



Disinfection Procedures and Their Effect on the Microorganism Colonization of Dental Impression Materials: A Systematic Review and Meta-Analysis of In Vitro Studies

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Abstract: Dental impressions are contaminated with potentially pathogenic microorganisms when they come into contact with patient blood, saliva, and plaque. Numerous disinfectants are used; however, no sole disinfectant can be designated as universal for all the impression materials. Thus, the aim of this study is to systemically review the literature to evaluate the effect of the existing disinfection procedures on the bacterial colonization of dental impression materials. This systematic review and meta-analysis was conducted according to the PRISMA statement. PubMed (MEDLINE), Web of Science, Scopus, EMBASE, and SciELO databases were screened up to April 2021. Eligibility criteria included in vitro studies reporting the antibacterial activity of disinfectant solutions in dental impression materials. The meta-analysis was performed using Review Manager (version 5.3.5). A global comparison was performed with the standardized mean difference based on random-effect models at a significance level of $\alpha = 0.05$. A total of seven studies were included in the meta-analysis. The included studies described the effect of disinfection processes with chlorhexidine gluconate, alcohol, sodium hypochlorite, glutaraldehyde, and hydrogen peroxide in alginate, polyvinyl siloxane, and polyether impression materials. The meta-analyses showed that the use of chlorhexidine, alcohol, glutaraldehyde, and sodium hypochlorite reduced the colony-forming units by a milliliter (CFU/mL) in alginate (p < 0.001). On the other hand, glutaraldehyde, sodium hypochlorite, and alcohol reduced the CFU/mL in polyvinyl siloxane (p < 0.001). Finally, alcohol and glutaraldehyde reduced the CFU/mL in polyether material (p < 0.001). High heterogenicity was observed for the alginate and polyvinyl siloxane materials ($I^2 = 74\%$; $I^2 = 90\%$). Based on these in vitro studies, the disinfection of impression materials with several disinfection agents reduces the CFU/mL count.

Keywords: antibacterial effect; dental impressions; disinfectant agents; disinfection; oral bacteria



Citation: Hardan, L.; Bourgi, R.; Cuevas-Suárez, C.E.; Lukomska-Szymanska, M.; Cornejo-Ríos, E.; Tosco, V.; Monterubbianesi, R.; Mancino, S.; Eid, A.; Mancino, D.; et al. Disinfection Procedures and Their Effect on the Microorganism Colonization of Dental Impression Materials: A Systematic Review and Meta-Analysis of In Vitro Studies. *Bioengineering* **2022**, *9*, 123. https://doi.org/10.3390/ bioengineering9030123

Academic Editor: Chengfei Zhang

Received: 15 February 2022 Accepted: 11 March 2022 Published: 16 March 2022

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1. Introduction

Dental impressions are certainly contaminated with possibly pathogenic microorganisms when they come into contact with patient blood, saliva, and plaque [1,2]. This could be the source of disease transmitters and cross-infections for dentists, dental assistants, and laboratory technicians [3,4]. Consequently, sanitizing the impressions efficiently before transportation to the laboratory technician ensembles is crucial [5]. Indeed, when the impressions are sterilized, this can avoid the transmission of disease, yet it is not the ideal way, since dimensional changes can occur [6].

Considering that, in some countries, tap water contains halogenated compounds, the Advisory British Dental Association Service recommends the rinsing of impression materials with tap water in daily dental practice; despite this, although some of the microorganisms adhered to the surface of a dental impression could be removed by this procedure, a high percentage still remains [7]. This has been exhibited to lessen the amounts of the bacteria on the surface of the impression presented by nearly 90% [8]. Nevertheless, a noteworthy number of bacteria would persist. More recent suggestions support the use of a disinfecting solution [9]. Knowledge evidently varies about the type, concentration, and immersion times of disinfection protocols, making it difficult to evaluate the most applicable method [10,11].

Numerous disinfectants are used regularly such as sodium hypochlorite, chlorhexidine, alcohol, glutaraldehyde, and hydrogen peroxide [12]. Since no sole disinfectant can be designated as a universal disinfectant for all impression materials, it is fundamental to select an ideal disinfectant agent with superior antimicrobial activity that does not disturb the recorded features, such as surface characteristics or dimensional stability of an impression materials [13,14].

Additionally, many combinations between impression materials and disinfectant could occur by knowing that a large range of branded impression materials (reversible and irreversible hydrocolloids, polyethers, polysulphides, and silicones) and gypsum-based casts existed in the marketplace. A disinfectant possesses a dual purpose: it needs to be an effective antimicrobial agent but produce no adverse effect on the dimensional accuracy of the impression material and resultant gypsum model. The latter is of significance in an attempt to deliver a functional and well-fitting finished appliance. Disagreement happens in the literature as to whether the disinfection procedure produces degradation or distortion of impressions [15–17].

The reaction of some specific brands of gypsum products and impression materials to disinfection process is diverse, advising a deficiency of compatibility between a given material and protocol. Hence, individual analysis of impression materials is needed to define the effectiveness of a specific disinfection method in different areas [18].

Accordingly, the aim of this study was to systemically review the literature of the existing disinfection procedures on the bacterial colonization of dental impression materials. The null hypothesis to be tested was that the use of disinfectant agents will not reduce the colony-forming units per milliliter (CFU/mL) adhered to the surface of impression materials used in dentistry.

2. Materials and Methods

This systematic review and meta-analysis was reported following the guidelines of the Preferred Reporting Items for Systematic Reviews and Meta-Analyses (PRISMA statement) [19]. The registration protocol was carried out in the Open Science Framework with the registration number 0000-0002-2759-8984. The following PICOS strategy was used: population, impression materials; intervention, use of disinfection materials; control, rinsing with tap water; outcome: antimicrobial activity; and type of study, in vitro studies. The research question was as follows: Does the use of disinfection procedures for impression materials in dental practice reduce the microbial count?

2.1. Search Strategy

The literature search was performed by two independent reviewers (E.C.R. and R.B.) up to April 22, 2021. The following databases were screened: PubMed (MEDLINE), Web of Science, Scopus, EMBASE, and SciELO. The search strategy was performed according to the keywords defined in Table 1. All studies were imported into Rayyan QCRI platform [20].

Table 1. Keywords used in search strategy.

Search Strategy	
#1	Dental models OR Dental impressions OR Irreversible hydrocolloid OR Alginate impressions OR Silicone impression OR Primary impression OR Polyvinyl siloxane
# 2	Disinfection OR Sodium hypochlorite OR Disinfection techniques OR Sterilization OR Chemical disinfection OR Disinfection protocol OR Immersion disinfection OR Cross contamination OR Ultraviolet disinfection OR Microbial activity OR Disinfectant solutions OR Autoclave OR Disinfectant agents
#3	#1 and #2

2.2. Eligibility Criteria

The title and abstract of each identified article were reviewed by two independent reviewers (E.C.R. and R.B.) to determine if the article should be considered for full-text review according to the following eligibility criteria: (1) in vitro studies reporting the antibacterial activity of disinfectant solutions in dental impression materials; (2) included mean and standard deviation (SD) in CFU/mL; (3) included a control group where tap water was used; and (4) published in the English language. Case reports, case series, pilot studies, expert opinions, conference abstracts, and reviews were excluded. In the case of disagreements at the time of the selection of the studies for the full-text review, they were resolved by discussion and consensus by a third reviewer (C.E.C.-S).

2.3. Data Extraction

The Microsoft Office Excel 2019 program (Microsoft Corporation, Redmond, Washington, DC, USA) was used to extract the data of interest from the included manuscripts. These were placed on a standardized form. Two reviewers (L.H. and R.B.), who received training in this software, performed the analysis. The data recovered from each manuscript were author, year, impression material evaluated, disinfection agents used, type of microorganism evaluated, main outcome, and main results.

2.4. Quality Assessment

The risk of bias of the selected articles was assessed by two reviewers (R.B. and E.C.R.) according to the parameters of the previous systematic review [21]. The risk of bias of each article was evaluated according to the description of the following parameters: specimen randomization, single-operator protocol implementation, blinding of the operator, the presence of a control group, complete outcome data, and description of the sample size calculation. If the authors reported the parameter, the study received a "YES" for that specific parameter. In case of missing information, the parameter received a "NO". The risk of bias was classified according to the sum of "YES" answers received: 1 to 2 indicated a high bias, 3 to 4 indicated a medium risk of bias, and 5 to 6 indicated a low risk of bias.

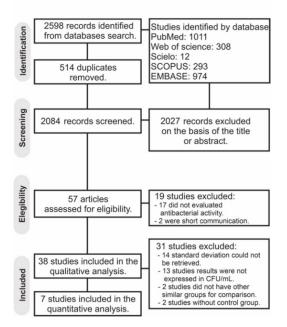
2.5. Statistical Analysis

The meta-analyses were performed using Review Manager Software version 5.1 (The Nordic Cochrane Centre, The Cochrane Collaboration, Copenhagen, Denmark). The analyses were carried out using a random-effect model, and pooled-effect estimates were obtained by comparing the standardized mean difference between CFU/mL values obtained when a disinfection agent was used; against a control group when tap water was

used. The standardized mean difference was performed since this statistic in meta-analysis is used when all the studies assess the same outcome but measure it in a variety of ways; for this to be appropriate, it must be assumed that between-study variation reflects only differences in measurement scales, such as the different scientific notation used among the studies included. Additionally, for comparison purposes, when a value of 0 was found in the data, this was replaced with "0.1" with a SD of "0.01" for the statistical analysis. The comparisons were made considering the type of impression material and the type of disinfection agent used. A *p*-value < 0.05 was considered statistically significant. Statistical heterogeneity of the treatment effect among studies was assessed using the Cochran Q test and the inconsistency I^2 test.

3. Results

The search resulted in the retrieval of 2598 records (Figure 1). After removal of duplicates, 2084 articles were screened, and 2027 were excluded based on the title or abstract. A total of 57 full-text articles were assessed for eligibility. Of these, nineteen were not considered for the qualitative analysis: seventeen did not evaluate the antibacterial activity and two were short communications, leaving thirty-eight studies for the qualitative analysis; from these, thirty-one were excluded from the quantitative analysis: in fourteen studies, the SD could not be retrieved, and in another thirteen studies, the results were not expressed in CFU/mL, two studies did not have any control group, and two studies did not have enough comparison groups. Finally, seven studies were considered for the meta-analysis.



PRISMA FLOW DIAGRAM

Figure 1. Search flowchart according to the PRISMA Statement.

The characteristics of the studies included in this systematic review are summarized in Table 2. Several disinfection agents were identified for the present review, including chlorhexidine, alcohol, sodium hypochlorite, glutaraldehyde, and hydrogen peroxide. Most of the studies included in this review evaluated the antibacterial activity to alginate and polyvinyl siloxane impressions, only two studies evaluated the effect of disinfection on polyether, while only one tested on condensation silicone. Utmost of the studies reported the effect of disinfection agents on CFU/mL, while a few reported inhibition halos.

Study	Impression Material	Disinfection Agent	Type of Microorganism	Main Outcome	Main Results	
Ahmed 2020 [22]	Alginate	Chlorhexidine Desident CaviCide Alcohol	Bacteria Fungi	Colony-forming units	Disinfectants killed the bacteria completely.	
Al-Enazi 2016 [23]	Polyvinyl siloxane Alginate	Sodium hypochlorite Glutaraldehyde	Streptococcus diphtheroid Neisseria	Colony-forming units	Use of 1% sodium hypochlorite yielded better results than did 2% glutaraldehyde.	
Al-Jabrah 2007 [24]	Alginate Polyether Polyvinyl siloxane	Dimenol Perform-ID [®] MD520 [®] Haz-tabs [®]	Does not specify	Colony-forming units	All four disinfectant solutions tested produced effective disinfection of the impression materials investigated.	
Alwahab 2012 [25]	Alginate	Chlorhexidine digluconate	Pseudomonas aeruginosa Staphylococcus aureus Candida albicans	Inhibition halos	The least antibacterial activity of chlorhexidine digluconate was observed against <i>Pseudomonas aeruginosa.</i>	
Azevedo 2019 [26]	Polyvinyl siloxane	Hydrogen peroxide MD520 [®] Sodium hypochlorite	Does not specify	Colony-forming units	All disinfectants tested showed high antimicrobial efficiency.	
Bal 2007 [27]	Polyvinyl siloxane Polyether	Sodium hypochlorite Gludex spray Mikrozid spray	Staphylococcus aureus Enterococcus faecalis	Colony-forming units	The disinfectant spray was less effective than sodium hypochlorite or Gludex.	
Benakatti 2017 [28]	Alginate	Chlorhexidine Gluconate solution	Staphylococcus aureus	Inhibition halos	This disinfection method was effective in the elimination of <i>S. aureus</i> .	
Beyerle 1994 [29]	Alginate	Sodium Hypochlorite	Bacillis subtilis Mycobacleriuiii bovis	Colony-forming units	One-minute exposure resulted in very inconsistent killing in all instances.	
Brauner 1990 [30]	Alginate	Blueprint asept [®]	Streptococcus mutans Streptococcus sanguis Streptococcus aureus Streptococcus pyogenes Staphylococcus aureus Actinomyces odontolyticus Escherichia coli Klebsiella pneumoniae Proteus mirabilis Enterobacter aerogenes Pseudomonas aeruginosa	Inhibition halos	Due to its bactericidal effect, Blueprint asept [®] can be recommended.	
Bustos 2010 [31]	Alginate Condensation silicone Alginate	Sodium Hypochlorite Glutaraldehyde	Gram (+) and (-) coccus and Gram (-) bacillus Candida	Colony-forming units	Alginate and silicone impressions can successfully be disinfected if they are immersed in either 0.5% NaOCl solution or 2% glutaraldehyde for 5 min.	
Choudhury 2018 [32]	Alginate	Sodium Hypochlorite Epimax®	Staphylococcus aureus Candida albicans Pseudomonas aeruginosa	Colony-forming units	Both Epimax and 0.525% sodium hypochlorite can disinfect the alginate impression material against <i>Candida albicans, Pseudomonas aeruginosa,</i> and <i>Staphylococcus aureus.</i>	

 Table 2. Characteristics of the included studies.

Table 2. Cont.

Study	Impression Material	Disinfection Agent	Type of Microorganism	Main Outcome	Main Results	
Cserna 1994 [33]	Cserna 1994 [33] Alginate		Lactobacillus Streptococcus mutans	Inhibition halos	Antimicrobial alginates are more effective than nonantimicrobial alginates in reducing the surface growth of the oral bacteria <i>Lactobacillus</i> and <i>Streptococcus mutans.</i>	
Cubas 2014 [34]	Alginate	Chlorhexidine	Streptococci Candida	Colony-forming units	Chlorhexidine as a water substitute during impression taking offers decreased microbial contamination with no negative alterations of the resulting casts, thus providing an easy method for controlling cross-infection.	
Demajo 2016 [35]	Alginate Polyvinyl siloxane	MD 520 [®] Minuten [®]	Does not specify	Colony-forming units	Glutaraldehyde is more effective than alcohol-based chemical disinfectants.	
Doddamani 2011 [36]	Alginate	Povidone Iodine Sodium Hypochlorite Glutaraldehyde Distilled Water	Staphylococcus aureus Bacillus subtilis Streptococcus viridans	Colony-forming units	Disinfectants work equally well on an irreversible hydrocolloid impression material.	
Estafanous 2012 [37]	Polyvinyl siloxane Polyether	EcoTru [EnviroSystems] ProSpray [Certol] Sodium hypochlorite	Pseudomonas aeruginosa Salmonella choleraesius Staphylococcus aureus	Colony-forming units	Disinfectants investigated in this study will effectively disinfect Polyvinyl siloxane and polyether elastomeric impression materials.	
Flanagan 1998 [38]	Alginate	Single quaternary ammonium compound Chlorhexidine Dual quaternary ammonium compound	Gram-positive cocci Gram-negative bacilli yeast	Colony-forming units	The alginate with chlorhexidine killed all the gram-negative bacilli and the majority (95–99%) of the gram-positive cocci and yeast.	
Gerhardt 1991 [39]	Alginate	Sodium hypochlorite	Staphylococcus aureus Pseudomonas aeruginosa Bacillus subtitis	Inhibition halos	The results indicated that chlorine disinfecting solutions of sufficient concentration can be retained for periods up to 1 week and still maintain their effectiveness.	
Ginjupalli 2016 [40]	Alginate	Silver nanoparticles	E. coli S. aureus C. albicans	Inhibition halos	The particles imparted significant antimicrobial activity to the alginate impression materials tested.	
Goel, 2014 [41]	Alginate	Sodium hypochlorite Microwave irradiation	Staphylococcus aureus Pseudomonas aeruginosa	Colony-forming units	The results suggested that the microwave irradiated Kala stone casts proved to be a better disinfection method when compared with 0.07% sodium hypochlorite chemically disinfected incorporated cast.	
Hiramine 2021 [42]	Alginate	Sodium dichloroisocyanurate NaClO	Streptococcus mutans Escherichia coli Staphylococcus aureus Candida albicans Dental plaque bacteria	Colony-forming units	The number of oral bacteria adhering to the surfaces of impressions markedly decreased following a 10 min immersion in the 1000 ppm sodium dichloroisocyanurate solution.	

Table 2. Cont.

Study	Impression Material	Disinfection Agent	Type of Microorganism	Main Outcome	Main Results
Ishida 1991 [43]	Ishida 1991 [43] Alginate Condensation silicone		Candida albicans C. glabrota C. tropicalis C. parupsilosis C. krusei C. guilliermondi	Colony-forming units	UV light is effective in disinfecting impression materials that are contaminated with candida organisms.
Ismail 2016 [44]	Alginate	Povidone iodine powder	Streptococcus mutans and Staphylococcus aureus	Inhibition halos	Modified alginate impression material with 15 weight % povidone-iodine powered gives the material self-disinfected properties
Ivanovski 1995 [45]	Alginate	Sterile Water Chlorhexidine Glutaraldehyde Povidone-iodine Sodium hypochlorite with sodium chloride	Escherichia coli Staphylococcus aureus Enterobacter cloacae Pseudomonas aeruginosa Klebsiella pneumoniae Actinobacter calcoaceticus Bacillus subtilis Mycobacterium phlei Candida albicans.	Colony-forming units	When glutaraldehyde was used, all the microorganisms tested were killed after 1 h. Chlorhexidine was ineffective against most microorganisms.
Jennings 1991 [3]	Polysulfide rubber Alginate Polyvinyl siloxane	Chlorhexidine gluconate	C albicans P. aeruginosa	Colony-forming units	Chlorhexidine gluconate (0.2%) was found to be less effective than either glutaraldehyde (2%) or sodium hypochlorite (0.0125%).
Jeyapalan 2018 [46]	Polyvinyl siloxane	Electrolyzed oxidizing water Glutaraldehyde Sodium hypochlorite	Streptococci Staphylococci Pseudomonas Candida Proteus Klebsiella E. coli	Colony-forming units	All three chemical disinfectants employed in this study showed acceptable mean log reduction values and kill rate % for antimicrobial efficacy.
Mathew 2017 [47]	Polyvinyl siloxane	Radio frequency glow discharge	Gram-negative bacilli Gram-positive cocci Escherichia coli Staphylococcus aureus	Inhibition halos	Ratio glow discharge is a very rapid and handy device, which can disinfect saliva contaminated elastomeric impression material surfaces.
McNeill 1992 [48]	Alginate	Glutaraldehyde Hypochlorite solution chlorine Hygojet system	Streptococcus sanguis poliovirus	Colony-forming units	Washing the impression for 15 s followed by immersion in 2% glutaraldehyde for 20.0 min or a hypochlorite solution for 7.5 min effectively disinfected the impression.
Moura 2010 [49]	Alginate	Sodium hypochlorite	Does not specify	Colony-forming units	5.25% sodium hypochlorite can be used with antimicrobial efficacy, using the humidifier box and nebulizer box methods, and 2.5% sodium hypochlorite was not effective in the nebulizer box method.
Nascimento 2015 [50]	Alginate	Sodium hypochlorite Chlorhexidine	S. mutans S. sanguis E. faecalis	Colony-forming units	4% chlorhexidine was the most suitable disinfectant.

Table 2. Cont.

Study	Impression Material	Disinfection Agent	Type of Microorganism	Main Outcome	Main Results	
Rweyendela 2009 [13]	Rweyendela 2009 [13] Alginate		Candida albicans Staphylococcus aureus Pseudomonas aeruginosa Streptococcus mutans Bacillus subtilis spores	Colony-forming units	The compounds effectively disinfected the alginate the presence of organic material, but Aseptrol did after an immersion time of only 1.5 min.	
Samra 2010 [51]	Alginate Polyvinyl siloxane	Glutaraldehyde Sodium hypochlorite Ultraviolet chamber	Streptococcus viridans Diphtheroids Streptococcus pneumoniae Candida albicans Pseudomonas aeruginosa Staphylococcus albus	Colony-forming units	All the disinfection systems were effective in reducing the microbial load with the ultraviolet chamber as the most effective.	
Savabi 2018 [52]	Alginate	Ozonated water	Pseudomonas aeruginosa Staphylococcus aureus Candida albicans	Colony-forming units	Immersion of alginate impression material in ozonated water for 10 min will not lead to complete disinfection but decreases the microorganisms to a level that can prevent infection transmission.	
Schwartz 1996 [53]	Alginate	Sodium hypochlorite	Staphylococcus aureus Salmonella choleraesuis Pseudomonas aeruginosa Mycobacterium bovis Bacillus subtilis	Colony-forming units	It was found that a 10 min immersion in solutions reduced to pH 7 to 11 consistently produced a 4-log (99.99%) or greater reduction in viable organisms.	
Singla 2018 [54]	Polyether	Disinfectant spray Deconex	Escherichia coli Staphylococcus aureus Pseudomonas aeruginosa Candida albicans	Colony-forming units	The disinfectant used was effective.	
Tanaka 1994 [55]	Alginate	Chlorhexidine	Streptococcus mitis Actinomyces naeslundii Staphylococcus aureus Veillonella parvula Porphyromonas gingivalis Candida albicans	Colony-forming units	The use of an impression material supplemented with 1% chlorhexidine, such as Coe Hydrophilic Gel, may protect clinical staff and dental technicians from at least some bacterial infections associated with impression procedures.	
Trivedi 2019 [56]	Alginate	Aloe Vera	Staphylococcus aureus Pseudomonas aeruginosa Candida albicans	Colony-forming units	The effectiveness of aloe vera as a disinfectant was demonstrated.	
Zhang 2017 [57]	Elastomer impression material	Glutaraldehyde Ultraviolet radiation	Human Immunodeficiency Virus Hepatitis B virus	Colony-forming units	Combined use of ultraviolet radiation and 2% glutaraldehyde immersion can eliminate both <i>Human</i> <i>Immunodeficiency Virus</i> and <i>Hepatitis B virus</i> .	

Figures 2–4 show the result from the meta-analyses. With regards to alginate, the use of disinfection agents such as chlorhexidine, alcohol, glutaraldehyde, and sodium hypochlorite significantly reduced the CFU/mL count (p < 0.001). It is worth mentioning that a high heterogenicity was observed ($I^2 = 74\%$) (Figure 2).

	C	ontrol		Antimi	crobial a	aent		Std. Mean Difference	Std. Mean Difference
Study or Subgroup	Mean	SD	Total	Mean	SD	-	Weight	IV, Random, 95% CI	IV, Random, 95% Cl
25.1.1 Chlorhexidine	•								<u>í</u>
Ahmed 2020	930	57	5	0.1	0.01	5	0.2%	20.84 [9.04, 32.64]	•
Cubas 2014	2.9	2.1	10	0.06	0.03	10	8.5%	1.83 [0.75, 2.91]	— •
Cubas 2014	2.9	3.6	10	0.75	1.8	10	9.3%	0.72 [-0.19, 1.64]	+
Cubas 2014	1	0.8	10	0.05	0.05	10	8.7%	1.61 [0.57, 2.64]	— —
Subtotal (95% CI)			35			35	26.7%	1.65 [0.22, 3.08]	
Heterogeneity: Tau ² :	= 1.35; CI	hi² = 1	3.29, df	= 3 (P =	0.004); P	'= 77%			
Test for overall effect	: Z = 2.26	(P = 0	1.02)						
25.1.2 Alcohol									
Ahmed 2020	930	57	5	0.1	0.01	5	0.2%	20.84 [9.04, 32.64]	· · ·
Ahmed 2020	930	57	5	0.1	0.01	5	0.2%	20.84 [9.04, 32.64]	· · ·
Al-Jabrah 2007	2.35	2.17	15	0.1	0.01	15	9.7%	1.43 [0.61, 2.24]	
Subtotal (95% CI)			25			25	10.2%	13.51 [-2.18, 29.21]	-
Heterogeneity: Tau ² =				df = 2 (F	< 0.000	1); I² = 9	0%		
Test for overall effect	. Z = 1.69	(P=0	1.09)						
25.1.3 Glutaraldehyd	le								
Al-Enazi 2016	4.2	2.5	60	1.2	1	60	11.3%	1.57 [1.16, 1.98]	
Al-Enazi 2016	3.8	2.5	60	1	1.5	60	11.4%	1.35 [0.95, 1.75]	
Al-Jabrah 2007	2.35	2.17	15	0.1	0.01	15	9.7%	1.43 [0.61, 2.24]	→
Demajo 2016	6.24	0.24	7	0.1	0.01	7	0.2%	33.84 [19.02, 48.67]	•
Subtotal (95% CI)			142			142	32.6%	1.56 [0.69, 2.42]	-
Heterogeneity: Tau² =				= 3 (P =	0.0003);	l² = 849	6		
Test for overall effect	: Z = 3.53	(P = 0	1.0004)						
05 4 4 0 - 15									
25.1.4 Sodium hypod									
Al-Enazi 2016	3.8	2.5	60	0.3	0.5	60	11.2%	1.93 [1.49, 2.36]	
Al-Enazi 2016 Moura 2010	4.2 45.35	2.5	60 5	0.5 11.45	0.5 12.49	60 5	11.2% 4.7%	2.04 [1.60, 2.48]	
Moura 2010 Moura 2010	40.30		5	0.2	0.52	5	4.7%	3.04 [0.93, 5.16] 8.21 [3.43, 13.00]	
Moura 2010 Moura 2010	40.75	7.9	5	1.05	1.39	5	1.3%	7.17 [2.95, 11.40]	
Moura 2010 Moura 2010	48.75		5	0.05	0.22	5	0.4%	16.74 [7.23, 26.24]	
Subtotal (95% CI)	40.7 J	3.71	140	0.05	0.22	140	30.5%	2.91 [1.83, 4.00]	
Heterogeneity: Tau ² :	- n 80 [.] CI	hi≊ – 2		- 5 (P -	0.0005)-			2101[1100]	
Test for overall effect					5.5003),	07	•		
Total (95% CI)			342			343	100.0%	2.05 [1.47, 2.63]	
	0.74.00			- 48.00				2.03[1.47, 2.03]	
Heterogeneity: Tau ² =					× 0.0000	1), 14 = 8	170		-4 -2 0 2 4
Test for overall effect Test for subgroup dif					- 0.11\	Z - 60 4	ov.		Antimicrobial agent Control
rest for subgroup dif	ierences	. Unite	= 0.01,	ui = 3 (P	= 0.11), I	-= 50.1	70		



	C	ontrol		Antimicrobial agent		:	Std. Mean Difference	Std. Mean Difference		
Study or Subgroup	Mean	SD	Total	Mean	SD	Total	Weight	IV, Random, 95% CI	IV, Random, 95% CI	
25.2.1 Glutaraldehyd	e									_
Al-Enazi 2016	3.7	2	60	0.9	1	60	15.3%	1.76 [1.34, 2.18]	-	
Al-Enazi 2016	3.7	2	60	0.8	1	60	15.3%	1.82 [1.39, 2.25]	-	
Al-Jabrah 2007	8.5	1.3	15	0.1	0.01	15	5.8%	8.89 [6.37, 11.41]	,	
Demajo 2016	2	0.4	7	0.1	0.01	7	4.7%	6.29 [3.35, 9.23]	,	,
Jeyapalan 2016	6.6	1	10	2.6	2	10	11.4%	2.42 [1.21, 3.64]		
Subtotal (95% CI)			152			152	52.5%	3.30 [2.10, 4.51]	-	
Heterogeneity: Tau ² =	1.34; Ch	ni² = 39	.25, df	= 4 (P < 0	0.00001)	; I ² = 909	%			
Test for overall effect:	Z = 5.37	(P < 0	.00001)						
25.2.2 Sodium hypot	chlorite									
Al-Enazi 2016	3.6	3.2	60	0.3	0.2	60	15.4%	1.45 [1.04, 1.85]		
Al-Enazi 2016	3.6	3.2	60	0.1	0.2	60	15.4%	1.53 [1.13, 1.94]	-	
Jeyapalan 2016	6.56	1	10	2.3	2	10	11.2%	2.58 [1.33, 3.83]		
Subtotal (95% CI)			130			130	41.9%	1.58 [1.21, 1.94]	•	
Heterogeneity: Tau ² =	0.03; Ch	ni ² = 2.8	87, df =	2 (P = 0.	24); I ² =	30%				
Test for overall effect:	Z = 8.47	(P < 0	.00001)						
25.2.3 Alcohol										
Al-Jabrah 2007	8.5	1.25	15	0.1	0.01	15	5.5%	9.25 [6.64, 11.86]		·
Subtotal (95% CI)			15			15	5.5%	9.25 [6.64, 11.86]	,	<u></u>
Heterogeneity: Not ap										
Test for overall effect:	Z = 6.94	(P < 0	.00001)						
Total (95% CI)			297			207	100.0%	2.90 [2.14, 3.66]		
. ,								2.90 [2.14, 3.00]		
Heterogeneity: Tau ² =					0.00001)	; 1* = 90%	70		-4 -2 0 2 4	
Test for overall effect:		·		,		A) 12 - A			Antimicrobial agent Control	
Test for subgroup diffe	erences:	$Chi^2 =$	38.61,	dt = 2 (P	< 0.0000	(1), 1 ² = §	94.8%			

Figure 3. Forest plot of the analysis of CFU/mL count in polyvinyl siloxane after disinfection.

	Control		Antimicrobial agent			:	Std. Mean Difference	Std. Mean Difference
Study or Subgroup Mean	SD '	Total	Mean	SD	Total	Weight	IV, Random, 95% CI	IV, Random, 95% CI
25.3.1 Alcohol								
Al-Jabrah 2007 1.8	5 1.42	15	0.1	0.01	15	50.0%	1.70 [0.84, 2.55]	
Subtotal (95% CI)		15			15	50.0%	1.70 [0.84, 2.55]	
Heterogeneity: Not applicable	9							
Test for overall effect: Z = 3.9	1 (P < 0.	0001)						
25.3.2 Glutarladehyde								
Al-Jabrah 2007 1.8	5 1.42	15	0.1	0.01	15	50.0%	1.70 [0.84, 2.55]	∎
Subtotal (95% CI)		15			15	50.0%	1.70 [0.84, 2.55]	
Heterogeneity: Not applicable	è							
Test for overall effect: Z = 3.9	1 (P < 0.	0001)						
		-						
Total (95% CI)		30			30	100.0%	1.70 [1.09, 2.30]	•
Heterogeneity: Tau ² = 0.00; 0	Chi² = 0.0	0, df = 1	(P = 1.0	0); I ² = (0%		-	
Test for overall effect: Z = 5.5	2 (P < 0.	00001)						-4 -2 0 2 4 Antimicrobial agent Control
Test for subgroup differences	Chi2 = 0	00 df-	- 1 /D - 1	1 001 12	- 0%			Anumicrobial agent Control

Test for overall effect: Z = 5.52 (P < 0.00001) Test for subgroup differences: $Chi^2 = 0.00$, df = 1 (P = 1.00), I² = 0%

Figure 4. Forest plot of the analysis of CFU/mL count in polyether after disinfection.

Figure 3 shows the effect of different disinfection agents on polyvinyl siloxane material. According to the meta-analysis, all the disinfection agents tested significantly reduced the CFU/mL count (p < 0.001). Again, a high heterogenicity was observed in the comparisons (90%).

Finally, Figure 4 shows the effect of different disinfection agents on polyether impression material. According to the meta-analysis, both alcohol and glutaraldehyde significantly reduced the CFU/mL count (p < 0.001). As only one study was included in this analysis, a 0% heterogenicity was found.

The risk of bias analysis was shown that most of the studies were categorized with high and medium risk of bias (Table 3). Utmost of the manuscripts examined failed to report the single operator, operator blinded, and sample size calculation factors.

Study	Specimen Randomization	Single Operator	Operator Blinded	Control Group	Complete Outcome Data	Sample Size Calculation	Risk of Bias
Ahmed 2020 [22]	NO	NO	NO	YES	NO	NO	High
Al-Enazi 2016 [23]	YES	NO	NO	YES	YES	NO	Medium
Al-Jabrah 2007 [24]	YES	NO	NO	YES	YES	NO	Medium
Alwahab 2012 [25]	NO	NO	NO	YES	YES	NO	High
Azevedo 2019 [26]	YES	NO	NO	YES	NO	NO	High
Bal 2007 [27]	NO	NO	NO	YES	NO	NO	High
Benakatti 2017 [28]	NO	YES	NO	YES	YES	NO	Medium
Beyerle 1994 [29]	NO	NO	NO	YES	NO	NO	High
Brauner 1990 [30]	YES	NO	NO	YES	NO	NO	High
Bustos 2010 [31]	YES	NO	NO	YES	YES	NO	Medium
Choudhury 2018 [32]	NO	NO	NO	YES	NO	NO	High
Cserna 1994 [33]	NO	NO	NO	YES	YES	NO	High
Cubas 2014 [34]	YES	NO	YES	YES	YES	YES	Low
Demajo 2016 [35]	NO	NO	NO	YES	YES	NO	High
Doddamani 2011 [36]	NO	NO	NO	YES	NO	NO	High
Estafanous 2012 [37]	NO	NO	NO	YES	NO	NO	High
Flanagan 1998 [38]	NO	NO	NO	YES	YES	NO	High
Gerhardt 1991 [39]	NO	NO	NO	YES	NO	NO	High
Ginjupalli 2016 [40]	NO	YES	NO	YES	YES	NO	Medium
Goel 2014 [41]	NO	NO	NO	YES	YES	NO	High
Hiramine 2021 [42]	NO	NO	NO	YES	YES	NO	High
Ishida 1991 [43]	NO	NO	NO	YES	YES	NO	High
Ismail 2016 [44]	NO	NO	NO	YES	NO	NO	High
Ivanovski 1995 [45]	NO	NO	NO	YES	YES	NO	High
Jennings 1991 [3]	YES	NO	NO	YES	YES	NO	Medium
Jeyapalan 2018 [46]	YES	NO	NO	YES	YES	NO	Medium
Mathew 2017 [47]	NO	NO	NO	YES	NO	YES	High
McNeill 1992 [48]	NO	NO	NO	YES	NO	NO	High
Moura 2010 [49]	YES	NO	NO	YES	YES	NO	Medium
Nascimento 2015 [50]	NO	NO	NO	YES	YES	NO	High
Rweyendela 2009 [13]	NO	NO	NO	YES	YES	NO	High
Samra 2010 [51]	NO	NO	NO	YES	NO	NO	High
Savabi 2018 [52]	NO	NO	NO	YES	YES	NO	High
Schwartz 1996 [53]	NO	NO	NO	YES	YES	NO	High
Singla 2018 [54]	NO	NO	NO	YES	NO	NO	High
Tanaka 1994 [55]	NO	NO	NO	YES	NO	NO	High
Trivedi 2019 [56]	NO	NO	NO	YES	YES	YES	Medium
Zhang 2017 [57]	YES	NO	NO	YES	NO	NO	High

Table 3. The results of the risk of bias assessment.

4. Discussion

This systematic review and meta-analysis was directed towards testing the effect of disinfection agents on the bacterial colonization of different impression materials. This review focused on the study of the CFU/mL measure, since this is the most common measure used to determine the antibacterial activity. To the best of the authors knowledge, this is the first approach to prove that the application of disinfectant agents is effective to reduce the count of some oral pathogens on the surface of alginate, polyvinyl siloxane, and polyether impression materials and that this procedure can certainly reduce the possibility of cross-contamination. Accordingly, the hypothesis tested in this study was rejected.

Normally, chemical disinfectant agents were generally used in dental exercise because of their easy application. For the alginate materials, the use of disinfection agents such as chlorhexidine, alcohol, glutaraldehyde, and sodium hypochlorite significantly reduced the CFU/mL count (p < 0.001). Irreversible hydrocolloids, the frequent material used in dentistry, tend to absorb both blood and saliva [23]. Thus, research was focused on a solution to inhibit the colonization of microbe on the surface of these materials [11].

Collected data were established on the CFU in a media culture. These were recorded by using a colony counter, and the counts were expressed by a standard technique of estimating microbial colony count known as the CFU count. The bacteriological examination evidently exhibited that the CFU recorded after disinfection were fewer than before disinfection [23], thus making the disinfection process an important issue to solve after taking an impression in the dental world.

It is highlighted in a previous study [58] that the use of tap water on the surface of alginate impression failed to kill *Streptococcus Mutans* and *Lactobacilli*; however, by using chlorhexidine, a positive antimicrobial activity has been shown [59]. This could be possible by the binding between the positive site of chlorhexidine and negative sites of the bacterial cell, which resulted in interference with osmosis and escapes the constituents that lead to cell death [22]. In addition, alcohol was able to kill all the detected bacteria in this study by inactivating the growth of the bacteria on the alginate impression, and this was deemed probable by alkylating the amino and sulf hydral groups of bacterial proteins [60–62]. Further, for the other disinfectants, it was demonstrated that by using 2% glutaraldehyde solution or 1% sodium hypochlorite, gram-positive organisms will be modified by reducing their growth [11]. Indeed, this effect was most noticeable for 1% sodium hypochlorite, as described in this research.

A previous study denoted that after immersion in sterile water for 10 min, for some of impression materials, including alginate impression, the number of microorganisms counted was diminished, though alginate material still retained some of these microorganisms in comparison to other materials [24]. The physical nature of alginate impression could affect the capacity of disinfectants for doing their biocidal activity. In the oral environment, microorganisms might become integrated into the gelling impression material since the presence of oral fluids or saliva [29]. The set-up of these microorganisms in the alginate material restricted the efficacy of the water rinse, and the alginate gel assembly could hinder the penetration of the disinfectant [29,63]. Thus, this idea explained the results of this study as tap water did not reduce the microorganism counts in comparison to the other disinfectant solution. Overall, for alginate impressions, the use of disinfectant agents would be of great interest, and the efficacy of the disinfection ranged between 92% and 99.97% in all the situations [23].

According to the meta-analysis, all the disinfection agents significantly reduced the CFU/mL count (p < 0.001) on polyvinyl siloxane material. Among the numerous available impression materials in prosthodontics, this material was considered the material of choice, due to their fine detail reproduction, excellent physical properties, remarkable dimensional stability, good acceptance by the patient, and elastic recovery feature [64–66]. In addition, these materials were tasteless and odorless [46]. As stated above in alginate impression, using 2% glutaraldehyde solution or 1% sodium hypochlorite could be recommended also for disinfecting the polyvinyl siloxane impression [67,68]. In this manner, it is advisable to

immerse these kinds of impressions in these solutions rather than spraying, as successful finding was observed in many previous studies, without harming the physical properties [15,18,31,69]. Seemingly, by putting the polyvinyl siloxane material in an alcohol-based disinfectant solution for a contact time of 15 min, a media free of microorganisms could be observed [24]. Accordingly, this can support the finding obtained in this study, as any kind of disinfectant solution tested showed promising results with polyvinyl siloxane impression material.

The present analysis noted that both alcohol and glutaraldehyde significantly reduced the CFU/mL count (p < 0.001) of polyether impression material. This could be explained by the fact that the contaminating bacteria could be reduced by 85% when soaking this kind of impression material in sterile water for 15 min; in addition, as found with polyvinyl siloxane impression, the use of alcohol-based solution produced effective disinfection of the polyether impression [24]. With regards to glutaraldehyde, the antimicrobial activity of this compound depends on the duration of dilution and its concentration. This could be elucidated by the fact that the biocidal activity of glutaraldehyde results from alkylation of sulfhydryl, hydroxyl, carboxyl, and amino groups of microorganisms, which alters RNA, DNA, and protein synthesis [70]. This conclusion seems to support the results in this meta-analysis.

From this review, various disinfectant agents were used to show the importance of reducing microorganisms on the surface of the impression materials used in dentistry. The results should be considered with caution since other brands of impression materials were available in the dental market and not included. In addition, there is the opportunity for slight changes in chemistry of these materials, causing significantly different reactions. Additionally, most of the studies included were classified as having high or medium risk of bias, and, therefore, better experimental designs should be conducted in order to obtain a higher degree of evidence. One of the limitations of this review relies on the fact that it is only focusing on the antibacterial efficacy of the application of a disinfectant on the surface of a dental impression; however, other variables should be taken into account, such as the effect of this procedure on the accuracy, precision, and surface quality of the resulting working models, especially when dental impressions are disinfected both in the dental office and in the dental laboratory. This led to controversy as to whether the disinfection process causes degradation or distortion of dental impressions and to what extent. Therefore, studying the effect of these disinfectants on the dimensional stability of the impression materials should be considered in further research. Additionally, viruses could be considered in future investigation by having the required equipment since their manipulation was considered dangerous for some researchers. Moreover, clinical studies were needed since testing of the efficacy of disinfectants from different patients derived impressions was scarce, knowing the differences in oral flora composition of individual person.

5. Conclusions

Based on in vitro studies, disinfection of alginate with chlorhexidine, alcohol, glutaraldehyde, and sodium hypochlorite reduced the CFU/mL count on the surface of alginate impressions. This trend was observed when polyvinyl siloxane impressions were disinfected with glutaraldehyde, sodium hypochlorite, and alcohol and when polyether was immersed in alcohol or glutaraldehyde. Therefore, these substances could be employed to reduce cross-contamination in the dental office.

Supplementary Materials: The following supporting information can be downloaded at: https://www.mdpi.com/article/10.3390/bioengineering9030123/s1, Table S1: Quantitative data extracted from studies included in the meta-analysis.

Author Contributions: Conceptualization, L.H., R.B. and C.E.C.-S.; methodology, L.H., R.B., N.K. and C.E.C.-S.; software, L.H., R.B. and C.E.C.-S.; validation, D.M., A.E., N.K., V.T. and Y.H.; formal analysis, L.H., R.B. and C.E.C.-S.; investigation, L.H., R.B., N.K., M.L.-S., E.C.-R., R.M., D.M. and

S.M.; resources, E.C.-R., Y.H., N.K., M.L.-S. and D.M.; data curation, L.H., R.B. and C.E.C.-S.; writing original draft preparation, L.H., R.B. and C.E.C.-S.; writing—review and editing, M.L.-S., L.H., N.K. and Y.H.; visualization, N.K., E.C.-R., S.M., A.E., V.T., L.H., R.B. and R.M.; supervision, L.H.; project administration, L.H. All authors have read and agreed to the published version of the manuscript.

Funding: This research received no external funding.

Institutional Review Board Statement: Not applicable.

Informed Consent Statement: Not applicable.

Data Availability Statement: The data presented in this study are available in the article.

Acknowledgments: Authors Louis Hardan and Rim Bourgi would like to acknowledge the Saint-Joseph University of Beirut, Lebanon. Furthermore, the referees would also recognize the University of Hidalgo State, Mexico, the Medical University of Lodz, and the University of Strasbourg for accompanying this research.

Conflicts of Interest: The authors declare no conflict of interest.

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