

Effects of aging and exercise training on apoptosis in the heart

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Aging is characterized by a progressive decline in cardiac function. A critical contributor to the age-related impairment in cardiac function is the loss of cardiac myocytes through "apoptosis", or programmed cell death. Structural remodeling in the heart with advancing age includes (a) loss of cardiomyocytes, (b) reactive hypertrophy of the remaining cardiomyocytes, and (c) increased connective tissue and altered geometry. The loss of cardiomyocytes with aging occurs through apoptosis. Particularly, mitochondrial-mediated apoptotic pathway is the best characterized and believed critical in regulating apoptosis with aging, suggesting that mitochondria are very important sites of programmed cell death. It has been also reported that mitochondrial dysfunction, oxidative stress, and impaired stress response contribute to age-induced mechanical remodeling as well as apoptosis. In contrast, exercise

training not only improves cardiac function, but also reduces the risk of heart disease. We recently found that aging increased mitochondrial-mediated apoptotic signaling and apoptosis in the left ventricle, while chronic exercise training was effective in diminishing mitochondrial-mediated apoptotic signaling pathways in the aging heart, as indicated by lower DNA fragmentation, terminal deoxynucleotidyl transferase-mediated dUTP nick end labeling (TUNEL)-positive staining, and caspase-3 cleavage, when compared with left ventricles from the age-matched sedentary group. In this review, we will provide a comprehensive update regarding the effects of aging and exercise training on apoptosis in the heart.

Keywords: Aging, Myocytes, Apoptosis, Mitochondria, Heart

INTRODUCTION

Aging is characterized by a progressive decline in cardiac function including stroke volume, cardiac output, blood flow, and oxygen consumption (Beere et al., 1999; Dinunno et al., 2001; Fiechter et al., 2013) and an increase in susceptibility to inflammation, oxidative stress, and disease (Higashi et al., 2012; Sastre et al., 2000; Zhang and Herman, 2002). Impaired cardiac function, increased inflammatory cytokines, and elevated oxidative stress are commonalities between aging and cardiovascular disease. A significant reduction in the number of myocytes is now believed to contribute directly to impaired contractile function, cardiomyopathy, heart disease, and heart failure (Favaloro et al., 2012; Kajstura et al., 1996; Pollack et al., 2002). Indeed, the typical 70 year old man has a 30% reduction in the number of cardiac myo-

cytes compared with young adults (Higami and Shimokawa, 2000; Olivetti et al., 1991).

Loss of myocytes with aging occurs through necrosis (i.e., accidental cell death) and apoptosis (i.e., programmed cell death), two distinct mechanisms leading to cell death (Konstantinidis et al., 2012). Necrosis results from cellular injury seen with infection or inflammatory disease, and is characterized by cellular swelling and rupturing with inflammation (Carraro and Franceschi, 1997; Mughal et al., 2012; Phaneuf and Leeuwenburgh, 2001; Zhang and Herman, 2002). Cell rupture with necrosis results in the release of stress and inflammatory proteins and substrates. In contrast, apoptosis is highly regulated cell death without injury and inflammation, and is characterized by cell shrinking/blebbing and condensation of the nucleus (Carraro and Franceschi, 1997; Mughal et al., 2012; Pollack and Leeuwenburgh, 2001; Zhang and Herman,

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2002). Apoptosis is a highly regulated, programmed means of cell death or elimination that plays an essential role in governing development, growth, and repair (Hengartner, 2000; Jacobson et al., 1997; Zhang and Herman, 2002). However, excessive apoptosis results in dysfunction and disease. A dramatic increase in the rate of apoptosis has been reported with aging in the rat left ventricle, while the rate of necrosis remained constant (Higami and Shimokawa, 2000; Kajstura et al., 1996; Konstantinidis et al., 2012; Liu et al., 2012). Phaneuf and Leeuwenburgh (2002) described age-related disruption of Bcl-2 family signaling in the rat heart. Progressive apoptosis from age in post-mitotic tissues such as the heart is dire, as lost myocytes are not replaced. Unfortunately, the mechanisms responsible for apoptotic signaling and apoptosis in the aging heart remain unclear and very limited.

In contrast, exercise training can reduce the risk of injury, oxidative stress, and inflammatory signaling from mechanical and oxidant perturbations, as long as overtraining does not occur (Fielding and Evans, 1997; Hammeren et al., 1992; Papathanasiou et al., 2012; Powers et al., 2002). Also, exercise training improves cardiovascular capacity and reduces cardiovascular disease risk in both young adults and the elderly (Booth et al., 2002; Gremeaux et al., 2012; Lawler et al., 1993). Exercise has the potential to reduce apoptosis through upregulation of protective stress-sensitive proteins including nuclear factor kappaB (NF-κB), insulin-like growth factor (IGF-1), and heat shock proteins (HSP90 and HSP70) (Frost and Lang, 2003; Milne and Noble, 2002; Morton et al., 2009; Naito et al., 2001). However, the mechanisms by which exercise training improves heart function and cardiovascular disease risk profile are not well delineated. Moreover, the ability of exercise training to modulate Bcl-2 family apoptotic signaling and apoptosis in the aging heart has not been clearly explored so far. Therefore, in the next section, an overview of aging and apoptosis in the heart will be provided, followed by a brief review of the literature supporting the effects of exercise in the aging heart.

APOPTOSIS AND MITOCHONDRIAL CONTROL

Apoptosis is regulated by genetic programming and plays an essential role in governing development, growth, and disease (Hengartner, 2000; Zhang and Herman, 2002). One of the most visible examples of cell death that occurs in normal animal development is the loss of the tadpole's tail (Jacobson et al., 1997). However, excessive apoptosis or dysregulation of apoptosis results in dysfunction and disease (Zhang and Herman, 2002).

Apoptosis is distinct from necrosis (Carraro and Franceschi, 1997; Fiers et al., 1998; Konstantinidis et al., 2012; Pollack and Leeuwenburgh, 2001). Necrosis or “accidental cell death” is the pathological process which is caused by injury, infection or inflammatory diseases such as high blood pressure and heart attacks (Hofmann, 1999). In contrast, apoptosis or “programmed cell death” is highly regulated cell death involving DNA fragmentation, blebbing, and dismantling of the cell without inflammation (Favaloro et al., 2012; Jacobson et al., 1997; Lemasters et al., 1999).

Apoptotic signaling induces apoptosis through caspase-dependent pathways (Budihardjo et al., 1999; Favaloro et al., 2012; Hengartner, 2000) (Fig. 1). They include (a) cytokines/Fas-driven pathways (Hofmann, 1999), (b) mitochondrial-driven pathways (Mignotte and Vayssiere, 1998), and (c) endoplasmic reticulum (ER)/Ca²⁺-driven pathways (Pollack and Leeuwenburgh, 2001). Binding of cytokines (e.g., TNF-α, IL-1β) or Fas to their cell membrane receptors initiate apoptotic pathway through the recruitment of adaptor proteins (e.g., Fas-associated death domain; FADD). The resulting death-inducing signaling complex activates caspase-8, which subsequently activates caspase-3 (Hofmann, 1999). Endoplasmic reticulum (ER)-mediated apoptotic signaling pathway is initiated by the involvement of caspase-12, which also subsequently activates caspase-3 (Pollack and Leeuwenburgh,

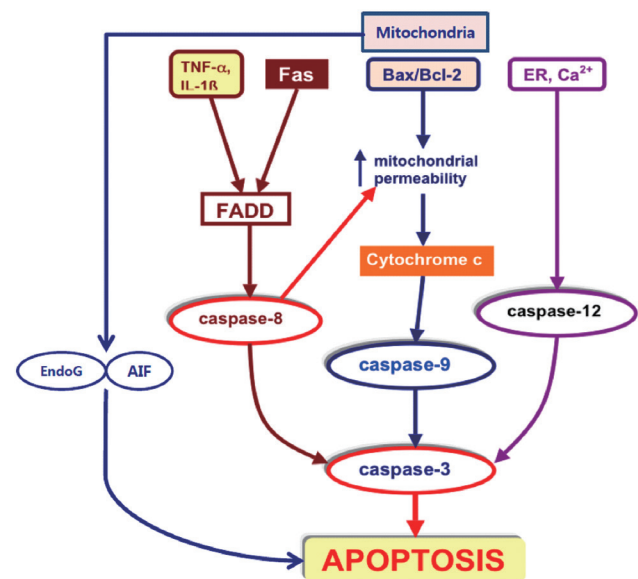


Fig. 1. Apoptotic signaling pathways. Schematic overview of three major caspases-dependent apoptotic signaling pathways including (a) cytokines/Fas-mediated caspase-8 pathways, (b) mitochondrial-mediated caspase-9 pathways, and (c) endoplasmic reticulum (ER)/Ca²⁺-mediated caspase-12 pathways. Alternatively, mitochondria can release AIF and EndoG, which perform apoptosis independent of caspases activation.

2001). Mitochondrial-mediated pathways including the Bcl-2 family is the best characterized, and Bcl-2 signaling is believed critical in regulating apoptosis with aging (Green and Reed, 1998; Hengartner, 2000; Konstantinidis et al., 2012; Mignotte and Vayssiere, 1998; Pollack and Leeuwenburgh, 2001). In other words, mitochondria are important sites of programmed cell death (Favaloro et al., 2012; Remmen and Richardson, 2001). A critical factor in mitochondrial-mediated apoptosis is that deficiency of survival stimuli such as growth factors increases mitochondrial permeability. Mitochondrial membrane permeability transition pore opening is regulated by a number of pro- and anti-apoptotic proteins such as Bcl-2 family of proteins (Green and Reed, 1998; Hengartner, 2000; Mignotte and Vayssiere, 1998).

Bcl-2 family has been classified into three functional groups (Hengartner, 2000). Group I such as Bcl-2, Bcl-X_L inhibits apoptosis. Group II such as Bax initiates apoptosis. And group III such as Bid also initiates apoptosis. In other words, Group I functions as gatekeepers at the outer surface of mitochondria, whereas Group II and III function as gatecrashers. Therefore, cell death or survival depends on the ratio of the pro- and anti-apoptotic proteins expressed. For example, high levels of Bcl-2 relative to Bax promote survival, whereas the reverse ratio promotes myonuclear cell death (Hengartner, 2000; Pawlowski and Kraft, 2000; Primeau et al., 2002). Thus the ratio of anti-apoptotic/pro-apoptotic proteins in the Bcl-2 family functions as an upstream modulator of apoptosis in the mitochondrial-mediated apoptotic pathways. The proteins of Bcl-2 family can regulate apoptosis by controlling permeability transition pore (PTP) opening and the release of cytochrome c from mitochondria (Mignotte and Vayssiere, 1998).

Caspases are a group of cysteine dependent aspartate-specific proteases (Favaloro et al., 2012; Nicholson, 1999; Zhang et al., 2003). In other words, caspases are a complex cascade of protein-cleaving enzymes. Fourteen caspases have now been identified in mammals, leading to apoptotic cell death (Kumar, 1999). Caspases play a pivotal role in apoptotic pathways and interact with the non-caspase apoptotic pathways including endonuclease G (Endo G) and apoptosis inducing factor (AIF) (Marzetti et al., 2012; Nicholson, 1999; Zhang et al., 2003). Cytochrome c associates with Apaf-1 (apoptotic protease-activating factor 1) and pro-caspase-9 to form the apoptosome, which activates caspase-9 and caspase-3, resulting in apoptosis through DNA fragmentation (Favaloro et al., 2012; Kumar et al., 2002; Mignotte and Vayssiere, 1998; Pollack et al., 2002; Susin et al., 2000; Zhang et al., 2002).

AGING AND APOPTOSIS

Aging is characterized by a general decline of physiological function (Dinenno et al., 2001; Liu et al., 2012). Aging enhances the susceptibility to apoptosis in several types of tissues. It has been shown that aging is associated with increased Bax protein as well as enhanced DNA fragmentation in brains (Higami and Shimokawa, 2000). Aging-induced apoptosis may contribute to a 30% loss of cardiac myocytes outside changes noted in cardiac diseases (Higami and Shimokawa, 2000; Liu et al., 2012; Olivetti et al., 1991). Myocyte cell death including both apoptosis and necrosis increased with aging in the heart of Fischer-344 rats, mediating ventricular dysfunction and failure (Kajstura et al., 1996). Liu et al. (1998) also reported that expression of Bcl-2 and Bax proteins both increased in the heart of elderly Fischer-344 rats under physiological conditions. Moreover, our research group demonstrated that aging increased pro-apoptotic Bax, cleaved caspase-3, and DNA fragmentation in rat skeletal muscle (Song et al. 2006) as well as heart (Kwak et al., 2006). In contrast, Nitahara et al. (1998) showed that there was an increase in cardiomyocyte apoptosis with age from Fischer-344 rats without significant changes in Bax and Bcl-2 protein levels with age. These findings suggest that age-related changes in the expression of Bcl-2 family proteins (Bcl-2 and Bax) are still debatable.

Progressive apoptosis with advancing age in post-mitotic tissues including heart, skeletal muscle, and brain is dire, because lost myocytes are not replaced. Phaneuf and Leeuwenburgh (2002) showed markers of apoptosis in hearts (left and right ventricles) of 6-, 16-, and 24-month-old male Fischer-344 rats. Cytosolic cytochrome c was significantly increased in the 16- and 24-month-old animals compared to the 6-month-old animals, indicating increased mitochondrial permeability transition pore (PTP) opening and activation of the apoptosome. Also, Bcl-2 levels were decreased with aging, whereas mitochondrial Bax levels remained unchanged.

In contrast, mitotic tissues such as the liver, colon mucosa, etc. display reduced pro-apoptotic protein levels, cytochrome c contents, caspase levels, and DNA fragmentation with aging. This dysregulation of apoptotic signaling from aging is postulated as a mechanism contributing to cancer risk (Suh et al., 2002). For example, it has been reported that caspase-3 and caspase-9 protein levels and activities were lower in colon mucosa in old (22-24 months) Fischer-344 rats compared to their young (4-5 months) and middle-age (12-14 months) counterparts (Xiao et al., 2001). These changes were accompanied by a reduced number of apoptotic cells in the colonic mucosa of 12-14 and 22-24 month old

rats compared with in 4-6 month old rats. In addition, the levels of pro-apoptotic Bak were decreased by about 50%, whereas anti-apoptotic Bcl-X_L levels were increased by about 50% in 22 month old rats compared to 4 or 13 month old rats (Xiao et al., 2001). Suh et al. (2002) also showed that apoptosis-induced DNA fragmentation was dramatically upregulated in the liver of young (2 months) female Fischer rats but not in the old (26 months) counterparts. Additionally, the levels of cytochrome c in livers from old (26 months) Fischer-344 rats were significantly lower than those from young (6 months) counterparts (Zhang et al., 2002). These opposite results could be tissue specific and possible characteristics of mitotic but not post-mitotic tissues.

APOPTOSIS AND HEART

The heart may undergo an increase in apoptosis during ischemia-reperfusion, heart failure, doxorubicin treatment, and aging (Childs et al., 2002; Higami and Shimokawa, 2000; Olivetti et al., 1997; Powers et al., 2002; Tacar et al., 2013). It has been reported that a number of cardiomyopathies are associated with mitochondrial DNA damage which leads to defects in electron transport chain and the upregulation of ROS (Anderson et al., 2011; Mignotte and Vayssiere, 1998), resulting in the apoptosis of cardiac myocytes (Li et al., 2012; Mughal et al., 2012; Von Harsdorf et al., 1999). Heart is an energetically-demanding tissue that requires a continuous production of ATP via mitochondrial respiration (Li et al., 2012; Primeau et al., 2002). In highly aerobic tissues such as heart, mitochondria occupy up to 25% of cell volume (Lemasters et al., 1999; Li et al., 2012). It was also shown that tumor necrosis factor- α (TNF- α) increased apoptosis via inducible nitric oxide synthase (iNOS) expression and nitric oxide (NO) in cardiac myocytes (Ing et al., 1999; Song et al., 2000). In both humans and animals, the aging heart is characterized by a decrease in the total number of myocytes resulting in reactive hypertrophy of the remaining cells and increased fibrosis (Kwak et al., 2006, 2011; Olivetti et al., 1991; Phaneuf and Leeuwenburgh, 2002).

Ischemia and reperfusion of myocardium induces apoptotic cell death (Powers et al., 2002), with direct relevance to heart disease and heart attacks (Quindry et al., 2012). Interestingly, brief periods of acute myocardial ischemia, or "preconditioning," protects the heart from the deleterious effects of longer periods of ischemia and reperfusion (Murry et al., 1986; Taylor and Starnes, 2003). The mechanisms of protection against apoptosis due to preconditioning may be mediated by NF- κ B, which inhibits the susceptibility to apoptosis and can promote upregulation of anti-apoptotic

Bcl-2 (Das et al., 1999; Maulik et al., 1998). Indeed, NF- κ B DNA binding activity progressively increased as a function of the preconditioning of ischemia (Maulik et al., 2000), and Bcl-2 levels were decreased in the ischemic/reperfused heart, but increased in the preconditioned myocardium (Maulik et al., 2000). In addition, overexpression of anti-apoptotic Bcl-2 was effective in reducing myocardial reperfusion injury and improving heart function (Brocheriou et al., 2000).

Indeed, the balance between Bax/Bcl-2 is postulated as a critically important factor in the increased rate of apoptosis in cardiac myocytes (Condorelli et al., 1999; Pollack et al., 2002). For example, Condorelli et al. (1999) indicated that left ventricular hypertrophy and left ventricular dysfunction in a rat model of chronic pressure overload were accompanied by upregulated pro-apoptotic Bax and Bax/Bcl-2 ratio, leading to cardiomyocyte apoptosis. Kang et al. (2000) also found that apoptosis predominated in cardiomyocytes after reoxygenation through a mitochondrial apoptotic pathway, and that Bcl-2 prevented reoxygenation-induced apoptosis by inhibiting the release of cytochrome c from mitochondria. In addition, Kirshenbaum and Moissac (1997) concluded that the anti-apoptotic Bcl-2 protein can prevent apoptosis of ventricular myocytes.

Growing evidence shows that cardiomyocyte apoptosis also contributes to congestive heart failure. For example, Olivetti et al. (1997) showed that cardiomyocyte apoptosis dramatically increased in the heart of patients with cardiac failure, although the level of Bcl-2 was about two times higher than that of the normal hearts, while the level of Bax remained unchanged. Saraste et al. (1999) also showed that the number of cardiomyocytes undergoing apoptosis was significantly increased in failing human hearts compared with control myocardium, and that the expression of Bcl-2 was increased in failing hearts. Enhanced expression of Bcl-2 in the failing heart suggested that compensatory mechanisms were activated to prevent apoptosis (Saraste et al., 1999). The expression of Bcl-2 protein was induced in human cardiac myocytes at the acute stage of infarction, but the expression of Bax protein was overexpressed at the old stage, which was related to myocyte death in human hearts (Misao et al., 1996).

EXERCISE AND APOPTOSIS IN THE AGING HEART

Our research group recently indicated that the Bax/Bcl-2 ratio was significantly upregulated with age and markedly reduced by exercise training in the left ventricles (Kwak et al., 2006). These

data imply that aging heart is more susceptible to apoptosis than young heart, and that chronic exercise training exerts anti-apoptotic action in the aging heart (Soufi et al., 2008). These data are consistent with previous reports showing that aging is associated with increased Bax protein as well as decreased Bcl-2 (Higami and Shimokawa, 2000; Liu et al., 1998), indicating that mitochondrial Bcl-2 family signaling is an important site of regulating apoptosis with aging and exercise training.

Based on our data (Kwak et al., 2006), the increased Bax/Bcl-2 ratio with aging heart was directly related to increases in downstream caspase signaling including caspase-9 and cleaved caspase-3, which promote DNA fragmentation and cell death (e.g., increased TUNEL-positive nuclei). Similarly, there were consistent reports that caspase-9 and caspase-3 activities were increased in aging rat liver (Zhang and Herman, 2002). In contrast, exercise training markedly decreased caspase-9 and cleaved caspase-3 levels as well as the Bax/Bcl-2 ratio in the aging heart, which presumably led to the decreased DNA fragmentation. These results are consistent with the hypotheses that aging increases Bcl-2 family pro-apoptotic signaling in the heart, and exercise training in the aging heart results in amelioration of age-induced changes in the mitochondrial-mediated apoptotic pathways. In particular, cleaved caspase-3 showed similar patterns to DNA fragmentation in the effect of aging and exercise training, suggesting that caspase-3 may be an essential hallmark of apoptosis (Fig. 2).

Aging is generally characterized by a decline of cardiac function and an upregulation of oxidative stress in the heart (Beere et al., 1999; Dinenn et al., 2001; Sastre et al., 2000; Soufi et al., 2008), which are major contributors to cell death through mitochondrial dysfunction (Dirks and Leeuwenburgh, 2002; Sastre et al., 2000). Previous work has demonstrated that aging promotes the suscep-

tibility to apoptosis in rat heart (Higami and Shimokawa, 2000; Kajstura et al., 1996; Nitahara et al. 1998; Olivetti et al., 1991; Soufi et al., 2008). While aging heart was vulnerable to apoptosis, exercise training was effective in diminishing apoptosis (DNA fragmentation) in the aging heart, which indicated the beneficial effect of long-term exercise training on apoptosis in the aging heart (Soufi et al., 2008). Interestingly we found that even if apoptosis was dramatically increased with aging, there was no significant changes in heart-to-body weight ratio between young seden-

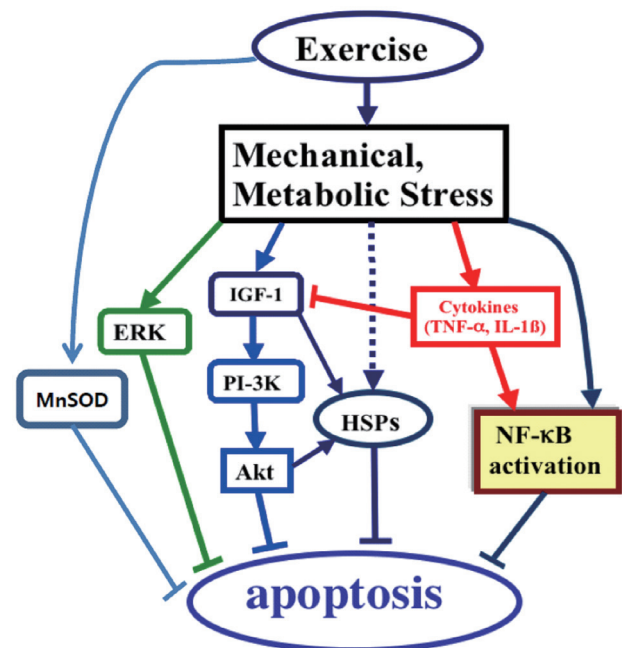


Fig. 3. Exercise protection model against apoptosis in the heart. An integrative model of exercise protection against apoptosis in the heart whereby exercise can potentially upregulate stress proteins such as MnSOD, ERK, IGF-1/Akt, HSPs, and NF-κB.

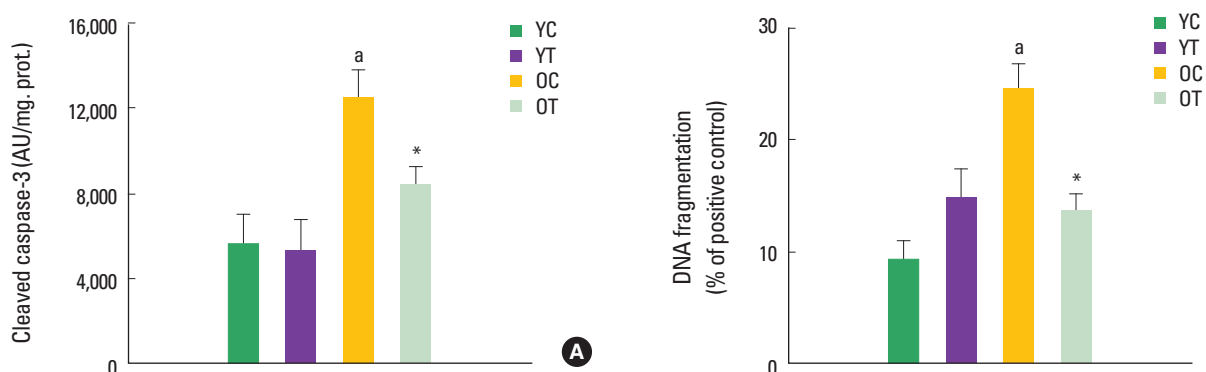


Fig. 2. Effects of aging and exercise training on apoptosis in the heart. (A) Cleaved caspase-3 protein levels and (B) DNA fragmentation through quantification of mono and oligonucleosomes via ELISA of left ventricles in young control (YC), young exercise trained (YE), old control (OC), and old exercise trained (OT) (Kwak et al., 2006).

tary rats and old sedentary rats (Kwak et al., 2006) because the aging process of the heart in humans and animals is characterized by a significant loss of myocytes and reactive hypertrophy of the remaining cells (Olivetti et al., 1991; Phaneuf and Leeuwenburgh, 2002). Thus, the loss of cardiac myocytes doesn't necessarily cause a reduction in mass. In addition, with aging there is an increase in heart connective tissue, increasing wall stiffness/thickness, and losing elasticity of the heart (Anversa et al., 1990; Kwak et al., 2011; Lakatta and Sollott, 2002). However, cardiac myocytes that hypertrophy often have poor contractile function due to increased left ventricle wall thickness (Lakatta and Sollott, 2002).

Based upon current findings regarding exercise and apoptosis in the aging heart, further investigations as future directions are required to determine the effects of exercise training on the upstream cell protective mechanisms in the aging heart (Fig. 3). For example, exercise training may promote cell-survival proteins including Mn isoform of superoxide dismutase (MnSOD), NF- κ B, extracellular receptor kinase (ERK), IGF-1/Akt pathway, and heat shock proteins (HSPs) in heart (Higashi et al., 2012; Powers et al., 2002; Soufi et al., 2008; Starnes et al., 2003; Taylor and Starnes, 2003), which may be potential upstream regulators of age-induced apoptosis in the aging heart.

CONCLUSIONS

In summary, aging resulted in increases in mitochondrial-mediated apoptotic pathways including pro-apoptotic protein levels such as Bax, Bax/Bcl-2 ratio, caspase-9, and cleaved caspase-3 and apoptosis. However, endurance exercise training reversed the elevation of apoptotic signaling and apoptosis, suggesting that exercise training protects the heart against apoptosis. However, further research is necessary to determine the cellular and molecular mechanisms by which exercise training protects against aging-induced apoptosis in the heart.

CONFLICT OF INTEREST

No potential conflict of interest relevant to this article was reported.

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REFERENCES

- Anderson EJ, Rodriguez E, Anderson CA, Thayne K, Chitwood WR, Kypson AP. Increased propensity for cell death in diabetic human heart is mediated by mitochondrial-dependent pathways. *Am J Physiol Heart Circ Physiol* 2011;300:118-124.
- Anversa P, Palackal T, Sonnenblick EH, Olivetti G, Meggs LG, Capasso JM. Myocyte cell loss and myocyte cellular hyperplasia in the hypertrophied aging rat heart. *Circ Res* 1990;67:871-885.
- Beere PA, Russell SD, Morey MC, Kitzman DW, Higginbotham MB. Aerobic exercise training can reverse age-related peripheral circulatory changes in healthy older men. *Circulation* 1999;100:1085-1094.
- Booth FW, Chakravarthy MV, Gordon SE, Spangenburg EE. Waging war on physical inactivity: using modern molecular ammunition against an ancient enemy. *J Appl Physiol* 2002;93:3-30.
- Brocheriou V, Hagege AA, Oubenaissa A, Lambert M, Mallet VO, Duriez M, Wassef M, Kahn A, Menasche P, Gilgenkrantz H. Cardiac functional improvement by a human Bcl-2 transgene in a mouse model of ischemic/reperfusion injury. *J Gene Med* 2000; 2:326-333.
- Budihardjo I, Oliver H, Lutter M, Luo X, Wang X. Biochemical pathways of caspase activation during apoptosis. *Annu Rev Cell Dev Biol* 1999; 15:269-290.
- Carraro U, Franceschi C. Apoptosis of skeletal and cardiac muscles and physical exercise. *Aging Clin Exp Res* 1997;9:19-34.
- Childs AC, Phaneuf SL, Dirks AJ, Phillips T, Leeuwenburgh C. Doxorubicin treatment in vivo causes cytochrome c release and cardiomyocyte apoptosis, as well as increased mitochondrial efficiency, superoxide dismutase activity, and Bcl-2:Bax ratio. *Can Res* 2002;62:4592-4598.
- Condorelli G, Morisco C, Stassi G, Notte A, Farina F, Sgarrella G, De Rienzo A, Roncarati R, Trimarco B, Lembo G. Increased cardiomyocyte apoptosis and changes in proapoptotic and antiapoptotic genes bax and bcl-2 during left ventricular adaptations to chronic pressure overload in the rat. *Circulation* 1999;99:3071-3078.
- Das DK, Maulik N, Sato M, Ray PS. Reactive oxygen species function as second messenger during ischemic preconditioning of heart. *Mol Cell Biochem* 1999;196:59-67.
- Dinenna FA, Seals DR, DeSouza CA, Tanaka H. Age-related decreases in basal limb blood flow in humans: time course, determinants and habitual exercise effects. *J Physiol* 2001;531.2:573-579.
- Dirks A, Leeuwenburgh C. Apoptosis in skeletal muscle with aging. *Am J Physiol Regul Integr Comp Physiol* 2002;282:519-527.
- Favaloro B, Allocati N, Graziano V, Di Ilio C, De Laurenzi V. Role of apop-

- tosis in disease. *Aging* (Albany NY) 2012;4:330-349.
- Fiechter M, Fuchs TA, Gebhard C, Stehli J, Klaeser B, Stähli BE, Manka R, Manes C, Tanner FC, Gaemperli O, Kaufmann PA. Age-related normal structural and functional ventricular values in cardiac function assessed by magnetic resonance. *BMC Med Imaging* 2013;13:6.
- Fielding RA, Evans WJ. Aging and the acute phase response to exercise: implications for the role of systemic factors on skeletal muscle protein turnover. *Int J Sports Med* 1997;18:22-27.
- Fiers W, Beyaert R, Declercq W, Vandenabeele P. More than one way to die: apoptosis, necrosis and reactive oxygen damage. *Oncogene* 1998; 18:7719-7730.
- Frost RA, Lang CH. Regulation of insulin-like growth factor-1 in skeletal muscle and muscle cells. *Minerv Endocrinol* 2003;28:53-73.
- Green DR, Reed JC. Mitochondria and apoptosis. *Science* 1998;281:1309-1312.
- Gremeaux V, Gayda M, Lepers R, Sosner P, Juneau M, Nigam A. Exercise and longevity. *Maturitas* 2012;73:312-317.
- Hammeren J, Powers S, Lawler J, Criswell D, Martin D, Lowenthal D, Pollock M. Exercise training-induced alterations in skeletal muscle oxidative and antioxidant enzyme activity in senescent rats. *Int J Sports Med* 1992;13:412-416.
- Hengartner MO. The biochemistry of apoptosis. *Nature* 2000;407:770-776.
- Higami Y, Shimokawa I. Apoptosis in the aging process. *Cell Tissue Res* 2000;301:125-132.
- Higashi Y, Sukhanov S, Anwar A, Shai SY, Delafontaine P. Aging, atherosclerosis, and IGF-1. *J Gerontol A Biol Sci Med Sci* 2012;67:626-639.
- Hofmann K. The modular nature of apoptotic signaling proteins. *Cell Mol Life Sci* 1999;55:1113-1128.
- Ing DJ, Zang J, Dzau VJ, Webster KA, Bishopric NH. Modulation of cytokine-induced myocyte apoptosis by nitric oxide, Bak, and Bcl-x. *Circ Res* 1999;84:21-33.
- Jacobson MD, Weil M, Raff MC. Programmed cell death in animal development. *Cell* 1997;88:347-354.
- Kajstura J, Cheng W, Sarangarajan R, Li P, Li B, Nitahara JA, Charnick S, Reiss K, Olivetti G, Anversa P. Necrotic and apoptotic myocyte cell death in the aging heart of Fischer 344 rats. *Am J Physiol Heart Circ Physiol* 1996;271:1215-1228.
- Kang PM, Haunstetter A, Aoki H, Usheva A, Izumo S. Morphological and molecular characterization of adult cardiomyocyte apoptosis during hypoxia and reoxygenation. *Circ Res* 2000;87:118-125.
- Kirshenbaum LA, De Moissac D. The bcl-2 gene product prevents programmed cell death of ventricular myocytes. *Circulation* 1997;96:1580-1585.
- Konstantinidis K, Whelan RS, Kitsis RN. Mechanisms of cell death in heart disease. *Arterioscler Thromb Vasc Biol* 2012;32:1552-1562.
- Kumar D, Lou H, Singal PK. Oxidative stress and apoptosis in heart dysfunction. *Herz* 2002;662-668.
- Kumar S. Mechanisms mediating caspase activation in cell death. *Cell Death Differ* 1999;6:1060-1066.
- Kwak HB, Kim JH, Joshi K, Yeh A, Martinez DA, Lawler JM. Exercise training reduces fibrosis and matrix metalloproteinase dysregulation in the aging rat heart. *FASEB J* 2011;25:1106-1117.
- Kwak HB, Song W, Lawler JM. Exercise training attenuates age-induced elevation in Bax/Bcl-2 ratio, apoptosis, and remodeling in the rat heart. *FASEB J* 2006;20:791-793.
- Lakatta EG, Sollott SJ. The heartbreak of older age. *Mol Interv* 2002;2:431-446.
- Lawler JM, Powers SK, Hammeren J, Martin AD. Oxygen cost of treadmill running in 24 month old Fisher-344 rats. *Med Sci Sports Exer* 1993; 25:1259-1264.
- Lemasters JJ, Qian T, Bradham CA, Brenner DA, Cascio WE, Trost LC, Nishimura Y, Nieminen AL, Herman B. Mitochondrial dysfunction in the pathogenesis of necrotic and apoptotic cell death. *J Bioener Biomem* 1999;31:305-319.
- Li Q, Zhou LY, Gao GF, Jiao JQ, Li PF. Mitochondrial network in the heart. *Protein Cell* 2012;3:410-418.
- Liu L, Azhar G, Gao W, Zhang X, Wei JY. Bcl-2 and Bax expression in adult rat hearts after coronary occlusion: age-associated differences. *Am J Physiol Regul Integr Comp Physiol* 1998;275:315-322.
- Liu M, Zhang P, Chen M, Zhang W, Yu L, Yang XC, Fan Q. Aging might increase myocardial ischemia/reperfusion-induced apoptosis in humans and rats. *Age (Dordr)* 2012;34:621-632.
- Marzetti E, Calvani R, Bernabei R, Leeuwenburgh C. Apoptosis in skeletal myocytes: a potential target for interventions against sarcopenia and physical frailty - a mini-review. *Gerontology* 2012;58:99-106.
- Maulik N, Sasaki H, Addya S, Das DK. Regulation of cardiomyocyte apoptosis by redox-sensitive transcription factors. *FEBS Lett* 2000;485:7-12.
- Maulik N, Yoshida T, Engelman RM, Deaton D, Flack JE, Rousou JA, Das DK. Ischemic preconditioning attenuates apoptotic cell death associated with ischemia/reperfusion. *Mol Cell Biochem* 1998;186:139-145.
- Mignotte B, Vayssiere JL. Mitochondria and apoptosis. *Eur J Biochem* 1998; 252:1-15.
- Milne KJ, Noble EG. Exercise-induced elevation of HSP70 is intensity dependent. *J Appl Physiol* 2002;93:561-568.
- Misao J, Hayakawa Y, Ohno M, Kato S, Fujiwara T, Fujiwara H. Coronary heart disease/myocardial infarction: expression of Bcl-2 protein, an inhibitor of apoptosis, and Bax, an accelerator of apoptosis, in ventricular myocytes of human hearts with myocardial infarction. *Circulation* 1996;94:1506-1512.
- Morton JP, Kayani AC, McArdle A, Drust B. The exercise-induced stress

- response of skeletal muscle, with specific emphasis on humans. *Sports Med* 2009;39:643-662.
- Mughal W, Dhingra R, Kirshenbaum LA. Striking a balance: autophagy, apoptosis, and necrosis in a normal and failing heart. *Curr Hypertens Rep* 2012;14:540-547.
- Murry CE, Jennings RB, Reimer KA. Preconditioning with ischemia: a delay of lethal cell injury in ischemic myocardium. *Circulation* 1986;74:1124-1136.
- Naito H, Powers SK, Demirel HA, Aoki J. Exercise training increases heat shock protein in skeletal muscles of old rats. *Med Sci Sports Exerc* 2001;33:729-734.
- Nicholson MW. Caspase structure, proteolytic substrates, and function during apoptotic cell death. *Cell Death Differ* 1999;6:1028-1042.
- Nitahara JA, Cheng W, Liu Y, Li B, Leri A, Li P, Mogul D, Gambert SR, Kajstura J, Anversa P. Intracellular calcium, DNase activity and myocyte apoptosis in aging Fischer 344 rats. *J Mol Cell Cardiol* 1998;30:519-535.
- Olivetti G, Abbi R, Quaini F, Kajstura J, Cheng W, Nitahara JA, Quaini E, Loreto CD, Beltrami CA, Krajewski S, Reed JC, Anversa P. Apoptosis in the failing human heart. *New Eng J Med* 1997;336:1131-1141.
- Olivetti G, Melissari M, Capasso JM, Anversa P. Cardiomyopathy of the aging human heart: myocyte loss and reactive cellular hypertrophy. *Circ Res* 1991;68:1560-1568.
- Papathanasiou JV, Ilieva EM, Nikolov FP. Exercise training modes in rehabilitation of patients with chronic heart failure. *Folia Med (Plovdiv)* 2012;54:22-28.
- Pawlowski J, Kraft AS. Bax-induced apoptotic cell death. *Proc Natl Acad Sci USA* 2000;97:529-531.
- Phaneuf S, Leeuwenburgh C. Apoptosis and exercise. *Med Sci Sports Exerc* 2001;33:393-396.
- Phaneuf S, Leeuwenburgh C. Cytochrome c release from mitochondria in the aging heart: a possible mechanism for apoptosis with age. *Am J Physiol Regul Integr Comp Physiol* 2002;282:423-430.
- Pollack M, Leeuwenburgh C. Apoptosis and aging: role of the mitochondria. *J Gerontol* 2001;56A:B475-B482.
- Pollack M, Phaneuf S, Dirks A, Leeuwenburgh C. The role of apoptosis in the normal aging brain, skeletal muscle, and heart. *Ann N Y Acad Sci* 2002;959:93-1072.
- Powers SK, Lennon SL, Quindry J, Mehta JL. Exercise and cardioprotection. *Curr Opin Cardiol* 2002;17:495-502.
- Primeau AJ, Adhithetty PJ, Hood DA. Apoptosis in heart and skeletal muscle. *Can J Appl Physiol* 2002;27:349-395.
- Quindry JC, Miller L, McGinnis G, Kliszczewicz B, Irwin JM, Landram M, Urbiztondo Z, Nanayakkara G, Amin R. Ischemia reperfusion injury, KATP channels, and exercise-induced cardioprotection against apoptosis. *J Appl Physiol* 2012;113:498-506.
- Remmen HV, Richardson A. Oxidative damage to mitochondria and aging. *Exp Gerontol* 2001;36:957-968.
- Saraste A, Pulkki K, Kallajoki M, Heikkila P, Laine P, Mattila S, Nieminen MS, Parvine M, Voipio-Pulkki LM. Cardiomyocyte apoptosis and progression of heart failure to transplantation. *Eur J Clin Inv* 1999;29:380-386.
- Sastre J, Pallardo FV, Vina J. Mitochondrial oxidative stress plays a key role in aging and apoptosis. *IUBMB Life* 2000;49:427-435.
- Song W, Kwak HB, Lawler JM. Exercise training attenuates age-induced changes in apoptotic signaling in rat skeletal muscle. *Antioxid Redox Signal* 2006;8:517-528.
- Song W, Lu X, Feng Q. Tumor necrosis factor- α induces apoptosis via inducible nitric oxide synthase in neonatal mouse cardiomyocytes. *Cardiovasc Res* 2000;45:595-602.
- Soufi FG, Farajnia S, Aslanabadi N, Ahmadiasl N, Alipour M, Alipour M, Doustar Y, Abdolalizadeh J, Sheikhzadeh F. Long-term exercise training affects age-induced changes in HSP70 and apoptosis in rat heart. *Gen Physiol Biophys* 2008;27:263-270.
- Starnes JW, Taylor RP, Park Y. Exercise improves postischemic function in aging heart. *Am J Physiol Heart Circ Physiol* 2003;285:347-351.
- Suh Y, Lee KA, Kim WH, Han BG, Vijg J, Park SC. Aging alters the apoptotic response to genotoxic stress. *Nat Med* 2002;8:3-4.
- Susin SA, Daugas E, Ravagnan L, Samejima K, Zamzami N, Loeffler M, Costantini P, Ferri KF, Irinopoulou T, Prevost MC, Brothers G, Mak TW, Penninger J, Earnshaw WC, Kroemer G. Two distinct pathways leading to nuclear apoptosis. *J Exp Med* 2000;192:571-579.
- Tacar O, Sriamornsak P, Dass CR. Doxorubicin: an update on anticancer molecular action, toxicity and novel drug delivery systems. *J Pharm Pharmacol* 2013;65:157-170.
- Taylor RP, Starnes JW. Age, cell signaling and cardioprotection. *Acta Physiol. Scand* 2003;178:107-163.
- Von Harsdorf R, Li PF, Dietz R. Signaling pathways in reactive oxygen species-induced cardiomyocyte apoptosis. *Circulation* 1999;99:2934-2941.
- Xiao Z, Moragoda L, Jaszewski R, Hatfield JA, Fligel SF, Majumdar AP. Aging is associated with increased proliferation and decreased apoptosis in the colonic mucosa. *Mech Age Dev* 2001;122:1849-1864.
- Zhang JH, Zhang Y, Herman B. Caspases, apoptosis and aging. *Ageing Res Rev* 2003;2:357-366.
- Zhang Y, Chong E, Herman B. Aging-associated increases in the activity of multiple caspases in Fischer 344 rat organs. *Exp Gerontol* 2002;37:777-789.
- Zhang Y, Herman B. Ageing and apoptosis. *Mech Age Dev* 2002;123:245-260.