# Changes in blood ion levels after removal of metal-on-metal hip replacements

16 patients followed for 0–12 months

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Submitted 13-02-07. Accepted 13-11-22

**Background and purpose** — In patients with metal-on-metal (MoM) hip prostheses, pain and joint effusions may be associated with elevated blood levels of cobalt and chromium ions. Since little is known about the kinetics of metal ion clearance from the body and the rate of resolution of elevated blood ion levels, we examined the time course of cobalt and chromium ion levels after revision of MoM hip replacements.

Patients and methods — We included 16 patients (13 female) who underwent revision of a painful MoM hip (large diameter, modern bearing) without fracture or infection, and who had a minimum of 4 blood metal ion measurements over an average period of 6.1 (0-12) months after revision.

**Results** — Average blood ion concentrations at the time of revision were 22 ppb for chromium and 43 ppb for cobalt. The change in ion levels after revision surgery varied extensively between patients. In many cases, over the second and third months after revision surgery ion levels decreased to 50% of the values measured at revision. Decay of chromium levels occurred more slowly than decay of cobalt levels, with a 9% lag in return to normal levels. The rate of decay of both metals followed second-order (exponential) kinetics more closely than first-order (linear) kinetics.

**Interpretation** — The elimination of cobalt and chromium from the blood of patients who have undergone revision of painful MoM hip arthroplasties follows an exponential decay curve with a half-life of approximately 50 days. Elevated blood levels of cobalt and chromium ions can persist for at least 1 year after revision, especially in patients with high levels of exposure.

After a dramatic rise in initial popularity, the use of metalon-metal (MoM) hip arthroplasties has declined precipitously, both in hip resurfacings and conventional hip replacement, due to an alarming incidence of adverse inflammatory reactions (van der Weegen et al. 2011, NJR 2012).

There are now serious concerns about potential adverse biological effects, both local and systemic, arising from wear debris generated by MoM articulations (Matthies et al. 2011). The nano-scale particles generated through wear of MoM bearings can enter the reticulo-endothelial system and cross over into the circulation as early as 5 days after implantation (Daniel et al. 2007). The specific surface area (surface area to mass ratio) of these particles makes them susceptible to corrosion in vivo (Hart et al. 2010), leading to elevated levels of cobalt and chromium ions in the blood, usually ranging from 5 to 10 times normal values (Jacobs et al. 1996, Brodner et al. 1997, Skipor et al. 2002, Dunstan et al. 2005, Daniel et al. 2009, Hart et al. 2009, van der Weegen et al. 2011). Possible complications from long-term elevated metal ion levels include immune reactions (Pandit et al. 2008), necrosis (Campbell et al. 2010), toxicity (Keegan et al. 2007, Tower et al. 2010, Corradi et al. 2011), chromosomal aberrations (Ladon et al. 2004), and carcinogenicity (Case et al. 1994). However, the most common short-term complication is joint pain associated with inflammatory reactions (Milosev et al. 2005). In many cases where there is pain with soft-tissue masses, often in association with elevated levels of cobalt and chromium ions, removal of the implanted components is necessary. This form of failure is more common in female patients than in male patients, and it has been reported in 1-20% of cases at 5 years (Schmidt et al. 1996) depending on the design of the prosthesis.

Although revision is performed in the hope that the elevation of ion levels and symptoms will resolve, little is known about the kinetics of storage and turnover of these ions in the body. We examined the kinetics of cobalt and chromium ion decay after revision of MoM hip replacements. Our principal goals were to characterize the decay curves of cobalt and chromium ions after revision procedures and to determine when blood concentrations return to levels below the MHRA action level of 7 ppb, in order to determine exposure risk (MHRA 2010). We hypothesized that: (1) removal of MoM components results in a rapid drop in the level of metal ions in the blood followed by a slow steady-state decline; (2) chromium concentrations will decrease more slowly than cobalt concentrations; (3) the total body exposure to ions is orders of magnitude higher with poorly functioning or loose implants than with well-functioning components.

## Patients and methods

With institutional review board approval, 16 patients (13 females) were enrolled in this study following revision hip surgeries performed during the period October 1999 to December 2008. Each patient underwent primary MoM total hip arthroplasty (13 resurfacing and 3 modular) at an average of 4.9 (2.1-10.9) years before revision. For inclusion in the study, patients were required to have had a painful MoM hip prosthesis without proven infection or fracture. The average age of the patients in the study group was 62 (47-74) years. The average head size was 47 (36–54) mm and the average cup size was 53 (48-60) mm. This cohort was a subgroup of a larger cohort (39 hips) of revision MoM hip arthroplasties that has been reported previously (Liddle et al. 2013). The patients in this cohort were those from the first cohort who were prepared to undergo the repeated blood ion measurements required for the present study.

Whole-blood samples were collected in trace-element blood tubes on successive days of each patient's hospitalization for the revision procedure, and then at follow-up visits performed at 6 weeks, 3 and 6 months, and 1 year after revision surgery. Blood samples were also collected for trace-element analysis if additional blood was drawn for other reasons during the follow-up period. 106 blood samples (on average 6.6 (4-13) samples per patient) were collected during the course of the study over an average follow-up period of 5.8 (0.1-20.5) months. Each sample was taken, stored, and processed in a standardized fashion, which has been described previously (Hart et al. 2011b). The concentrations of cobalt and chromium ions in each blood sample were measured using standard operating procedures using dynamic reaction-cell inductively coupled plasma mass spectrometry (De Smet et al. 2008, Heisel et al. 2008, Sampson et al. 2012).

In 2010, the Medicines and Healthcare Products Regulatory Agency in the UK issued a safety alert which recommended on-going surveillance of patients with blood concentrations of cobalt or chromium, or both, of 7 ppb or greater (MHRA 2010). Based on this recommendation, we divided our study group into 2 subgroups. The first subgroup comprised 12 patients with initial post-revision metal concentrations of 7 ppb or greater; they were followed for an average of 7 months with an average of 7.2 blood measurements during the post-revision surveillance period. The second group, consisting of 4 patients with initial concentrations of less than 7 ppb, was followed for an average of 3 months with an average of 5 blood draws during that period.

The metal ion concentrations of each blood sample were plotted for each patient as a function of the time since revision. To standardize comparisons between different patients, the data on each plot were interpolated to yield concentrations of cobalt and chromium present at 7, 30, and 180 days post-revision. In the case of the high-concentration subgroup, the time required for ion concentrations to drop to 7 ppb was also recorded. Plots were also prepared expressing ion levels as a percentage of the initial values measured at revision and interpolated to calculate the period until blood levels dropped to 50% of the levels at revision. 10 of the cases in the highconcentration subgroup had 4 or more blood draws distributed over more than 50 days after revision. For these cases, plots of ln[ion] against time and 1/[ion] against time were also generated to test for first- and second-order decay kinetics.

The area under the curve for ion concentration vs. time was calculated for the patients with very high blood ion levels who entered steady-state decay well above 7 ppb, in order to compare metal ion load to that in a well-functioning implant. The line of best fit was fitted to each data set, and the integrated value of cumulative exposure (in ppb-years) was calculated and compared to that in a typical patient with a blood ion concentration of 2 ppb. This threshold value of 2 ppb was based on the work of Hart et al. (2009), who measured the ion concentrations of 88 unilateral BHR hips 3–5 years postoperatively.

## Results

16 patients with a mean age of 62 (47–74) years were enrolled into the study (Table 1). 12 of the 16 were resurfacing prostheses, and 9 of the 12 were Birmingham Hip Resurfacings. The other resurfacing prostheses were ASR (3 patients) and XL Magnum (1 patient). The remaining modular cases were Polarstem/BHR, Durom and 36-mm Pinnacle. All patients underwent revision to a non-MoM bearing couple; ceramic on ceramic in 9 of the16 cases and metal on ultra-high molecular weight polyethylene in 7 of the 16. 1 patient (with a stemmed prosthesis) was revised to a modular revision system; the remainder received "primary" cemented (Exeter; Stryker, Newbury, UK) or cementless (Furlong HAC; JRI, Sheffield, UK) femoral stems. In this cohort, there have been no readmissions, reoperations, or significant complications.

MRI findings were available for 9 patients, 4 of whom showed fluid-filled masses. Loose components without gross

Table 1. Patient demographics and	information regarding prostheses
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Patient	Age	Sex	First	Modular	Head	Cup	Inclination	Version	Survival	Revision	orosthesis
no.			prosthesis		size	size			time	Articulation	Cemented
1	47	F	BHR	Ν	42	50	43	39	65	CoC	Ν
2	58	F	BHR	Ν	46	52	42	20	25	CoC	Ν
3	60	F	BHR	Ν	46	52	36	26	68	MoP	Ν
4	61	Μ	BHR	Ν	46	52	43	22	131	MoP	Ν
5	72	F	XL Magnum	N	44	50	38	-5	22	CoC	N
6	73	F	Durom	Y	50	56	55	37	38	CoC	Y
7	74	F	Polarstem/BHR	Y	50	56	37	19	49	MoP <sup>a</sup>	N
8	69	F	ASR	N	43	48	55	27	62	MoP	N
9	63	F	ASR	N	47	54	55	48	29	CoC	Y
10	69	М	BHR	N	54	60	73	41	63	MoP	Y
11	50	F	BHR	Ν	50	56	71	42	31	CoC	Y
12	66	F	BHR	N	42	50	66	34	68	CoC	N
13	64	Μ	BHR	Ν	50	56	57	43	92	CoC	Y
14	61	F	Pinnacle	Y	36	52	68	-3	56	MoP	Ν
15	55	F	BHR	Ν	46	50	64	30	72	MoP	Ν
16	57	F	ASR	Ν	53	60	70	43	45	CoC	Ν

CoC: ceramic on ceramic; MoP: metal on polyethylene.

<sup>a</sup> Revision stem (all other are primary stems)

#### Table 2. Whole-blood concentrations of chromium and cobalt ions

		Ch	romium, ppl	h				Cobalt, ppb			
Case	Initial	7	30	180	Final	Initial	7	30	180	Final	Observation
no.	value	days	days	days	value	value	days	days	days	value	(days postop.)
Lowma	etal ion cas	95									
1	0.78	na	na	na	0.26	0.53	na	na	na	0.47	3
2	1.7	3.0	1.8	na	1.0	1.4	1.3	1.1	na	0.9	35
3	3	3.3	1.7	1.4	1.4	1.4	1.7	1.1	1.2	1.2	183
4	2.9	3.5			1.4	3.8	0.9			0.9	7
		1	na	na	I	3.0	0.9	na	na	0.9	1
	etal ion cas		0.4	- 4	0.4	0.0	<u> </u>	0.5	0.7	0.0	000
5	1.9	2.5	2.4	1.4	0.4	8.8	6.9	6.5	3.7	0.3	363
6	1.5	1.0	na	na	1.0	7.7	6.0	na	na	6.0	7
7	3.1	3.1	2.6	na	0.5	8.9	10.9	9.2	na	1.7	135
8	6.6	5.8	7.7	4.9	4.7	11	7.5	10	5.6	5	196
9	21.3	4.3	6.1	na	5	24	22.1	14	na	5	77
10	13.8	6.7	6.1	2.4	0.7	33	15.4	14	2.1	2.1	340
11	23	19.5	10.8	7.1	6.3	42	32	19	8.4	8.2	224
12	41.9	40	28.8	na	24.3	70	41.9	37	na	7.2	165
13	36.0	23	16.8	18.8	16.1	71	14	11	4.3	3.0	270
14	37.5	na	na	na	32.1	102	na	na	na	89	3
15	53	30	9.2	na	7.8	141	55	38	na	11.9	85
16	101.3	79.5	53	54.1	36.3	156	99.4	61	5	1.2	322
Mean (high ion cases)											
	28	20	14	15	11	56	28	22	4	12	182
95% C		6.2–33	5.1–24	0.0–30	4.3–18	27.8–85	12.3–44	11.6–32	3.3–6.4	-1.6 to 2	

Reported at the time of revision (initial value), at 7, 30, and 180 days after revision, and at the final blood draw (last observation). Data for fixed time points have been interpolated from the nearest available measurements.

osteolysis were present in 2 of 9 patients, osteolysis in 4 patients, synovitis in 5 patients, pseudotumor in 5 patients, and destructive muscle loss in 3 patients.

## Metal ion decay

On the day of revision of the MoM implants, the average blood ion concentrations of chromium and cobalt were 22 ppb and 43 ppb, respectively, for all 16 patients (Table 2). For the 12 patients with initial cobalt and/or chromium levels in excess of 7 ppb, the average whole-blood ion concentration at the time of revision was 28 ppb for chromium (95% CI: 12–45) and 56 ppb for cobalt (95% CI: 27–86). These values are 106 and 157 times higher than established controls for concentrations of chromium and cobalt in serum (Muñiz et al. 2001). In this subgroup, average chromium and cobalt levels fell to 20 ppb and 28 ppb after 7 days, and to 14 ppb and 22 ppb at 30 days,



Figure 1. Variation in blood chromium concentration as a function of time since revision for the 16 patients enrolled in the study.



Figure 3. Change in blood chromium as a percentage of the initial value at revision.

based on data available from 11 and 10 patients (Figures 1 and 2). This corresponds to 69% and 50% of initial levels at 7 days, and 48% and 39% at 30 days (Figures 3 and 4). In the 6 patients with blood ion values at 180 days or more, chromium ions remained at 15 ppb at 180 days postoperatively (56% of the initial value), while cobalt ion levels dropped to an average of 5 ppb (corresponding to 10% of initial levels).

The half-life of chromium decay was 57 days (95% CI: 6-108) (Figures 1 and 3). A temporary increase in chromium levels occurred in 6 patients, 5 of which occurred between postoperative days 53 and 113. 8 of the 16 patients had chromium ion levels in excess of 7 ppb. In 4 of these cases, chromium ion levels had not dropped below 7 ppb by the last blood draw at an average of 190 days after revision. In the other 4 cases, chromium levels dropped to 7 ppb after an average of 12 (1–27) weeks.

Blood cobalt concentration (ppb)



Figure 2. Variation in blood cobalt concentration as a function of time since revision.



Figure 4. Change in blood cobalt as a percentage of the initial value at revision.

In 12 of the 16 patients, cobalt ion concentrations had dropped to less than 50% of the initial values at the time of the final blood draw (Figure 4). In these cases, the half-life of decay was 44 days (95% CI: 11–77). In 8 of the 12 patients with high initial ion levels, cobalt concentrations dropped to the 7 ppb level at an average of 63 days after revision (95% CI: 36–90 days). 1 of the remaining 4 patients had been followed for only 3 days after revision, while in the other 3 cases cobalt levels averaged 14 ppb at 158 days post-revision, corresponding to 16% of the initial values.

When we compared cobalt and chromium levels at each time point (a total of 96 observations), the drop in cobalt ion concentration exceeded the corresponding value for chromium for 71% of observations (Figure 5). Averaged over all time points, the average decrease in cobalt levels was 45%—as opposed to 36% for chromium, a difference of 9%.



Percent reduction in blood cobalt after revision

Figure 5. Plot of chromium concentration vs. cobalt concentration (% of initial value) for each patient and each time point. Solid line and equation correspond to the line of best fit. The red dashed line corresponds to Cr% = Co%.

Table 3. Ion exposure data for 6 patients with high metal ion levels

Patient. no	Years of cobalt exposure	Years of chromium exposure	Days postoperatively
5	5.3	5.4	165
6	4.1	1.4	85
7	9.1	47	707
10	5.5	1.9	82
15	3.4	2.7	224
16	2.7	7.2	270
Median valu	ues 4.7	4.1	195

We plotted  $\ln[c(t)/cO]$  vs. time and 1/[c(t)/cO] vs. time for the subset of 8 patients with multiple blood draws and longer post-revision follow-up in our cohort. A linear relationship between  $\ln[c(t)/cO]$  and time indicates first-order decay kinetics, while a linear relationship between 1/[c(t)/cO] and time is indicative of second-order kinetics. Cobalt decay followed second-order kinetics more closely than first-order kinetics, although the correlation coefficients were similar (average values of r<sup>2</sup> for 8 cases: 0.87 and 0.76, respectively). Chromium decay was less accurately described by either secondorder or first-order kinetics, with both models generating similar correlation coefficients (r<sup>2</sup> = 0.59 and 0.58).

Between postoperative day 0 and a median time of 6.5 months after revision, the blood ion load of these patients was comparable to 4 (1.4–47) years of chromium exposure in a well-functioning implant and 5 (3–9) years of cobalt exposure (Table 3).

### Discussion

Using data collected from the London Implant Retrieval Centre (LIRC), a tertiary center specializing in revision of MoM implants (Hart et al. 2009, 2010, 2011a, Matthies et al. 2011, Sampson et al. 2012), the present study provides new insights to aid in the management of patients after revision of painful MoM prostheses. These patients had multiple sequential blood draws to measure decline in their whole-blood metal ion concentrations. Close surveillance of metal ion decay after revision has not yet become standard in evaluation of problematic implants, but it may be able to guide treatment (Brodner et al. 1997, MacDonald et al. 2004, De Smet et al. 2008, Sampson et al. 2012).

Our study had several shortcomings, including the limited number of patients. Measurement of blood trace metal ion levels requires special preparation and testing compared to common laboratory tests, and as such is very expensive. Additionally, studies of this type in which revision patients must agree to frequent blood sampling often have a high attrition rate, which limits the number of cases analyzed. There was a large degree of variability in our patients, both in their initial metal concentrations and in the pattern of decay. Thus, it was not possible to define a "normal" pattern of decay representative of all patients. This may be due to the numerous differences between patients, both in their physiology and in the wear characteristics of their original implants. It also suggests that a larger sample may be required before a typical response becomes clear. When calculating the total-body metal ion exposure in patients with high metal concentrations, we used 2 ppb as a control. Although this value has been used as a reference level for well-functioning implants, its use may lack the rigorous precision needed for a scientific study (Skipor et al. 2002, Dunstan et al. 2005, Vendittoli et al. 2007, Daniel et al. 2009). Until the widespread acceptance of 2 ppb as "normal", this remains a weakness of the study (MacDonald et al. 2004).

Postoperatively, there was a dramatic decrease in metal ion concentrations in the blood compared to values at the time of revision, with cobalt and chromium levels dropping by 50% and 31% at 1 week, and 61% and 52% at 1 month after the revision surgery. During the period of surveillance in this study, 8 of 12 patients had their cobalt ion levels dropping below 7 ppb at an average of 2 months. Only 4 patients had a drop in chromium ion levels to 7 ppb, in this case at an average of 3 months after revision. After the initial exponential decrease in ion concentration, the decay curve entered a steady state with abnormal elevations of metal ion levels in the blood for periods of up to 2 years after revision. Although in most patients metal ion concentrations dropped below 7 ppb within 3 months, in a minority of cases (4 of 12) chromium ion concentrations remained between 12 and 36 ppb at an average of 1 year after revision.

Chromium concentrations decreased more slowly than cobalt concentrations at both early and late time points after revision surgery.

In our patient population, average blood ion concentrations on the day of revision were 43 ppb for cobalt and 22 ppb for chromium, with large inter-patient variability. We attribute these differences to variations in the debris burden remaining in the tissues after debridement at revision (and its rate of corrosion in vivo), to differences between patients in the rate of renal clearance of ions in the blood, and to variations in the extent to which debris was bound and encapsulated in tissues and organs and therefore available for dissolution and excretion (Case et al. 1994, Urban et al. 2004, Hart et al. 2010). It is important to note that blood ion concentrations are singular pieces of information that may be of diagnostic significance when evaluating the performance of hip implants, but they cannot take the place of a thorough history and physical examination or radiographic evaluation. In fact, 1 of 3 patients with destructive muscle loss and 1 of 4 patients with massive osteolysis had cobalt and chromium blood levels below 7 ppb.

The average time in situ was 5 (1-11) years before revision. Although the average survivorship of MoM implants is much longer, the overall systemic exposure to cobalt and chromium at the time of revision was 156 and 107 times larger than control values. Interestingly, in half of our study population with ion measurements beyond 30 days, the levels of chromium ions increased between 2 and 4 months postoperatively. One possible cause of this increase could be mobilization of chromium ions stored in the organs triggered by a decrease in circulating chromium levels. Urban et al. (2004) noted accumulation of wear particles in the spleen and liver at autopsy or biopsy in 7 of 8 patients after revision of their hip replacement. Exposure to high doses of metallic salts in animals showed that urinary excretion of chromium lagged behind that of cobalt and nickel, and it accumulated in the lung, liver, kidney, spleen, and red blood cells (Merritt et al. 1989). Other possible explanations include a mobilization of ions from the operative joint to the blood in response to healing, reduced renal clearance of chromium, or an increase in exogenous chromium from food or supplements.

Based on the values of the regression coefficients for decay of cobalt and chromium ions, a second-order model best fitted the kinetics of elution of cobalt ions after implant removal, indicating that the change in cobalt ion levels occurs through 2 independent processes. Cobalt is usually conserved by the kidneys in the normal population, but little is known about the kinetics of renal transport and excretion of metal ions. Both first- and second-order kinetics approximated the decay in chromium ion concentration. Our decay curves and the lag in chromium excretion compared to cobalt excretion shows that a single independent transport mechanism is unlikely. It is unknown whether second-order kinetics also describes excretion of ions from patients with well-functioning implants or from patients without implants, or whether the ion levels are sufficiently low for secretion to occur via a single-stage mechanism.

Patients with the highest initial ion levels were also exposed to markedly elevated concentrations of blood metal ions for extended periods after revision of their MoM components. Based on our measurements of the area under the curves of ion concentration vs. time, cumulative exposures in these patients in the first few months after revision were equivalent to several years in patients with well-functioning implants. It is important to note that this exposure comes even after removal of the source of MoM debris from the body, and that the metal ion exposure is presumed to be considerably higher while in situ. Despite the rapid decay in blood ion levels in many of our patients, the sheer magnitude of ion exposure from these poorly functioning implants is cause for concern. The longterm sequelae of metal ion exposure are unknown; however, given the level of chronic exposure, long-term follow-up of patients after revision of MoM implants appears to be warranted in order to detect any medical complications (Tower 2010).

The work presented here was a collaboration between all the authors, who contributed to and approved the manuscript.

The authors wish to acknowledge the assistance of Ms Denise Leon and Ms Sophie Fuller in preparing the manuscript.

The study was performed at the London Implant Retrieval Centre (LIRC) and the Institute of Orthopedic Research and Education (IORE) in Houston. All activities were funded using the internal resources of both organizations. The LIRC is funded by the British Orthopaedic Association (BOA) and is a joint venture between Imperial College London, the Royal National Orthopaedic Hospital, the British Orthopaedic Association, and 9 orthopedic companies (JRI, Corin, Mathys, Zimmer, Depuy, Finsbury, Stryker, Smith and Nephew, and Biomet).

No competing interests declared.

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