP), Western blot, luciferase assay, RNA pull-down assay and RNA-binding protein immunoprecipitation (RIP) assay were performed.

Result: As measured by qPCR, HOTAIR expression in K562 cells, KCL22 cells, and samples from cases of advanced-stage CML increased with levels of several DNA methyltransferases and histone deacetylases, including DNMT1, DNMT3A, HDAC1, EZH2, and LSD1, and miR-143 levels were decreased and HOTAIR levels were increased. Treatment with 5-azacytidine, a DNA methylation inhibitor, decreased DNMT1, DNMT3A, HDAC1, EZH2, LSD1 mRNA, protein levels, and HOTAIR mRNA levels but increased miR-143 levels. HOTAIR knockdown and miR-143 overexpression both inhibited proliferation and promoted apoptosis in KCL22 and K562 cells through the PI3K/AKT pathway. RNA pull-down, mass spectrometry, and RIP assays showed that HOTAIR interacted with EZH2 and LSD1. A dual-luciferase assay demonstrated that HOTAIR interacted with miR-143.

Conclusion: Our findings demonstrate the key epigenetic interactions of HOTAIR related to CML progression and suggest HOTAIR as a potential therapeutic target for advanced CML. Furthermore, our results support the use of demethylation drugs as a CML treatment strategy.

Keywords: lncRNA HOTAIR, miR-143, epigenetic, PI3K/AKT, EZH2

Introduction

Chronic myeloid leukemia (CML) is a clonal disease characterized by the Philadelphia chromosome, which contains the *BCR-ABL1* fusion gene that encodes the P210 *BCR-ABL1* protein. Typically, patients with CML experience three phases: a chronic phase (CP), an accelerated phase (AP), and a blast phase (BP), where the AP and BP involve progression of the disease. Patients who progress to BP receive standard treatment similar to that used for acute leukemia; but the effect is insufficient and the patients have a poor prognosis.^{1,2} The development of CML is very complex and involves gene mutations, chromosomal variations, and epigenetic regulation of genes.³

In contrast to genetic changes, epigenetic changes refer to hereditary changes in gene expression without changes in DNA sequence, such as DNA methylation, chroma-

Correspondence: Jianmin Luo
Department of Hematology, The Second Hospital of Hebei Medical University, Shijiazhuang, Hebei 050000, People's Republic of China
Tel +86 6 600 3927
Fax +86 6 600 2841
Email luojm315@163.com

tin remodeling, and histone modification.⁴ The DNMT family, which includes DNMT1 and DNMT3A, and the HDAC family, which includes HDAC1 and HDAC2, play important roles in DNA methylation and histone deacetylation, respectively.^{5–7} DNA methylation and histone deacetylation are known to play a role in the occurrence and progression of cancers.^{8,9} However, the relationship between DNMT1, DNMT3A, HDAC1, EZH2, LSD1, and CML is unknown.

Long noncoding RNAs (lncRNAs) are longer than 200 nt, and they do not encode proteins.¹⁰ Recent studies have reported that the lncRNA HOTAIR plays an important role in the development of not only solid tumors, such as breast cancer and non-small-cell lung cancer,^{11–14} but also hematopoietic malignancies, such as acute myeloid leukemia.^{15,16} However, the epigenetic regulation mechanisms of HOTAIR in advanced CML are unclear.

Similar to lncRNAs, microRNAs (miRNAs) do not encode proteins,¹⁷ but they are <200 nt in length.^{18,19} Low levels of miR-143-3p are associated with decreased risk of ovarian cancer.²⁰ In breast cancer, miR-143-3p regulates proliferation and apoptosis by targeting MYBL2.²¹ However, the relationship between miRNA and CML blast crisis is largely unknown.

Patients and methods

Specimen collection

Bone marrow samples were collected from 70 patients with CML admitted to the Department of Hematology of the Second Hospital of Hebei Medical University between May 2016 and June 2017 (Table 1). Bone marrow samples from 10 healthy donors were used as controls. Peripheral blood mononuclear cells were isolated via lymphocyte separation. This study was approved by the ethics committee of the Department of Hematology of the Second Hospital of Hebei Medical University, and each patient signed informed consent. The inclusion criteria were as follows: 1) diagnosis of CML via bone marrow morphology, immunology, molecular biology, and cytogenetic analysis; 2) clear pathological stag-

ing; and 3) availability of intact clinical data. The exclusion criteria were as follows: 1) significant organ dysfunction; 2) pregnancy; and 3) failure to provide informed consent. No chemotherapy was administered before specimens were collected.

Cell culture

KCL22 and K562 cells were from Shanghai Hong Shun Biotechnology Co., Ltd. (Shanghai, China). The use of K562 and KCL22 cells was confirmed by the ethics committee of the Department of Hematology of the Second Hospital of Hebei Medical University. KCL22 cells were cultured in Iscove's Modified Dulbecco's Medium (IMDM; Thermo Fisher Scientific, Waltham, MA, USA) supplemented with 10% fetal bovine serum (FBS; Clark Bioscience, Claymont, DE, USA), 100 units/mL penicillin, and 100 µg/mL streptomycin. K562 cells were maintained in the RPMI 1640 medium (Thermo Fisher Scientific) supplemented with 10% FBS, 100 units/mL penicillin, and 100 µg/mL streptomycin.

Cell treatment

5-Azacytidine was purchased from ApexBio (Houston, TX, USA). KCL22 and K562 cells were seeded in 6-well plates at 5×10^6 cells/well. MTT assays were performed to measure the EC50 concentrations of 5-azacytidine. The K562 and KCL22 cells were treated with 5-azacytidine at the EC50 values. KCL22 cells were treated with 60, 80, and 100 µmol/L 5-azacytidine; K562 cells were treated with 40, 60, and 80 µmol/L 5-azacytidine. The cells were treated for 48 hours.

MTT assays

We seeded KCL22 and K562 cells into 96-well plates (1×10^4 cells/well) after transfection and cultured them for 0, 24, 48, 72, 96, and 120 hours using IMDM with 10% FBS at 37°C and RPMI 1640 with 10% FBS. Proliferation of the KCL22 and K562 cells was determined with an MTT assay. Briefly, following cell culture, 10 µL of MTT reagent (Sigma-Aldrich Co., St Louis, MO, USA) was added to

Table 1 Characteristics of the patients included in the study

Item	CML-CP (n=40)	CML-AP (n=15)	CML-BP (n=15)
Age (years), median (range)	41.4 (9–65)	49.1 (13–69)	51.9 (20–69)
Male/female, (n/n)	26/14	9/6	10/5
WBCs $\times 10^9$ /median (range)	221.4 (30.2–517)	263.5 (47.4–396)	69.5 (27.4–224)
Hemoglobin level (g/L)	94 (76–120)	75 (61–105)	62.4 (52–79)
Platelet count, 10^9 /median (range)	518 (99–809)	305 (52–725)	35.5 (19–71)

Abbreviations: AP, accelerated phase; BP, blast phase; CML, chronic myeloid leukemia; CP, chronic phase; WBC, white blood cell.

each well, and the 96-well plates were incubated at 37°C in a humidified 5% CO₂ atmosphere for 4 hours.

Apoptosis assay

KCL22 and K562 cells were seeded into six-well plates after transfection and treated according to the manufacturer's instructions. The cells were stained with 5 µL of Annexin V-FITC and 5 µL of propidium iodide (BD Biosciences, Franklin Lakes, NJ, USA), and then analyzed with a BD FAC-SCanto II system (BD Biosciences). The apoptosis results were analyzed using the BD FACSDiva (BD Biosciences)

Real-time quantitative reverse transcription polymerase chain reaction (RT-PCR)

Total RNA was isolated using the RNA easy Mini Kit (Qiagen NV, Venlo, the Netherlands), and the RevertAid First Strand cDNA Synthesis Kit (Thermo Fisher Scientific) was used to synthesize cDNA according to the manufacturer's instructions. The reaction was performed at 42°C for 60 minutes, followed by 25°C for 5 minutes and finally 70°C for 5 minutes. Polymerase chain reaction (PCR) amplification was performed in 20 µL final volumes containing 1 µL of template cDNA, 1 µL of sense primer, 1 µL of antisense primer, 10 µL of SYBR Green Mix (Thermo Fisher Scientific) and 8 µL of DEPC-treated water. Real-time quantitative RT-PCR was performed

using an ABI 7500 real-time rotary analysis system (American BioInnovations, Sparks, MD, USA). Real-time PCR primer sequences are shown in Table 2. For each run, each reaction was repeated independently at least three times. RT-PCR was performed under the following conditions: enzyme activation at 95°C for 5 minutes, followed by 45 cycles of 95°C for 15 seconds, 60°C for 35 seconds, and 72°C for 20 seconds. The 2^{-ΔΔC_t} method was used to calculate relative gene expression.

Methylation-specific PCR

DNA extraction kit (Shanghai Generay Biotech Co., Ltd, Shanghai, China). DNA was stored at -80°C. Sulfite conversion of DNA was performed using the EZ DNA methylation-Gold Kit (Zymo USA, Inc., Fleming Island, FL, USA) according to the manufacturer's instructions. For all reactions, the amplification reaction contained bisulfite-modified DNA (2 µL), ZymoTaq PreMix (12.5 µL), H₂O (8.5 µL), upstream primer (1 µL), and downstream primer (1 µL). The PCR conditions were 95°C for 10 minutes, followed by 35 cycles (30 seconds at 95°C, 45 seconds at 54°C for annealing, and 45 seconds at 72°C) and a final extension step of 7 minutes at 72°C. The primers were designed according to reference documents (Oka et al, 2001), and the original cDNA sequences were checked in the gene pool. The PCR products after 2% agarose gel electrophoresis (GoldView; Saibaisheng Bioengineering Co., Ltd., Beijing, China) were as follows:

Table 2 Primer sequences for RT-PCR

Gene	Primer	Product (bp)
HOTAIR	Forward: 5'-CTCCCCTTCTAGCGCTCACG-3'	132
	Reverse: 5'-CTAGCCGCCGTCTATACTACCGGCT-3'	
DNMT1	Forward: 5'-AGACACTCGCTCAGCTTCTTG-3'	116
	Reverse: 5'-CAATTGCTGCTGGGATTCATC-3'	
DNMT3A	Forward: 5'-CTGCTTTGTATTCCCTTTTGCA-3'	141
	Reverse: 5'-TTGATTTCTCCTGGCTGTCTC-3'	
HDAC1	Forward: 5'-CCTCCCAGTGAAGTCATCGTGG-3'	181
	Reverse: 5'-GGACAGGTGCTTCATCAGCTCG-3'	
PI3K	Forward: 5'-ACCACACGTTCCCTAAGCTGG-3'	124
	Reverse: 5'-TCCCTGCACGCAGAGATTTT-3'	
AKT	Forward: 5'-AACTACAACCTCTTCCCTCGAA-3'	109
	Reverse: 5'-CAAAGTTATGTCCACTGTCTCT-3'	
LSD1	Forward: 5'-CATCACCGTAGTCCCATTG-3'	175
	Reverse: 5'-AAATCCCCACTTCGCTTGG-3'	
EZH2	Forward: 5'-CCACCCTTGTGCTCTTCCCTG-3'	100
	Reverse: 5'-TCTGCCACCCGAGTGTAACCA-3'	
mTOR	Forward: 5'-TCCCTGCACGCAGAGATTTT-3'	131
	Reverse: 5'-TTGATTTCTCCTGGCTGTCTC-3'	
ACTB	Forward: 5'-GAGCTACGAGCTGCCTGAC-3'	121
	Reverse: 5'-GGTAGTTTCGTGGATGCCACAG-3'	

Abbreviation: RT-PCR, reverse transcription polymerase chain reaction.

HOTAIR methylated (M) and HOTAIR unmethylated (U) positive–negative methylation, HOTAIR M and HOTAIR U partial methylation, and HOTAIR M and HOTAIR U positive–negative unmethylation. The primers were synthesized by Saibaisheng Bioengineering Co., Ltd., and their sequences are shown in Table 3.

Western blot (WB)

Cells were lysed using radioimmunoprecipitation assay (RIPA) buffer, and protein was obtained. The protein concentration was determined using a BCA protein assay kit (Thermo Fisher Scientific). Proteins were separated on 8%–15% SDS-PAGE gels, and the separated bands were electrotransferred to polyvinylidene fluoride (PVDF) membranes (Amersham Pharmacia, Piscataway, NJ, USA). The PVDF membranes were blocked with 5% non-fat skim milk at room temperature and probed with antibodies targeting PI3K (1:1,000), AKT (1:3,000), mTOR (1:2,000), pPI3K (1:3,000), pAKT (1:3,000), pmTOR (1:1,000), EZH2 (1:2,000), LSD1 (1:2,000), DNMT1 (1:2,000), DNMT3A (1:3,000), HDAC1 (1:2,000) and ACTB (1:5,000) (Abcam, Cambridge, MA, USA). The diluted corresponding secondary antibody was incubated with the membranes at room temperature for 1 hour, and the membranes were then rinsed three times for 5 minutes each with tris-buffered saline. Protein expression was examined using a BioSpectrum Imaging System (UVP, LLC, Upland, CA, USA).

RNA pull-down assay

HOTAIR-binding proteins were determined using a Pierce Magnetic RNA-Protein Pull-Down Kit (Thermo Fisher Scientific) according to the manufacturer's protocols. HOTAIR sense and HOTAIR antisense lncRNAs were constructed and incubated with the protein from KCL22 cells, and 100 mL of silver staining solution was added to the HOTAIR sense and HOTAIR antisense lncRNA and protein mixture. The

total protein was then collected for mass spectrometry sequencing.

RNA immunoprecipitation

RNA immunoprecipitation was performed using the Magna RIP Kit (EMD Millipore, Billerica, MA, USA). Complete KCL22 cell lysates were prepared according to the manufacturer's instructions and contained phosphatase and proteinase inhibitors. Next, we evaluated the connection and configuration of the antibody beads, antibody beads suspension was mixed with the sample after thawing and placed on a table at 4°C for overnight incubation. Immunoprecipitation was completed after the suspension was placed on the magnetic frame and washed with washing buffer at least six times. Finally, the immune co-precipitation products were collected, and RNA was extracted and purified to determine the abundance of the target RNA.

Luciferase assay

In the dual-luciferase assay, we seeded HOTAIR and negative control (NC) cells into 24-well plates and co-transfected the cells with 0.4 mg of firefly luciferase reporter vector, 0.08 mg of Renilla luciferase pRL-TK (Promega Corporation, Fitchburg, WI, USA), and control vector according to the manufacturer's instructions. At 48 hours after transfection, the cells were lysed. Luciferase activity was measured by a dual-luciferase reporter gene assay system (Promega Corporation). Renilla luciferase activity was determined relative to the standardized fluorescence activity of firefly luciferase. All assays were repeated at least three times.

Statistical analysis

The data are expressed as mean±SD and were analyzed using SPSS 19.0 software (IBM Corporation, Armonk, NY, USA). Significant differences between groups were analyzed

Table 3 Primer sequences of the methylated HOTAIR and miRNA-143 gene

Gene	Primer	Product (bp)
HOTAIR, M-MSP	Forward: 5'-GCGAATTATTATTATATAGCCTTC-3' Reverse: 5'-TCACGCGCTACGAAGGGAAACG-3'	162
HOTAIR, U-MSP	Forward: 5'-GTGAGGTGTTATTACCGTATAGTTTGG-3' Reverse: 5'-TTCACACATACAGGTTCCAAACAAT-3'	162
miR-143, M-MSP	Forward: 5'-CGTAGCTACTGCTATTATCGTTC-3' Reverse: 5'-TTATGCCTTGGGCTACGTA-3'	174
miR-143, U-MSP	Forward: 5'-GGTACGTATGCTTGGGCTTATG-3' Reverse: 5'-ATTGGCGCTTAGCTAGGCTA-3'	174

Abbreviations: M, methylated; MSP, methylation-specific polymerase chain reaction; U, unmethylated.

using Student's *t*-test or one-way ANOVA for more than two subgroups. The chi-squared test was applied to compare rates, and $P < 0.05$ was considered to indicate a statistically significant difference.

Results

mRNA expression and protein levels in patients with CML

Expression levels of DNMT1, DNMT3A, HDAC1, HOTAIR, EZH2, and LSD1 were higher in AP and BP

than in CP and healthy donors ($P < 0.05$), whereas miR-143 expression was lower in patients with advanced-stage CML ($P < 0.05$). The highest levels of DNMT1, DNMT3A, HDAC1, HOTAIR, EZH2, and LSD1 were found in BP CML (Figures 1A–E and 3A and F). The advanced stage of CML was associated with a high mRNA level of DNMT1, DNMT3A, HDAC1, HOTAIR, EZH2, and LSD1 and a low level of miR-143.

Protein levels of DNMT1, DNMT3A, HDAC1, EZH2, and LSD1 were determined in the bone marrow mononuclear

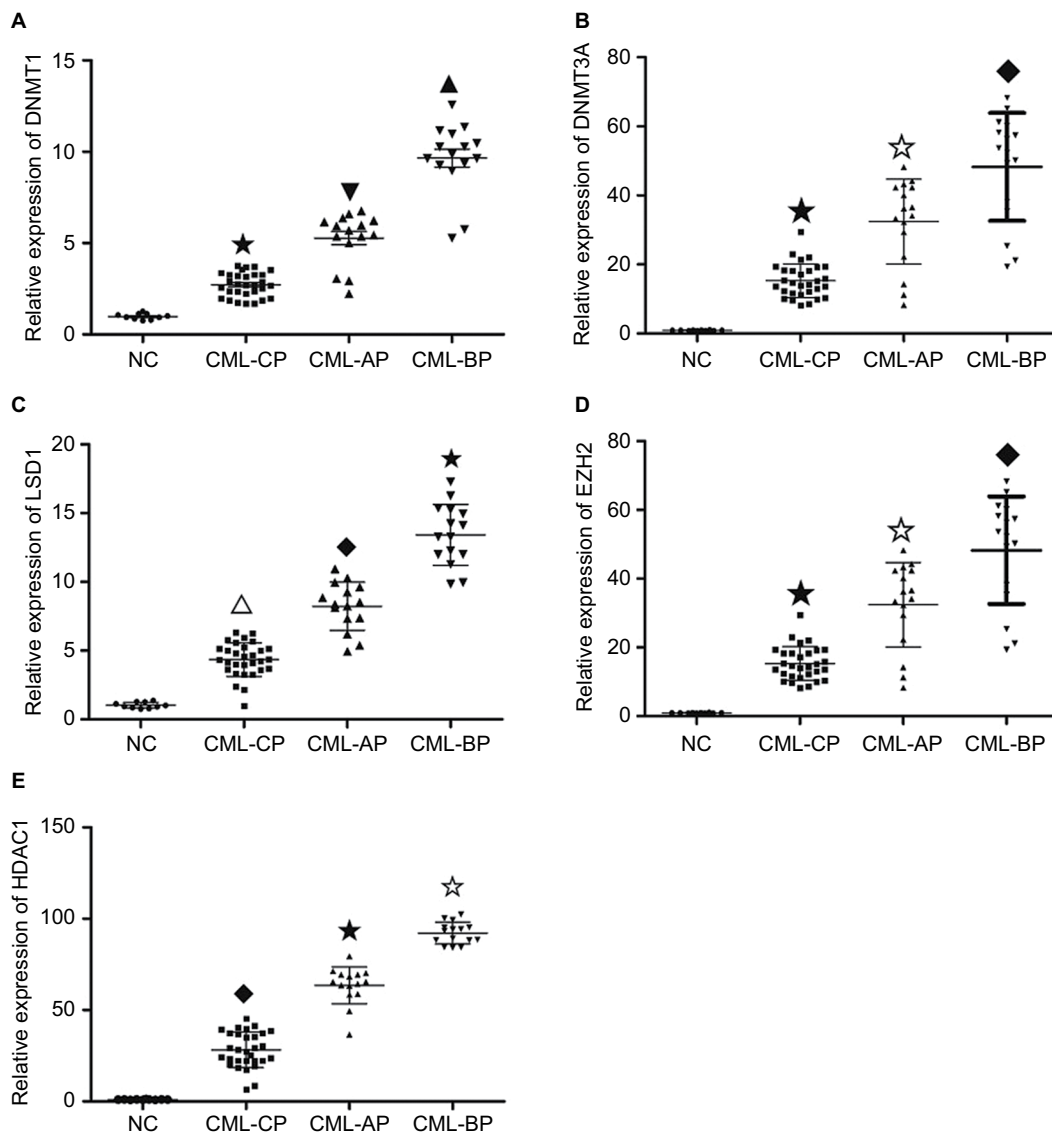


Figure 1 mRNA level in different phases of CML and healthy donors.

Notes: mRNA levels were assayed by RT-qPCR. (A) DNMT1 level was higher in AP and BP than in CP and healthy donors ($\star\blacktriangledown\blacktriangle P < 0.05$, $\star\blacktriangledown P < 0.05$). (B) DNMT3A mRNA level was higher in AP and BP than in CP and healthy donors ($\star\blacklozenge P < 0.05$, $\blacktriangledown\blacklozenge P < 0.05$). (C) The mRNA level of LSD1 was higher in AP and BP than in CP and healthy donors ($\blacktriangle\blacklozenge P < 0.05$). (D) The level of EZH2 was higher in AP and BP than in CP and healthy donors ($\star\blacklozenge P < 0.05$). (E) The HDAC1 mRNA level was higher in AP and BP than in CP and healthy donors ($\blacklozenge\blackstar P < 0.05$).

Abbreviations: AP, accelerated phase; BP, blast phase; CML, chronic myeloid leukemia; CP, chronic phase; NC, negative control; RT-qPCR, quantitative reverse transcription polymerase chain reaction.

cells (BMBCs) of patients with different phases of CML and healthy donors. Patients with chronic-phase CML exhibited lower DNMT1, DNMT3A, HDAC1, EZH2, and LSD1 levels than patients with accelerated or BP CML ($P < 0.05$). The highest DNMT1, DNMT3A, HDAC1, EZH2, and LSD1 protein levels were found in BP CML (Figure 2). The advanced stage of CML was associated with a high protein level of DNMT1, DNMT3A, HDAC1, EZH2, and LSD1.

Methylation status of the HOTAIR promoter in different stages of CML

HOTAIR promoters were detected by methylation-specific PCR in different phases of CML and healthy donors. HOTAIR promoters were not detected in healthy donors and were detected in only four of 20 patients with CP CML and in all patients with AP and BP CML. The rate of methylation was higher in AP and BP CML than in CP CML or in

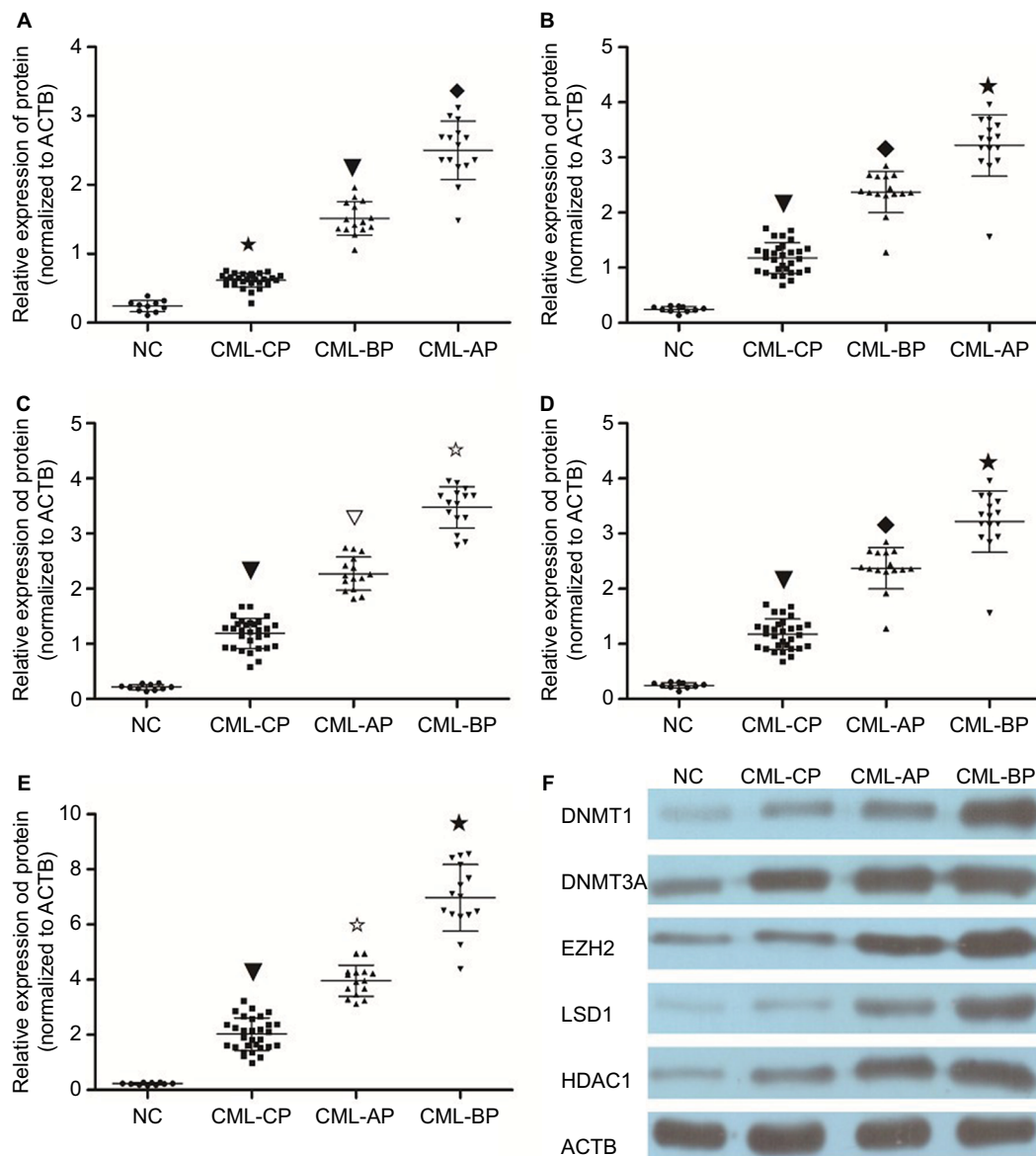


Figure 2 Protein level in different phases of CML and healthy donors.

Notes: Protein levels were assayed by WB. (A) DNMT1 protein level was higher in AP and BP than in CP and healthy donors ($\star \blacktriangledown \blacklozenge P < 0.05$). (B) DNMT3A protein level was higher in AP and BP than in CP and healthy donors ($\blacktriangledown \blacklozenge \star P < 0.05$). (C) Protein level of HDAC1 was higher in AP and BP than in CP and healthy donors ($\blacktriangledown \blacktriangledown \star P < 0.05$). (D) Protein level of LSD1 was higher in AP and BP than in CP and healthy donors ($\blacktriangledown \blacklozenge \star P < 0.05$). (E) EZH2 protein level was higher in AP and BP than in CP and healthy donors ($\blacktriangledown \star P < 0.05$). (F) Protein expression in bone marrow (BM) cells from CML patients and normal controls. Protein expression level was determined by immunoblotting with ACTB (b-actin) as a control.

Abbreviations: AP, accelerated phase; BP, blast phase; CML, chronic myeloid leukemia; CP, chronic phase; NC, negative control; WB, Western blot.

healthy donors ($P < 0.05$), and there was no significant difference in methylation rates between CP CML and healthy donors ($P > 0.05$; Figure 3). The advanced stages of CML were associated with a high methylation rate.

mRNA and protein levels after drug treatment

K562 and KCL22 cells were treated with different concentrations of 5-azacytidine according to the EC50 values of 5-azacytidine in these cells. K562 cells received 40, 60, and 80 $\mu\text{mol/L}$ 5-azacytidine, and KCL22 cells received 60, 80, and 100 $\mu\text{mol/L}$. The mRNA and protein levels of DNMT1, DNMT3A, HDAC1, EZH2, and LSD1, as well as HOTAIR mRNA levels, decreased with 5-azacytidine treatment in a concentration-dependent fashion. The maximum mRNA levels were found in the cells treated with the lowest drug concentrations ($P < 0.05$). miRNA-143 levels increased with treatment in a concentration-dependent manner ($P < 0.05$;

Figure 4). 5-azacytidine decreased the levels of DNMT1, DNMT3A, HDAC1, HOTAIR, EZH2, and LSD1 and increased the level of miR-143.

HOTAIR knockdown inhibited proliferation and promoted apoptosis

To examine the biological role of lncRNA HOTAIR in CML blast crisis, we first determined the expression of HOTAIR in BMMCs and K562 and KCL22 cells; the result indicated that the expression of HOTAIR was higher in KCL22 and K562 cells than in BMMCs ($P < 0.05$; Figure S1A). Next, sh-HOTAIR and sh-control were synthesized and transfected into KCL22 and K562 cells; the results indicated that the expression of HOTAIR was lower in the knockdown group than in the control group ($P < 0.05$; Figure S1B and C). HOTAIR knockdown significantly inhibited KCL22 and K562 cell proliferation ($P < 0.05$) and promoted apoptosis ($P < 0.05$) compared to transfection with control shRNA

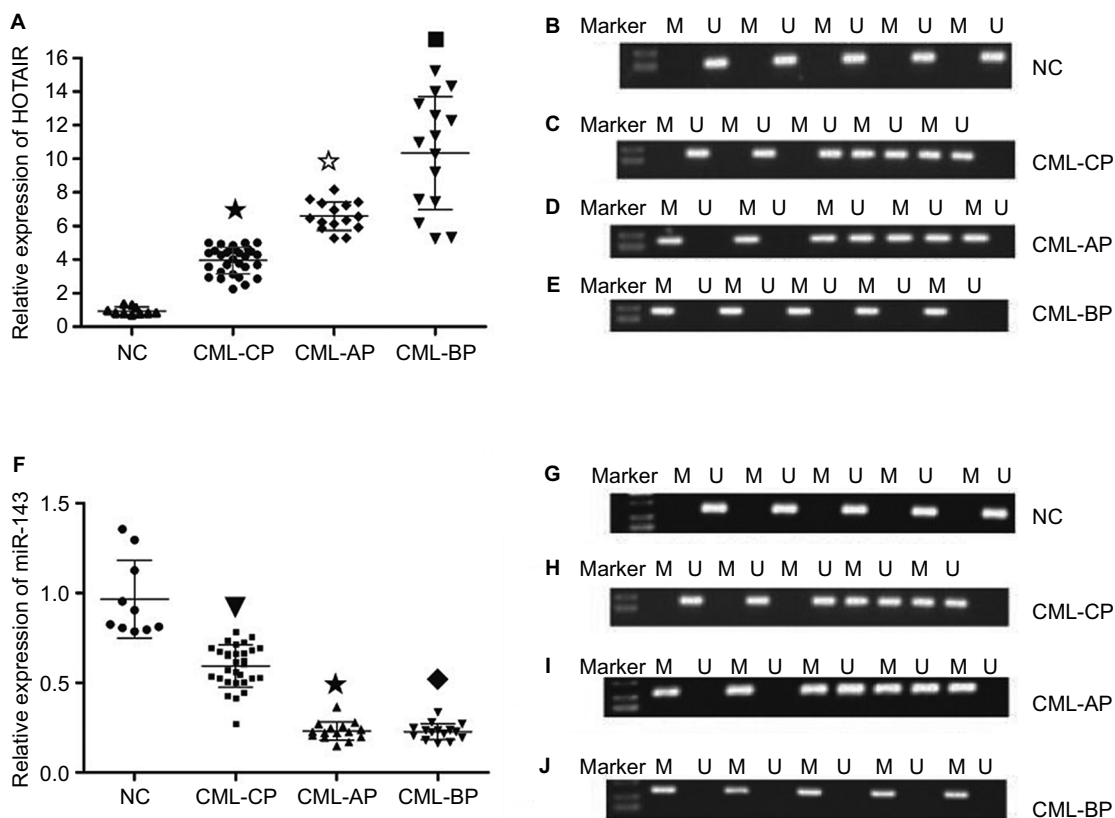


Figure 3 The mRNA level and methylation status of HOTAIR and miR-143 in different phases of CML.

Notes: (A) mRNA level of HOTAIR was higher in AP and BP than in CP and healthy donors ($\star \blacktriangle \blacksquare P < 0.05$). (B) The methylation rate of HOTAIR in the NC group was 0. (C) The methylation rate of HOTAIR in CML-CP was 20%. (D and E) All (100%) of the samples for patients with disease progression (CML-AP + CML-BP) were methylated compared with the normal controls and CML-CP patients, $P < 0.01$. (F) miR-143 mRNA level was lower in AP and BP than in CP and healthy donors ($\blacktriangledown \blacklozenge P < 0.05$). (G) Methylation of miR-143 in the NC group could not be detected. (H) The methylation rate of miR-143 in CML-CP was 25%. (I and J) Methylation of miR-143 was detected in all patients with disease progression (CML-AP + CML-BP).

Abbreviations: AP, accelerated phase; BP, blast phase; CML, chronic myeloid leukemia; CP, chronic phase; NC, negative control.

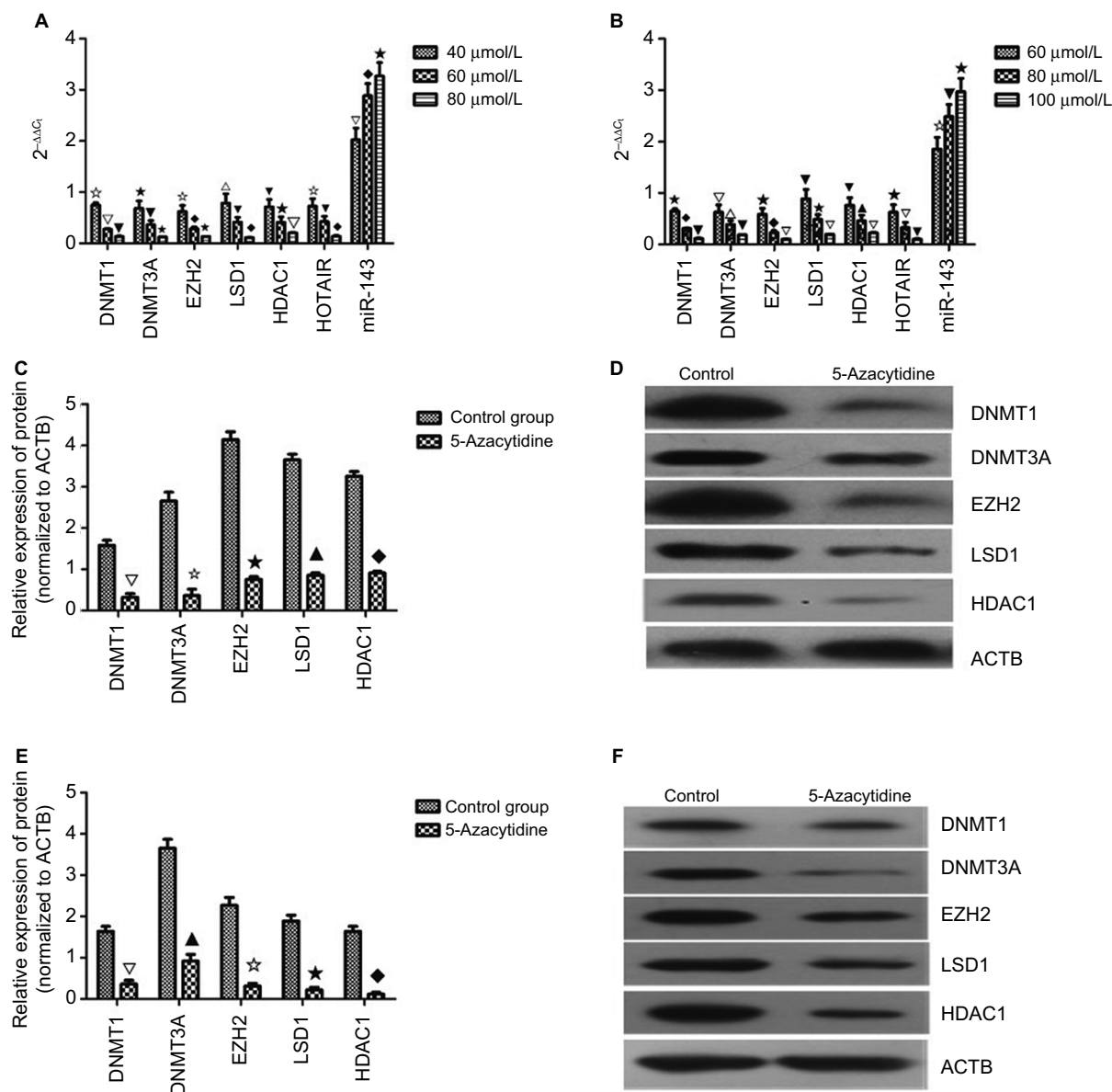


Figure 4 mRNA and protein level after drug treatment. **Notes:** (A) In K562 cells, the mRNA level of DNMT1, DNMT3A, EZH2, LSD1, HDAC1, HOTAIR, and miR-143 was decreased in a concentration-dependent manner (☆▽▽ P<0.05, ★▽☆ P<0.05, ☆◆★ P<0.05, △▽◆ P<0.05, ▼★▽ P<0.05, ☆▽◆ P<0.05, ▽◆★ P<0.05). (B) In KCL22 cells, the mRNA level of DNMT1, DNMT3A, EZH2, LSD1, HDAC1, HOTAIR, and miR-143 was decreased in a concentration-dependent manner (★◆▽ P<0.05, ▽△▽ P<0.05, ★◆▽ P<0.05, ▼★▽ P<0.05, ▼★▽ P<0.05, ★▽▽ P<0.05, ☆▽★ P<0.05). (C and D) In K562 cells, the protein level of DNMT1, DNMT3A, EZH2, LSD1, and HDAC1 was decreased in a concentration-dependent manner (▽☆★◆ P<0.05). (E and F) In KCL22 cells, the protein level of DNMT1, DNMT3A, EZH2, LSD1, and HDAC1 was decreased in a concentration-dependent manner (▽△★☆◆ P<0.05).

(Figure 5). HOTAIR knockdown inhibited proliferation and promoted apoptosis.

HOTAIR interacts with miR-143

Competing endogenous RNAs are lncRNAs that act as sponges to bind miRNAs and prevent them from performing normal regulatory activities. To investigate whether HOTAIR acts in this way, a luciferase reporter assay was performed to determine whether HOTAIR binds miR-143. The results

indicated that transfection with miR-143 mimic decreased the luciferase activity of HOTAIR wild-type plasmids in KCL22 cells (*P*<0.05; Figure 6B) but had no effect on mutant-type plasmids (*P*>0.05; Figure 6A). HOTAIR was able to interact with miR-143 (Figure 6D).

HOTAIR interacts with EZH2 and LSD1

To determine whether the lncRNA HOTAIR binds protein, RNA pull-down and RIP assays were performed. The results

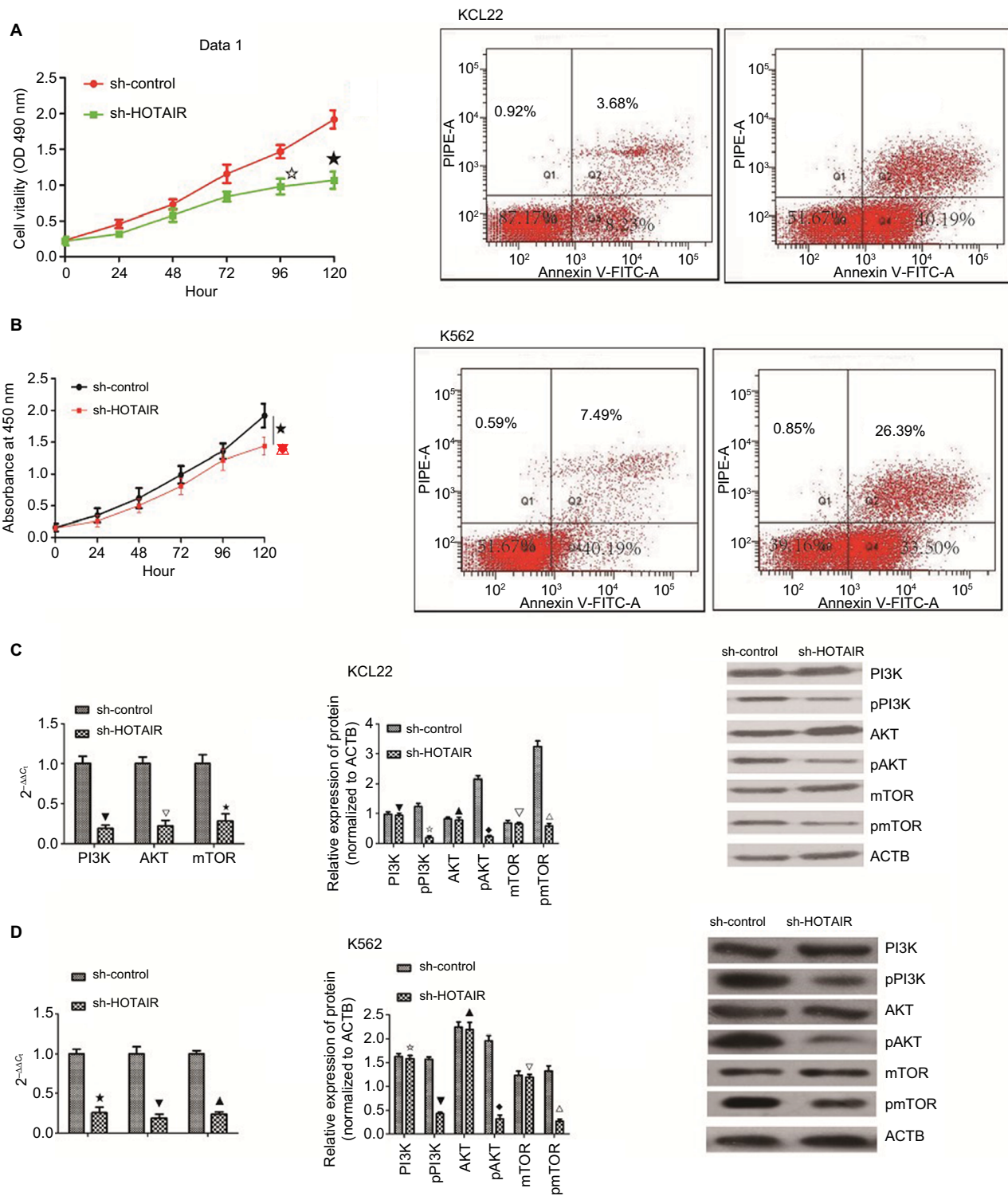


Figure 5 Biological role of HOTAIR in K562 and KCL22.

Notes: (A) K562 cells were transfected with sh-HOTAIR and sh-control, and MTT was performed. Knockdown of HOTAIR inhibited proliferation of cells (★☆ $P < 0.05$). Silencing of HOTAIR promoted apoptosis of KCL22 cells ($P < 0.05$). (B) KCL22 was transfected with sh-HOTAIR and sh-control, and MTT was performed. Knockdown of HOTAIR inhibited proliferation of cells (★ $P < 0.05$). Silencing of HOTAIR promoted apoptosis of K562 cells ($P < 0.05$). (C) In KCL22 cells, the PI3K, AKT, and mTOR mRNA level was decreased in the knockdown group (▼▽★ $P < 0.05$). In KCL22 cells, silencing of HOTAIR had no effect on the total protein level of PI3K, AKT, and mTOR (▼▲▽ $P > 0.05$) but decreased phosphorylated protein levels of PI3K, AKT, and mTOR (☆◆△ $P < 0.05$). (D) In K562 cells, the PI3K, AKT, and mTOR mRNA levels were decreased in the knockdown group (★▼▲ $P < 0.05$). In K562 cells, silencing of HOTAIR had no effect on the total protein level of PI3K, AKT, and mTOR (☆▲▽ $P > 0.05$) but decreased phosphorylated protein levels of PI3K, AKT, and mTOR (▼◆△ $P < 0.05$).

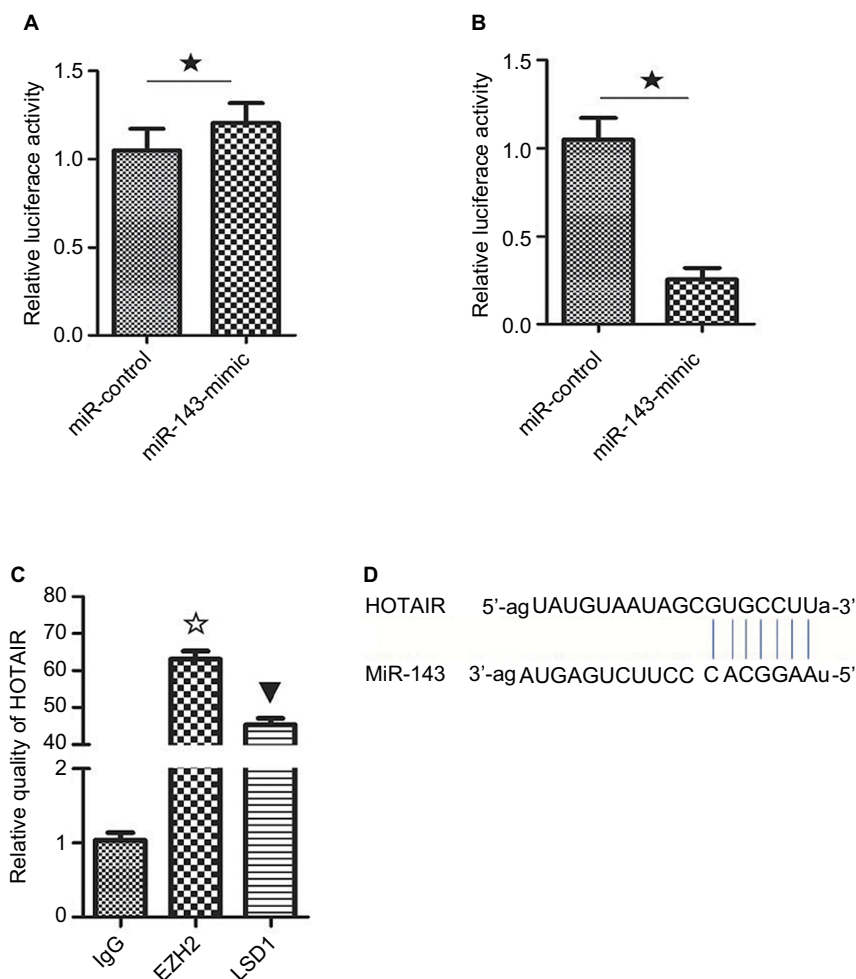


Figure 6 HOTAIR act as a ceRNA for miR143.

Notes: (A) Luciferase reporter plasmids containing the mutant HOTAIR (MUT-HOTAIR) sequence were co-transfected into HEK-293T cells with miR-143 mimics or their corresponding NCs. There was no significant difference in luciferase activity between the miR-143 mimic group and the negative control group ($\star P > 0.05$). (B) WT-HOTAIR was co-transfected into HEK-293T cells with miR-143 mimic or negative control. Luciferase activity was lower in the miR-143 mimic group compared with that in the negative control group ($\star P < 0.05$). (C) RIP-PCR was performed to verify that LSD1 and EZH2 bind HOTAIR ($\star \blacktriangledown P < 0.05$). (D) miR-143 was a target of HOTAIR.

Abbreviations: MUT-HOTAIR, mutant HOTAIR; NC, negative control; PCR, polymerase chain reaction; RIP, RNA-binding protein immunoprecipitation; WT, wild type.

showed that EZH2 and LSD1 interacted with the HOTAIR lncRNA ($P < 0.05$; Figure 6C). EZH2 and LSD1 were able to interact with HOTAIR.

Overexpression of miR-143 inhibited proliferation and promoted apoptosis

To examine the influence of miR-143 on cell proliferation and apoptosis of KCL22 and K562, we first measured miR-143 expression levels in KCL22, K562, and BMMCs. The result showed that miR-143 expression levels were lower in KCL22 and K562 cells than in BMMCs (Figure S1D). Next, the miR-143 mimic was transfected into KCL22 and K562 cells (Figure S1E and F); proliferation assays were performed to assess the influence of miR-143 on proliferation of KCL22 and K562 cells. We found that cell viability of KCL22 cells decreased 96 and 120 hours after

transfection with the miR-143 mimic ($P < 0.05$; Figure 7A and B), whereas that of K562 cells was reduced 120 hours after transfection with the miR-143 mimic ($P < 0.05$; Figure 7A and B). Flow cytometry was then performed to determine the influence of miR-143 on KCL22 and K562 apoptosis; the results indicated that miR-143 overexpression promoted apoptosis ($P < 0.05$; Figure 7A and B). Overexpression of miR-143 inhibited proliferation and promoted apoptosis

Both knockdown of HOTAIR and overexpression of miR-143 influence the PI3K/AKT pathway

PCR and WB were performed to assess the influence of miR-143 and HOTAIR on the PI3K/AKT signaling pathway. The results demonstrated that silencing of HOTAIR and overex-

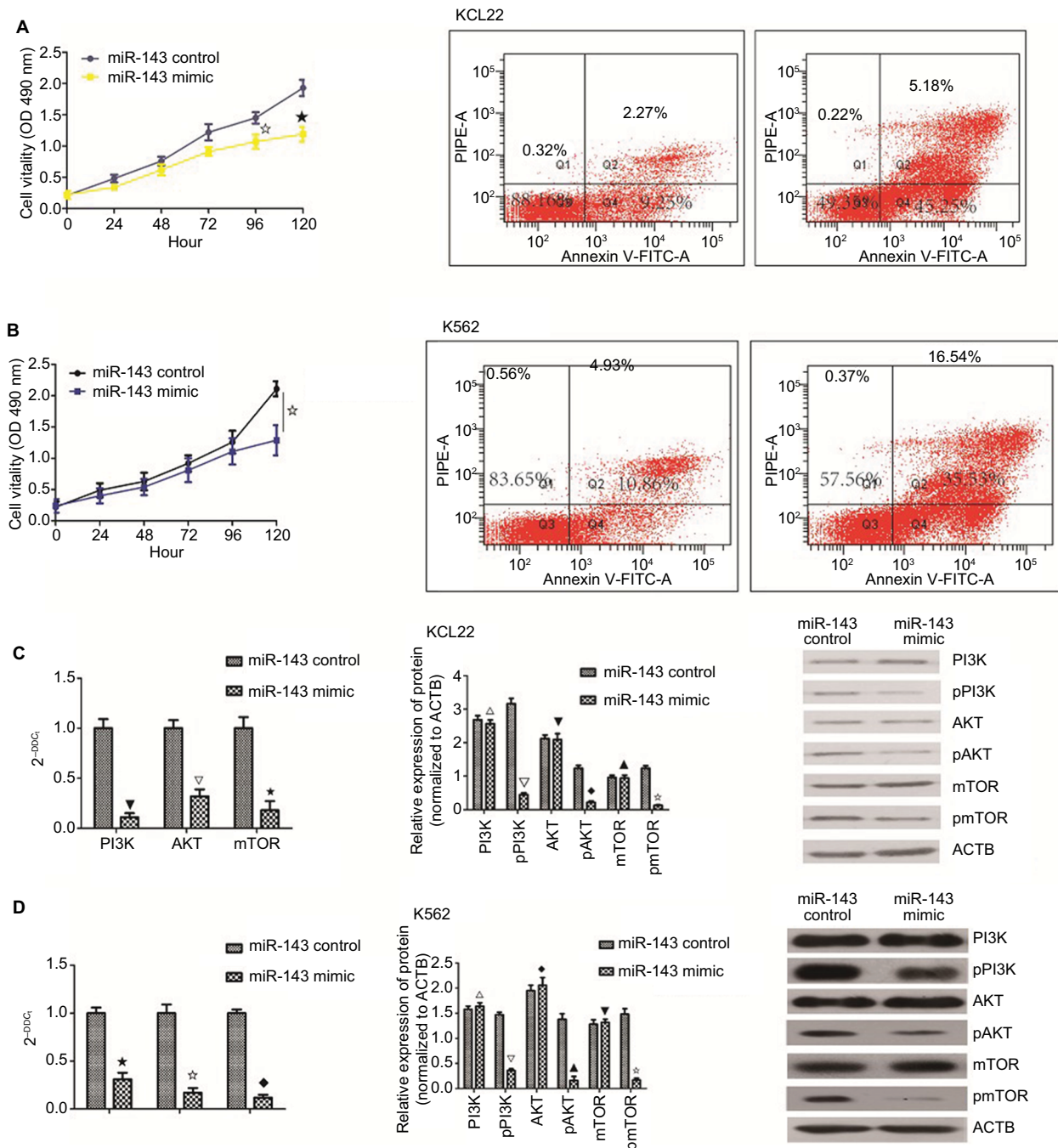


Figure 7 The biological role of miR143 in K562 and KCL22. **Notes:** (A) K562 cells were transfected with miR-143 mimic and miR-143 control. MTT assay was performed, and overexpression of miR-143 inhibited cell proliferation (☆★ $P < 0.05$). Overexpression of miR-143 promoted apoptosis in KCL22 cells ($P < 0.05$). (B) KCL22 cells were transfected with miR-143 mimic and miR-143 control. MTT assay was performed; overexpression of miR-143 inhibited cell proliferation (☆ $P < 0.05$). In K562 cells, overexpression of miR-143 promoted apoptosis ($P < 0.05$). (C) In KCL22 cells, PI3K, AKT, and mTOR mRNA levels were decreased in the overexpression group (▽▼☆ $P < 0.05$). In KCL22 cells, overexpression of miR-143 had no effect on total PI3K, AKT, and mTOR protein levels (△▽▲ $P > 0.05$) but decreased phosphorylated PI3K, AKT, and mTOR protein levels (▽◆☆ $P < 0.05$). (D) In K562 cells, PI3K, AKT, and mTOR mRNA levels were decreased in the overexpression group (★☆☆ $P < 0.05$); in K562 cells, overexpression of miR-143 had no effect on total PI3K, AKT, and mTOR protein levels (△◆▼ $P > 0.05$) but decreased phosphorylated PI3K, AKT, and mTOR protein levels (▽▲☆ $P < 0.05$).

pression of miR-143 decreased PI3K/AKT pathway mRNA levels ($P < 0.05$) and had no effect on total PI3K/AKT pathway protein levels ($P > 0.05$), but decreased levels of phosphorylated PI3K/AKT pathway proteins ($P < 0.05$; Figures 5 and 7). Both knockdown of HOTAIR and overexpression of miR-143 influenced phosphorylated PI3K/AKT pathway proteins.

Discussion

CML accounts for ~10% of all types of leukemia, and although it can occur in any age group, it most often occurs in patients aged 55–70 years. The incidence of CML is higher in males than in females.¹ The Philadelphia chromosome, characterized by the presence of the BCR–ABL1 fusion gene, is the most common chromosomal abnormality in CML. This fusion gene encodes a constitutively active tyrosine kinase signaling protein; this uncontrolled activity leads to CML.²² Although tyrosine kinase inhibitors have improved the 5-year survival rate of CML, some patients, due to drug resistance or an inability to tolerate tyrosine kinase inhibitors, still progress to AP and BP of CML.²³ Epigenetic gene regulation and noncoding RNA take part in this progression.²⁴

As a noncoding RNA, the lncRNA HOTAIR encodes no protein. It promotes proliferation of cervical cancer by targeting miR-143 and proliferation of breast cancer by upregulating Bcl-w.^{25,26} HOTAIR-mediated DNA methylation contributes to the progress of osteosarcoma by targeting miR126.²⁷ In our study, high levels of the HOTAIR were found in advanced stages of CML and were inversely proportional to miR-143 levels. We also found higher methylation promoter rates of HOTAIR and miR-143 in advanced CML. Thus, we speculated that high levels of HOTAIR lncRNA, low levels of miR-143, and higher methylation promoter rates of HOTAIR and miR-143 were associated with CML progression.

Methylation of DNA at CpG islands and histone modification are two mechanisms that lead to gene silencing, which affects the gene promoter.⁴ The DNMT family, which includes DNMT1, DNMT3A, and DNMT3B, mediates CpG island methylation,⁷ and the HDAC family regulates histone deacetylation.²⁸ The discovery of LSD1 was an important step in epigenetics research; methylation and other chemical modifications of histone lysine are dynamic regulatory processes. LSD1 has been accepted as an important prognostic factor in hepatocellular carcinoma and breast cancer.^{29,30} EZH2 belongs to polycomb repressive complex 2,³¹ is highly conserved, and can inhibit the transcription of downstream target genes. High levels of EZH2 are associated with poor prognosis in many types of cancer, such as hepatocellular carcinoma and pancreatic cancer.^{32,33}

Therefore, we examined DNMT1, DNMT3A, HDAC1, EZH2, and LSD1 mRNA and protein levels in different phases of CML and in healthy donors. We also measured the mRNA levels of HOTAIR and miR-143 in different phases of CML and in healthy donors. The results indicated that mRNA and protein levels of DNMT1, DNMT3A, HDAC1, EZH2, and LSD1 were higher in advanced CML and that mRNA levels of HOTAIR were higher and levels of miR-143 were lower in advanced CML. Thus, we concluded that high levels of EZH2 and LSD1 at least partly account for CML progression.

Next, we treated K562 and KCL22 cells with different concentrations of 5-azacytidine. Treatment increased miR-143 levels and decreased levels of DNMT1, DNMT3A, HDCA1, EZH2, LSD1, and HOTAIR in both KCL22 and K562 cell lines.

To further examine the role of HOTAIR and miR-143 in CML blast crisis, we knocked down HOTAIR and overexpressed miR-143. Both modifications inhibited proliferation and promoted apoptosis in KCL22 and K562 cells. We then performed RNA pull-down and RIP assays to study the regulation of HOTAIR in KCL22 cells and observed that EZH2 and LSD1 interacted with HOTAIR. Thus, we speculated that EZH2 and LSD1 may regulate HOTAIR in CML blast crisis.

PI3K/AKT signaling plays a critical role in many hematological diseases, including acute myeloid leukemia and CML.^{34–36} Our results showed that silencing of HOTAIR and overexpression of miR-143 decreased phosphorylation of PI3K/AKT/mTOR proteins; thus, we concluded that the PI3K/AKT pathway is a target for HOTAIR and miR-143.

Conclusion

HOTAIR is a potential therapeutic target for CML blast crisis, and demethylation drugs may represent a new treatment strategy.

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Author contributions

ZL conceived and designed the study and drafted the manuscript. JL reviewed the manuscript. Both authors read and approved the final manuscript. All authors contributed toward data analysis, drafting and revising the paper and agree to be accountable for all aspects of the work.

Disclosure

The authors report no conflicts of interest in this work.

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Supplementary material

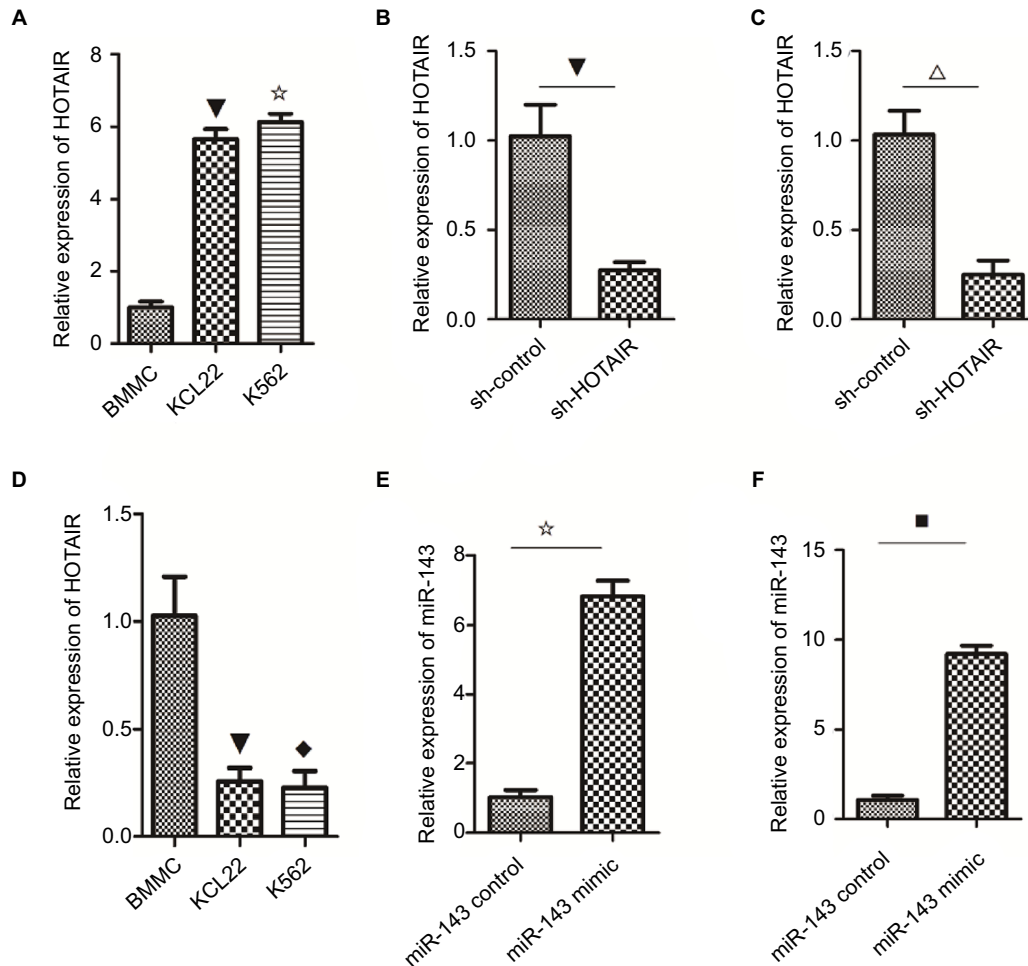


Figure S1 The level of HOTAIR and miR143 in k562 and KCL22.

Notes: (A) The expression of HOTAIR was higher in K562 and KCL22 cells than that in BMMCs (☆, ▼ $P < 0.05$). (B) In KCL22 cells, the expression of HOTAIR was downregulated in the knockdown group compared with the empty group (▼ $P < 0.05$). (C) In K562 cells, the expression of HOTAIR was decreased in the knockdown group compared with the empty group (△ $P < 0.05$). (D) The expression of miR-143 in K562 and KCL22 cells was lower than that in BMMCs (◆, ▼ $P < 0.05$). (E) In K562 cells, the expression of miR-143 was upregulated in the overexpression group compared with the control group (☆ $P < 0.05$). (F) The expression of miR-143 was higher in the overexpression group compared with the control group (■ $P < 0.05$).

Abbreviation: BMMC, bone marrow mononuclear cell.

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