

Citation: Vetrivel I, Mahajan S, Tyagi M, Hoffmann L, Sanejouand Y-H, Srinivasan N, et al. (2017) Knowledge-based prediction of protein backbone conformation using a structural alphabet. PLoS ONE 12(11): e0186215. https://doi.org/10.1371/ journal.pone.0186215

Editor: Yang Zhang, University of Michigan, UNITED STATES

Received: May 30, 2017

Accepted: September 27, 2017

Published: November 21, 2017

Copyright: This is an open access article, free of all copyright, and may be freely reproduced, distributed, transmitted, modified, built upon, or otherwise used by anyone for any lawful purpose. The work is made available under the <u>Creative</u> Commons CC0 public domain dedication.

Data Availability Statement: All relevant data has been deposited in Dryad (doi:10.5061/dryad. 3f5q5).

Funding: This work was supported by the Région Réunion and the Fond Social Européen [grant no. 20131528] to IV. This work was in part supported by Conseil Régional des Pays de la Loire in the framework of GRIOTE project. AdB and FC acknowledge grants from the Ministry of Research (France), National Institute for Blood Transfusion (INTS, France), National Institute for Health and **RESEARCH ARTICLE**

Knowledge-based prediction of protein backbone conformation using a structural alphabet

Iyanar Vetrivel¹, Swapnil Mahajan^{1,2}, Manoj Tyagi³^a, Lionel Hoffmann¹, Yves-Henri Sanejouand¹, Narayanaswamy Srinivasan⁴, Alexandre G. de Brevern⁵, Frédéric Cadet^{2,6}, Bernard Offmann¹*

1 Université de Nantes, Unité Fonctionnalité et Ingénierie des Protéines (UFIP), UMR 6286 CNRS, UFR Sciences et Techniques, 2, chemin de la Houssinière, France, 2 DSIMB, INSERM, UMR S-1134, Laboratory of Excellence, GR-Ex, Université de La Réunion, Faculty of Sciences and Technology, Saint Denis Cedex, La Réunion, France, 3 Université de La Réunion, Saint Denis Cedex, La Réunion, France, 4 Molecular Biophysics Unit, Indian Institute of Science, Bangalore, India, 5 INSERM UMR_S 1134, DSIMB team, Laboratory of Excellence, GR-Ex, Univ Paris Diderot, Univ Sorbonne Paris Cité, INTS, rue Alexandre Cabanel, Paris, France, 6 PEACCEL SAS, Paris, France

¤ Current address: National Cancer Institute, Bethesda, MA, United States of America * bernard.offmann@univ-nantes.fr

Abstract

Libraries of structural prototypes that abstract protein local structures are known as structural alphabets and have proven to be very useful in various aspects of protein structure analyses and predictions. One such library, Protein Blocks, is composed of 16 standard 5residues long structural prototypes. This form of analyzing proteins involves drafting its structure as a string of Protein Blocks. Predicting the local structure of a protein in terms of protein blocks is the general objective of this work. A new approach, PB-kPRED is proposed towards this aim. It involves (i) organizing the structural knowledge in the form of a database of pentapeptide fragments extracted from all protein structures in the PDB and (ii) applying a knowledge-based algorithm that does not rely on any secondary structure predictions and/ or sequence alignment profiles, to scan this database and predict most probable backbone conformations for the protein local structures. Though PB-kPRED uses the structural information from homologues in preference, if available. The predictions were evaluated rigorously on 15,544 query proteins representing a non-redundant subset of the PDB filtered at 30% sequence identity cut-off. We have shown that the kPRED method was able to achieve mean accuracies ranging from 40.8% to 66.3% depending on the availability of homologues. The impact of the different strategies for scanning the database on the prediction was evaluated and is discussed. Our results highlight the usefulness of the method in the context of proteins without any known structural homologues. A scoring function that gives a good estimate of the accuracy of prediction was further developed. This score estimates very well the accuracy of the algorithm (R² of 0.82). An online version of the tool is provided freely for non-commercial usage at http://www.bo-protscience.fr/kpred/.



Medical Research (INSERM, France) and labex GR-Ex. The labex GR-Ex, reference ANR-11-LABX-0051 is funded by the program "Investissements d'avenir" of the French National Research Agency, reference ANR-11-IDEX-0005-02. AdB acknowledge supports by University Paris Diderot, Sorbonne, Paris Cité (France), FC acknowledge supports by Université de La Réunion, Faculty of Sciences and Technology. NS and AdB acknowledge to Indo-French Centre for the Promotion of Advanced Research / CEFIPRA for collaborative grant (number 5302-2). Research in NS laboratory is also supported by Department of Biotechnology, Government of India. NS is a J.C. Bose National Fellow.

Competing interests: BO and FC are co-founders of PEACCEL SAS. This does not alter our adherence to PLOS ONE policies on sharing data and materials.

Introduction

Knowledge of protein structure considerably helps towards understanding protein function. The Protein Data Bank (PDB) that serves as the central repository of knowledge for the protein structural biology community contains more than 125,000 protein structures and its growth has been considerable in the past decade [1]. This number is however still far below the ~70 million protein sequences referenced in UniProt database [2]. Hence it is at stake to find methods to bridge this considerable gap. Computational methods for predicting protein secondary and tertiary structure have persistently tried to fill it. In this paper, we explore the ability of a structural alphabet based prediction method to fulfill in part this role.

Since the seminal works by Kabsch and Sander in 1984 [3], one of the most popular and rewarding computational method to predict and analyze protein structures is by breaking them down to their constituent parts in the so-called fragment-based approach. Multiple fragment libraries have been developed so far and they differ in the number of fragments, the length of the fragments, the methods used for clustering and the criteria used for clustering. The first fragment library was developed by Unger and co-workers [4]. There are reviews that give a good overview of the different fragment libraries developed since then [5,6]. Also referred to as structural alphabets (SAs), these have shed some light on the sub-secondary structure level intricacies in proteins [7]. By identifying redundant structural fragments found in proteins, structural alphabets help in abstracting protein structures accurately. Such collections of fragments have been used in information theory based methods that attempt to predict secondary structure (see for example [8] and to reconstitute protein structures [9–11].

In that respect, a SA called *protein blocks* (PBs) was developed for the purpose of describing and predicting the local backbone structure of proteins [12,13]. This SA accounts for all local backbone conformations in protein structures available in the Protein Data Bank (PDB). Since then, PBs have been used in various applications [12]: for structural motif identification [14–16], structural alignments [17,18] and fold recognition [19,20]. There have also been various efforts to use PBs to predict protein local structure. These approaches are based on the Bayes theorem [10,13] support vector machines [21–23] and neural networks [24]. Some of these methods have used prior predictions of classical three state secondary structures (svmPRAT [23] uses YASSPP [25], SVM-PB-Pred [22] uses GOR [26] and Etchebest et. al. [27] explored the utility of PSI-PRED [28]) and sequence alignment profiles like position specific scoring matrices (PSSMs) are used by LOCUSTRA [21], SVM-PB-Pred [22] and Dong and coworkers methodology [24]. The currently available web-based tools that can predict local structure in terms of protein blocks are LocPred [29] and SVM-PB-Pred [22]. The former implements a Bayesian methodology and the latter is SVM-based.

In this work we describe PB-kPRED, a fragment and knowledge-based approach to predict local backbone structure of proteins in terms of protein blocks and a web-based tool that implements the method. In essence, it takes no other inputs than the amino acid sequence of a query and interrogates a database of pentapeptides extracted from protein structures, relying on evolutionary information only when available, hence useful in the context of proteins without any known structural homologues. It returns the predicted local structures of the polypeptide chain in the form of a sequence of protein blocks. Very importantly, PB-kPRED also implements a scoring function that efficiently auto-evaluates the quality of the prediction.

Results

PENTAdb, a database of pentapeptides from protein structures

A total of 68.84 million pentapeptides obtained from the 0.26 million protein chains and their corresponding local structure represented as one of the 16 PBs were obtained and stored in a dedicated database named PENTAdb. Out of the total number of theoretically possible 3.2 million (20⁵) pentapeptides, 2.26 million (70.9%) are found in PENTAdb. These 2.26 million pentapeptides occur in different counts amounting to a total of 68.84 million pentapeptides. PENTAdb had 8 different threshold of sequence identity, and can be directly used by PB-kPRED (see Table 1). A 32-fold decrease (from 68.62 to 5.13 million) in the number of pentapeptides in PENTAdb is observed when PDB chains not sharing more than 30% sequence identity are considered. Nonetheless, the decrease in the number of unique pentapeptides present in PENTAdb is only of 1.3 fold at this threshold.

Shown here are the total number of pentapeptides and unique pentapeptides for the full PDB and for subsets of PDB filtered at different sequence identity cut-off values.

Not all possible tri-PB combinations are observed in known protein structures

Out of all the theoretically possible 4,096 (16³) tri-PBs, a total of 1,375 (i.e 33.5%) were never observed in a non-redundant subset of PDB filtered at 30% sequence identity, subsequently termed as PDB30 dataset (see <u>Methods</u> section). Likewise, out of all the 1.04 million (16⁵) theoretically possible penta-PB motifs, only 40,130 (3.8%) were observed in the PDB30 dataset. These results are indicative of the possibility that many combinations of three or five consecutive PBs are stereochemically unfavorable. The distributions of penta-PB motifs at other sequence identity cut-offs i.e. 40%, 50%, 70%, 90%, 95%, 100% and the entire PDB were also computed (see Table A in <u>S1 File</u>). Towards higher sequence identity cut-offs, a continuous increase in the penta-PB coverage is observed, but it comes at the price of the addition of redundant data. Still the entire PDB covered less than 10% of the total penta-PB space. The penta-PB frequency table derived from the PDB30 dataset was used to develop a scoring function (see <u>Methods</u> section), further termed as accuracy score, even if it contained only 40,130 penta-PB motifs. This is a small fraction but was sufficient to efficiently score PB sequences (see below).

Completeness of PENTAdb for knowledge-based prediction

A quantitative assessment of how often the correct PB can be found in the list of all possible PBs reported for every query protein was performed. This assessment gives the highest

| Seq. identity cut-off values (%) | Total number of chains | Total number of unique pentapeptides | Total number of pentapeptides |
|----------------------------------|------------------------|--------------------------------------|-------------------------------|
| <30 | 24,564 | 1,742,890 | 5,126,423 |
| <40 | 28,590 | 1,881,813 | 6,153,846 |
| <50 | 32,588 | 1,985,203 | 7,074,571 |
| <70 | 37,741 | 2,095,663 | 8,336,277 |
| <90 | 42,594 | 2,148,064 | 9,351,888 |
| <95 | 44,714 | 2,157,239 | 9,768,748 |
| <100 | 64,129 | 2,189,924 | 14,791,285 |
| Full PDB | 274,920 | 2,268,307 | 68,621,454 |

Table 1. Content of the different subsets of PDB in terms of pentapeptides accessible to PB-kPRED.

https://doi.org/10.1371/journal.pone.0186215.t001

accuracy attainable for a query protein using the proposed knowledge-based approach. To this end, for every query protein sequence, different portions of the pentapeptide database were made accessible to the prediction algorithm. This was possible thanks to the hierarchical clustering at different sequence identity levels by the BLASTClust algorithm [30]. PDB30 dataset was used to benchmark the approach. For each of the 15,544 unrelated query protein sequences of this dataset, only pentapeptides coming from a subset of the PDB that shared sequence identities below an indicated cut-off values (from 30% to 100%) and excluding the query itself were used by PB-kPRED (see Table 1 for size of the database for each subset). The results of the predictions are detailed in Table 2. At 30% sequence identity cut-off, the correct PB was found in the list of all possible PBs for 71.4% of the case and the success rate increased to 77.3% when only "homologues" sharing 100% sequence identity to the queries were filtered out. When full PDB was used (but excluding the query) as a database, the percentage times the correct PB is in the list of all possible PBs topped to 99.93% (see Table B in S1 File).

Shown is the percentage of pentapeptide queries for which the correct local conformation was found in the list of all possible PBs reported by the PB-kPRED algorithm after querying PENTAdb. A total number of 15,544 query proteins not sharing more that 30% sequence identity (PDB30 dataset) was used in this assessment.

Prediction accuracies

Two methods, further termed as *majority rule method* and *hybrid method*, were explored to predict the optimal PB sequence within the list of all the possible PBs obtained after querying the database (see Methods section for details). The average prediction accuracies for the PDB30 query proteins using the *majority rule method* and the *hybrid method* using the classic scheme for querying the database are given in Table 3. When homologues sharing \geq 30% sequence identity with each of the queries were removed from the database, PB-kPRED performed with an average accuracy of 39.2% and 40.8% for the *majority rule method* and *hybrid method* and *hybrid method* and *hybrid method* and *hybrid sequence* identity (Table 3). Surprisingly, the effect of enlarging the database to include closer homologues sharing <95% sequence identity with the queries improved only marginally the prediction accuracies reaching on average 40.4% and 42.4% for *majority rule method* and *hybrid method* respectively. Accuracy topped to 58.0% and 54.6% respectively when full PDB (excluding the query itself) was used as database for prediction. This overall gain in accuracy is due to an incremental increase of accuracy across all the 16 PBs.

Shown are the accuracies for the PDB30 dataset using both the *majority rule method* and *hybrid method*. For each of the 15,544 query protein sequences, the portion of PENTAdb accessible for prediction was dynamically determined using MySQL queries: only pentapeptides

| Sequence identity cut-off values (%) | Correct PB found in the list of all possible PBs (%) |
|--------------------------------------|--|
| <30 | 71.4% |
| <40 | 71.5% |
| <50 | 71.6% |
| <70 | 71.9% |
| <90 | 72.5% |
| <95 | 72.8% |
| <100 | 77.3% |
| FULL PDB (excluding query) | 99.93% |

Table 2. Assessment of the richness of PENTAdb towards knowledge-based prediction of protein backbone in terms of protein blocks.

https://doi.org/10.1371/journal.pone.0186215.t002



| Experiment number | Sequence identity cut-off values (%) | Majority rule method | Hybrid method without noise filtering | |
|----------------------|--------------------------------------|----------------------|--|--|
| A1 | FULL PDB (excluding query) | 58.0%±12.9 | 54.6%±22.0 | |
| A2 | <100 | 44.0%±14.7 | 48.0%±18.7 | |
| A3 | <95 40.4%±12.7 | | 42.4%±16.1 | |
| A4 | <90 | 40.1%±12.5 | 42.0%±15.9 | |
| A5 | <70 | 39.7%±12.5 | 41.4%±15.8 | |
| A6 | <50 | 39.4%±12.5 | 41.0%±15.9 | |
| A7 | <40 | 39.3%±12.5 | 40.9%±15.9 | |
| A8 | <30 | 39.2%±12.5 | 40.8%±15.9 | |

Table 3. Evaluation of performance of PB-kPRED knowledge-based approach to predict local conformations of protein backbone in terms of protein blocks.

https://doi.org/10.1371/journal.pone.0186215.t003

coming from protein chains in PDB that shared sequence identities below the indicated cut-off values were accessible to PB-kPRED for prediction of local structures in terms of PBs.

To improve the prediction rates, the *hybrid method* was tested using the *noise filtering scheme* whereby, for each query pentapeptide, data in PENTAdb coming from closest homologues was queried in preference to data from more distant homologues (see also Text B in <u>S1</u> File). Results are detailed in <u>Table 4</u>. When compared to the *without noise filtering scheme* (<u>Table 3</u>), the prediction rates improved to reach a maximum of 66.3%. Interestingly, for experiments B2 to B7 where closest homologues to be queried first are in the range of <40% to <95% sequence identities, the prediction accuracies remained high at a level of about 61.6%. Only in experiment B8 the prediction accuracy rate dropped to 40.8%. It must be noticed that this experiment is identical to the one featured for <30% threshold (see <u>Table 3</u>) using the *hybrid method* and the *without noise filtering scheme* for querying the database.

For each query protein, the portion of the database accessible to the algorithm is first restricted to the closest homologues and if no hits were found, only then the more distant homologues are made accessible progressively. Eight results shown here correspond to the eight experiments described further below (see <u>Methods</u> section). Shown are the prediction rates averaged over 15,544 query proteins from the PDB30 dataset that was used in this assessment.

All results further detailed hereafter are concerned with data obtained in experiment B1 where *hybrid method* was applied using the *noise filtering scheme* for querying PENTAdb and where best predictions were obtained.

The distribution of the prediction accuracies for experiment B1 (<u>Table 4</u>) is bimodal (Fig 1). A spike in frequency is observed at the >80% range representing the set of queries which

| Experiment | Closest homologues to be queried first | Average prediction rate (%) | | | | | |
|------------|--|-----------------------------|--|--|--|--|--|
| B1 | 100 % | 66.31±27.62 | | | | | |
| B2 | <100 % | 61.61±24.50 | | | | | |
| B3 | <95 % | 61.60±24.49 | | | | | |
| B4 | <90 % | 61.59±24.48 | | | | | |
| B5 | <70 % | 61.59±24.48 | | | | | |
| B6 | <50 % | 61.59±24.47 | | | | | |
| B7 | <40 % | 61.58±24.47 | | | | | |
| B8 | <30 % | 40.79±15.90 | | | | | |

Table 4. Assessment of the performance of PB-kPRED using the *hybrid method with noise filtering scheme* for guerying the database.

https://doi.org/10.1371/journal.pone.0186215.t004





https://doi.org/10.1371/journal.pone.0186215.g001

have closely related proteins of known structure available in the PDB and for which the method is able to perform extremely well. Many queries in this set indeed had prediction accuracy values above 90%. At the other end of the spectrum, there is an almost normal distribution with an average around the 35%-40% accuracy range. Hence, the mean falls in between these two at 66.31% accuracy. This distribution did not substantially vary when homologues sharing less than 100% to 40% sequence identity to the query corresponding to experiments B2 to B7 respectively were queried first (data not shown). However, once the twilight zone of 30% sequence identity is crossed, the accuracy distribution drastically changes to that of a unimodal distribution with a very sharp peak at the 40% range and gradually tapering tail towards the higher accuracies (see Fig A in S1 File).

The accuracy by the *hybrid method* using the *noise filtering scheme* was compared to the *majority rule method* (Fig 2). As shown by the data points below the diagonal, the *hybrid method* performed significantly better than the *majority rule method* for a total of 8,195 cases (52.7%) out of the 15,544 protein queries. For remaining 7,245 cases, the *majority rule method* performed slightly better than the *hybrid method*.





https://doi.org/10.1371/journal.pone.0186215.g002

PB predictions

Results from the best performing condition (experiment B1 featured in Table 4) were further analyzed for the PB-wise prediction rates and compared with published rates from other methods (Table 5). The rates are heterogeneous across the 16 PBs. Top two best-predicted PBs by PB-kPRED were PB *m* and PB *a*, with accuracies of 75.9% and 67.2%, respectively. On the other hand, the two most badly predicted PBs by PB-kPRED were PB *j* and PB *g* with prediction rates of 49.9% and 43.5% respectively. Analysis of the corresponding confusion matrix (Table C in S1 File) shows that, the prediction algorithm frequently gets confused between the PBs *c* and *d*. PB *c* is wrongly predicted as PB *d* almost 31,000 times (22.4%). The vice-versa, PB *d* being predicted as PB *c* is more than 42,000 times (16.9%). These PBs are in fact highly related (i) as seen from a pure structural point of view, low Root Mean Squared Deviation on Angular values (RMSDA) and similar transitions [28] and (ii) as they have been seen to be highly interchangeable thanks to PB substitution matrix [31,32]. As some PBs are highly similar, it is possible to relax the assessment, *i.e.* considering two PB series as equivalent. With such



| PBs | | Majority method | Hybrid method | | | | | Other methods | | |
|-----|--------------|-----------------|---------------|------------|------------|--------|-------------|---------------|------------|--------------|
| | PB frequency | Expt A8 | Expt B8 | Expt B4 | Expt B2 | | Expt B1 | | LOCUSTRA | Bayes method |
| | | | Αςςι | ıracies | | | Specificity | мсс | Accuracies | |
| а | 3.68% | 34.40% | 45.93% | 64.60% | 64.60% | 67.20% | 98.15% | 0.69 | 58.16% | 56.60% |
| b | 4.29% | 15.19% | 18.99% | 44.52% | 44.58% | 52.15% | 97.72% | 0.56 | 26.14% | 20.90% |
| С | 8.31% | 23.45% | 28.76% | 51.62% | 51.66% | 58.53% | 95.95% | 0.58 | 44.81% | 32.90% |
| d | 18.68% | 39.24% | 40.09% | 61.83% | 61.85% | 67.00% | 94.12% | 0.63 | 71.58% | 54.00% |
| е | 2.18% | 21.78% | 28.59% | 52.30% | 52.38% | 57.45% | 98.96% | 0.62 | 44.74% | 38.60% |
| f | 6.45% | 24.87% | 29.49% | 54.64% | 54.64% | 60.30% | 97.21% | 0.61 | 41.45% | 30.90% |
| g | 1.14% | 10.33% | 14.94% | 36.79% | 36.87% | 43.45% | 99.19% | 0.51 | 26.84% | 30.10% |
| h | 2.10% | 24.25% | 33.02% | 56.90% | 56.89% | 61.05% | 98.82% | 0.64 | 38.45% | 40.90% |
| i | 1.49% | 20.69% | 30.45% | 55.36% | 55.35% | 59.17% | 99.18% | 0.63 | 36.87% | 38.10% |
| j | 0.83% | 15.48% | 20.81% | 42.57% | 42.68% | 49.98% | 99.40% | 0.56 | 48.19% | 49.70% |
| k | 5.25% | 30.46% | 35.59% | 59.30% | 59.32% | 63.93% | 97.67% | 0.65 | 46.46% | 33.40% |
| 1 | 5.20% | 26.56% | 31.88% | 54.87% | 54.93% | 59.99% | 97.69% | 0.62 | 42.71% | 35.50% |
| m | 32.89% | 60.96% | 55.68% | 72.38% | 72.40% | 75.89% | 91.03% | 0.67 | 83.76% | 70.60% |
| n | 1.78% | 26.48% | 35.40% | 58.16% | 58.21% | 62.15% | 99.05% | 0.65 | 52.08% | 50.00% |
| 0 | 2.44% | 29.62% | 38.40% | 59.49% | 59.52% | 63.19% | 98.70% | 0.66 | 55.10% | 48.10% |
| р | 3.29% | 23.22% | 31.72% | 54.27% | 54.28% | 59.24% | 98.25% | 0.62 | 40.80% | 29.20% |

Table 5. Assessment of the performance of PB-kPRED and comparison with other previously reported methods.

https://doi.org/10.1371/journal.pone.0186215.t005

relaxed criteria, the accuracy increases from 66.31% to 68.87% (a 2.56% gain on average). Interestingly, significant increases in accuracies were observed for PB g (from 43.5% to 67.4%) and for PB j (from 49.98% to 67.2%).

Shown are the PB-wise prediction accuracies for experiment A8 of the majority method and four different experiments of *hybrid method* with noise filtering. These are compared with PB-wise results from LOCUSTRA [20] and Bayesian approach [26]. Experiment B1 PB-wise accuracies were compared with the other two methods and corresponding cell values in bold represent the best accuracy achieved between the experiment B1 of *hybrid method*, LOCUS-TRA and Bayes method.



Fig 3. Assessment of the ability of the scoring function to estimate the prediction accuracy of PB**kPRED.** (a) Score versus accuracy and (b) Scores for predicted PB versus real PB sequences for the 15,544 query proteins of PDB30 dataset. Data points are coloured based on level of accuracy of predictions.

https://doi.org/10.1371/journal.pone.0186215.g003



| | Queries wi | th known homologues in | Queries with no known homologues in PDB | | | |
|----------------------------------|---|---|---|---|--|--|
| Sequence identity thresholds (%) | 2HX0_A (hypothetical DNA binding protein) | 4HUQ_T (energy-coupling factor transporter EcfT) | 2HXV_A (deminase/ reductase) | 1A27_A (hydroxysteroid dehydrogenase) | 4HZU_S (transmembrane protein associated with Ecf transporter) | |
| 100 | 2 | 3 | 1 | 1 | 1 | |
| 95 | 2 | 3 | 1 | 1 | 1 | |
| 90 | 2 | 3 | 1 | 1 | 1 | |
| 70 | 2 | 3 | 1 | 1 | 1 | |
| 50 | 2 | 3 | 1 | 1 | 1 | |
| 40 | 2 | 5 | 1 | 1 | 1 | |
| 30 | 2 | 5 | 10 | 1 | 1 | |
| Accuracy (%) | 100% | 73.41% | 39.13% | 75.44% | 9.37% | |
| Accuracy score | 2.81 | 1.31 | 0.30 | 1.99 | -0.28 | |

Table 6. Impact of the availability of known homologues on the accuracy of PB-kPRED.

https://doi.org/10.1371/journal.pone.0186215.t006

PB-kPRED globally outperformed two other PB prediction methods (Table 5). Its predictions were better for all the 16 PBs when compared to the Bayes method and better than almost all PBs when compared to LOCUSTRA. Only PBs *d* and *m* were better predicted by this latter method²⁰. kPRED predictions were also evaluated against secondary structure assignments from DSSP. This is not a straightforward activity as the 16 PBs are not a categorization of the secondary structure elements. Nonetheless, the results of the comparison are presented in Table D in S1 File and the results are favorable when compared to popular secondary structure prediction algorithms, at 80.94%. It is important to note that, secondary structure is useful mainly for a good description of repetitive structures, i.e. alpha-helix and beta-sheet and not for the remaining approximately 50% of the protein structure that are not defined, namely the coil.

A Mathews correlation coefficient (MCC) close to +1 indicates a good agreement between the observed and the predicted outcomes and a MCC of close to -1 otherwise. For our analysis all the 16 PBs had MCCs between 0.5 and 0.7. PBs *a* and *m* were close to 0.7, PB *g* at 0.51 and the remaining fluctuated around the 0.6 mark. The sensitivity and specificity ranges were 0.4– 0.7 and 0.9–1.0 respectively. A common pattern is observed in the case of PBs corresponding to the regular secondary structure elements (PBs *d* and *m*): in both these cases, the sensitivity values peak while the specificity values plummet. Although the sensitivity values varied between 0.4 and 0.7, the specificity values were consistently above 0.9 indicating that the method was able to achieve a very high true negative rate. High specificity may be explained by the binary nature of the test while sensitivity is a sixteen state test.

Measure of accuracy

A probabilistic scoring function was developed for the *a posteriori* analysis of the predicted PB sequences so as to provide a measure of how accurate the *hybrid method* using the *noise filter-ing scheme* was performing. An assessment of the scoring function is provided in Fig 3. It shows that the score is correlated with the accuracy of the prediction with a Pearson's correlation coefficient of 0.82 (Fig 3A). The two distinct clusters of data points correspond to those featured in the histogram in Fig 1. As a further assessment of the scoring function, the scores for the predicted PB sequences were compared with the scores for the real PB sequences (Fig 3B). It shows that in case of more accurate predictions (rates above 60%), the two scores correlated very well (red points along the diagonal in Fig 3B) with both score values mostly ranging between +1 and +3. In the case of less accurate predictions (rates below 60%), the two scores

were no more correlated (green dots below the diagonal in Fig 3B) and scores for predicted PB sequences ranged mostly between -2 and +1.

Case studies

Here were considered the predictions for 5 specific cases to look at the strengths and limitations of the PB-kPRED algorithm namely in presence or absence of homologues of known structure (see <u>Table 6</u>). These case studies correspond to counter intuitive prediction instances where (i) prediction accuracy is high despite not having any close homologues and (ii) prediction accuracy is low despite having sequences of PBs of closely-related proteins in PENTAdb.

Query PDB chains with known homologues and with no known homologues are featured. The *hybrid method with noise filtering* scheme for querying the database was applied for the prediction whereby the conditions were identical to experiment B1 (see <u>Methods</u> section) as featured in <u>Table 6</u>. The queries themselves were excluded from the database prior to prediction. Shown here are the accuracies of the predictions and the numbers of known homologues for different sequence identity thresholds.

Regarding prediction in employing information from homologues of known structure, three contrasting cases were studied. The first case relates to chain A of a hypothetical DNA binding protein from Salmonella cholera (PDB id 2HX0_A) which has a homologue from Salmonella typhimurium (PDB id 2NMU) that is 100% identical, 100% accuracy was achieved as shown by the high accuracy score of 2.81. Both structures aligned very well with a Root Mean Squared Deviation (RMSD) of 0.14 Å (Figure B in S1 File). The second case relates to an energy-coupling factor transporter transmembrane protein EcfT from Lactobacillus brevis (PDB id 4HUQ_T) which has two other "homologues" (PDB id 4RFS_T and 4HZU_T) that are 100% identical to the query. Here, the prediction accuracy is 73.4% only with an accuracy score of 1.31. 3-D structural alignment with these two "homologues" resulted in RMSDs 1.41 Å and 1.90 Å respectively displaying some structural variations (Figure B in <u>S1 File</u>) despite being 100% identical at the amino acid sequence level. These structural variations were due to rigid body movement. The third case is chain A of a pyrimidine deaminase / uracil reductase from Thermotoga maritima (PDB id 2HXV_A) which had only ten very distantly related proteins that shared less than 30% sequence identity in the PDB. Prediction rate is even lower here with accuracy reaching a value of 39.1% as shown by the low accuracy score of 0.30.

As for predictions in absence of homologues of known structure, two contrasting cases were studied. The first case is about a human hydroxysteroid dehydrogenase and the second case is a membrane protein associated with Ecf transporter from *Lactobacillus brevis*. The prediction performed quite well in the first case with an accuracy of 75.4% as shown by the high accuracy score of 1.99 while in the second case, the prediction almost completely failed with the accuracy of only 9.37% and also shown by the unfavorable accuracy score of -0.28.

Implementation of the PB-kPRED methods as a web-tool

The PB-kPRED methodology has been implemented as a web-tool that is freely available to the community at http://www.bo-protscience.fr/kpred/. Both *majority rule* and *hybrid method without the noise filtering scheme* for querying the database have been implemented. The tool provides a predicted PB sequence for each query amino acid sequence and also provides the *accuracy score* that serves as an *a posteriori* estimation of the prediction accuracy. In case the prediction score value is below -1, the prediction accuracy cannot be estimated and the user is notified. Users can provide multiple query protein sequences. All results are downloadable as FASTA formatted flat files. Optionally when submitting numerous query sequences, the user can provide an email address to which a notification will be sent when the job is completed.

Discussion

The exponential growth in the structural knowledge of proteins has warranted the necessity of competent knowledge-based prediction algorithms for local structure prediction. At the level of short protein segments like pentapeptides, this increase in structural knowledge invariably brings with it an unprecedented signal to noise ratio for deciding on the most probable local conformations. Indeed, it is well established that similar pentapeptides can adopt different local conformations [3,33]. This is verified when the content of PENTAdb is inspected.

Hazout's team along with defining the protein blocks also predicted the local structure in terms of PBs using a Bayesian approach [13]. They achieved an accuracy of 34.4% using a 15-residue window and this increased to 40.7% upon supplementing the Bayesian predictor with sequence profiles in the form of sequence families. In 2005, Etchebest and colleagues [27] used a combination of statistical optimization procedure and improved sequence family data to bump up the accuracy to 48.7%. Incorporating secondary structure predictions from PSI-PRED into the PB prediction process did not contribute much to improve the accuracy *i.e.* only 1% gain resulting in 49.9%. Machine learning techniques have also been used to predict protein local structure in terms of PBs. Support vector machine based methods like LOCUS-TRA [21], symPRAT [23] and SVM-PB-Pred [22] achieve mean accuracies of 61.0%, 67.0% and 53.0% respectively. A dual layer neural network based prediction method achieved 58.5% accuracy [25]. The most refined version of the PB-kPRED method proposed here, i.e hybrid method with noise filtering scheme, outperformed most of the previously developed methods for PB prediction except for svmPRAT where it performed equivalently. Although all the methods evaluated their accuracies on non-redundant sets of proteins, an even comparison is hindered by difference in datasets, varying training regimes for the machine learning methods and different levels of sequence identity used as input in the prediction process. This motivated us to perform a battery of tests on the algorithm to estimate the prediction accuracy when incremental levels of sequence identities are made available in PENTAdb for the prediction (see Table 4). Importantly, to our knowledge, this is the first report of a querying scheme that dynamically filters out, on a per query basis, homologues at different cut-off values so that the portion of the PENTAdb that is made accessible for prediction is calculated on the fly. For each of the 15,544 query sequences of PDB30, 16 experiments were performed, hence amounting to a total dataset size of 248,704 proteins. Using the noise filtering strategy, PB-kPRED was able to efficiently weed out the noise present in the database due to redundancy and hence to narrow down the search in the database to find the most appropriate local structure for a given pentapeptide. Therefore, filtering out from the database the pentapeptides from proteins that shared less than 30% sequence identity with the query indeed improves the prediction efficiency. In general, in those cases where pentapeptides from very close homologues were indeed present in the database, kPRED performed extremely well.

Remarkably, the reported average accuracy (66.31%) is very high considering the fact that for every pentapeptide, there are 16 possibilities. Indeed, if random PB predictions were made taking into account respective PB frequencies, prediction rate (Q16 value) would be very poor with an average value of about 7 to 8%. Similarly, when predicting secondary structure, there are only 3 possibilities and random Q3 prediction accuracies would be of about 30%.

When the *majority method* and the *hybrid method* (Fig 2) were compared, two distinct clusters were noticed. Upon further investigating the reason for this distinct clustering, we note that, irrespective of the sequence identity cut-off, the points below the diagonal were found in more populated clusters while the points above the diagonal were found in least populated clusters Hence the *hybrid method* using the *noise filtering scheme* will perform better when there are some closely-related protein structures to look-up to in PENTAdb. In a real-life

scenario, this will not be always the case. Indeed, proteins for which we want to predict the structure and which do not have any homologues even at 30% sequence identity are not so uncommon. This brings us to the conclusion that even though overall the *hybrid method* performs better, we cannot ignore the *majority rule method* all together.

Nonetheless, this method still has room for improvement as it can be seen from the values in Table 2. The list of all possible PBs reported by the PB-kPRED algorithm after querying PEN-TAdb database indeed shows that the good PB was present in more than 70% of the cases. However, owing to the scoring functions S1 and S2 (see Methods for more details), both *majority rule* and *hybrid methods* failed to pick up these good PBs as predictions in several instances.

Interestingly, once the local backbone of a protein was predicted in the form of a PB sequence, we were able to provide an *a posteriori* assessment of how accurate was the prediction. The method used here to achieve this relied on the simple idea that successions of PBs should follow the rule that not all combinations of PBs would be allowed. This intuition turned out to be correct since there was a remarkable correlation between the score and the accuracy of the predictions. Noteworthy, the *accuracy scores* for actual (native) PB sequences are overwhelmingly distributed between +1 and +3, while poorly predicted PB sequences have scores below +1. This scoring of PB sequences could also serve as an indicator towards improving predictions. Because the calculation of the score of a PB sequence is very fast, one could imagine implementing a score-guided optimization procedure to climb the prediction accuracy gradient using Monte-Carlo or genetic algorithms for example.

The case studies documented in this work (Table 6) indicate that the relationship between local structure predictability and the number of homologues of the query available in PDB are not very straightforward. Optimistically, in spite of not having any homologues, the PBkPRED algorithm can perform a good prediction if the pentapeptides constituting the query adopt consensus local structures for the respective pentapeptides. Two such examples were provided but with contrasting outcomes, one achieving good accuracies and the other failing to predict correctly the PB sequence. Interestingly, the accuracy scores provided by our scoring function helped to reliably differentiate one prediction from the other. On the other hand, even if a query has multiple homologues in the PDB, its prediction accuracy will take a hit if the homologues are contrasting structural analogues of the query. For example, the activation of human pancreatic lipase involves considerable conformational transition in the form of a 'lid movement'. The hypothetical prediction case when the query is the 'lid open form' and PENTAdb has pentapeptides from the 'lid closed form' would confuse the prediction algorithm despite both the forms of lipase being identical in amino acid sequences. Hence these case studies establish two take home messages: (i) there are exceptions to the general observation that the presence of homologues improves the prediction accuracy of PB-kPRED and (ii) the accuracy score used to evaluate the predictions is a reliable gauge for estimating the accuracy of the method as illustrated in Fig 3.

PB-kPRED web-server could form a vital link in the pipeline of PB based structure analysis tools. Namely, it can be bridged with PB-based fast structure comparison tools like iPBA [18] and PB-ALIGN [17] and help to mine for similar structures and map the fold space. It can also be used to predict the occurrence of structural motifs in protein sequences. Indeed, the alpha version of the server which was made available on-line earlier, has already been used by some research groups for the structural characterization of RNA binding sites in protein structures and predicting proteins sequences that contain RNA binding sites [34,35] and also in predicting β -turns and their types [36].

The web-based tool currently does not feature the *hybrid method with noise filtering scheme* because it would require running an instance of BLASTClust on every query. We plan to implement this functionality in a future version of the tool.

Our objective in this work was to predict protein backbone represented as protein blocks. We believe that the assembly of protein blocks fragments can ultimately lead to 3D structure prediction using sophisticated algorithms like the one implemented in Rosetta. Indeed, Rosetta achieves this by leveraging on the I-Sites fragment libraries developed by Bystroff and Baker [10]. These fragment libraries, including PB-based fragment libraries, can be of great interest in the research of protein fragments in the twilight zone when doing 3D-modeling. We are currently developing a method and tool towards this end that takes advantage of existing algorithms implemented in Modeller [37] and Rosetta [10].

Methods

Dataset

All the protein chains from PDB [1] were segregated into clusters culled at 30% sequence identity using the BLASTClust algorithm [30] resulting in a collection of 15,544 clusters. The dataset comprises of 15,554 protein chains each corresponding to the best representative structure available from each of these clusters. It is hereafter termed as "PDB30 dataset". Preference was given to crystallographic structures over NMR and electron microscopy structures and also preferring better resolution and lowest R-value structures. Out of these 15,544 structures, 14,207 are crystallographic structures, 1,128 are from NMR experiments and 209 are solved by electron microscopy. Further, chains smaller than 100 residues were filtered out. We preferred to keep the NMR and EM structures, as we wanted to investigate if the experimental method impact on the quality of the predictions. For each of these 15,544 proteins the subsets of PDB that were homologous at 30%, 40%, 50%, 70%, 90%, 95% and 100% as reported by BLASTClust were also calculated in order to implement the "*Hybrid method with noise filtering*" scheme.

Protein blocks

The set of protein blocks (PBs) is a structural alphabet composed of 16 structural prototypes each representing backbone conformation of a fragment of 5 contiguous residues [12,13]. The 16 PBs are represented by the letters *a* to *p* and were identified from a collection of 228 nonredundant proteins. Clustering these pentapeptides was based on the 8 dihedral angles (ψ_{i-2} , φ_{i-1} , ψ_{i-1} , φ_{i} , ψ_{i} , φ_{i+1} , ψ_{i+1} , φ_{i+2}) that define their local backbone conformation. An unsupervised learning algorithm (Kohonen algorithm [38]) was used to arrive to an unbiased classification of the dihedral vectors and to the definition of standard dihedral angles for each PB. Protein blocks are assigned on the basis of the dissimilarity measure called RMSDA between observed dihedral angles and the standard dihedral angles for the 16 PBs. The PB with lowest RMSDA is assigned to the central residue of the pentapeptide region. The choice of fragment size as 5 and library size as 16 for the PBs was because 5 consecutive residues capture well the local contacts in regular secondary structures (α -helices and β -strands) and 16-library size is a good balance between the specificity and sensitivity of predictions [13].

All the 15,544 protein chains from PDB30 dataset were encoded into their corresponding protein blocks sequences (PB sequences) after comparing their backbone φ and ψ torsion angles with the corresponding standard torsion angles for the 16 PBs [12] using an in-house developed Perl script. Sequence of PBs as observed in crystal and NMR structures were later used as a reference to assess the accuracy of predicted PB sequences.

Database of pentapeptide conformations from protein structures

A database of pentapeptide conformations (PENTAdb) was developed using known 3-D structures of proteins. PENTAdb is essentially the entire structural information contained in the PDB, broken down into chunks of pentapeptides. A sliding window of 5 residues was used to extract structural features for every overlapping pentapeptide of a polypeptide chain. The dihedral vector associated with the five consecutive residues that is required to assign PBs as described in the previous section was obtained from the DSSP [39] program. All the information was stored as a MySQL relational database. PENTAdb is maintained up-to-date; the update frequency corresponds to the weekly updates of PDB. The protein chains from which the pentapeptides are extracted are filterable at 30%, 40%, 50%, 70%, 90%, 95% and 100% sequence identity thresholds.

Prediction scheme

The overall scheme for predicting the local structure in terms of PBs is based on querying the PENTAdb database for every constitutive pentapeptides of a query protein sequence using a sliding window of 5 residues (Fig 4A). Hits from the database are reported as predicted protein blocks (PBs). Predicted PBs are assigned to the central residue of each query pentapeptide. The prediction results are presented at different levels of refinement. The prediction in the coarsest form consists of the list of all the possible PBs for a particular pentapeptide of the query protein sequence. This is the case when multiple hits from PENTAdb database are obtained for a particular query pentapeptide (Fig 4B). The multiple hits correspond to the different conformations, which the pentapeptide has been seen to adopt in protein structures (Fig 4B). When the query pentapeptide is not found in PENTAdb, the information available for the tetrapeptides covering the first four residues with a wildcard for the fifth position was used (Fig 4B) to identify the list of possible PBs with first 4 amino acid residues matching this query. The position of wildcard did not influence the outcome of the results (data not shown). The list of hits thus obtained is referred as *all possible PBs*. This list serves as a framework from which the most probable PB sequence is predicted.

Two methods were explored to predict the optimal PB sequence within the list of all the possible PBs obtained after querying the database (Fig 5). The first method, termed as *majority*



Fig 4. The knowledge-based methodology behind PB-kPRED. (a) Overview of the scheme followed by PB-kPRED for the prediction process. (b) The different outcomes possible when PENTAdb database is queried for a pentapeptide sequence: hits are reported as a single PB or multiple PBs.

https://doi.org/10.1371/journal.pone.0186215.g004



max (S1), here PB = a

PLOS ONE

Fig 5. Details of the scoring schemes underlying the *majority rule method* and the *hybrid method*. S1 scores are simply the raw counts of all possible PBs reported by PENTAdb database for a given query pentapeptide. S2 scores are calculated through the summation of the odds of tri-PBs that have a common PB in the central position. For the majority rule method, predictions are based only on the ranking of the S1 scores. For the *hybrid method*, predictions are based on the ranking of the product of scores S1 and S2.

https://doi.org/10.1371/journal.pone.0186215.g005

rule method, is purely probabilistic and consists of simply picking up the most frequently observed PB for each query pentapeptide. It corresponds to the PB that has highest S1 score, where S1 scores are simply the raw counts of all possible PBs reported by PENTAdb database for the query pentapeptide. In cases when there is no decisive majority (two or more equiprobable PB), both of them are reported as predictions.

However, it is known that the structure adopted by a short peptide can be highly dependent on its local environment³. A second method that integrates contextual information was hence developed and is hereafter termed as *hybrid method*. Here, to predict the local structure of a pentapeptide, the information about the structural status (in terms of PBs) of the two immediately adjacent and overlapping pentapeptides (preceding and succeeding) is also taken into account (Fig 5). It requires a normalized frequency look-up table for observed motifs of 3 consecutive PBs also termed as tri-PBs (see Text A in S1 File for more details). For each query pentapeptide, in complement to the calculated S1 score, an additional S2 score is calculated as follows. A list of all possible combinations of three successive PBs (tri-PB motifs) is built. This is derived from the list of *all possible PBs* for the query pentapeptide and for its two adjacent pentapeptides (Fig 5). For each possible tri-PB motif, their normalized frequencies ("odds" in Fig 5) are looked up in the tri-PB normalized frequency table. S2 scores are calculated through the summation of the odds of tri-PB motifs that have a common PB in the central position (Fig 5). The predicted PB for the query pentapeptide is determined after multiplying S1 scores by their corresponding S2 scores and taking the highest value among these products (Fig 5). This approach is called the *hybrid method* because it combines the *majority rule method* with contextual information in the prediction process.

Evaluating PB-kPRED using different subsets of PENTAdb

Two evaluation schemes were developed to benchmark the PB-kPRED methodology. As mentioned above, the query dataset used here constituted of the 15,544 proteins from the PDB30 dataset. The schemes relied on the ability to control which subsection of PENTAdb will be accessible to the prediction algorithm for every query. For example, allowing only pentapeptides in PENTAdb from non-homologues to be accessible by the prediction algorithm emulates a scenario of attempting to predict the local structure of a protein with no known homologues. On the other hand, as in the case of other local structure prediction methods [13, 21–23, 27], it can be advantageous to have the ability to privilege information from homologous structures when these are available to predict the local structure of a query protein. Such a scheme can be emulated by allowing only pentapeptides in PENTAdb from closest detectable homologues to be accessible by the search algorithm.

In first instance, the prediction methodology was assessed with increasing sequence identity cut-offs ranging from 30%, 40%, 50%, 70%, 90%, 95% to 100%, named experiments A1-A8 (Fig 6A). This scheme is subsequently termed as "*without noise filtering scheme*". In second instance, an alternative assessment scheme hereby called the "*with noise filtering scheme*" was applied to further assess the PB-kPRED methodology (experiments B1-B8, see Fig 6B). It aimed at evaluating how privileging information from close homologues, when available, contributed to improve the quality of the predictions. In brief, the algorithm initially searches for a pentapeptide among the closest homologues first. If the search finds a hit, then the hit is used for the prediction; otherwise the search space is increased to include the immediately next level of more distantly related homologues. This process is repeated until a hit is obtained. Due to this process of introducing more distant homologues in a conditional fashion, wrong pentapeptides (noise) from PENTAdb were potentially filtered out, hence the name *with noise filtering scheme*. In all the cases, care was taken to exclude the pentapeptides from the query proteins themselves.

Reducing the PB predictions into a binary outcome permits the use of classical Mathews correlation coefficient (MCC) to compare our predictions to a random choice. MCCs for the 16 PBs were evaluated based on a confusion matrix similar. For each PB, MCC was calculated according to Eq 1.

$$MCC = \frac{TP * TN - FP * FN}{\sqrt{(TP + FP)(TP + FN)(TN + FP)(TN + FN)}}$$
(1)



Fig 6. Diagrammatic representations of the schemes used by PB-kPRED for querying PENTAdb with (a) representing the so-called "classic" or "without noise filtering scheme" and (b) representing the "with noise filtering scheme". Sections of the database accessible are indicated in green and those not accessible in red. The sections are delimited by sequence identity thresholds. In both schemes, eight different experiments (A1 to A8 and B1 to B8) were performed. See Text B in S1 File for detailed legend of this figure.

https://doi.org/10.1371/journal.pone.0186215.g006

A scoring function to estimate the accuracy of the predictions

A probabilistic scoring function was developed for the *a posteriori* analysis of the predicted PB sequence through namely the analysis of its content in penta-PB motifs, with the objective of providing a measure of how accurate PB-kPRED was performing. The principle of the analysis relies on the fact that not all penta-PBs are found in proteins at the same frequency. Indeed, many successions of 5 consecutive PBs are highly improbable because they are geometrically not allowed as explicated by the Ramachandran rules. The probabilistic function is hence based on the look-up table of normalized frequencies of successive penta-PB motifs observed in a non-redundant set of protein structures (see Text A in S1 File). In brief, using a sliding window of 5 consecutive PBs (penta-PB motif) along the predicted PB sequence, the normalized frequencies of all penta-PB motifs were looked-up in the penta-PB frequency table. The logarithm of these normalized frequencies were then summed and divided by the length of the predicted PB sequence to generate an *accuracy score* (*A*) as shown here:

$$A = \frac{\sum_{i=1}^{l-4} \log(N_i)}{l} \tag{2}$$

where *A* is the *accuracy score* for a predicted PB sequence, *l* is the length of the PB sequence, *N* is the normalized frequency of the penta-PB motif observed at window position *i* in the PB sequence. Since an overlapping sliding window of five consecutive PBs is used, the total number of penta-PB motifs (*i.e* the number of windows) is *l*-4. In the case a particular penta-PB motif has a null value in the frequency table (*i.e* it is never observed), a penalty of -5 was instead added to the score.

Supporting information

S1 File. Supplementary tables, figures and text. (PDF)

Acknowledgments

Authors thank Tristan Rialland and Sara Bachiri for providing technical support in the development of the PB-kPRED web server.

Author Contributions

Conceptualization: Iyanar Vetrivel, Manoj Tyagi, Alexandre G. de Brevern.

Data curation: Lionel Hoffmann.

Formal analysis: Iyanar Vetrivel, Swapnil Mahajan.

Funding acquisition: Frédéric Cadet.

Investigation: Iyanar Vetrivel, Swapnil Mahajan.

Methodology: Iyanar Vetrivel, Swapnil Mahajan, Yves-Henri Sanejouand, Narayanaswamy Srinivasan, Alexandre G. de Brevern, Bernard Offmann.

Project administration: Frédéric Cadet.

Resources: Frédéric Cadet.

Software: Swapnil Mahajan, Manoj Tyagi, Lionel Hoffmann.

Supervision: Narayanaswamy Srinivasan, Frédéric Cadet, Bernard Offmann.

Validation: Narayanaswamy Srinivasan, Alexandre G. de Brevern, Bernard Offmann.

Writing - original draft: Iyanar Vetrivel, Bernard Offmann.

Writing – review & editing: Swapnil Mahajan, Yves-Henri Sanejouand, Narayanaswamy Srinivasan, Alexandre G. de Brevern, Frédéric Cadet.

References

- Rose PW, Prlić A, Bi C, Bluhm WF, Christie CH, Dutta S, et al. The RCSB Protein Data Bank: views of structural biology for basic and applied research and education. Nucleic Acids Res. 2015; 43:D345– D356. https://doi.org/10.1093/nar/gku1214 PMID: 25428375
- Consortium Uniprot. UniProt: a hub for protein information. Nucleic Acids Res. 2015; 43:D204–D212. https://doi.org/10.1093/nar/gku989 PMID: 25348405
- Kabsch W, Sander C. On the use of sequence homologies to predict protein structure: identical pentapeptides can have completely different conformations. Proc Natl Acad Sci USA. 1984; 81:1075–1078. PMID: 6422466
- Unger R, Harel D, Wherland S, Sussman JL. A 3D building blocks approach to analyzing and predicting structure of proteins. Proteins. 1989; 5:355–373. https://doi.org/10.1002/prot.340050410 PMID: 2798411
- Karchin R, Cline M, Karplus K. Evaluation of local structure alphabets based on residue burial. Proteins. 2004; 55:508–518. https://doi.org/10.1002/prot.20008 PMID: 15103615
- Offmann B, Tyagi M, de Brevern AG. Local Protein Structures. Current Bioinformatics. 2007; 33:165– 202.
- Tyagi M, Bornot A, Offmann B, de Brevern AG. Protein short loop prediction in terms of a structural alphabet. Comput Biol Chem. 2009; 33:329–333. https://doi.org/10.1016/j.compbiolchem.2009.06.002 PMID: 19625218
- Cheng H, Sen TZ, Jernigan RL, Kloczkowski A. Consensus data mining (CDM) protein secondary structure prediction server: Combining GOR V and fragment database mining (FDM). Bioinformatics. 2007; 23(19): 2628–2630. https://doi.org/10.1093/bioinformatics/btm379 PMID: 17660202
- 9. Bystroff C, Simons KT, Han KF, Baker D. Local sequence-structure correlations in proteins. Curr Opin Biotechnol. 1996; 7:417–421. PMID: 8768900
- Simons KT, Kooperberg C, Huang E, Baker D. Assembly of protein tertiary structures from fragments with similar local sequences using simulated annealing and Bayesian scoring functions. J Mol Biol. 1997; 268:209–225. https://doi.org/10.1006/jmbi.1997.0959 PMID: 9149153
- Kolodny R, Koehl P, Guibas L, Levitt M. Small libraries of protein fragments model native protein structures accurately. J Mol Biol. 2002; 323:297–307. PMID: 12381322
- Joseph AP, Agarwal G, Mahajan S, Gelly J-C, Swapna LS, Offmann B, et al. A short survey on protein blocks. Biophys Rev. 2010; 2:137–147. https://doi.org/10.1007/s12551-010-0036-1 PMID: 21731588
- de Brevern AG, Etchebest C, Hazout S. Bayesian Probabilistic Approach for Predicting Backbone. Proteins. 2000; 287:271–287.
- Dudev M, Lim C. Discovering structural motifs using a structural alphabet: application to magnesiumbinding sites. BMC Bioinformatics. 2007; 8:106. https://doi.org/10.1186/1471-2105-8-106 PMID: 17389049
- Wu CY, Chen YC, Lim C. A structural-alphabet-based strategy for finding structural motifs across protein families. Nucleic Acids Res. 2010; 38:e150. https://doi.org/10.1093/nar/gkq478 PMID: 20525797
- Schneider B, Cerný J, Svozil D, Cech P, Gelly J-C, de Brevern AG. Bioinformatic analysis of the protein/ DNA interface. Nucleic Acids Res. 2014; 42:3381–3394. <u>https://doi.org/10.1093/nar/gkt1273</u> PMID: 24335080
- 17. Tyagi M, de Brevern AG, Srinivasan N, Offmann B. Protein structure mining using a structural alphabet. Proteins. 2008; 71:920–937. https://doi.org/10.1002/prot.21776 PMID: 18004784
- Joseph AP, Srinivasan N, de Brevern AG. Improvement of protein structure comparison using a structural alphabet. Biochimie. 2011; 93:1434–1445. https://doi.org/10.1016/j.biochi.2011.04.010 PMID: 21569819
- Mahajan S, de Brevern AG, Sanejouand Y-H, Srinivasan N, Offmann B. Use of a structural alphabet to find compatible folds for amino acid sequences. Protein Sci. 2015; 24:145–153. <u>https://doi.org/10.1002/pro.2581 PMID: 25297700</u>
- Ghouzam Y, Postic G, de Brevern AG, Gelly J-C. Improving protein fold recognition with hybrid profiles combining sequence and structure evolution. Bioinformatics. 2015; 31:3782–9. https://doi.org/10.1093/ bioinformatics/btv462 PMID: 26254434

- Zimmermann O, Ulrich HEH. LOCUSTRA: Accurate Prediction of Local Protein Structure Using a Two-Layer Support Vector Machine Approach. J Chem Inf Model. 2008; 48:1903–1908. https://doi.org/10. 1021/ci800178a PMID: 18763837
- Suresh V, Parthasarathy S. SVM-PB-Pred: SVM based protein block prediction method using sequence profiles and secondary structures. Protein Pept Lett. 2014; 21(8):736–742. PMID: 23855661
- Rangwala H, Kauffman C, Karypis G. svmPRAT: SVM-based protein residue annotation toolkit. BMC Bioinformatics. 2009; 10:439. https://doi.org/10.1186/1471-2105-10-439 PMID: 20028521
- Dong Q, Wang X, Lin L, Wang Y. Analysis and prediction of protein local structure based on structure alphabets. Proteins. 2008; 72:163–172. https://doi.org/10.1002/prot.21904 PMID: 18214985
- Karypis G. YASSPP: Better kernels and coding schemes lead to improvements in protein secondary structure prediction. Proteins. 2006; 64(3):575–586. https://doi.org/10.1002/prot.21036 PMID: 16763996
- Garnier J, Gibrat JF, Robson B. GOR secondary structure prediction method version IV. Methods Enzym. 1996; 266:540–553.
- Etchebest C, Benros C, Hazout S, de Brevern AG. A structural alphabet for local protein structures: Improved prediction methods. Proteins. 2005; 59:810–827. https://doi.org/10.1002/prot.20458 PMID: 15822101
- Jones DT. Protein secondary structure prediction based on position-specific scoring matrices. J Mol Biol. 1999; 292:195–202. https://doi.org/10.1006/jmbi.1999.3091 PMID: 10493868
- de Brevern AG, Benros C, Gautier R, Valadié H, Hazout S, Etchebest C. Local backbone structure prediction of proteins. In Silico Biol. 2004; 4:381–386. PMID: 15724288
- Altschul SF, Gish W, Miller W, Myers EW, Lipman DJ. Basic local alignment search tool. J Mol Biol. 1990; 215:403–410. https://doi.org/10.1016/S0022-2836(05)80360-2 PMID: 2231712
- Tyagi M, Gowri VS, Srinivasan N, de Brevern AG De, Offmann B. A Substitution Matrix for Structural Alphabet Based on Structural Alignment of Homologous Proteins and its Applications. Proteins. 2006; 39:32–39.
- Joseph AP, Srinivasan N, de Brevern AG. Improvement of protein structure comparison using a structural alphabet. Biochimie. 2011; 93:1434–1445. https://doi.org/10.1016/j.biochi.2011.04.010 PMID: 21569819
- Argos P. Analysis of sequence-similar pentapeptides in unrelated protein tertiary structures. Strategies for protein folding and a guide for site-directed mutagenesis. J Mol Biol. 1987; 197:331–48. PMID: 3681998
- Suresh V, Liu L, Adjeroh D, Zhou X. RPI-Pred: predicting ncRNA-protein interaction using sequence and structural information. Nucleic Acids Res. 2015; 43:1370–1379. <u>https://doi.org/10.1093/nar/</u> gkv020 PMID: 25609700
- Luo J, Liu L, Venkateswaran S, Song Q, Zhou X. RPI-Bind: a structure-based method for accurate identification of RNA-protein binding sites. Sci Rep. 2017; 7:614. <u>https://doi.org/10.1038/s41598-017-</u> 00795-4 PMID: 28377624
- Nguyen L, Dang X, Le T, Saethang T, Tran V, Ngo D, et al. Predicting Beta-Turns and Beta-Turn Types Using a Novel Over-Sampling Approach. Journal of Biomedical Science and Engineering. 2014; 7:927–940.
- **37.** Webb B, Sali A. Comparative protein structure modeling using MODELLER. Curr Protoc Bioinforma 2014; 5.6.1–32.
- Kohonen T. Self-organized formation of topologically correct feature maps. Biol Cybern 1982; 43(1):59– 69.
- Kabsch W, Sander C. Dictionary of protein secondary structure: Pattern recognition of hydrogenbonded and geometrical features. Biopolymers. 1983; 22(12):2577–2637. https://doi.org/10.1002/bip. 360221211 PMID: 6667333