



Review

Harnessing Tumor Necrosis Factor Alpha to Achieve Effective Cancer Immunotherapy

María Florencia Mercogliano ¹, Sofía Bruni ², Florencia Mauro ², Patricia Virginia Elizalde ² and Roxana Schillaci ^{2,*}

¹ Laboratorio de Biofísicoquímica de Proteínas, Instituto de Química Biológica de la Facultad de Ciencias Exactas y Naturales-Consejo Nacional de Investigaciones Científicas y Técnicas (IQUIBICEN-CONICET), Buenos Aires 1428, Argentina; florenciamercogliano@qb.fcen.uba.ar

² Laboratory of Molecular Mechanisms of Carcinogenesis, Instituto de Biología y Medicina Experimental (IBYME-CONICET), Buenos Aires 1428, Argentina; sbruni@dna.uba.ar (S.B.); fmauro@dna.uba.ar (F.M.); patriciaelizalde@ibyme.conicet.gov.ar (P.V.E.)

* Correspondence: rschillaci@ibyme.conicet.gov.ar; Tel.: +54-11-4783-2869; Fax: +54-11-4786-2564

Simple Summary: Inflammation has been acknowledged as one of the causes of increased cancer risk. Among the pro-inflammatory mediators, tumor necrosis factor alpha (TNF α) has been identified as an important player in cancer progression and metastasis. On the other hand, TNF α has a central role in promoting innate and adaptive immune responses. These apparently controversial effects are now starting to be uncovered through different studies on TNF α isoforms and distinct mechanisms of action of TNF α receptors. The use of immunotherapies for cancer treatment such as monoclonal antibodies against cancer cells or immune checkpoints and adoptive cell therapy, are beginning to broaden our understanding of TNF α 's actions and its potential therapeutic role. This work describes TNF α participation as a source of treatment resistance and its implication in side effects to immunotherapy, as well as its participation in different cancer types, where TNF α can be a suitable target to improve therapy outcome.

Abstract: Tumor necrosis factor alpha (TNF α) is a pleiotropic cytokine known to have contradictory roles in oncoimmunology. Indeed, TNF α has a central role in the onset of the immune response, inducing both activation and the effector function of macrophages, dendritic cells, natural killer (NK) cells, and B and T lymphocytes. Within the tumor microenvironment, however, TNF α is one of the main mediators of cancer-related inflammation. It is involved in the recruitment and differentiation of immune suppressor cells, leading to evasion of tumor immune surveillance. These characteristics turn TNF α into an attractive target to overcome therapy resistance and tackle cancer. This review focuses on the diverse molecular mechanisms that place TNF α as a source of resistance to immunotherapy such as monoclonal antibodies against cancer cells or immune checkpoints and adoptive cell therapy. We also expose the benefits of TNF α blocking strategies in combination with immunotherapy to improve the antitumor effect and prevent or treat adverse immune-related effects.

Keywords: TNF α ; immunotherapy; adoptive cell therapy; monoclonal antibody; immune checkpoint inhibitor; cancer



Citation: Mercogliano, M.F.; Bruni, S.; Mauro, F.; Elizalde, P.V.; Schillaci, R. Harnessing Tumor Necrosis Factor Alpha to Achieve Effective Cancer Immunotherapy. *Cancers* **2021**, *13*, 564. <https://doi.org/10.3390/cancers13030564>

Academic Editors: Michael Kershaw and George Mosialos
Received: 7 December 2020
Accepted: 22 January 2021
Published: 2 February 2021

Publisher's Note: MDPI stays neutral with regard to jurisdictional claims in published maps and institutional affiliations.



Copyright: © 2021 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (<https://creativecommons.org/licenses/by/4.0/>).

1. Introduction

It is well known that tumor necrosis factor alpha (TNF α) participates as a proinflammatory cytokine, increasing the risk of several cancers, such as colorectal, esophageal, pancreatic, liver, and breast cancer [1]. However, another layer of complexity in TNF α functions was added with the emergence of immunotherapy. In this review we highlight recent data pointing out TNF α participation in the effectiveness of monoclonal antibodies (mAbs) targeting cancer cells, immune checkpoint inhibitors, and adoptive cell therapy (ACT), as well as its involvement in the adverse immune effects of immunotherapy.

2. TNF α Overview

TNF α was identified in 1975 as a molecule capable of causing tumor necrosis at high concentration [2–4]. Many studies on TNF α showed that it is a pleiotropic proinflammatory cytokine involved in a wide variety of cellular processes and, moreover, has contradictory effects ranging from cell proliferation to cell death. First, it was described that TNF α was involved in immune system regulation and was mainly secreted by cells such as monocytes, macrophages, natural killer (NK) cells, T lymphocytes, mast cells, and neutrophils, but later several works showed that it is also produced by non-immune cells like endothelial cells, adipocytes, neurons, fibroblasts, and smooth muscle, among others [5–9].

The human TNF α gene consists of a single copy located in chromosome 6 near the major histocompatibility complex genes [10]. It comprises four exons and three introns. The first exon contains the leader peptide sequence and the last ones the information for the protein. TNF α transcription is stimulated by NF- κ B [11], AP-1, c-Jun, and Nuclear Factor of Activated T-cells (NFAT) [6]. TNF α is present in either of two forms: transmembrane (tmTNF α) or soluble TNF α (sTNF α). tmTNF α is classified as a type II membrane protein, like many of the TNF-related ligands; it has a molecular weight of 26 kDa and forms a homotrimer that can also act as a receptor. TNF α can regulate several pathological and physiological processes beyond the immune system. The duality of many members of the TNF superfamily, as ligand and receptor, gives rise to the particular phenomenon of reverse signaling [12]: when acting as a receptor, tmTNF α can signal outside-to-inside back to the tmTNF α expressing cell. This mechanism has been mostly described in the regulation of the immune system but has not yet been completely characterized. On the other hand, sTNF α of 17 kDa is generated through proteolytic cleavage of tmTNF α by the TNF α Converting Enzyme (TACE/ADAM17) [13]. The active mature sTNF α also forms a homotrimer of 52 kDa that exerts a powerful autocrine, paracrine, and endocrine effect [14].

There are two membrane receptors for TNF α , also classified as type I membrane proteins, TNF α receptor 1 (TNFR1/CD120a, 55 kDa) and 2 (TNFR2/CD120b, 75 kDa) [15], and tm- and sTNF α can bind to them. Both isoforms of TNF α trigger receptor trimerization and subsequent recruitment of scaffold proteins to the cytoplasmic domain to induce different signaling pathways depending on the receptor involved, the type of TNF α that activated the receptor, the cell type, and the cellular context [16]. Pleiotropic effects of TNF α can be due not only to its two forms but also to the low homology of the ligand binding domain and no homology in the intracellular domain of the receptors, which have no enzymatic activity and therefore have to recruit scaffold proteins to unleash the signaling cascade [16]. Another particularity of the TNF α pathway, which explains its contradictory and varied effects, is that both TNFR1 and TNFR2 have soluble forms that are cleaved by TACE/ADAM17. The function of soluble receptors is to regulate TNF α availability and protect this cytokine from degradation to accomplish a sustained signal [17,18]. Most nucleated cells of the body express TNFR1, which can be activated by both forms of TNF α [19]. On the contrary, TNFR2 is expressed mainly in immune cells and in limited cell types like neurons, oligodendrocytes, astrocytes, and endothelial cells, among others, and can only be fully activated by tmTNF α [20,21].

Regarding the signaling of each receptor, TNFR1 has a cytoplasmic death domain, which can recruit TNFR1-Associated Death Domain (TRADD) protein and TNF Receptor-Associated Factor 2 (TRAF2), which can form two complexes: complex I, which stimulates cell survival and proliferation through JNK, NF- κ B, AP-1, and MAPK pathways [22], and complex II, which, on the contrary, recruits Fas-Associated protein with Dead Domain (FADD) and pro-caspases that constitute a death-inducing signaling complex [23], which ends in apoptosis [24]. Which of these pathways prevails is determined by the signaling molecules of the scaffold, signal strength, and crosstalk with other pathways [25]. TNFR2, instead, lacks the death domain and mainly regulates cell activation, migration, and proliferation [26]. Nonetheless, TNFR2 can also bind TRAF2 through TRAF1, concluding in the activation of both the canonical and non-canonical NF- κ B pathway like TNFR1, but activation is slower and more sustained [27,28]. It has also been reported that TNFR2 can

activate the abovementioned TNFR1 pathways through recruitment of Receptor Interacting Protein 1 (RIP-1) and TRADD via TRAF2, resulting in apoptosis.

Briefly, the convoluted pathway of TNF α involves a transmembrane and a soluble form, as well as two distinct receptors, which also exist in a soluble form. The combination of these elements activates distinct and unique signaling pathways that account for the pleiotropic effects of this cytokine. To add to the complexity, the pathways stimulated by each receptor can converge depending on different factors such as the adaptor proteins, TNF α concentration, cell type, and cellular context.

3. TNF α and the Immune System

Regulation of the innate immune system is the main role of TNF α , and it has been reviewed extensively throughout time. In particular, it is a major protagonist in immunity against intracellular organisms [29–32], has been intensively studied in *Mycobacterium* infection [33], and is responsible for the proliferation of thymocytes [34]. TNF α is also the main player in the initiation of inflammatory reactions characterizing the onset of the immune response.

Neither the TNF α nor TNFRs knockout model is lethal, but lymphoid organs and the immune response are affected. TNF α and its receptors are essential for the regulation of pro- and anti-inflammatory processes [30], the formation of Peyer's patches [35], and the adaptive B cell immune response [36], since it is involved in the generation of B cell follicles and germinal centers, and consequently, they affect the humoral immune response, among others.

TNF α also has contradictory effects in the immune system, since it can act as an immunosuppressor or an immunostimulant [2,37]. TNF α activates macrophages that produce more TNF α , generating a feed-forward loop, and is essential in guiding proliferation and proper effector function of several cell populations of the immune system, such as T, B, NK, and dendritic cells (DC). TNF α immunosuppressor effects encompass the regulation of suppressor cell populations like regulatory T and B cells (Tregs and Bregs, respectively) [38–40] and myeloid-derived suppressor cells (MDSCs) [41,42].

The central role of TNF α as an immunostimulant is to initiate the inflammatory response of the innate immune system and stimulate the Th1 profile. When a pathogen enters the organism, TNF α expression is induced. The elevated level of TNF α induces a chemokine/cytokine signaling cascade which, at the site of injury, induces certain adhesion molecule expression on the endothelial cells and immune cells, which allow neutrophil extravasation and the recruitment of macrophages and lymphocytes. It is noteworthy that TNF α generates a positive autocrine feedback loop that activates NF- κ B, which increases GM-CSF, IL-8, and TNF α itself [43].

As stated before, TNFR2 is mainly expressed in immune cells, and when TNF α binds to it, TRAF1, 2, and 3 are recruited together with cIAP1/2 to activate canonical and non-canonical NF- κ B and PI3K-Akt pathways, which consequently guides cell proliferation and survival. TNFR2 expression is higher in Tregs with respect to the rest of the T cell population, and in humans, this set of Tregs also expresses higher levels of cytotoxic T lymphocyte antigen 4 (CTLA-4), a well-known immunomodulator. TNFR2 has also been found to be involved in the suppressive activity of Tregs, but the mechanisms behind this process remain to be elucidated. Tregs also produce TNF α in certain inflammatory pathologies, and their function depends on the context, indicating that TNF α could be an attractive target to treat these inflammatory diseases. This proves once again the pleiotropic activity of TNF α , since it can promote the inhibition of Treg function in co-culture conditions with effector T lymphocytes but can also stimulate their immunosuppressive role, promoting Treg proliferation and survival, depending on the context [44–46]. Unstimulated CD4⁺ T lymphocytes increase MDSC accumulation [47] through tmTNF α via TNFR2 [48] and through 17- β -estradiol [49], and enhance their immunosuppressive activity through Nos2 [42].

4. TNF α in Cancer

TNF α has a plethora of functions and implications, and this also applies to cancer cells. TNF α has been described as having contradictory effects on almost every type of can-

cer. In high concentrations, TNF α is able to eliminate methylcolanthrene (MCA)-induced sarcomas, as first described by Carswell [2], and approximately 28% of cancers are sensitive to sTNF α [50]. TNF α antitumor mechanisms are varied and include the following: mediating cellular apoptosis extensively reviewed by Rath et al. [51]; directing tumor-associated macrophages (TAMs) to the M1 profile (antitumoral phenotype) [52]; guiding neutrophils and monocytes to tumor sites [53,54], activating macrophages and inhibiting monocyte differentiation to immunosuppressive phenotypes [55]; and inducing disruption of tumor vasculature [56,57]. Despite the above, TNF α expression at low levels can be pro-tumorigenic, an effect broadly reviewed by Balkwill [37,58].

There has been a large amount of evidence linking pro-inflammatory cytokines to cancer and the association with poor prognosis (reviewed by Mantovani) [59]. TNF α is one of the major pro-inflammatory cytokines of the immune system and has been found in several human cancers, such as breast [60], gastric [61], pancreatic [62], ovarian [63,64], endometrial [65], prostate [45], bladder [66], colorectal [67], oral [68], and liver [69]. It has also been detected in leukemias and lymphomas. Even so, there has been disagreement in considering TNF α expression as a biomarker, since the cytokine is increased in numerous other pathologies as well.

The distinct and opposing effects of TNF α in cancer depend on cytokine concentration and s- TNF α or tm- TNF α isoforms, distinct caspase activation, varied expression of adaptor proteins, different expression levels of members of the Bcl-2 family, among others [70]. TNF α acts as a pro-tumoral cytokine involved in different processes, such as cell proliferation, tumor progression, migration, epithelial-to-mesenchymal transition (EMT), angiogenesis and metastasis in several cancer types. The pro- and anti-tumorigenic/tumoral effects of TNF α are shown in Table 1 for different types of cancers.

Concerning breast cancer, our group has extensively reviewed TNF α impact/role on the different subtypes [71]. Regarding TNF α involvement in resistance to therapy, we have described TNF α involvement in trastuzumab resistance in HER2+ breast and gastric cancer. In the case of gastric cancer, the HER2 expressing gastric cancer cell line sensitive to trastuzumab NCI-N87 becomes refractory to the antibody after TNF α exposure [72]. In pancreatic cancer, blocking TNF α strategies proved to be effective in animal models [62] and in patients [73].

In melanoma, TNF α induces cell invasion [74] and aggressiveness [75], extravascular migration of cancer cells [76] and impairs CD8 T lymphocytes accumulation in the TME [77], moreover blocking TNF α prevents metastasis formation in the lungs in pre-clinical models [78]. TNF α is also overexpressed in oral squamous cell carcinoma (OSCC) [79], promotes the sphere-forming abilities of its cells maintaining a cancer stem cell-like phenotype [80], and increases proliferation in leukemia stem cells [81]. Interestingly, TNF α at low doses increases CD20 expression in B chronic lymphocytic leukemia, which can take advantage of the proven anti-CD20 therapy [82].

There are reports pointing to TNF α having no effect in endometrial cancer [83,84]. On the other hand, elevated pre-diagnostic concentrations of TNF α and its soluble receptors and the activation of TNF α -related pathways have been related to higher risk and poorer survival in endometrial cancer [85], prostate cancer [86] where could induce a shift to an untreatable phenotype [87]. In OSCC correlates with progression [88,89] and with relapse in children with B-lineage acute lymphoblastic leukemia (ALL) [90], but not with response to treatment [91] and in patients with non-Hodgkin's lymphoma [92] and diffuse large B cell lymphoma, TNF α is useful to differentiate risk groups [93]. In the latter TNFR1 expression in the tumor is also a good biomarker for prognosis [94].

Regarding TNF α as a potential biomarker, it was shown that TNF α polymorphisms in the gene promoter or coding region are associated with a risk of progression in patients with gastric lesions [95,96], with worse prognosis in prostate cancer patients [97], with tumor stage in bladder cancer [98], with risk of recurrence in hepatocellular carcinoma [99] and with higher risk in non-Hodgkin's lymphoma [100], T cell lymphoma [101], and gastric B cell lymphoma [102]. In ovarian cancer, TNF α gene polymorphisms are associated with pathogenesis but remains to be validated [103].

Table 1. Dual role of tumor necrosis factor alpha (TNF α) in cancer.

Cancer Type	Pro-Tumorigenic	References	Anti-Tumorigenic	References
Breast	Promotes proliferation, progression, and metastasis	[70]	Apoptosis and inhibition of proliferation	[70]
Gastric	Proliferation, progression and metastasis	[104–108]	Apoptosis acting together with TGF β	[109]
Pancreatic	Promotes tumor progression Generates a immune evasive microenvironment	[110,111] [115]	Apoptosis -	[112–114] -
Ovarian	Tumor promotion through TNFR1 and IL-17 Generates a immunosuppressor microenvironment Contributes to the EMT process through the NF- κ B pathway Tumor proliferation, progression, and invasion.	[116] [117] [64] [118–120]	- -	- - - -
Prostate	Survival and proliferation, progression, angiogenesis and metastasis	[121–126]	Apoptosis	[127,128]
Bladder	Migration and invasion through the p38 MAPK pathway	[129–131]	Apoptosis	[132,133]
Colorectal	Together with Th17-cytokines promotes immune escape, proliferation, survival, progression, and metastasis	[134–136]	-	-
Oral	Promotes immune evasion Promotes cell viability Promotes angiogenesis, invasion and metastasis	[137] [138] [139,140]	- -	- - -
Liver	Induces PTTG1, which in turn upregulates c-myc Promotes proliferation and metastasis in HCC through p38 MAPK, Erk1/2 and β -catenin Promotes resistance to the adaptive immune response through PD-L1 and PD-L2	[141] [143–145] [146]	In combination with IFN- γ showed reduction of liver tumors - -	[142] - -
Melanoma	Induces cell invasion and metastasis Increases aggressiveness	[74,76] [75]	Reduces tumor growth Apoptosis	[147,148] [149]
Hematological	Cell survival	[150–152]	Apoptosis thorough TNFR1, iNOS and PKC	[153,154]
	Promotes progression through the NF- κ B pathway and proliferation thorough GM-CSF	[81,155,156]	Increases efficacy of anti-CD20 therapy	[82]
	Promotes cell survival in Burkitt's lymphoma through reverse signaling	[157]	Induces maturation of AML generating specific cytotoxic CD8+ lymphocytes targeting leukemic disease	[158]
	-	-	Activate B cells to fight again lymphoma cells	[159]
	-	-	Combined with IL-1 and IFN- γ has an antiproliferative effect	[160]
-	-	Participates in the crosstalk between DC and NK cells	[161]	
-	-	Promotes cell death in Burkitt's lymphoma through forward signaling	[157]	

TGF β : Transforming Growth Factor beta; EMT: epithelial-to-mesenchymal transition; p38MAPK: p38 Mitogen-Activated Protein Kinase; PTTG1: Pituitary Tumor Transforming Gene 1; IFN- γ : Interferon gamma; HCC: hepatocellular carcinoma; PD-L1: Programmed Death Ligand 1; PD-L2: Programmed Death Ligand 2; iNOS: Inducible Nitric Oxide Synthase; PKC: Protein Kinase C; GM-CSF: Granulocyte-Macrophage Colony-Stimulating Factor; AML: Acute Myeloid Leukemia; DC: dendritic cells; NK: natural killer cells.

Comprehensively, the data presented in this section point to the central role of TNF α in cancer initiation, progression, and metastasis, despite its potential to activate cell death when present in high concentrations. A plethora of accumulated evidence highlights TNF α as a pro-tumoral cytokine, which stresses its appeal as a potential target to treat different cancers.

5. Immunotherapy Overview

The concept of using immune response specificity to target cancer cells has been investigated for a long time and has given rise to different strategies. So-called passive immunotherapy is based on the administration of antibodies or adoptive cell therapy, including chimeric antigen receptor (CAR)-T cells. Active immunotherapy, on the other hand, relies on several approaches, including the use of cancer vaccines, which can, for example, enhance antigen uptake and presentation, and the administration of antibodies that release the brakes of the immune response, known as immune checkpoint inhibitors. TNF α participation in these immunotherapies, either by hampering their success or mediating side effects, is discussed below and summarized in Table 2. For cancer vaccines, we refer to several recent reviews [162–164].

Table 2. Impact of anti-TNF α drugs in cancer immunotherapies.

IT	Target/Cell Type	Drug Name	Anti-TNF α	Effect on Cancer	Side Effects of IT	Ref.	
Monoclonal antibodies	HER2	Trastuzumab	Etanercept	Overcomes trastuzumab resistance in HER2+ breast cancer	NT	[72]	
			INB03	Overcomes trastuzumab resistance in HER2+ breast cancer	-	[165]	
	CD20	Rituximab	Etanercept	Improves disease-related symptoms and increases OS in chronic lymphocytic leukemia patients	NT	[166,167]	
	PD-1	anti-PD-1	Anti-TNFR1 or anti-TNF α	Prevents T lymphocytes exhaustion and death by anti PD-1 treatment in melanoma	Prevents immune-related adverse effect	[168]	
			Infliximab	NT	Treatment of immune-related adverse effects	[169–171]	
	PD-1+ CTLA-4	anti-PD-1 + anti CTL-4	Etanercept	Improves antitumor effect of anti PD-1+ anti CTL-4 antibodies in colon cancer	Prevents immune-related adverse effect	[172]	
	CTLA-4	Ipilimumab	Infliximab	NT	Treatment of immune-related adverse effects	[169,170, 173,174]	
	PD-L1	Atezolizumab, duvalumab and avelumab	Infliximab	NT	Treatment of immune-related adverse effects	[170]	
	CAR-T cells	CD19	-	Etanercept	NT	Treatment of systemic inflammatory response syndrome	[175]

IT: immunotherapy; OS: overall survival; NT: not tested; TNF α : Tumor necrosis factor alpha; TNFR1: TNF α receptor 1; PD-1: programmed cell death protein 1; PD-L1: PD-ligand 1; CTLA-4: cytotoxic T-lymphocyte-associated protein 4; CAR-T cells: chimeric antigen receptor T cell.

5.1. Monoclonal Antibodies

The mAbs, widely used to treat cancer and inflammatory diseases, are either chimeric, humanized, or fully human mAbs [176–178]. In this section, we outline the different cancer therapies based on mAbs targeting cancer cells and, in the following section, the mAbs directed to immune checkpoints, highlighting the implications of combining them with TNF α blocking agents.

Anti-TNF α Drugs

The first attempts to target TNF α were made decades ago, with the understanding that this cytokine was the major mediator of inflammation and its deregulation was implicated in a variety of autoimmune diseases, such as rheumatoid arthritis (RA), multiple sclerosis, psoriasis, Crohn's disease, scleroderma, systemic lupus erythematosus, ankylosing spondylitis, and diabetes. The pro-inflammatory effects of TNF α are mediated mainly by the activation of the NF- κ B pathway, which, in turn, promotes the transcription of inflammatory proteins, generating a positive feedback loop.

In several models of experimental metastasis in mice, both endogenous and exogenous administration of TNF α increased the development and number of metastatic lesions [179–181]. Additionally, TNF α is known to be a major inducer of chemokines [182] such as CCL2 and IL-6 in the TME, thus increasing monocyte and macrophage infiltration [183] as well as tumor growth and angiogenesis [184], respectively. On the other hand, increasing evidence has been accumulating about the positive impact of TNF α -blocking strategies in cancer treatment. Balkwill and collaborators demonstrated that neutralization of TNF α during early stages of skin carcinogenesis is sufficient to inhibit tumor formation and set the basis of the rationale of anti-TNF α therapy for cancer treatment [185]. Given the mentioned effects of TNF α , several blocking agents have been developed against it for use in the clinical setting. In this review, we will address the well-known etanercept, infliximab, and adalimumab and the new blocking agent, INB03.

Etanercept is a fusion protein that consists of two extracellular portions of human TNFR2 linked to the Fc portion of human immunoglobulin 1 (IgG1) [19] and exerts its anti-inflammatory properties by competitively binding sTNF α and tmTNF α , preventing their interaction with their receptors and therefore inhibiting the activation of important inflammatory pathways. Its use in cancer is limited and certainly poorly explored. Kai Sha and collaborators proved that a TNF α –CCL2 paracrine loop is induced in response to androgen deprivation therapy with enzalutamide in prostate cancer patients and might account for some forms of prostate cancer therapy resistance. Moreover, they showed that TNF α inhibition with etanercept in castration-resistant prostate cancer cells blocked enzalutamide-induced CCL2 protein secretion and mRNA expression. These data suggest that TNF α blockade would be a suitable therapy combined with androgen deprivation therapy in prostate cancer patients with primary tumors prior to the onset of castration-resistant prostate cancer and metastasis [186]. Almost two decades ago, etanercept was evaluated in a phase II clinical trial on patients with advanced metastatic breast cancer [187] who had shown incomplete or partial response, and a decrease in TNF α and CCL2 concentration in plasma samples was shown. A phase I trial assessing the clinical benefit of infliximab in patients with advanced cancer also reported no objective responses (either complete or partial). However, several patients achieved disease stabilization, which correlated with undetectable TNF α , CCL2, and IL-6 plasma levels [188]. These trials highlighted the need to further explore the use of TNF α -blocking agents in combination with radiotherapy and chemotherapy for advanced cancer treatment, yet scarce progress has been made in this direction.

Another TNF α blocking agent is the chimeric human-murine mAb infliximab, initially approved by the FDA in 1999 to treat patients with Crohn's disease who failed to respond to conventional therapy. Its structure consists of human constant regions and murine variable regions that specifically bind to human TNF α [189]. Like etanercept, this mAb binds both sTNF α and tmTNF α molecules and interferes with their activity. Moreover, the drug lyses cells bearing tmTNF α . However, infliximab contains 25% murine sequences in its structure, leading to the secretion of human anti-infliximab antibodies, which generates adverse reactions or a gradually increasing lack of efficacy [190].

The beneficial use of mAbs against TNF α has also been demonstrated in ovarian cancer xenografts; treatment of tumor-bearing mice with infliximab twice a week for 4 weeks resulted in reduced tumor burden, a significantly decreased proportion of infiltrating macrophages, and a marked reduction of IL-6 in the TME [191]. The authors suggested that targeting predominant cytokines like TNF α in the TME would be more useful in combination with

conventional chemotherapy regimens or treatments that target malignant cells directly, and better tolerated as well, than simply addressing tumor cells with targeted therapy.

Another way in which this pro-inflammatory cytokine can orchestrate the TME in ovarian cancer was described by Charles and collaborators, who demonstrated that TNF α is able to bind to TNFR1 and maintain the production of IL-17 in CD4⁺ leukocytes [116]. This sustained TNFR1-dependent IL-17 production and secretion leads to recruitment of the myeloid cell population to the TME and increased tumor growth [116]. These data were confirmed after blocking TNF α with infliximab in a mouse model of ovarian cancer and were also consistent with clinical results; patients with advanced ovarian cancer treated with infliximab exhibited substantially reduced plasma and ascitic levels of IL-17. Additionally, the authors found an association between high activation of TNF α signaling and expression of genes related to Th17 cell activation and expansion [116]. Unfortunately, some tumor types showed no benefit from the combination regime of the gold standard with TNF α blocking agents. Such is the case with advanced renal cell carcinoma (RCC), where phase I and II clinical trials demonstrated that the combined administration of sorafenib and infliximab, whose acceptable safety and tolerability were duly reported, was not more efficient than sorafenib alone [192]. In line with these data, recent results indicate that TNF α pathway activation would play a crucial role in resistance to tyrosine kinase inhibitors (TKIs) in patients with clear RCC [193]. Moreover, the authors suggest that TNFR1 could be a predictive biomarker for patient responsiveness to TKI treatment since it is augmented in TKI-resistant RCC tumors. In addition, the potential antitumor activity of infliximab has been reported in advanced RCC patients who progressed on cytokine therapy [194,195]. Therefore, a combination using infliximab or other TNF α inhibitors still holds promise as a therapeutic strategy for patients with RCC.

Adalimumab is a fully human recombinant mAb that binds and neutralizes both TNF α isoforms. Moreover, this mAb also induces apoptosis in immune cells bearing TNF α receptors. It was first approved by the FDA in 2008 for psoriasis treatment, but it is currently used in many other inflammatory diseases, such as RA, ankylosing spondylitis, Crohn's disease, ulcerative colitis and certain types of uveitis [196]. Its role in clinical oncology is not certain, but there is evidence that proves its efficacy in inhibiting TNF α tumor-promoting properties. In colorectal cancer cells, treatment with adalimumab hindered the induction of the Metastasis-Associated in Colon Cancer 1 (MACC1), a crucial oncogene that promotes cell proliferation, motility, and survival, increasing metastasis in preclinical models [197]. The authors proved that the expression of MACC1 in inflamed tissues from ulcerative colitis and Crohn's disease patients is upregulated by TNF α through NF- κ B signaling pathway, via TNFR1, which lead to an increase in cell migration. These effects were abolished using anti-TNFR1 antibodies or adalimumab, suggesting a potential role of this mAb in MACC1 driven colorectal tumors. Moreover, adalimumab has demonstrated a high efficacy to delay the acquisition of the senescence associated secretory phenotype (SASP) in endothelial cells, which is strictly related to inflammation and cancer progression [198]. Treatment of HUVEC cells with adalimumab generated a decrease in the release of the SASP marker IL-6, together with an upregulation of eNOS, indicating an enhanced endothelial function. Interestingly, TNF α inhibition by adalimumab in senescent endothelial cells diminished the tumor-promoting and pro-metastatic properties of their conditioned medium since the authors observed a decreased migration rate and mammospheres formation of MCF-7 breast cancer cells in the presence of such senescent secretome. These data highlight the potential role of adalimumab in restraining the SASP and delaying the consequent age-related diseases onset and progression in patients with a chronic inflammation background.

Etanercept, infliximab and adalimumab are based on the structure and function of mAbs, but there are other approaches to neutralize this cytokine. An example is INB03, a dominant-negative TNF α biologic that selectively neutralizes sTNF α without affecting the tmTNF α variant [199]. INB03 consists of a sTNF α mutant that forms inactive heterotrimers with the native cytokine. This differential blockade of TNF α isoforms is critical for activating the immune system in cancer patients, since it is known that the tmTNF α -TNFR2 inter-

action is necessary for the crosstalk between DC and NK cells, which does not depend on the sTNF α -TNFR1 axis [200–202]. This crosstalk acts as an immunomodulatory mechanism inducing an increase in Th1-type cytokines and promotes antitumor response [203]. There is evidence, including results from our own team, that confirms the pivotal role of sTNF α in the recruitment and expansion of MDSCs in the tumor bed, generating immunosuppression and favoring tumor progression [71,165,200]. Sobo-Vujanovic and collaborators proved that selectively blocking sTNF α with INB03 reduced tumor incidence and growth rate in mice with chemically induced carcinogenesis, compared to MCA-injected mice treated with etanercept or with vehicle [200]. Moreover, the authors demonstrated that wild-type mice and TNFR2 knockout mice treated with MCA exhibited significantly higher tumor incidence and poorer survival than TNFR1 knockout mice. These results suggest that sTNF α is the one that drives tumor progression and is critical for MCA-induced carcinogenesis, while tmTNF α is dispensable for tumor growth but has a pivotal role in immune system activation and promotion of its antitumor activity. In addition, they propose that tmTNF α could have a protective role in cancer and should therefore not be inhibited during treatment regimens [200]. These data place INB03 as the most appealing treatment option to block TNF α and avoid compromising the immune system in order to mount an antitumor response.

5.2. Monoclonal Antibodies Targeting Cancer Cells

5.2.1. HER2

HER2 tyrosine kinase receptor is overexpressed in 13–20% of human breast cancer cases and in 60% of metastases to bone and is associated with poor outcome. Additionally, it is amplified in 80% and 12% of urinary bladder and ovarian tumors, respectively, as well as in pancreatic adenocarcinoma and gastric cancer [204,205]. Moreover, when small-cell lung cancer (SCLC) cells acquire chemoresistance, HER2 is frequently upregulated and acts as a biomarker of poor prognosis in advanced cases [206–208]. As it constitutes an interesting target for directed therapy, several mAbs have been developed against it.

Trastuzumab is a humanized mAb that recognizes the fourth domain of the extracellular region of HER2 [177] and was approved in 1998 by the FDA as the first mAb for solid tumors, particularly for breast cancer treatment. We have also demonstrated that blockade of tmTNF α and sTNF α with etanercept downregulates the membrane glycoprotein mucin 4 (MUC4) expression and overcomes trastuzumab de novo or acquired resistance in HER2+MUC4+ breast cancer cells and xenografts. Moreover, we disclosed that it is sTNF α and not tmTNF α that drives MUC4 expression; we observed that HER2+MUC4+ breast cancer cells and tumors were also sensitized to trastuzumab in combination with INB03 [165]. TNF α blockade overcomes trastuzumab resistance in HER2+ breast cancer tumors not only by downregulating MUC4 expression, but also by transforming the TME to a less immunosuppressive state, characterized by increased NK cell activation and degranulation, a higher M1/M2 ratio, and decreased MDSC infiltration [71]. Tumor heterogeneity poses an immense challenge, which is why current therapeutic research is intended to develop several strategies to tackle HER2. sTNF α is certainly an interesting target for HER2+ breast cancer, and its combination with HER2 blocking agents should be further investigated to offer better treatment for patients.

T-DM1 is an antibody–drug conjugate (ADC) that combines trastuzumab with maytansine, a cytotoxic agent that inhibits microtubule polymerization. T-DM1 has also shown efficacy in women with progressive disease as second-line HER2-targeted therapy for metastatic breast cancer [209]. Results from our team demonstrate that TNF α expression and secretion by tumor cells is implicated in the resistance of HER2+ breast cancer cells to T-DM1 therapy by the upregulation of MUC4 [72].

5.2.2. EGFR/HER1

It is common knowledge that chemokines play a substantial role in cancer progression and metastasis, as they regulate cell migration in and out of the TME, among other

cellular processes that promote metastasis formation [210,211]. TNF α has been implicated in the transactivation of EGFR signaling to promote survival of colon epithelial cells [212]. It has been demonstrated that TNF α signaling, through its receptors, stimulates EGFR phosphorylation and promotes cellular proliferation, migration, and survival [213,214], and both TNF α and EGF can induce expression and secretion of cyclooxygenase 2 (COX-2), a prostaglandin synthase implicated in several biologic responses through prostaglandins [215,216]. Chronically elevated COX-2 levels correlate with increased risk for colorectal adenocarcinomas, and the use of chronic nonsteroidal anti-inflammatory drugs and administration of TNF α blocking antibodies have been associated with a decreased risk of developing colorectal cancer [217,218]. In this respect, it has been demonstrated that the induction of COX-2 expression by TNF α in gastrointestinal epithelial cells is dependent on TNF α -induced EGFR transactivation, promoting cell survival and proliferation. Furthermore, it has been elucidated that COX-2 expression is driven by the TNFR1 signaling pathway and not by TNFR2, by means of an EGFR-, Src-, and MAPK-dependent mechanism. These results add to accumulating evidence in favor of a critical role of sTNF α in colorectal cancer, which should be addressed by TNF α -blocking agents [191].

Interestingly, it has recently been demonstrated that in several ovarian cancer cell lines, cytokines like CCL20, CXCL1-3, and CXCL8 are the primary cytokines induced by EGFR activation or TNF α , through the NF- κ B and PI3K-Akt signaling pathways [219], indicating that TNF α could be a suitable target in ovarian cancer. We speculate that it would be beneficial for patients with EGFR+ ovarian tumors that secrete TNF α to consider a combination regime of anti-EGFR mAb, like cetuximab, and TNF α blocking agents. Considering all the above, it seems that TNF α is the driving force of the increased expression of pro-inflammatory cytokines in several cancer types and that it promotes this increase by transactivating the EGFR molecule and the consequent autocrine and paracrine loop with its ligands, EGF and TGF α . These data suggest the potential use of TNF α blocking agents in combination with anti-EGFR therapies to overcome resistance and target the pro-inflammatory and tumor promoting TME for better outcomes for said patients.

5.2.3. CD20

It is known that TNF α inhibits CLL cell death by upregulating Bcl-2, among other anti-apoptotic proteins, while it increases the proliferation of malignant cells [220]. In addition, TNF α is one of the main cytokines released as part of the toxicity in CLL patients receiving weekly treatment with rituximab [221,222].

Several clinical trials have been carried out to test the potential improvement of the anti-CD20 mAb rituximab treatment in combination with TNF α blocking agents, such as etanercept. Administration of etanercept has been shown to be safe in patients with CLL and other hematologic malignancies, whose disease-related symptoms also improved [166]. Particularly, in a phase I/II clinical trial, Woyach and collaborators showed that 75% of patients treated with rituximab in combination with etanercept exhibited a response, either complete or partial (29%), or had stable disease (56%) and did not require further treatment for 12 months after trial completion [167]. Moreover, the combination of rituximab and etanercept showed increased OS in responder patients, suggesting an improved outcome when compared to historical cytotoxic agent-based therapies. Furthermore, the addition of anti-TNF α mAb mitigated the toxicity of rituximab treatment [167]. The authors claimed that this combination regime would benefit fludarabine-refractory patients and people who are not eligible for more aggressive therapy, such as chemoimmunotherapy or rituximab alone, due to their high infusion toxicity.

Another fact that favors the study of TNF α in hematologic malignancies is that TNF α concentration is higher in the serum of patients with progressive CLL compared to healthy donors or patients with indolent disease [223]. Furthermore, it has been identified that TNF α overproduction in progressive CLL patients and CLL mouse models induces a decrease of plasmacytoid dendritic cells (pDCs), an immune cell population crucial for antiviral immunity and antitumor responses. The reduction in number and functionality

of pDCs causes impaired $\text{INF}\alpha$ production due to the decreased expression of FMS-like tyrosine kinase 3 receptor (Flt3) and Toll-like receptor 9 (TLR9). This effect was reverted when splenocytes from progressing CLL mice were treated with anti-TNF α mAbs, upon which increased pDC numbers and restored Flt3 expression were observed [223]. Similar results, along with reduced splenic tumor burden and increased splenic pDCs, were obtained by injection of anti-TNF α mAbs in mice with progressive CLL compared to control mice. In addition, anti-TNF α therapy promoted an increase in serum $\text{INF}\alpha$ production and augmented CD8+ T lymphocytes [223]. Blocking TNF α may be a potential strategy for immune reactivation in CLL patients. These results confirm the role of TNF α in CLL and the importance of addressing this pro-inflammatory cytokine as a therapeutic target in combination regimes with targeted therapies such as rituximab or standard cytotoxic agents like chemotherapy.

5.3. Monoclonal Antibodies against Immune Checkpoints

One of the shutdown mechanisms that are triggered after T lymphocyte activation operates through checkpoint inhibitors. The most well-known checkpoints in the context of cancer immunotherapy are CTLA-4 and programmed cell death protein 1 (PD-1, CD279), which are transmembrane molecules expressed by T lymphocytes after their activation. CTLA-4 binds to CD80 (B7-1) and CD86 (B7-2) expressed in DC, and PD-1 interacts with PD-ligand 1 (PD-L1, CD274) present in T lymphocytes, B lymphocytes, APCs, and tissues with immunological tolerance such as placenta and pancreatic islets, and with PD-L2 (CD273), expressed in APCs, thus mediating T lymphocyte inhibition [224–228]. Both immune checkpoints are hijacked by cancer cells, which promote CTLA-4 induction in T lymphocytes and induce PD-L1 expression in tumor cells as a mechanism of immune evasion. Thus, the interest in preventing CTLA-4/CD80/86 and PD-1/PD-L1 interactions derived in the development of antibodies against them as T cell-targeted immunomodulators [229,230] whose action is based on reinvigoration of the antitumor immune response. The impressive clinical benefit of this strategy, obtained first in melanoma patients [231], triggered a large number of clinical trials for the treatment of almost all types of cancer (Table S1).

There is plenty of evidence that TNF α upregulates PD-L1 expression in several cancer types. In prostate cancer cell lines HCT116 and LNCaP, TNF α increase PD-L1 mRNA and protein expression. In the case of LNCaP cells [232], the pathways involved in PD-L1 upregulation, dependent on ERK1/2 activation in HCT116 and in Akt and NF- κ B. In ovarian cancer cell lines HO8910 and SKOV3, it was demonstrated that TAMs or cytokines released from them, like $\text{INF-}\gamma$, TNF α , IL-10, and IL-6, are responsible for the upregulation of PD-L1 expression in the surface of these cells, but no modification in its mRNA was observed. The increase in PD-L1 levels produced by $\text{INF-}\gamma$ and TNF α was due to the activation of PI-3K and ERK1/2 pathways, respectively. In a preclinical model, treatment with anti-PD-1 or anti-PD-L1 was able to inhibit SKOV3 tumor growth [233] and was associated with decreased PD-1+ CD8+ T lymphocytes infiltration. A study demonstrated a progressive increase in PD-L1 levels ranging from immature bone marrow monocytes in tumor to circulating monocytes and to tumor tissue macrophages, the latter exhibiting the highest expression.

TNF α has been identified as the cytokine present in tumor-conditioned medium from B16 melanoma cells and 4T1 breast cancer cells that causes upregulation of PD-L1 in monocytes. In addition, tumor cells secrete versican, which stimulates TNF α production by monocytes via activation of TLR2 [234]. The role of adipocytes in PD-L1 expression was also addressed. Using an obese mouse model, it was demonstrated that B16-F10 melanomas and Hep-G2 hepatomas grew faster in the treated mice than in control animals, which was correlated with PD-L1 expression in cancer cells. Conditioned medium of adipocytes was able to increase PD-L1 levels due to the presence of TNF α and IL-6, both regulating the NF- κ B and STAT3 pathways (Figure 1) [235].

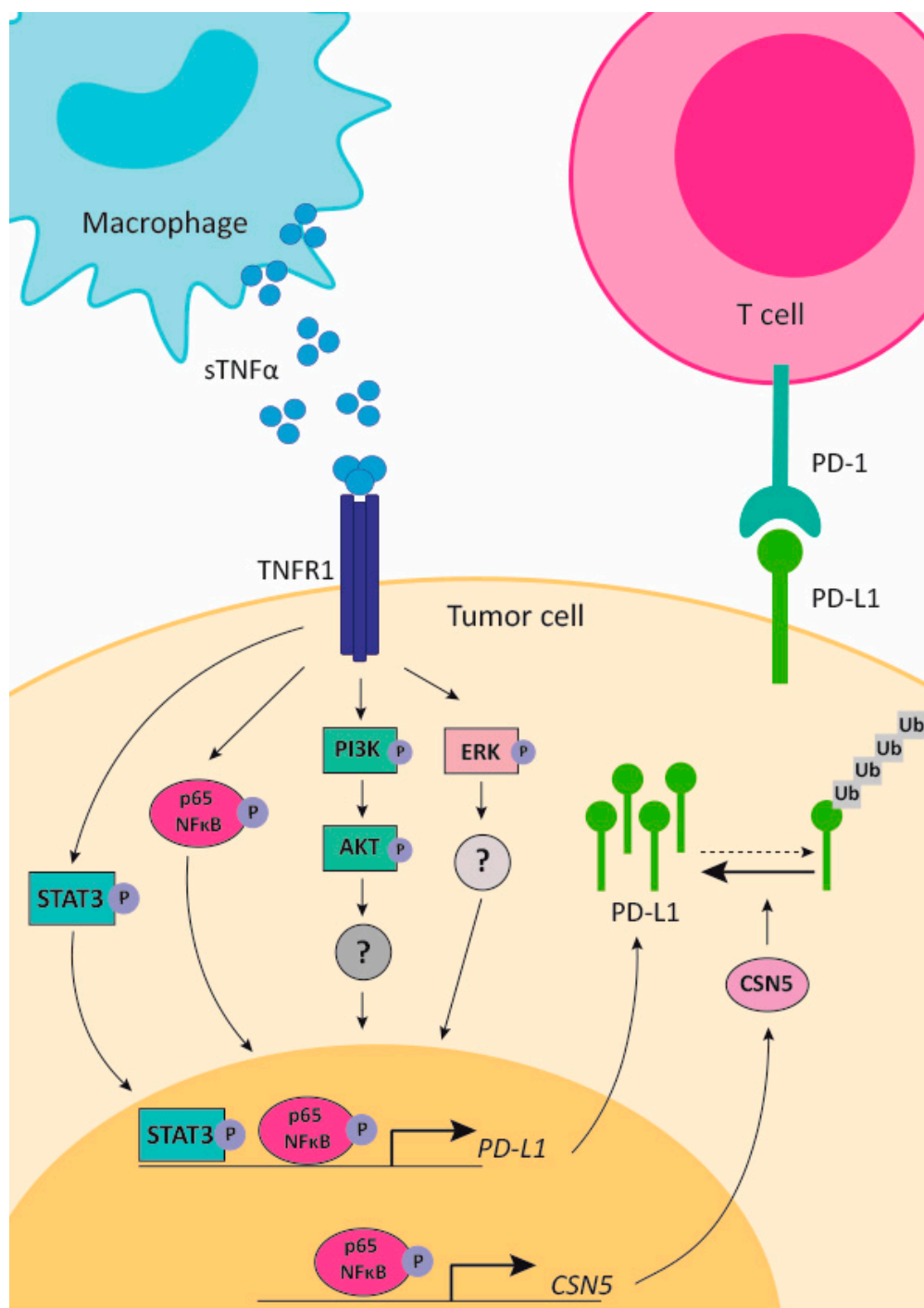


Figure 1. Tumor necrosis factor alpha (TNF α) modulates programmed death ligand 1 (PD-L1) expression transcriptionally and post-transcriptionally. TNF α , acting through TNF α receptor 1 (TNFR1), activates extracellular signal-regulated kinase (ERK) and phosphatidylinositol-3-kinase (PI-3K/AKT) pathways and nuclear factor kappa B (NF- κ B) and signal transducer and activator of transcription 3 (STAT3) transcription factors that promote PD-L1 gene transcription. In addition, NF- κ B also induces transcription of the deubiquitinase COP9 signalosome 5 (CSN5), which promotes PD-L1 protein stability. Ub: ubiquitin, sTNF α : soluble TNF α .

In addition, PD-L1 expression induced by TNF α was also proved in gastric cancer, where mast cell infiltration was directly related to its progression and reduced overall survival. A direct correlation was demonstrated between PD-L1+ mast cells and TNF α in gastric cancer specimens. TNF α secreted from gastric cancer cells induces PD-L1 expression in mast cells via activation of the NF- κ B signaling pathway [236]. It was recently demonstrated that PD-L1

expression in gastric cancer was dependent on TNF α and IL-6 produced by infiltrating macrophages. These cytokines promote PD-L1 expression through the activation of NF- κ B and STAT3 signaling [237]. Similar findings were observed in pancreatic cancer, where TNF α was the macrophage-secreted cytokine responsible for upregulation of PD-L1 in pancreatic ductal adenocarcinoma cells. In pancreatic cancer specimens, PD-L1 expression in tumor cells directly correlated with macrophage infiltration and poor survival [115].

PD-L1 expression can also be regulated at the posttranscriptional level. Seminal work by Hung's lab demonstrated that TNF α can increase PD-L1 expression in breast cancer cells by posttranscriptional regulation. TNF α stabilizes PD-L1 protein by inducing the expression of the deubiquitinating enzyme COP9 signalosome 5 (CSN5) via NF- κ B activation. This PD-L1 stabilization by TNF α also affects dendritic and T lymphocytes, inducing an immunosuppressive response (Figure 1) [238].

5.4. TNF α in Resistance to Anti-PD-1/PD-L1 and Anti-CTLA-4 Therapies

Several antibodies were designed to interfere with the PD-1/PD-L1 interaction and have been approved by the FDA for the treatment of different types of cancer at different stages (Table S1). These are the anti-PD-1 antibodies nivolumab, pembrolizumab, cemiplimab, and sintinimab and the anti-PD-L1 antibodies atezolizumab, durvalumab, and avelumab. In addition, CTLA-4 was effectively targeted by ipilimumab. Nivolumab, cemiplimab, sintinimab, avelumab, and ipilimumab are human monoclonal antibodies, while pembrolizumab, atezolizumab, and durvalumab are humanized monoclonal antibodies. The impressive clinical impact of these antibodies in the oncology arena was recognized by the 2018 Nobel Prize in Physiology or Medicine awarded to Dr. James Allison (MD Anderson Cancer Center at the University of Texas, Houston, TX, USA) and Dr. Tasuku Honjo (Kyoto University, Kyoto, Japan), for their contributions to the research on CTLA-4 and PD-1, respectively [239]. However, some patients exhibit resistance to anti-immune checkpoint treatment, depending on their cancer type and stage. Here, we highlight TNF α involvement in treatment failure based on immune checkpoint blockade.

In a preclinical melanoma model, TNF α , acting through TNFR1, impaired the infiltration of CD8+T lymphocytes into the TME and promoted their activation-induced cell death, facilitating tumor growth [77]. In addition, TNFR1 blockade improved the efficacy of anti-PD-1 treatment. Preventing TNF α upregulation of PD-L1 and TIM-3 expression by CD8+ tumor infiltrating lymphocytes (TILs) causes reinvigoration of the antitumor immune response, consequently overcoming anti-PD-1 resistance (Figure 2). These findings were validated using TCGA melanoma data, where a direct correlation was observed between TNF α and an immune escape signature, particularly with genes encoding PD-L1, PD-L2, and TIM-3 [168].

In an experimental melanoma, it was determined that anti-CTLA-4 treatment increased the production of TNF α associated with T lymphocytes infiltration, which in turn upregulated Ezh2, silencing tumor cell immunogenicity and antigen presentation. The inhibition of Ezh2 improved the effectiveness of anti-CTLA-4 and IL-2 immunotherapy [240].

The metabolic status of the tumor also conditions the efficacy of PD-L1 antibodies. In NSCLC, it was found that TNF α -induced aerobic glycolysis of TAMs was associated with tumor hypoxia in preclinical and clinical settings. TAM depletion facilitates the upregulation of PD-L1 in tumor cells, which can then be effectively targeted by anti-PD-L1 antibodies [241]. In addition, NSCLC patients with increased IFN- γ , TNF α , IL-1 β , IL-2, IL-4, IL-5, IL-6, IL-8, IL-10, and IL-12 serum levels at diagnosis and at 3 months post initiation of anti-PD-1 treatment exhibited longer OS [242]. Determination of these cytokines was proposed as a biomarker of patient selection for anti-PD-1 treatment.

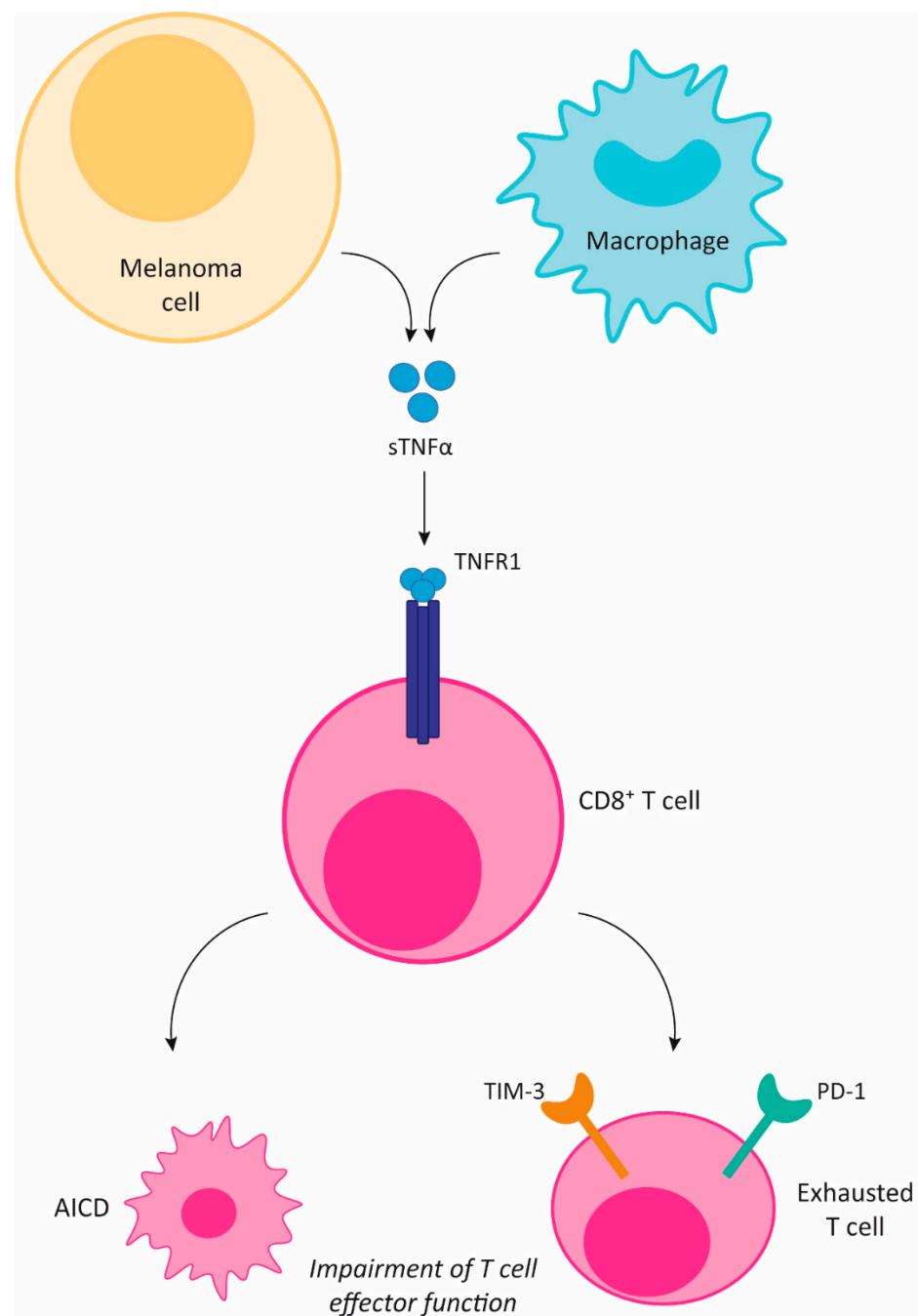


Figure 2. TNF α induces anti-immune checkpoint therapy resistance acting on CD8⁺ T lymphocytes. TNF α , produced by either tumor cells or macrophages from the tumor microenvironment, induces activation-induced cell death (AICD) and exhaustion of CD8⁺ T lymphocytes, impairing the effectiveness of anti-immune checkpoint therapy. TIM3: T-cell immunoglobulin and mucin-domain containing-3.

5.5. TNF α Involvement in the Adverse Effects of Immune Checkpoint Inhibitors

Releasing the brakes of the immune system through immune checkpoint blockade can trigger nonspecific immunologic activation that resembles autoimmune disease. These secondary effects, known as immune-related adverse effects (irAEs), can compromise the liver and skin (rash, pruritus, and vitiligo) and the endocrine (hypophysitis, hypothyroidism, and thyroiditis) and gastrointestinal (diarrhea and colitis) systems, among others [243]. About 50% of patients treated with anti-immune checkpoint therapy experienced some form of irAE and 20% suffered grade 3 or 4 toxicity, limiting the implementation of this treatment [244–246]. irAEs sometimes lead to discontinuation of treatment or administration of

corticosteroids or other immunosuppressive agents or TNF α antagonists [169–171,173,174]. The combination of anti-CTLA-4 and anti-PD-1 or PD-L1 antibodies is now more frequently used because of its increased clinical benefit compared to monotherapy regimens [247], but it also increases the severity of irAEs.

While in several cancers, such as lung and bladder cancer, there is an association between clinical benefit and irAEs, in melanoma the results are contradictory. In melanoma, the presence of irAEs does not guarantee tumor response, whereas the absence of side effects can be accompanied by clinical benefit [248,249]. In this respect, Perez-Ruiz et al., using melanoma and colon carcinoma models, demonstrated that the combined administration of anti-CTLA-4 and anti-PD-1 antibodies with anti-TNF α or etanercept reduced colitis and hepatitis in mice [172]. Importantly, they also showed that TNF α blockade enhanced the antitumor effect of immune checkpoint inhibitor treatment in melanoma and colon cancer, revealing that TNF α mediates irAEs [172]. This is an important piece of evidence indicating that preventing irAEs with TNF blocking agents allows the antitumor effect of immune checkpoint blockade.

Another work analyzed the effect of anti-TNF α treatment concomitant with or after anti-CTL-4 administration on irAEs and the antitumor effect. Results showed that although the antitumor effect on breast and colon cancer models of anti-CD40 decreased, the most suitable combination was simultaneous rather than delayed treatment with anti-TNF α administration. In this way, irAEs were prevented [250]. In the clinical setting, a recent report on patients from the Dutch Melanoma Treatment Registry showed that those treated with ipilimumab and anti-PD1 with severe irAEs had longer survival. However, treatment with infliximab blunted this clinical benefit [251]. Using large cohorts of 225,090 and 188,420 patients with Crohn's disease or ulcerative colitis, respectively, it was demonstrated that those treated with anti-TNF α agents were less likely to develop colorectal cancer. Further studies in different cancer types are needed to define the clinical benefit of TNF α blockade in terms of dose and administration in patients undergoing anti-immune checkpoint treatment [252].

5.6. Adoptive Cell Therapies

The development of ACTs has increased greatly in the last four years. Hundreds of new cell therapies have been added since 2017, quadrupling in 2020. Even in the current year, despite the COVID-19 pandemic, the number of cellular therapies has outgrown that of all existing types of immunotherapy [253].

In recent years, many advances have been made in immunotherapy for ALL [254]. ACTs have been developed with CAR-T cells, which consist of genetically modified T lymphocytes obtained from patients themselves, resulting in cells that combine an extracellular antigen-binding domain with one or more intracellular T lymphocytes signaling domains, leading to the activation of T lymphocytes and finally the elimination of lymphoblasts. In other words, these modified T lymphocytes are redirected to target specific antigens on the surface of lymphoblasts [255–257]. The CD19 antigen is a transmembrane protein expressed in all cells of the B lineage and is thus an attractive target for CAR-T cell therapy toward ALL B lymphoblasts [258,259]. Indeed, in 2017 the FDA approved an anti-CD19 CAR-T called CTL019 for the treatment of B cell ALL that is refractory to treatment or for second or later relapse of patients up to 25 years of age. The future of ACTs with CAR-T cells for B cell ALL is promising. Currently, various groups are working on addressing different targets such as CD22 for patients with CD19 negative relapses, optimizing the dose of CAR-T cells, and standardizing the management of neurological toxicity and systemic inflammatory response syndrome (SIRS) [254].

SIRS is the most common toxicity associated with CAR-T cell therapies. SIRS is generated due to the release of proinflammatory cytokines such as IL-6, IL-10, and IFN- γ (and possibly TNF- α and IL-1 α) after the activation of CAR-T cells. SIRS causes symptoms that range from myalgia, fever, and flu-like symptoms to capillary leak, vascular collapse, pulmonary edema, coagulopathy, and multiple organ failure [260]. Another highly unwanted possible adverse effect of CAR-T cell infusion is anaphylactic shock [261]. Treatment of

SIRS still remains challenging, and management is not well established. Corticosteroids have been used to treat severe SIRS with some success, but such treatment may interfere with the efficacy of CAR-T cell therapy itself [262]. Other anti-inflammatory agents have also been proposed, such as the IL-1 receptor antagonist anakinra or etanercept [175].

A barrier to ACTs in solid tumors is the formation of abnormal blood vessels, which hinders tumor infiltration of T lymphocytes [263]. Hypoxia can lead to the formation of new, tortuous, and leaky vessels, thus generating irregular blood flow and increased interstitial tumor pressure. Furthermore, endothelial cells fail to express leukocyte adhesion molecules correctly, an event known as endothelial anergy [264]. Therefore, crossing the abnormal endothelial barrier and interstitium in solid tumors is a major obstacle for cells of the immune system and CAR-T cell therapy [265]. This may also explain the resistance of some solid tumors to immune checkpoint inhibitors [266,267]. It has been shown that minimizing the amount of TNF α targeting the vascular endothelium with Cys-Asn-Gly-Arg-Cys-Gly-TNF α (NGR-TNF), a fusion protein targeting the tumor vasculature [147], can activate the endothelial cells and enhance tumor infiltration by cytotoxic T lymphocytes [268]. This approach has also been shown to enhance ACT with TCR redirected T lymphocytes [269]. Based on this, Elia et al. proposed using low doses of TNF α directed toward the tumor vasculature in association with ACT, which may represent a novel strategy to improve the infiltration of T cells in solid tumors and overcome the resistance to CAR-T cells and anti-immune checkpoint inhibitor therapy [263].

Anti-cancer ACT with tumor-specific cytotoxic T lymphocytes has been well documented in animal models, where infusion of modified T lymphocytes into mice resulted in tumor eradication [270–272]. The results of Ye et al. show that cytotoxic T lymphocytes transfected with adenovirus genetically modified to express TNF α , cytotoxicity, and survival of lymphocytes were improved [273]. Furthermore, ACT induces long-term anti-tumor immunity by generating memory T lymphocytes after ACT. Therefore, cytotoxic T lymphocytes designed to secrete TNF α may be useful when designing strategies for ACT in solid tumors [273].

Induction of antitumor immunity by DC vaccines correlates with their maturation stage. TNF α appears to have profound effects on DC function, as it contributes to activation [274], maturation [275], subsequent migration and accumulation in lymph nodes [276], and significantly reduces inhibition of these processes mediated by IL-10 [277]. Based on this, and the previously mentioned characteristics of TNF α as an antitumor cytokine, Liu et al. proposed the use of combination immunotherapy [278]. Gene therapy with adenoviruses expressing TNF α and DC vaccines genetically modified to overexpress TNF α were used to treat well-established tumors in animal models. The modified DCs stimulated cytotoxic T lymphocytes *in vitro* and *in vivo* and produced more efficient antitumor immune responses than wild-type DCs [278].

Lymphodepletion is a preconditioning strategy carried out by high-dose chemotherapy and is commonly used to increase the clinical efficacy of adoptive T cell therapy. Suppression of the host's immune system ensures that the transferred immune cells will be capable of surviving and proliferating, since they would otherwise be suppressed or deprived of key cytokines for their functioning [279]. However, as might be expected, this type of treatment can become highly toxic to patients, causing severe cytopenias [280,281]. In contrast, oncolytic adenoviruses are safer and, when engineered to express IL-2 and TNF α , can achieve lymphodepletion-like antitumor immunomodulatory effects [282]. When produced from these adenoviruses, IL-2 and TNF α can recruit NK and T lymphocytes into the tumor bed [283]. Studies in patients and mice revealed that toxicity was minimal. These findings demonstrate that ACT can be facilitated by adenoviruses that encode cytokines, thus avoiding lymphodepletion and its consequences [282].

In the case of melanoma, ACT with cytotoxic T lymphocytes that target melanocytic antigens can achieve remission in patients with metastatic melanomas, but tumors often relapse [284,285]. Landsberg et al. demonstrated that melanoma cells can resist ACT through a reversible dedifferentiation process in response to the inflammatory microenvironment in-

duced by T lymphocytes [286]. TNF α secreted by macrophages induces dedifferentiation of human melanoma cells, leading to impaired recognition by cytotoxic T lymphocytes specific for melanocytic antigens. These results demonstrate that an inflammatory microenvironment is responsible for the phenotypic plasticity of melanoma cells, contributing to tumor relapse after initially successful T lymphocyte immunotherapy [286]. This inflammation-induced dedifferentiation mechanism from tumor cells to precursor cells was also shown in a case report of a 60-year-old male patient with metastatic melanoma who received specific ACT against the melanocytic antigen MART-1 and developed resistance to therapy in association with a dedifferentiated tumor phenotype lacking conventional melanocytic antigens [287]. In vitro assays showed that TNF α treatment led to dedifferentiation of tumor cells. The dedifferentiation process was proved to be reversible upon removal of inflammatory media from cultures. The RNA of different melanoma cell lines treated with TNF α was also sequenced, and it was seen that the pathways of dedifferentiation induced by inflammation may overlap with those of innate resistance to anti-PD-1 gene signature [288], which includes genes related to EMT transition, hypoxia, and angiogenesis, and suggests that dedifferentiation may reflect a more invasive phenotype. The data exposed here highlight the need to deepen studies of the underlying mechanisms of ACT resistance in humans [287].

6. Clinical Implications

The administration of anti-TNF α drugs was originally limited to inflammatory and autoimmune pathologies, where they proved to be beneficial for patients [289]. Nevertheless, about 40% of patients did not respond to anti-TNF α treatment [290]. The different anti-TNF α biologics show no differences in the treatment of inflammatory pathologies such as RA and spondyloarthritis [291]. Regarding treatment effectiveness, it has been shown that polymorphisms in the TNF α promoter or the gene region can predict response to TNF α inhibition therapy. Meta-analyses showed that TNF α -308 G and -238 G alleles predicted good response to anti-TNF α therapy, and this prediction was more powerful for etanercept than for infliximab in patients with spondyloarthritis [292] or refractory sarcoidosis. Regarding patients with psoriatic arthritis, another study found that the polymorphism in +489 A exhibited a trend of association with better response to etanercept [293].

One of the main concerns of anti-TNF α treatment is the increased risk of infection upon therapy administration, a matter extensively studied in patients with inflammatory diseases. In these patients, TNF α -blocking therapies are administered alone or in combination with disease-modifying anti-rheumatic drugs (DMARDs) [294–296]. Concerning opportunistic intracellular bacterial infections, tuberculosis (TB) is one of the most studied, since TNF α is responsible for the recruitment and effector function of neutrophils and lymphocytes to battle the infection [297,298]. It was shown that TNF α or TNFR1 knockout mice, as well as those treated with TNF α inhibitors, cannot fight TB infection [299]. There are controversial studies about the adverse effects of TNF α inhibitor administration [300]. While several works show no correlation between adverse effects and TNF α inhibitor treatment [301–303], a substantial number show the opposite [304–307]. These works underline the importance of appropriate TB screening before TNF α inhibitor administration. Similar results were found for *L. monocytogenes* infection [304]. Something similar also occurs in viral and fungal infections. While there are reports that show no correlation between herpes zoster infection and TNF α blockage [308], others show the opposite [304,309,310]. Other detrimental effects of TNF α inhibitors were described regarding the nervous system, with headache as the most common event. Other serious neurological events [311,312] were also reported: multiple sclerosis [313], central and peripheral demyelinating events, vasculitis, and transverse myelitis, among others [314–316]. Regarding cardiac disease, it was demonstrated that anti-TNF α therapy is injurious [317]. Surprisingly, TNF α inhibition can cause de novo disease or reactivation of inflammatory disease, such as psoriasis, arthritis, colitis, uveitis, etc. [318].

Given the mentioned pivotal role of TNF α in the immune system, another important concern regarding its blockade is related to cancer development. Numerous studies have addressed this issue in patients with inflammatory diseases. At present, there is increasing evidence that TNF α inhibition does not correlate with an augmented incidence of cancer [319–321], but there are a few reports that show the opposite [322], including two reports that indicate an elevated risk of hematological malignancies and nonmelanoma skin cancers in patients with RA [298,323]. It is noteworthy that these studies also claim that an increased risk of lymphoma is associated with RA regardless of the anti-TNF α therapy, which inhibits any conclusion regarding the treatment. Instead, more recent studies show that, plausibly, the underlying inflammation caused by the pathology is more likely to promote the development of malignancy than the therapy itself [321,324]. Moreover, they claim the treatment can have positive implications in preventing cancer. There is a study on inflammatory bowel disease in patients with a prior history of cancer showing that TNF α inhibition poses a mild risk of acquiring cancer. This report poses a conundrum, since it is an established fact that former cancer patients have a higher probability of developing new cancers, once again highlighting that the results cannot be ascribed to the anti-TNF α therapy. These data indicate that TNF α -blocking treatments should not be administered to patients with cancer in their clinical history [325].

There are limited reports in the field of cancer and TNF α inhibitors. Only one phase II clinical trial studied the effect of etanercept in breast cancer patients, and the results showed that there was no objective response to treatment, which could, however, be due to the advanced tumor stage of the cohort [187]. Polymorphisms in the TNF α gene have also been studied related to cancer incidence. In breast cancer, it was reported that the TNF α -308 G>A allele is associated with higher expression of TNF α , but no predisposition for any breast cancer subtype was found, although this polymorphism is associated with an increased risk of metastasis in triple-negative breast cancer [326]. Another study showed that the same polymorphism was correlated with vascular invasion in breast cancer [327], while another group found a possible association between the -308 G>A polymorphism and lower OS in cancer patients [328]. However, these polymorphisms could be meaningful regarding responsiveness to anti-TNF α therapy. The matter remains to be explored in cohorts of patients with malignancies receiving anti-TNF α treatment.

Various reports have acknowledged the pro-tumorigenic role of TNFR1, which indicates that hindering sTNF α action could be a potential new strategy to tackle cancer [37,329]. In this regard, it has been shown that targeting sTNF α prevents skin carcinogenesis [203] and overcomes trastuzumab resistance in HER2+ breast cancer [72].

Besides targeting TNF α , another interesting approach is the development of therapies directed to TNFRs. One of the strategies is based on the fact that soluble TNFRs (sTNFR) are immunosuppressive because they impede TNF- α activity. Therefore, a selective apheresis to remove sTNFRs from systemic circulation can release TNF α and reactivate an effective antitumor immune response. In the beginnings, the apheresis column contained anti-TNFR and antiIL-2R antibodies and the treatment was effective in reducing patient's tumor burden [330]. Now an improvement was achieved using single-chain TNF α as bait [331] and this strategy has shown to be effective in the treatment of canine cancers [332]. On the other hand, TNFR2 is expressed in Tregs and particularly in a subset that present the most immunosuppressive characteristics [333]. Tregs in general have been a potential target for cancer therapy [334], but delivery to the TME has been challenging [335]. TNFR2 induces activation of NF- κ B and PI3K/Akt pathways, which finalizes in cell proliferation, augmenting the number of Tregs [336], positioning TNFR2 as an attractive target. In addition, MDSCs also express TNFR2 in mice, and their inhibition diminished metastasis in a liver cancer model [337]. Furthermore, TNFR2 expression has been proved in different cancers, such as RCC [338], colorectal cancer [339], Hodgkin's lymphoma [340], multiple myeloma [341], and ovarian cancer [342]. Interestingly, several studies show that tumors can escape immune checkpoint inhibitor therapy by upregulating TNFR2 expression in Tregs [343]. Moreover, TNF α /TNFR2 axis supports angiogenesis promoting VEGF secre-

tion and neovascularization via endothelial colony forming cells [344]. Recently, it has been demonstrated that endothelial progenitor cells secrete immunosuppressive cytokines in a TNFR2-dependent manner and inhibit T lymphocytes proliferation [345]. All this evidence points to the need to develop TNFR2-targeted therapy to diminish tumor-infiltrating Tregs, to impair MDSCs differentiation and endothelial cell neovascularization and to directly attack TNFR2-expressing tumor cells. In this respect, Vanamee et al. postulated that TNFR2 inhibitors could be safer than immune checkpoint inhibitors for cancer treatment, given the restricted expression of the receptor [346]. TNFR2 antagonistic antibodies were successfully tested in the OVCAR3 preclinical model and proved to be effective in killing Tregs from ascites and ovarian cells [347]. Finally, an IgG2 antibody targeting TNFR2 proved to be effective in killing cancer cells in direct correlation to their TNFR2 expression density. It was also shown that this antibody modified the TME eliminating Tregs while preserving viability of effector T cells [348]. Therefore, a new horizon of specific treatment targeting immunosuppressive cells is open with anti-TNFR2 strategies.

7. Conclusions

The clinical relevance of TNF α to either fostering or hindering the success of immunotherapy has not yet been fully elucidated. In practice, however, the clinical application of anti-TNF α drugs to prevent irAEs produced by immune checkpoint inhibitors and ACTs has provided interesting results, showing that neutralizing this cytokine has potential antitumor benefit. In addition, several clinical trials have demonstrated the importance of TNF α blockade in prostate and renal cancer and in hematologic malignancies, as it promotes higher OS. Furthermore, there is plenty of preclinical evidence showing that TNF α is able to induce immunotherapy resistance. For example, TNF α can induce PD-L1 overexpression in a large variety of tumors, rendering an immunosuppressive TME, impairing inhibition of immune checkpoints, and inducing resistance to targeted therapies [72,165–168,172,232–238]. All of the above therefore suggest that the use of TNF α inhibitors should be considered as a novel strategy in cancer treatment, particularly in combination with the gold standard therapy for each particular cancer.

On the other hand, due to its potent antitumor activity, the production of TNF α by DC and cytotoxic T lymphocytes is important in ACT. In addition, the administration of a fusion protein of TNF α targeting tumor vessels can rescue their normal permeability and promote tumor infiltration by cytotoxic T lymphocytes, enhancing the effectiveness of immune checkpoint inhibitors.

In conclusion, the administration of TNF α -blocking agents emerges as a promising option in the oncology arena, but their combination with other therapies in specific tumor types needs to be further studied to attain optimal clinical results.

Supplementary Materials: The following are available online at <https://www.mdpi.com/2072-6694/13/3/564/s1>, Table S1: FDA approved drugs targeting PD-1, PD-L1 and CTLA-4 (current as November 2020).

Author Contributions: M.F.M. and P.V.E. wrote and edited the manuscript, S.B. wrote, revised the manuscript and prepared the figures, F.M. wrote, revised the manuscript and prepared the table, R.S. conceptualized, wrote, and edited the manuscript. All authors have read and agreed to the published version of the manuscript.

Funding: This work was supported by grants from National Cancer Institute (Argentina) 2018, IDB/PICT 2017-1517 and 2018-2086 from the National Agency of Scientific Promotion of Argentina (ANPCyT), Florencio Fiorini Foundation and Alberto J. Roemmers Foundation awarded to RS; a grant from Oncomed-Reno, CONICET 1819/03, awarded to PVE and RS; grants from National Cancer Institute (Argentina) 2018, IDB/PICT 2015-1587, IDB/PICT 2017-1587, from ANPCyT, PID 2012-066 from Consejo Nacional de Investigaciones Científicas y Técnicas and from the National Institute of Cancer from Argentina (Argentina) 2018-2019, all of them awarded to PVE. We are grateful to Fundación René Baron and Fundación Williams for their institutional support to IBYME-CONICET.

Conflicts of Interest: The authors declare no conflict of interest.

References

1. Coussens, L.M.L.M.; Werb, Z. Inflammation and Cancer. *Nature* **2002**, *420*, 860–867. [[CrossRef](#)]
2. Carswell, E.A.; Old, L.J.; Kassel, R.L.; Green, S.; Fiore, N.; Williamson, B. An Endotoxin-Induced Serum Factor That Causes Necrosis of Tumors. *Proc. Natl. Acad. Sci. USA* **1975**, *72*, 3666–3670. [[CrossRef](#)]
3. Green, S.; Dobrjansky, A.; Carswell, E.A.; Kassel, R.L.; Old, L.J.; Fiore, N.; Schwartz, M.K. Partial Purification of a Serum Factor That Causes Necrosis of Tumors. *Proc. Natl. Acad. Sci. USA* **1976**, *73*, 381–385. [[CrossRef](#)] [[PubMed](#)]
4. Pennica, D.; Nedwin, G.E.; Hayflick, J.S.; Seeburg, P.H.; Derynck, R.; Palladino, M.A.; Kohr, W.J.; Aggarwal, B.B.; Goeddel, D.V. Human Tumour Necrosis Factor: Precursor Structure, Expression and Homology to Lymphotoxin. *Nature* **1984**, *312*, 724–729. [[CrossRef](#)] [[PubMed](#)]
5. Falvo, J.V.; Tsytsykova, A.V.; Goldfeld, A.E. Transcriptional Control of the TNF Gene. *Curr. Dir. Autoimmun.* **2010**, *11*, 27–60. [[CrossRef](#)] [[PubMed](#)]
6. Tsai, E.Y.; Yie, J.; Thanos, D.; Goldfeld, A.E. Cell-Type-Specific Regulation of the Human Tumor Necrosis Factor Alpha Gene in B Cells and T Cells by NFATp and ATF-2/JUN. *Mol. Cell. Biol.* **1996**, *16*, 5232–5244. [[CrossRef](#)] [[PubMed](#)]
7. Gahring, L.C.; Carlson, N.G.; Kulmar, R.A.; Rogers, S.W. Neuronal Expression of Tumor Necrosis Factor Alpha in the Murine Brain. *Neuroimmunomodulation* **1996**, *3*, 289–303. [[CrossRef](#)]
8. Ranta, V.; Orpana, A.; Carpén, O.; Turpeinen, U.; Ylikorkala, O.; Viinikka, L. Human Vascular Endothelial Cells Produce Tumor Necrosis Factor-Alpha in Response to Proinflammatory Cytokine Stimulation. *Crit. Care Med.* **1999**, *27*, 2184–2187. [[CrossRef](#)]
9. Rydén, M.; Arner, P. Tumour Necrosis Factor-Alpha in Human Adipose Tissue—from Signalling Mechanisms to Clinical Implications. *J. Intern. Med.* **2007**, *262*, 431–438. [[CrossRef](#)]
10. Spriggs, D.R.; Deutsch, S.; Kufe, D.W. Genomic Structure, Induction, and Production of TNF-Alpha. *Immunol. Ser.* **1992**, *56*, 3–34.
11. Shakhov, A.N.; Collart, M.A.; Vassalli, P.; Nedospasov, S.A.; Jongeneel, C.V. Kappa B-Type Enhancers Are Involved in Lipopolysaccharide-Mediated Transcriptional Activation of the Tumor Necrosis Factor Alpha Gene in Primary Macrophages. *J. Exp. Med.* **1990**, *171*, 35–47. [[CrossRef](#)] [[PubMed](#)]
12. Qu, Y.; Zhao, G.; Li, H. Forward and Reverse Signaling Mediated by Transmembrane Tumor Necrosis Factor-Alpha and TNF Receptor 2: Potential Roles in an Immunosuppressive Tumor Microenvironment. *Front. Immunol.* **2017**, *8*. [[CrossRef](#)] [[PubMed](#)]
13. Kriegler, M.; Perez, C.; DeFay, K.; Albert, I.; Lu, S.D. A Novel Form of TNF/Cachectin Is a Cell Surface Cytotoxic Transmembrane Protein: Ramifications for the Complex Physiology of TNF. *Cell* **2018**, *53*, 45–53. [[CrossRef](#)]
14. Perez, C.; Albert, I.; DeFay, K.; Zachariades, N.; Gooding, L.; Kriegler, M. A Nonsecretable Cell Surface Mutant of Tumor Necrosis Factor (TNF) Kills by Cell-to-Cell Contact. *Cell* **1990**, *63*, 251–258. [[CrossRef](#)]
15. Tartaglia, L.A.; Weber, R.F.; Figari, I.S.; Reynolds, C.; Palladino, M.A., Jr.; Goeddel, D.V. The Two Different Receptors for Tumor Necrosis Factor Mediate Distinct Cellular Responses. *Proc. Natl. Acad. Sci. USA* **1991**, *88*, 9292–9296. [[CrossRef](#)]
16. Ledgerwood, E.C.; Pober, J.S.; Bradley, J.R. Recent Advances in the Molecular Basis of TNF Signal Transduction. *Lab. Invest.* **1999**, *79*, 1041–1050.
17. Aderka, D.; Engelmann, H.; Maor, Y.; Brakebusch, C.; Wallach, D. Stabilization of the Bioactivity of Tumor Necrosis Factor by Its Soluble Receptors. *J. Exp. Med.* **1992**, *175*, 323–329. [[CrossRef](#)]
18. Watts, A.D.; Hunt, N.H.; Madigan, M.C.; Chaudhri, G. Soluble TNF-Alpha Receptors Bind and Neutralize over-Expressed Transmembrane TNF-Alpha on Macrophages, but Do Not Inhibit Its Processing. *J. Leukoc. Biol.* **1999**, *66*, 1005–1013. [[CrossRef](#)]
19. Tracey, D.; Klareskog, L.; Sasso, E.H.; Salfeld, J.G.; Tak, P.P. Tumor Necrosis Factor Antagonist Mechanisms of Action: A Comprehensive Review. *Pharmacol. Ther.* **2008**, *117*, 244–279. [[CrossRef](#)]
20. Choi, S.J.; Lee, K.H.; Park, H.S.; Kim, S.K.; Koh, C.M.; Park, J.Y. Differential Expression, Shedding, Cytokine Regulation and Function of TNFR1 and TNFR2 in Human Fetal Astrocytes. *Yonsei Med. J.* **2005**, *46*, 818–826. [[CrossRef](#)]
21. Faustman, D.; Davis, M. TNF Receptor 2 Pathway: Drug Target for Autoimmune Diseases. *Nat. Rev. Drug Discov.* **2010**, *9*, 482. [[CrossRef](#)] [[PubMed](#)]
22. Liu, Z.G.; Hsu, H.; Goeddel, D.V.; Karin, M. Dissection of TNF Receptor 1 Effector Functions: JNK Activation Is Not Linked to Apoptosis While NF-KappaB Activation Prevents Cell Death. *Cell* **1996**, *87*, 565–576. [[CrossRef](#)]
23. Chinnaiyan, A.M.; Tepper, C.G.; Seldin, M.F.; O'Rourke, K.; Kischkel, F.C.; Hellbardt, S.; Krammer, P.H.; Peter, M.E.; Dixit, V.M. FADD/MORT1 Is a Common Mediator of CD95 (Fas/APO-1) and Tumor Necrosis Factor Receptor-Induced Apoptosis. *J. Biol. Chem.* **1996**, *271*, 4961–4965. [[CrossRef](#)]
24. Kischkel, F.C.; Hellbardt, S.; Behrmann, I.; Germer, M.; Pawlita, M.; Krammer, P.H.; Peter, M.E. Cytotoxicity-Dependent APO-1 (Fas/CD95)-Associated Proteins Form a Death-Inducing Signaling Complex (DISC) with the Receptor. *Embo J.* **1995**, *14*, 5579–5588. [[CrossRef](#)] [[PubMed](#)]
25. Ihnatko, R.; Kubes, M. TNF Signaling: Early Events and Phosphorylation. *Gen. Physiol. Biophys.* **2007**, *26*, 159–167. [[PubMed](#)]
26. Bradley, J.R. TNF-Mediated Inflammatory Disease. *J. Pathol.* **2008**, *214*, 149–160. [[CrossRef](#)]
27. Rothe, M.; Wong, S.C.; Henzel, W.J.; Goeddel, D.V. A Novel Family of Putative Signal Transducers Associated with the Cytoplasmic Domain of the 75 KDa Tumor Necrosis Factor Receptor. *Cell* **1994**, *78*, 681–692. [[CrossRef](#)]
28. Naudé, P.J.W.; den Boer, J.A.; Luiten, P.G.M.; Eisel, U.L.M. Tumor Necrosis Factor Receptor Cross-Talk. *FEBS J.* **2011**, *278*, 888–898. [[CrossRef](#)]
29. Nunes-Alves, C.; Booty, M.G.; Carpenter, S.M.; Jayaraman, P.; Rothchild, A.C.; Behar, S.M. In Search of a New Paradigm for Protective Immunity to TB. *Nat. Rev. Microbiol.* **2014**, *12*, 289–299. [[CrossRef](#)]

30. Marino, M.W.; Dunn, A.; Grail, D.; Inglese, M.; Noguchi, Y.; Richards, E.; Jungbluth, A.; Wada, H.; Moore, M.; Williamson, B.; et al. Characterization of Tumor Necrosis Factor-Deficient Mice. *Proc. Natl. Acad. Sci. USA* **1997**, *94*, 8093–8098. [[CrossRef](#)]
31. Lin, P.L.; Myers, A.; Smith, L.; Bigbee, C.; Bigbee, M.; Fuhrman, C.; Grieser, H.; Chiosea, I.; Voitenek, N.N.; Capuano, S.V.; et al. Tumor Necrosis Factor Neutralization Results in Disseminated Disease in Acute and Latent Mycobacterium Tuberculosis Infection with Normal Granuloma Structure in a Cynomolgus Macaque Model. *Arthritis Rheum.* **2010**, *62*, 340–350. [[CrossRef](#)] [[PubMed](#)]
32. Bruns, H.; Meinken, C.; Schauenberg, P.; Härter, G.; Kern, P.; Modlin, R.L.; Antoni, C.; Stenger, S. Anti-TNF Immunotherapy Reduces CD8+ T Cell-Mediated Antimicrobial Activity against Mycobacterium Tuberculosis in Humans. *J. Clin. Investig.* **2009**, *119*, 1167–1177. [[CrossRef](#)] [[PubMed](#)]
33. Matucci, A.; Maggi, E.; Vultaggio, A. Cellular and Humoral Immune Responses during Tuberculosis Infection: Useful Knowledge in the Era of Biological Agents. *J. Rheumatol. Suppl.* **2014**, *91*, 17–23. [[CrossRef](#)] [[PubMed](#)]
34. Baseta, J.G.; Stutman, O. TNF Regulates Thymocyte Production by Apoptosis and Proliferation of the Triple Negative (CD3-CD4-CD8-) Subset. *J. Immunol.* **2000**, *165*, 5621–5630. [[CrossRef](#)] [[PubMed](#)]
35. Neumann, B.; Luz, A.; Pfeffer, K.; Holzmann, B. Defective Peyer's Patch Organogenesis in Mice Lacking the 55-KD Receptor for Tumor Necrosis Factor. *J. Exp. Med.* **1996**, *184*, 259–264. [[CrossRef](#)]
36. Pasparakis, M.; Alexopoulou, L.; Episkopou, V.; Kollias, G. Immune and Inflammatory Responses in TNF Alpha-Deficient Mice: A Critical Requirement for TNF Alpha in the Formation of Primary B Cell Follicles, Follicular Dendritic Cell Networks and Germinal Centers, and in the Maturation of the Humoral Immune Respons. *J. Exp. Med.* **1996**, *184*, 1397–1411. [[CrossRef](#)]
37. Balkwill, F. Tumour Necrosis Factor and Cancer. *Nat. Rev. Cancer* **2009**, *9*, 361–371. [[CrossRef](#)]
38. Jung, M.K.; Lee, J.S.; Kwak, J.E.; Shin, E.C. Tumor Necrosis Factor and Regulatory T Cells. *Yonsei Med. J.* **2019**, *60*, 126–131. [[CrossRef](#)]
39. Schioppa, T.; Moore, R.; Thompson, R.G.; Rosser, E.C.; Kulbe, H.; Nedospasov, S.; Mauri, C.; Coussens, L.M.; Balkwill, F.R. B Regulatory Cells and the Tumor-Promoting Actions of TNF- α during Squamous Carcinogenesis. *Proc. Natl. Acad. Sci. USA* **2011**, *108*, 10662–10667. [[CrossRef](#)]
40. Mauri, C.; Menon, M. Human Regulatory B Cells in Health and Disease: Therapeutic Potential. *J. Clin. Investig.* **2017**, *127*, 772–779. [[CrossRef](#)]
41. Zhao, X.X.; Rong, L.; Zhao, X.X.; Li, X.; Liu, X.; Deng, J.; Wu, H.; Xu, X.; Erben, U.; Wu, P.; et al. TNF Signaling Drives Myeloid-Derived Suppressor Cell Accumulation. *J. Clin. Investig.* **2012**, *122*, 4094–4104. [[CrossRef](#)] [[PubMed](#)]
42. Schröder, M.; Krötschel, M.; Conrad, L.; Naumann, S.K.; Bachran, C.; Rolfe, A.; Umansky, V.; Helming, L.; Swee, L.K. Genetic Screen in Myeloid Cells Identifies TNF- α Autocrine Secretion as a Factor Increasing MDSC Suppressive Activity via Nos2 up-Regulation. *Sci. Rep.* **2018**, *8*, 13399. [[CrossRef](#)] [[PubMed](#)]
43. Xue, Q.; Lu, Y.; Eisele, M.R.; Sulistijo, E.S.; Khan, N.; Fan, R.; Miller-Jensen, K. Analysis of Single-Cell Cytokine Secretion Reveals a Role for Paracrine Signaling in Coordinating Macrophage Responses to TLR4 Stimulation. *Sci. Signal.* **2015**, *8*, ra59. [[CrossRef](#)] [[PubMed](#)]
44. Nagar, M.; Jacob-Hirsch, J.; Vernitsky, H.; Berkun, Y.; Ben-Horin, S.; Amariglio, N.; Bank, I.; Kloog, Y.; Rechavi, G.; Goldstein, I. TNF Activates a NF-KappaB-Regulated Cellular Program in Human CD45RA-Regulatory T Cells That Modulates Their Suppressive Function. *J. Immunol.* **2010**, *184*, 3570–3581. [[CrossRef](#)]
45. Balkwill, F.; Mantovani, A. Inflammation and Cancer: Back to Virchow? *Lancet* **2018**, *357*, 539–545. [[CrossRef](#)]
46. Waters, J.P.; Pober, J.S.; Bradley, J.R. Tumour Necrosis Factor and Cancer. *J. Pathol.* **2013**, *230*, 241–248. [[CrossRef](#)]
47. Bauswein, M.; Singh, A.; Ralhan, A.; Neri, D.; Fuchs, K.; Blanz, K.D.; Schäfer, I.; Hector, A.; Handgretinger, R.; Hartl, D.; et al. Human T Cells Modulate Myeloid-Derived Suppressor Cells through a TNF- α -Mediated Mechanism. *Immunol. Lett.* **2018**, *202*, 31–37. [[CrossRef](#)]
48. Hu, X.; Li, B.; Li, X.; Zhao, X.; Wan, L.; Lin, G.; Yu, M.; Wang, J.; Jiang, X.; Feng, W.; et al. Transmembrane TNF- α Promotes Suppressive Activities of Myeloid-Derived Suppressor Cells via TNFR2. *J. Immunol.* **2014**, *192*, 1320–1331. [[CrossRef](#)]
49. Dong, G.; You, M.; Fan, H.; Ji, J.; Ding, L.; Li, P.; Hou, Y. 17 β -Estradiol Contributes to the Accumulation of Myeloid-Derived Suppressor Cells in Blood by Promoting TNF- α Secretion. *Acta Biochim. Biophys. Sin.* **2015**, *47*, 620–629. [[CrossRef](#)]
50. Greish, K.; Taurin, S.; Morsy, M.A. The Effect of Adjuvant Therapy with TNF- α on Animal Model of Triple-Negative Breast Cancer. *Ther. Deliv.* **2018**, *9*, 333–342. [[CrossRef](#)]
51. Rath, P.C.; Aggarwal, B.B. TNF-Induced Signaling in Apoptosis. *J. Clin. Immunol.* **1999**, *19*, 350–364. [[CrossRef](#)] [[PubMed](#)]
52. Benoit, M.; Desnues, B.; Mege, J.-L. Macrophage Polarization in Bacterial Infections. *J. Immunol.* **2008**, *181*, 3733–3739. [[CrossRef](#)] [[PubMed](#)]
53. Lejeune, F.J.; Liénard, D.; Matter, M.; Rüegg, C. Efficiency of Recombinant Human TNF in Human Cancer Therapy. *Cancer Immun.* **2006**, *6*, 6. [[PubMed](#)]
54. Qiao, Y.; Huang, X.; Nimmagadda, S.; Bai, R.; Staedtke, V.; Foss, C.A.; Cheong, I.; Holdhoff, M.; Kato, Y.; Pomper, M.G.; et al. A Robust Approach to Enhance Tumor-Selective Accumulation of Nanoparticles. *Oncotarget* **2011**, *2*, 59–68. [[CrossRef](#)]
55. Kratochvill, F.; Neale, G.; Haverkamp, J.M.; Van de Velde, L.-A.; Smith, A.M.; Kawauchi, D.; McEvoy, J.; Roussel, M.F.; Dyer, M.A.; Qualls, J.E.; et al. TNF Counterbalances the Emergence of M2 Tumor Macrophages. *Cell Rep.* **2015**, *12*, 1902–1914. [[CrossRef](#)]
56. Hoving, S.; Seynhaeve, A.L.B.; van Tiel, S.T.; Aan de Wiel-Ambagtsheer, G.; de Bruijn, E.A.; Eggermont, A.M.M.; ten Hagen, T.L.M. Early Destruction of Tumor Vasculature in Tumor Necrosis Factor-Alpha-Based Isolated Limb Perfusion Is Responsible for Tumor Response. *Anticancer Drugs* **2006**, *17*, 949–959. [[CrossRef](#)]
57. Mackay, F.; Loetscher, H.; Stueber, D.; Gehr, G.; Lesslauer, W. Tumor Necrosis Factor Alpha (TNF-Alpha)-Induced Cell Adhesion to Human Endothelial Cells Is under Dominant Control of One TNF Receptor Type, TNF-R55. *J. Exp. Med.* **1993**, *177*, 1277–1286. [[CrossRef](#)]
58. Balkwill, F. TNF-Alpha in Promotion and Progression of Cancer. *Cancer Metastasis Rev.* **2006**, *25*, 409–416. [[CrossRef](#)]
59. Mantovani, A.; Allavena, P.; Sica, A.; Balkwill, F. Cancer-Related Inflammation. *Nature* **2008**, *454*, 436–444. [[CrossRef](#)]

60. Cruceriu, D.; Baldasici, O.; Balacescu, O. The Dual Role of Tumor Necrosis Factor-Alpha (TNF- α) in Breast Cancer: Molecular Insights and Therapeutic Approaches. *Cell. Oncol.* **2020**, 1–18. [[CrossRef](#)]
61. Zhao, C.; Lu, X.; Bu, X.; Zhang, N.; Wang, W. Involvement of Tumor Necrosis Factor-Alpha in the Upregulation of CXCR4 Expression in Gastric Cancer Induced by Helicobacter Pylori. *BMC Cancer* **2010**, *10*, 419. [[CrossRef](#)] [[PubMed](#)]
62. Egberts, J.-H.; Cloosters, V.; Noack, A.; Schniewind, B.; Thon, L.; Klose, S.; Kettler, B.; von Forstner, C.; Kneitz, C.; Tepel, J.; et al. Anti-Tumor Necrosis Factor Therapy Inhibits Pancreatic Tumor Growth and Metastasis. *Cancer Res.* **2008**, *68*, 1443–1450. [[CrossRef](#)] [[PubMed](#)]
63. Gupta, M.; Babic, A.; Beck, A.H.; Terry, K. TNF- α Expression, Risk Factors, and Inflammatory Exposures in Ovarian Cancer: Evidence for an Inflammatory Pathway of Ovarian Carcinogenesis? *Hum. Pathol.* **2016**, *54*, 82–91. [[CrossRef](#)] [[PubMed](#)]
64. Naylor, M.S.; Malik, S.T.; Stamp, G.W.; Jobling, T.; Balkwill, F.R. In Situ Detection of Tumour Necrosis Factor in Human Ovarian Cancer Specimens. *Eur. J. Cancer* **1990**, *26*, 1027–1030. [[CrossRef](#)]
65. Morgado, M.; Sutton, M.N.; Simmons, M.; Warren, C.R.; Lu, Z.; Constantinou, P.E.; Liu, J.; Francis, L.L.W.; Steven Conlan, R.; Bast, R.C.; et al. Tumor Necrosis Factor- α and Interferon- γ Stimulate MUC16 (CA125) Expression in Breast, Endometrial and Ovarian Cancers through NF κ B. *Oncotarget* **2016**, *7*, 14871–14884. [[CrossRef](#)]
66. Sethi, G.; Shanmugam, M.K.; Ramachandran, L.; Kumar, A.P.; Tergaonkar, V. Multifaceted Link between Cancer and Inflammation. *Biosci. Rep.* **2012**, *32*, 1–15. [[CrossRef](#)]
67. Li, X.; Wang, S.; Ren, H.J.; Ma, J.; Sun, X.; Li, N.; Liu, C.; Huang, K.; Xu, M.; Ming, L. Molecular Correlates and Prognostic Value of TmTNF- α Expression in Colorectal Cancer of 5-Fluorouracil-Based Adjuvant Therapy. *Cancer Biol. Ther.* **2016**, *17*, 684–692. [[CrossRef](#)]
68. Tang, D.; Tao, D.; Fang, Y.; Deng, C.; Xu, Q.; Zhou, J. TNF-Alpha Promotes Invasion and Metastasis via NF-Kappa B Pathway in Oral Squamous Cell Carcinoma. *Med. Sci. Monit. Basic Res.* **2017**, *23*, 141–149. [[CrossRef](#)]
69. Roberts, R.A.; Kimber, I. Cytokines in Non-Genotoxic Hepatocarcinogenesis. *Carcinogenesis* **1999**, *20*, 1397–1401. [[CrossRef](#)]
70. Burow, M.E.; Weldon, C.B.; Tang, Y.; Navar, G.L.; Krajewski, S.; Reed, J.C.; Hammond, T.G.; Clejan, S.; Beckman, B.S. Differences in Susceptibility to Tumor Necrosis Factor α -Induced Apoptosis among MCF-7 Breast Cancer Cell Variants. *Cancer Res.* **1998**, *58*, 4940–4946.
71. Mercogliano, M.F.; Bruni, S.; Elizalde, P.V.; Schillaci, R. Tumor Necrosis Factor α Blockade: An Opportunity to Tackle Breast Cancer. *Front. Oncol.* **2020**, *10*. [[CrossRef](#)] [[PubMed](#)]
72. Mercogliano, M.F.; De Martino, M.; Venturutti, L.; Rivas, M.A.; Proietti, C.J.; Inurriagarro, G.; Frahm, I.; Allemand, D.H.; Deza, E.G.; Ares, S.; et al. TNF α -Induced Mucin 4 Expression Elicits Trastuzumab Resistance in HER2-Positive Breast Cancer. *Clin. Cancer Res.* **2017**, *23*, 636–648. [[CrossRef](#)] [[PubMed](#)]
73. Wu, C.; Fernandez, S.A.; Criswell, T.; Chidiac, T.A.; Guttridge, D.; Villalona-Calero, M.; Bekaii-Saab, T.S. Disrupting Cytokine Signaling in Pancreatic Cancer: A Phase I/II Study of Etanercept in Combination with Gemcitabine in Patients with Advanced Disease. *Pancreas* **2013**, *42*, 813–818. [[CrossRef](#)]
74. Katerinaki, E.; Evans, G.S.; Lorigan, P.C.; MacNeil, S. TNF-Alpha Increases Human Melanoma Cell Invasion and Migration in Vitro: The Role of Proteolytic Enzymes. *Br. J. Cancer* **2003**, *89*, 1123–1129. [[CrossRef](#)] [[PubMed](#)]
75. Rossi, S.; Cordella, M.; Tabolacci, C.; Nassa, G.; D'Arcangelo, D.; Senatore, C.; Pagnotto, P.; Magliozzi, R.; Salvati, A.; Weisz, A.; et al. TNF-Alpha and Metalloproteases as Key Players in Melanoma Cells Aggressiveness. *J. Exp. Clin. Cancer Res.* **2018**, *37*, 326. [[CrossRef](#)]
76. Bald, T.; Quast, T.; Landsberg, J.; Rogava, M.; Glodde, N.; Lopez-Ramos, D.; Kohlmeyer, J.; Riesenberger, S.; van den Boorn-Konijnenberg, D.; Hömig-Hölzel, C.; et al. Ultraviolet-Radiation-Induced Inflammation Promotes Angiotropism and Metastasis in Melanoma. *Nature* **2014**, *507*, 109–113. [[CrossRef](#)]
77. Bertrand, F.; Rochotte, J.; Colacios, C.; Montfort, A.; Tilkin-Mariamé, A.-F.; Touriol, C.; Rochaix, P.; Lajoie-Mazenc, I.; Andrieu-Abadie, N.; Levade, T.; et al. Blocking Tumor Necrosis Factor α Enhances CD8 T-Cell-Dependent Immunity in Experimental Melanoma. *Cancer Res.* **2015**, *75*, 2619–2628. [[CrossRef](#)]
78. Waterston, A.M.; Salway, F.; Andreacos, E.; Butler, D.M.; Feldmann, M.; Coombes, R.C. TNF Autovaccination Induces Self Anti-TNF Antibodies and Inhibits Metastasis in a Murine Melanoma Model. *Br. J. Cancer* **2004**, *90*, 1279–1284. [[CrossRef](#)]
79. St John, M.A.R.; Li, Y.; Zhou, X.; Denny, P.; Ho, C.-M.; Montemagno, C.; Shi, W.; Qi, F.; Wu, B.; Sinha, U.; et al. Interleukin 6 and Interleukin 8 as Potential Biomarkers for Oral Cavity and Oropharyngeal Squamous Cell Carcinoma. *Arch. Otolaryngol. Head Neck Surg.* **2004**, *130*, 929–935. [[CrossRef](#)]
80. Lee, S.H.; Hong, H.S.; Liu, Z.X.; Kim, R.H.; Kang, M.K.; Park, N.-H.; Shin, K.-H. TNF α Enhances Cancer Stem Cell-like Phenotype via Notch-Hes1 Activation in Oral Squamous Cell Carcinoma Cells. *Biochem. Biophys. Res. Commun.* **2012**, *424*, 58–64. [[CrossRef](#)]
81. Zhou, X.; Zhou, S.; Li, B.; Li, Q.; Gao, L.; Li, D.; Gong, Q.; Zhu, L.; Wang, J.; Wang, N.; et al. Transmembrane TNF- α Preferentially Expressed by Leukemia Stem Cells and Blasts Is a Potent Target for Antibody Therapy. *Blood* **2015**, *126*, 1433–1442. [[CrossRef](#)] [[PubMed](#)]
82. Sivaraman, S.; Deshpande, C.G.; Ranganathan, R.; Huang, X.; Jajeh, A.; O'Brien, T.; Huang, R.W.; Gregory, S.A.; Venugopal, P.; Preisler, H.D. Tumor Necrosis Factor Modulates CD 20 Expression on Cells from Chronic Lymphocytic Leukemia: A New Role for TNF Alpha? *Microsc. Res. Tech.* **2000**, *50*, 251–257. [[CrossRef](#)]
83. Chopra, V.; Dinh, T.V.; Hannigan, E.V. Serum Levels of Interleukins, Growth Factors and Angiogenin in Patients with Endometrial Cancer. *J. Cancer Res. Clin. Oncol.* **1997**, *123*, 167–172. [[CrossRef](#)] [[PubMed](#)]
84. Ellis, P.E.; Barron, G.A.; Bermano, G. Adipocytokines and Their Relationship to Endometrial Cancer Risk: A Systematic Review and Meta-Analysis. *Gynecol. Oncol.* **2020**, *158*, 507–516. [[CrossRef](#)] [[PubMed](#)]

85. Dossus, L.; Becker, S.; Rinaldi, S.; Lukanova, A.; Tjønneland, A.; Olsen, A.; Overvad, K.; Chabbert-Buffet, N.; Boutron-Ruault, M.-C.; Clavel-Chapelon, F.; et al. Tumor Necrosis Factor (TNF)- α , Soluble TNF Receptors and Endometrial Cancer Risk: The EPIC Study. *Int. J. Cancer* **2011**, *129*, 2032–2037. [[CrossRef](#)] [[PubMed](#)]
86. Sharma, J.; Gray, K.P.; Harshman, L.C.; Evan, C.; Nakabayashi, M.; Fichorova, R.; Rider, J.; Mucci, L.; Kantoff, P.W.; Sweeney, C.J. Elevated IL-8, TNF- α , and MCP-1 in Men with Metastatic Prostate Cancer Starting Androgen-Deprivation Therapy (ADT) Are Associated with Shorter Time to Castration-Resistance and Overall Survival. *Prostate* **2014**, *74*, 820–828. [[CrossRef](#)]
87. Banzola, I.; Mengus, C.; Wyler, S.; Hudolin, T.; Manzella, G.; Chiarugi, A.; Boldorini, R.; Sais, G.; Schmidli, T.S.; Chiffi, G.; et al. Expression of Indoleamine 2,3-Dioxygenase Induced by IFN- γ and TNF- α as Potential Biomarker of Prostate Cancer Progression. *Front. Immunol.* **2018**, *9*, 1051. [[CrossRef](#)]
88. Sahibzada, H.A.; Khurshid, Z.; Khan, R.S.; Naseem, M.; Siddique, K.M.; Mali, M.; Zafar, M.S. Salivary IL-8, IL-6 and TNF- α as Potential Diagnostic Biomarkers for Oral Cancer. *Diagnostics* **2017**, *7*, 21. [[CrossRef](#)]
89. Dantas, T.S.; de Barros Silva, P.G.; Lima Verde, M.E.Q.; de Ribeiro Junior, A.L.; do Cunha, M.P.S.S.; Mota, M.R.L.; Alves, A.P.N.N.; de Leitão, R.F.C.; Sousa, F.B. Role of Inflammatory Markers in Prognosis of Oral Squamous Cell Carcinoma. *Asian Pac. J. Cancer Prev.* **2019**, *20*, 3635–3642. [[CrossRef](#)]
90. Jaime-Pérez, J.C.; Gamboa-Alonso, C.M.; Jiménez-Castillo, R.A.; López-Silva, L.J.; Pinzón-Uresti, M.A.; Gómez-De León, A.; Gómez-Almaguer, D. TNF- α Increases in the CSF of Children with Acute Lymphoblastic Leukemia before CNS Relapse. *Blood Cells Mol. Dis.* **2017**, *63*, 27–31. [[CrossRef](#)]
91. Potapnev, M.P.; Petyovka, N.V.; Belevtsev, M.V.; Savitskiy, V.P.; Migal, N.V. Plasma Level of Tumor Necrosis Factor-Alpha (TNF-Alpha) Correlates with Leukocytosis and Biological Features of Leukemic Cells, but Not Treatment Response of Children with Acute Lymphoblastic Leukemia. *Leuk. Lymphoma* **2003**, 1077–1079. [[CrossRef](#)] [[PubMed](#)]
92. Macia, J.; Gomez, X.; Esquerda, A.; Perez, B.; Callao, V.; Marzo, C. Value of the Determination of TNF-Alpha in the Plasma of Patients with Non-Hodgkins Lymphoma. *Leuk. Lymphoma* **1996**, *20*, 481–486. [[CrossRef](#)] [[PubMed](#)]
93. Lech-Maranda, E.; Bienvenu, J.; Broussais-Guillaumot, F.; Warzocha, K.; Michallet, A.-S.; Robak, T.; Coiffier, B.; Salles, G. Plasma TNF-Alpha and IL-10 Level-Based Prognostic Model Predicts Outcome of Patients with Diffuse Large B-Cell Lymphoma in Different Risk Groups Defined by the International Prognostic Index. *Arch. Immunol. Exp.* **2010**, *58*, 131–141. [[CrossRef](#)]
94. Nakayama, S.; Yokote, T.; Tsuji, M.; Akioka, T.; Miyoshi, T.; Hirata, Y.; Hiraoka, N.; Iwaki, K.; Takayama, A.; Nishiwaki, U.; et al. TNF- α Receptor 1 Expression Predicts Poor Prognosis of Diffuse Large B-Cell Lymphoma, Not Otherwise Specified. *Am. J. Surg. Pathol.* **2014**, *38*, 1138–1146. [[CrossRef](#)] [[PubMed](#)]
95. Dantas, R.N.; de Souza, A.M.; Herrero, S.S.T.; Kassab, P.; Malheiros, C.A.; Lima, E.M. Association between PSCA, TNF- α , PARP1 and TP53 Gene Polymorphisms and Gastric Cancer Susceptibility in the Brazilian Population. *Asian Pac. J. Cancer Prev.* **2020**, *21*, 43–48. [[CrossRef](#)]
96. Du, L.C.; Gao, R. Role of TNF- α -308G/A Gene Polymorphism in Gastric Cancer Risk: A Case Control Study and Meta-Analysis. *J. Turk. Soc. Gastroenterol.* **2017**, *28*, 272–282. [[CrossRef](#)]
97. Pardo, T.; Salcedo, P.; Quintero, J.M.; Borjas, L.; Fernández-Mestre, M.; Sánchez, Y.; Carrillo, Z.; Rivera, S. Study of the association between the polymorphism of the TNF- α gene and prostate cancer. *Rev. Alerg. Mex.* **2019**, *66*, 154–162. [[CrossRef](#)]
98. Shi, H.-Z.; Ren, P.; Lu, Q.-J.; Niedrgethmann, M.; Wu, G.-Y. Association between EGF, TGF-B1 and TNF- α Gene Polymorphisms and Hepatocellular Carcinoma. *Asian Pac. J. Cancer Prev.* **2012**, *13*, 6217–6220. [[CrossRef](#)]
99. Verma, H.K.; Merchant, N.; Bhaskar, L.V.K.S. Tumor Necrosis Factor-Alpha Gene Promoter (TNF- α G-308A) Polymorphisms Increase the Risk of Hepatocellular Carcinoma in Asians: A Meta-Analysis. *Crit. Rev. Oncog.* **2020**, *25*, 11–20. [[CrossRef](#)]
100. He, Y.-Q.; Zhu, J.-H.; Huang, S.-Y.; Cui, Z.; He, J.; Jia, W.-H. The Association between the Polymorphisms of TNF- α and Non-Hodgkin Lymphoma: A Meta-Analysis. *Tumour Biol. J. Int. Soc. Oncodevelopmental Biol. Med.* **2014**, *35*, 12509–12517. [[CrossRef](#)]
101. Liu, J.; Liu, J.; Song, B.; Wang, T.; Liu, Y.; Hao, J.; Yu, J. Genetic Variations in CTLA-4, TNF- α , and LTA and Susceptibility to T-Cell Lymphoma in a Chinese Population. *Cancer Epidemiol.* **2013**, *37*, 930–934. [[CrossRef](#)] [[PubMed](#)]
102. Hellmig, S.; Fischbach, W.; Goebeler-Kolve, M.-E.; Fölsch, U.R.; Hampe, J.; Schreiber, S. A Functional Promotor Polymorphism of TNF-Alpha Is Associated with Primary Gastric B-Cell Lymphoma. *Am. J. Gastroenterol.* **2005**, *100*, 2644–2649. [[CrossRef](#)] [[PubMed](#)]
103. Ahmed, A.B.; Zidi, S.; Sghaier, I.; Ghazouani, E.; Mezlini, A.; Almawi, W.; Loueslati, B.Y. Common Variants in IL-1RN, IL-1 β and TNF- α and the Risk of Ovarian Cancer: A Case Control Study. *Cent. J. Immunol.* **2017**, *42*, 150–155. [[CrossRef](#)] [[PubMed](#)]
104. Kanda, K.; Komekado, H.; Sawabu, T.; Ishizu, S.; Nakanishi, Y.; Nakatsuji, M.; Akitake-Kawano, R.; Ohno, M.; Hiraoka, Y.; Kawada, M.; et al. Nardilysin and ADAM Proteases Promote Gastric Cancer Cell Growth by Activating Intrinsic Cytokine Signalling via Enhanced Ectodomain Shedding of TNF- α . *Embo Mol. Med.* **2012**, *4*, 396–411. [[CrossRef](#)]
105. Mochizuki, Y.; Nakanishi, H.; Kodera, Y.; Ito, S.; Yamamura, Y.; Kato, T.; Hibi, K.; Akiyama, S.; Nakao, A.; Tatematsu, M. TNF-Alpha Promotes Progression of Peritoneal Metastasis as Demonstrated Using a Green Fluorescence Protein (GFP)-Tagged Human Gastric Cancer Cell Line. *Clin. Exp. Metastasis* **2004**, *21*, 39–47. [[CrossRef](#)]
106. Cui, X.; Zhang, H.; Cao, A.; Cao, L.; Hu, X. Cytokine TNF- α Promotes Invasion and Metastasis of Gastric Cancer by down-Regulating Pentraxin3. *J. Cancer* **2020**, *11*, 1800–1807. [[CrossRef](#)]
107. Zhou, Q.; Wu, X.; Wang, X.; Yu, Z.; Pan, T.; Li, Z.; Chang, X.; Jin, Z.; Li, J.; Zhu, Z.; et al. The Reciprocal Interaction between Tumor Cells and Activated Fibroblasts Mediated by TNF- α /IL-33/ST2L Signaling Promotes Gastric Cancer Metastasis. *Oncogene* **2020**, *39*, 1414–1428. [[CrossRef](#)]

108. Kim, S.; Choi, M.G.; Lee, H.S.; Lee, S.K.; Kim, S.H.; Kim, W.W.; Hur, S.M.; Kim, J.-H.; Choe, J.-H.; Nam, S.J.; et al. Silibinin Suppresses TNF-Alpha-Induced MMP-9 Expression in Gastric Cancer Cells through Inhibition of the MAPK Pathway. *Molecules* **2009**, *14*, 4300–4311. [[CrossRef](#)]
109. Ha Thi, H.T.; Lim, H.-S.; Kim, J.; Kim, Y.-M.; Kim, H.-Y.; Hong, S. Transcriptional and Post-Translational Regulation of Bim Is Essential for TGF- β and TNF- α -Induced Apoptosis of Gastric Cancer Cell. *Biochim. Biophys. Acta* **2013**, *1830*, 3584–3592. [[CrossRef](#)]
110. Alam, M.S.; Gaida, M.M.; Bergmann, F.; Lasitschka, F.; Giese, T.; Giese, N.A.; Hackert, T.; Hinz, U.; Hussain, S.P.; Kozlov, S.V.; et al. Selective Inhibition of the P38 Alternative Activation Pathway in Infiltrating T Cells Inhibits Pancreatic Cancer Progression. *Nat. Med.* **2015**, *21*, 1337–1343. [[CrossRef](#)]
111. Aida, K.; Miyakawa, R.; Suzuki, K.; Narumi, K.; Udagawa, T.; Yamamoto, Y.; Chikaraishi, T.; Yoshida, T.; Aoki, K. Suppression of Tregs by Anti-Glucocorticoid Induced TNF Receptor Antibody Enhances the Antitumor Immunity of Interferon- α Gene Therapy for Pancreatic Cancer. *Cancer Sci.* **2014**, *105*, 159–167. [[CrossRef](#)] [[PubMed](#)]
112. Bharadwaj, U.; Marin-Muller, C.; Li, M.; Chen, C.; Yao, Q. Mesothelin Confers Pancreatic Cancer Cell Resistance to TNF- α -Induced Apoptosis through Akt/PI3K/NF-KB Activation and IL-6/Mcl-1 Overexpression. *Mol. Cancer* **2011**, *10*, 106. [[CrossRef](#)] [[PubMed](#)]
113. Guo, X.; Li, T.; Wang, Y.; Shao, L.; Zhang, Y.; Ma, D.; Han, W. CMTM5 Induces Apoptosis of Pancreatic Cancer Cells and Has Synergistic Effects with TNF-Alpha. *Biochem. Biophys. Res. Commun.* **2009**, *387*, 139–142. [[CrossRef](#)] [[PubMed](#)]
114. Murugesan, S.R.; King, C.R.; Osborn, R.; Fairweather, W.R.; O'Reilly, E.M.; Thornton, M.O.; Wei, L.L. Combination of Human Tumor Necrosis Factor-Alpha (HTNF-Alpha) Gene Delivery with Gemcitabine Is Effective in Models of Pancreatic Cancer. *Cancer Gene Ther.* **2009**, *16*, 841–847. [[CrossRef](#)] [[PubMed](#)]
115. Tsukamoto, M.; Imai, K.; Ishimoto, T.; Komohara, Y.; Yamashita, Y.-I.; Nakagawa, S.; Umezaki, N.; Yamao, T.; Kitano, Y.; Miyata, T.; et al. PD-L1 Expression Enhancement by Infiltrating Macrophage-Derived Tumor Necrosis Factor- α Leads to Poor Pancreatic Cancer Prognosis. *Cancer Sci.* **2019**, *110*, 310–320. [[CrossRef](#)]
116. Charles, K.A.; Kulbe, H.; Soper, R.; Escorcio-Correia, M.; Lawrence, T.; Schultheis, A.; Chakravarty, P.; Thompson, R.G.; Kollias, G.; Smyth, J.F.; et al. The Tumor-Promoting Actions of TNF-Alpha Involve TNFR1 and IL-17 in Ovarian Cancer in Mice and Humans. *J. Clin. Investig.* **2009**, *119*, 3011–3023. [[CrossRef](#)] [[PubMed](#)]
117. Hassan, M.I.; Kassim, S.K.; Saeda, L.; Laban, M.; Khalifa, A. Ovarian Cancer-Induced Immunosuppression: Relationship to Tumor Necrosis Factor-Alpha (TNF-Alpha) Release from Ovarian Tissue. *Anticancer Res.* **1999**, *19*, 5657–5662.
118. Wu, S.; Rodabaugh, K.; Martinez-Maza, O.; Watson, J.M.; Silberstein, D.S.; Boyer, C.M.; Peters, W.P.; Weinberg, J.B.; Berek, J.S.; Bast, R.C.J. Stimulation of Ovarian Tumor Cell Proliferation with Monocyte Products Including Interleukin-1, Interleukin-6, and Tumor Necrosis Factor-Alpha. *Am. J. Obstet. Gynecol.* **1992**, *166*, 997–1007. [[CrossRef](#)]
119. Li, H.; Chen, A.; Yuan, Q.; Chen, W.; Zhong, H.; Teng, M.; Xu, C.; Qiu, Y.; Cao, J. NF-KB/Twist Axis Is Involved in Chysin Inhibition of Ovarian Cancer Stem Cell Features Induced by Co-Treatment of TNF- α and TGF- β . *Int. J. Clin. Exp. Pathol.* **2019**, *12*, 101–112.
120. Kim, D.S.; Jang, Y.-J.; Jeon, O.-H.; Kim, D.-S. Saxatilin, a Snake Venom Disintegrin, Suppresses TNF-Alpha-Induced Ovarian Cancer Cell Invasion. *J. Biochem. Mol. Biol.* **2007**, *40*, 290–294. [[CrossRef](#)]
121. Schröder, S.K.; Asimakopoulou, A.; Tillmann, S.; Koschmieder, S.; Weiskirchen, R. TNF- α Controls Lipocalin-2 Expression in PC-3 Prostate Cancer Cells. *Cytokine* **2020**, *135*, 155214. [[CrossRef](#)] [[PubMed](#)]
122. Safari, H.; Zabihi, E.; Pouramir, M.; Morakabati, P.; Abedian, Z.; Karkhah, A.; Nouri, H.R. Decrease of Intracellular ROS by Arbutin Is Associated with Apoptosis Induction and Downregulation of IL-1 β and TNF- α in LNCaP; Prostate Cancer. *J. Food Biochem.* **2020**, e13360. [[CrossRef](#)] [[PubMed](#)]
123. Mu, H.Q.; He, Y.H.; Wang, S.B.; Yang, S.; Wang, Y.J.; Nan, C.J.; Bao, Y.F.; Xie, Q.P.; Chen, Y.H. MiR-130b/TNF- α /NF-KB/VEGFA Loop Inhibits Prostate Cancer Angiogenesis. *Clin. Transl. Oncol.* **2020**, *22*, 111–121. [[CrossRef](#)] [[PubMed](#)]
124. Lv, L.; Yuan, J.; Huang, T.; Zhang, C.; Zhu, Z.; Wang, L.; Jiang, G.; Zeng, F. Stabilization of Snail by HIF-1 α and TNF- α Is Required for Hypoxia-Induced Invasion in Prostate Cancer PC3 Cells. *Mol. Biol. Rep.* **2014**, *41*, 4573–4582. [[CrossRef](#)]
125. Maolake, A.; Izumi, K.; Natsagdorj, A.; Iwamoto, H.; Kadomoto, S.; Makino, T.; Naito, R.; Shigehara, K.; Kadono, Y.; Hiratsuka, K.; et al. Tumor Necrosis Factor- α Induces Prostate Cancer Cell Migration in Lymphatic Metastasis through CCR7 Upregulation. *Cancer Sci.* **2018**, *109*, 1524–1531. [[CrossRef](#)]
126. Wang, H.; Fang, R.; Wang, X.-F.; Zhang, F.; Chen, D.-Y.; Zhou, B.; Wang, H.-S.; Cai, S.-H.; Du, J. Stabilization of Snail through AKT/GSK-3 β Signaling Pathway Is Required for TNF- α -Induced Epithelial-Mesenchymal Transition in Prostate Cancer PC3 Cells. *Eur. J. Pharm.* **2013**, *714*, 48–55. [[CrossRef](#)]
127. Pilling, A.B.; Hwang, O.; Boudreault, A.; Laurent, A.; Hwang, C. IAP Antagonists Enhance Apoptotic Response to Enzalutamide in Castration-Resistant Prostate Cancer Cells via Autocrine TNF- α Signaling. *Prostate* **2017**, *77*, 866–877. [[CrossRef](#)]
128. Shi, J.; Chen, J.; Serradji, N.; Xu, X.; Zhou, H.; Ma, Y.; Sun, Z.; Jiang, P.; Du, Y.; Yang, J.; et al. PMS1077 Sensitizes TNF- α Induced Apoptosis in Human Prostate Cancer Cells by Blocking NF-KB Signaling Pathway. *PLoS ONE* **2013**, *8*, e61132. [[CrossRef](#)]
129. Lee, E.-J.; Kim, W.-J.; Moon, S.-K. Cordycepin Suppresses TNF-Alpha-Induced Invasion, Migration and Matrix Metalloproteinase-9 Expression in Human Bladder Cancer Cells. *Phytother. Res.* **2010**, *24*, 1755–1761. [[CrossRef](#)]
130. Lee, S.-J.; Park, S.-S.; Lee, U.-S.; Kim, W.-J.; Moon, S.-K. Signaling Pathway for TNF-Alpha-Induced MMP-9 Expression: Mediation through P38 MAP Kinase, and Inhibition by Anti-Cancer Molecule Magnolol in Human Urinary Bladder Cancer 5637 Cells. *Int. Immunopharmacol.* **2008**, *8*, 1821–1826. [[CrossRef](#)]

131. Lee, S.-J.; Park, S.-S.; Cho, Y.-H.; Park, K.; Kim, E.-J.; Jung, K.-H.; Kim, S.-K.; Kim, W.-J.; Moon, S.-K. Activation of Matrix Metalloproteinase-9 by TNF-Alpha in Human Urinary Bladder Cancer HT1376 Cells: The Role of MAP Kinase Signaling Pathways. *Oncol. Rep.* **2008**, *19*, 1007–1013. [[PubMed](#)]
132. Yang, T.; Shi, R.; Chang, L.; Tang, K.; Chen, K.; Yu, G.; Tian, Y.; Guo, Y.; He, W.; Song, X.; et al. Huachansu Suppresses Human Bladder Cancer Cell Growth through the Fas/FasL and TNF-Alpha/TNFR1 Pathway in Vitro and in Vivo. *J. Exp. Clin. Cancer Res.* **2015**, *34*, 21. [[CrossRef](#)] [[PubMed](#)]
133. Wang, Z.; Cheng, Y.; Zheng, R.; Qin, D.; Liu, G. Effect of TNF-Alpha and IFN-Alpha on the Proliferation and Cytotoxicity of Lymphokine-Activated Killer Cells in Patients with Bladder Cancer. *Chin. Med. J.* **1997**, *110*, 180–183. [[PubMed](#)]
134. Grimm, M.; Lazariotou, M.; Kircher, S.; Höfelmayer, A.; Germer, C.T.; von Rahden, B.H.A.; Waaga-Gasser, A.M.; Gasser, M. Tumor Necrosis Factor- α Is Associated with Positive Lymph Node Status in Patients with Recurrence of Colorectal Cancer—Indications for Anti-TNF- α Agents in Cancer Treatment. *Anal. Cell. Pathol.* **2010**, *33*, 151–163. [[CrossRef](#)]
135. Shen, Z.; Zhou, R.; Liu, C.; Wang, Y.; Zhan, W.; Shao, Z.; Liu, J.; Zhang, F.; Xu, L.; Zhou, X.; et al. MicroRNA-105 Is Involved in TNF- α -Related Tumor Microenvironment Enhanced Colorectal Cancer Progression. *Cell Death Dis.* **2017**, *8*, 3213. [[CrossRef](#)]
136. Møller, T.; James, J.P.; Holmstrøm, K.; Sørensen, F.B.; Lindebjerg, J.; Nielsen, B.S. Co-Detection of MiR-21 and TNF- α mRNA in Budding Cancer Cells in Colorectal Cancer. *Int. J. Mol. Sci.* **2019**, *20*, 1907. [[CrossRef](#)]
137. Kassouf, N.; Thornhill, M.H. Oral Cancer Cell Lines Can Use Multiple Ligands, Including Fas-L, TRAIL and TNF-Alpha, to Induce Apoptosis in Jurkat T Cells: Possible Mechanisms for Immune Escape by Head and Neck Cancers. *Oral Oncol.* **2008**, *44*, 672–682. [[CrossRef](#)]
138. Iulia Irimie, A.; Braicu, C.; Zanoaga, O.; Pileczki, V.; Soritau, O.; Berindan-Neagoe, I.; Septimiu Campian, R. Inhibition of Tumor Necrosis Factor Alpha Using RNA Interference in Oral Squamous Cell Carcinoma. *J. Buon.* **2015**, *20*, 1107–1114.
139. Lai, K.-C.; Liu, C.-J.; Lin, T.-J.; Mar, A.-C.; Wang, H.-H.; Chen, C.-W.; Hong, Z.-X.; Lee, T.-C. Blocking TNF- α Inhibits Angiogenesis and Growth of IFIT2-Depleted Metastatic Oral Squamous Cell Carcinoma Cells. *Cancer Lett.* **2016**, *370*, 207–215. [[CrossRef](#)]
140. Hsing, E.-W.; Shiah, S.-G.; Peng, H.-Y.; Chen, Y.-W.; Chuu, C.-P.; Hsiao, J.-R.; Lyu, P.-C.; Chang, J.-Y. TNF- α -Induced MiR-450a Mediates TMEM182 Expression to Promote Oral Squamous Cell Carcinoma Motility. *PLoS ONE* **2019**, *14*, e0213463. [[CrossRef](#)]
141. Lin, X.; Yang, Y.; Guo, Y.; Liu, H.; Jiang, J.; Zheng, F.; Wu, B. PTTG1 Is Involved in TNF- α -Related Hepatocellular Carcinoma via the Induction of c-Myc. *Cancer Med.* **2019**, *8*, 5702–5715. [[CrossRef](#)] [[PubMed](#)]
142. Yang, R.; Liu, Q.; Rescorla, F.J.; Grosfeld, J.L. Experimental Liver Cancer: Improved Response after Hepatic Artery Ligation and Infusion of Tumor Necrosis Factor-Alpha and Interferon-Gamma. *Surgery* **1995**, *118*, 764–768. [[CrossRef](#)]
143. Zhang, Y.H.; Yan, H.Q.; Wang, F.; Wang, Y.Y.; Jiang, Y.N.; Wang, Y.N.; Gao, F.G. TIPE2 Inhibits TNF- α -Induced Hepatocellular Carcinoma Cell Metastasis via Erk1/2 Downregulation and NF-KB Activation. *Int. J. Oncol.* **2015**, *46*, 254–264. [[CrossRef](#)] [[PubMed](#)]
144. Zhang, G.-P.; Yue, X.; Li, S.-Q. Cathepsin C Interacts with TNF- α /P38 MAPK Signaling Pathway to Promote Proliferation and Metastasis in Hepatocellular Carcinoma. *Cancer Res. Treat.* **2020**, *52*, 10–23. [[CrossRef](#)] [[PubMed](#)]
145. Chen, Y.; Wen, H.; Zhou, C.; Su, Q.; Lin, Y.; Xie, Y.; Huang, Y.; Qiu, Q.; Lin, J.; Huang, X.; et al. TNF- α Derived from M2 Tumor-Associated Macrophages Promotes Epithelial-Mesenchymal Transition and Cancer Stemness through the Wnt/ β -Catenin Pathway in SMMC-7721 Hepatocellular Carcinoma Cells. *Exp. Cell Res.* **2019**, *378*, 41–50. [[CrossRef](#)]
146. Li, N.; Wang, J.; Zhang, N.; Zhuang, M.; Zong, Z.; Zou, J.; Li, G.; Wang, X.; Zhou, H.; Zhang, L.; et al. Cross-Talk between TNF- α and IFN- γ Signaling in Induction of B7-H1 Expression in Hepatocellular Carcinoma Cells. *Cancer Immunol. Immunother.* **2018**, *67*, 271–283. [[CrossRef](#)]
147. Curnis, F.; Sacchi, A.; Borgna, L.; Magni, F.; Gasparri, A.; Corti, A. Enhancement of Tumor Necrosis Factor Alpha Antitumor Immunotherapeutic Properties by Targeted Delivery to Aminopeptidase N (CD13). *Nat. Biotechnol.* **2000**, *18*, 1185–1190. [[CrossRef](#)]
148. Cervera-Carrascon, V.; Siurala, M.; Santos, J.M.; Havunen, R.; Tähtinen, S.; Karell, P.; Sorsa, S.; Kanerva, A.; Hemminki, A. TNF α and IL-2 Armed Adenoviruses Enable Complete Responses by Anti-PD-1 Checkpoint Blockade. *Oncoimmunology* **2018**, *7*, e1412902. [[CrossRef](#)]
149. Broussard, L.; Howland, A.; Ryu, S.; Song, K.; Norris, D.; Armstrong, C.A.; Song, P.I. Melanoma Cell Death Mechanisms. *Chonnam Med. J.* **2018**, *54*, 135–142. [[CrossRef](#)]
150. Warzocha, K.; Robak, T. Antileukemic Effects of Recombinant Human Tumor Necrosis Factor Alpha (Rh-TNF Alpha) with Cyclophosphamide or Methotrexate on Leukemia L1210 and Leukemia P388 in Mice. *Acta Haematol. Pol.* **1992**, *23*, 55–62.
151. Kitajima, I.; Nakajima, T.; Imamura, T.; Takasaki, I.; Kawahara, K.; Okano, T.; Tokioka, T.; Soejima, Y.; Abeyama, K.; Maruyama, I. Induction of Apoptosis in Murine Clonal Osteoblasts Expressed by Human T-Cell Leukemia Virus Type I Tax by NF-Kappa B and TNF-Alpha. *J. Bone Miner. Res.* **1996**, *11*, 200–210. [[CrossRef](#)]
152. Gautam, S.C.; Pindolia, K.R.; Xu, Y.X.; Janakiraman, N.; Chapman, R.A.; Freytag, S.O. Antileukemic Activity of TNF-Alpha Gene Therapy with Myeloid Progenitor Cells against Minimal Leukemia. *J. Hematother.* **1998**, *7*, 115–125. [[CrossRef](#)] [[PubMed](#)]
153. Qin, Y.; Auh, S.; Blokh, L.; Long, C.; Gagnon, I.; Hamann, K.J. TNF-Alpha Induces Transient Resistance to Fas-Induced Apoptosis in Eosinophilic Acute Myeloid Leukemia Cells. *Cell. Mol. Immunol.* **2007**, *4*, 43–52.
154. D’Alessandro, N.; Flugy, A.; Tolomeo, M.; Dusonchet, L. The Apoptotic Signaling of TNF-Alpha in Multidrug Resistant Friend Leukemia Cells. *Anticancer Res.* **1998**, *18*, 3065–3072. [[PubMed](#)]
155. Kagoya, Y.; Yoshimi, A.; Kataoka, K.; Nakagawa, M.; Kumano, K.; Arai, S.; Kobayashi, H.; Saito, T.; Iwakura, Y.; Kurokawa, M. Positive Feedback between NF-KB and TNF- α Promotes Leukemia-Initiating Cell Capacity. *J. Clin. Investig.* **2014**, *124*, 528–542. [[CrossRef](#)] [[PubMed](#)]

156. Brailly, H.; Pebusque, M.J.; Tabilio, A.; Mannoni, P. TNF Alpha Acts in Synergy with GM-CSF to Induce Proliferation of Acute Myeloid Leukemia Cells by up-Regulating the GM-CSF Receptor and GM-CSF Gene Expression. *Leukemia* **1993**, *7*, 1557–1563.
157. Zhang, H.; Yan, D.; Shi, X.; Liang, H.; Pang, Y.; Qin, N.; Chen, H.; Wang, J.; Yin, B.; Jiang, X.; et al. Transmembrane TNF-Alpha Mediates “Forward” and “Reverse” Signaling, Inducing Cell Death or Survival via the NF-KappaB Pathway in Raji Burkitt Lymphoma Cells. *J. Leukoc. Biol.* **2008**, *84*, 789–797. [[CrossRef](#)] [[PubMed](#)]
158. Saudemont, A.; Corm, S.; Wickham, T.; Hetuin, D.; Quesnel, B. Induction of Leukemia-Specific CD8+ Cytotoxic T Cells with Autologous Myeloid Leukemic Cells Maturated with a Fiber-Modified Adenovirus Encoding TNF-Alpha. *Mol. Ther.* **2005**, *11*, 950–959. [[CrossRef](#)] [[PubMed](#)]
159. Heilig, B.; Mapara, M.; Bargou, R.; Fiehn, C.; Dörken, B. TNF Alpha Therapy Activates Human B-Lymphoma Cells in Vivo and May Protect Myelopoiesis. *Leuk. Res.* **1992**, *16*, 769–773. [[CrossRef](#)]
160. Di Pietro, R.; Robuffo, I.; Pucci, A.M.; Bosco, D.; Santavenere, E. Effects of TNF-Alpha/Colchicine Combined Treatment on Burkitt Lymphoma Cells: Molecular and Ultrastructural Changes. *Cytokine* **1999**, *11*, 144–150. [[CrossRef](#)]
161. Gupta, U.; Hira, S.K.; Singh, R.; Paladhi, A.; Srivastava, P.; Pratim Manna, P. Essential Role of TNF- α in Gamma c Cytokine Aided Crosstalk between Dendritic Cells and Natural Killer Cells in Experimental Murine Lymphoma. *Int. Immunopharmacol.* **2020**, *78*, 106031. [[CrossRef](#)] [[PubMed](#)]
162. Perez, C.R.; De Palma, M. Engineering Dendritic Cell Vaccines to Improve Cancer Immunotherapy. *Nat. Commun.* **2019**, *10*, 5408. [[CrossRef](#)] [[PubMed](#)]
163. Bowen, W.S.; Srivastava, A.K.; Batra, L.; Barsoumian, H.; Shirwan, H. Current Challenges for Cancer Vaccine Adjuvant Development. *Expert Rev. Vaccines* **2018**, *17*, 207–215. [[CrossRef](#)] [[PubMed](#)]
164. Aurisicchio, L.; Pallocca, M.; Ciliberto, G.; Palombo, F. The Perfect Personalized Cancer Therapy: Cancer Vaccines against Neoantigens. *J. Exp. Clin. Cancer Res.* **2018**, *37*, 86. [[CrossRef](#)] [[PubMed](#)]
165. Schillaci, R.; Bruni, S.; De Martino, M.; Mercogliano, M.F.; Inurrigarro, G.; Frahm, I.; Proietti, C.J.; Elizalde, P.V. Abstract P6-20-14: Neutralizing Soluble Tumor Necrosis Factor Alpha Overcomes Trastuzumab-Resistant Breast Cancer Immune Evasion by Down-regulating Mucin 4, Improving NK Cell Function and Decreasing Myeloid-Derived Suppressor Cells in Tumor Microenvironmen. *Cancer Res.* **2019**, *79* (Suppl. S4), P6-20-14. [[CrossRef](#)]
166. Tsimberidou, A.-M.; Thomas, D.; O’Brien, S.; Andreeff, M.; Kurzrock, R.; Keating, M.; Albitar, M.; Kantarjian, H.; Giles, F. Recombinant Human Soluble Tumor Necrosis Factor (TNF) Receptor (P75) Fusion Protein Enbrel in Patients with Refractory Hematologic Malignancies. *Cancer Chemother. Pharm.* **2002**, *50*, 237–242. [[CrossRef](#)]
167. Woyach, J.A.; Lin, T.S.; Lucas, M.S.; Heerema, N.; Moran, M.E.; Cheney, C.; Lucas, D.M.; Wei, L.; Caligiuri, M.A.; Byrd, J.C. A Phase I/II Study of Rituximab and Etanercept in Patients with Chronic Lymphocytic Leukemia and Small Lymphocytic Lymphoma. *Leukemia* **2009**, *23*, 912–918. [[CrossRef](#)]
168. Bertrand, F.; Montfort, A.; Marcheteau, E.; Imbert, C.; Gilhodes, J.; Filleron, T.; Rochaix, P.; Andrieu-Abadie, N.; Levade, T.; Meyer, N.; et al. TNF α Blockade Overcomes Resistance to Anti-PD-1 in Experimental Melanoma. *Nat. Commun.* **2017**, *8*. [[CrossRef](#)]
169. Stucci, S.; Palmirotta, R.; Passarelli, A.; Silvestris, E.; Argentiero, A.; Lanotte, L.; Acquafredda, S.; Todisco, A.; Silvestris, F. Immune-Related Adverse Events during Anticancer Immunotherapy: Pathogenesis and Management. *Oncol. Lett.* **2017**, *14*, 5671–5680. [[CrossRef](#)]
170. Puzanov, I.; Diab, A.; Abdallah, K.; Bingham III, C.O.; Brogdon, C.; Dadu, R.; Hamad, L.; Kim, S.; Lacouture, M.E.; LeBoeuf, N.R.; et al. Managing Toxicities Associated with Immune Checkpoint Inhibitors: Consensus Recommendations from the Society for Immunotherapy of Cancer (SITC) Toxicity Management Working Group. *J. Immunother. Cancer* **2017**, *5*, 95. [[CrossRef](#)]
171. Badran, Y.R.; Cohen, J.V.; Brastianos, P.K.; Parikh, A.R.; Hong, T.S.; Dougan, M. Concurrent Therapy with Immune Checkpoint Inhibitors and TNF α Blockade in Patients with Gastrointestinal Immune-Related Adverse Events. *J. Immunother. Cancer* **2019**, *7*, 226. [[CrossRef](#)] [[PubMed](#)]
172. Perez-Ruiz, E.; Minute, L.; Otano, I.; Alvarez, M.; Ochoa, M.C.; Belsue, V.; de Andrea, C.; Rodriguez-Ruiz, M.E.; Perez-Gracia, J.L.; Marquez-Rodas, I.; et al. Prophylactic TNF Blockade Uncouples Efficacy and Toxicity in Dual CTLA-4 and PD-1 Immunotherapy. *Nature* **2019**, *569*, 428–432. [[CrossRef](#)] [[PubMed](#)]
173. Arriola, E.; Wheeler, M.; Karydis, I.; Thomas, G.; Ottensmeier, C. Infliximab for IPILIMUMAB-Related Colitis-Letter. *Clin. Cancer Res.* **2015**, *21*, 5642–5643. [[CrossRef](#)] [[PubMed](#)]
174. Horvat, T.Z.; Adel, N.G.; Dang, T.-O.; Momtaz, P.; Postow, M.A.; Callahan, M.K.; Carvajal, R.D.; Dickson, M.A.; D’Angelo, S.P.; Woo, K.M.; et al. Immune-Related Adverse Events, Need for Systemic Immunosuppression, and Effects on Survival and Time to Treatment Failure in Patients With Melanoma Treated With Ipilimumab at Memorial Sloan Kettering Cancer Center. *J. Clin. Oncol.* **2015**, *33*, 3193–3198. [[CrossRef](#)]
175. Figueroa, J.A.; Reidy, A.; Mirandola, L.; Trotter, K.; Suvorava, N.; Figueroa, A.; Konala, V.; Aulakh, A.; Littlefield, L.; Grizzi, F.; et al. Chimeric Antigen Receptor Engineering: A Right Step in the Evolution of Adoptive Cellular Immunotherapy. *Int. Rev. Immunol.* **2015**, *34*, 154–187. [[CrossRef](#)]
176. von Mehren, M.; Adams, G.P.; Weiner, L.M. Monoclonal Antibody Therapy for Cancer. *Annu. Rev. Med.* **2003**, *54*, 343–369. [[CrossRef](#)]
177. Adams, G.P.; Weiner, L.M. Monoclonal Antibody Therapy of Cancer. *Nat. Biotechnol.* **2005**, *23*, 1147–1157. [[CrossRef](#)]
178. Harding, F.A.; Stickler, M.M.; Razo, J.; DuBridge, R.B. The Immunogenicity of Humanized and Fully Human Antibodies: Residual Immunogenicity Resides in the CDR Regions. *MAbs* **2010**, *2*, 256–265. [[CrossRef](#)]

179. Luo, J.-L.; Maeda, S.; Hsu, L.-C.; Yagita, H.; Karin, M. Inhibition of NF-KappaB in Cancer Cells Converts Inflammation- Induced Tumor Growth Mediated by TNFalpha to TRAIL-Mediated Tumor Regression. *Cancer Cell* **2004**, *6*, 297–305. [[CrossRef](#)]
180. Orosz, P.; Echtenacher, B.; Falk, W.; Rüschoff, J.; Weber, D.; Männel, D.N. Enhancement of Experimental Metastasis by Tumor Necrosis Factor. *J. Exp. Med.* **1993**, *177*, 1391–1398. [[CrossRef](#)]
181. Kitakata, H.; Nemoto-Sasaki, Y.; Takahashi, Y.; Kondo, T.; Mai, M.; Mukaida, N. Essential Roles of Tumor Necrosis Factor Receptor P55 in Liver Metastasis of Intrasplenic Administration of Colon 26 Cells. *Cancer Res.* **2002**, *62*, 6682–6687. [[PubMed](#)]
182. Wilson, J.; Balkwill, F. The Role of Cytokines in the Epithelial Cancer Microenvironment. *Semin. Cancer Biol.* **2002**, *12*, 113–120. [[CrossRef](#)] [[PubMed](#)]
183. Ueno, T.; Toi, M.; Saji, H.; Muta, M.; Bando, H.; Kuroi, K.; Koike, M.; Inadera, H.; Matsushima, K. Significance of Macrophage Chemoattractant Protein-1 in Macrophage Recruitment, Angiogenesis, and Survival in Human Breast Cancer. *Clin. Cancer Res.* **2000**, *6*, 3282–3289. [[PubMed](#)]
184. Kishimoto, T.; Akira, S.; Narazaki, M.; Taga, T. Interleukin-6 Family of Cytokines and Gp130. *Blood* **1995**, *86*, 1243–1254. [[CrossRef](#)] [[PubMed](#)]
185. Scott, K.A.; Moore, R.J.; Arnott, C.H.; East, N.; Thompson, R.G.; Scallan, B.J.; Shealy, D.J.; Balkwill, F.R. An Anti-Tumor Necrosis Factor-Alpha Antibody Inhibits the Development of Experimental Skin Tumors. *Mol. Cancer Ther.* **2003**, *2*, 445–451.
186. Sha, K.; Yeh, S.; Chang, C.; Nastiuk, K.L.; Krolewski, J.J. TNF Signaling Mediates an Enzalutamide-Induced Metastatic Phenotype of Prostate Cancer and Microenvironment Cell Co-Cultures. *Oncotarget* **2015**, *6*, 25726–25740. [[CrossRef](#)]
187. Madhusudan, S.; Foster, M.; Muthuramalingam, S.R.; Braybrooke, J.P.; Wilner, S.; Kaur, K.; Han, C.; Hoare, S.; Balkwill, F.; Talbot, D.C.; et al. A Phase II Study of Etanercept (Enbrel), a Tumor Necrosis Factor α Inhibitor in Patients with Metastatic Breast Cancer. *Clin. Cancer Res.* **2004**, *10*, 6528–6534. [[CrossRef](#)] [[PubMed](#)]
188. Brown, E.R.; Charles, K.A.; Hoare, S.A.; Rye, R.L.; Jodrell, D.I.; Aird, R.E.; Vora, R.; Prabhakar, U.; Nakada, M.; Corringham, R.E.; et al. A Clinical Study Assessing the Tolerability and Biological Effects of Infliximab, a TNF-Alpha Inhibitor, in Patients with Advanced Cancer. *Ann. Oncol.* **2008**, *19*, 1340–1346. [[CrossRef](#)]
189. Rutgeerts, P.; Van Assche, G.; Vermeire, S. Optimizing Anti-TNF Treatment in Inflammatory Bowel Disease. *Gastroenterology* **2004**, *126*, 1593–1610. [[CrossRef](#)]
190. Atzeni, F.; Sarzi-Puttini, P. *Autoimmunity and the Newer Biopharmaceuticals*; Chapter 92; Shoenfeld, Y., Meroni, P.L., Gershwin, M.E.B.T.-A., Third, E., Eds.; Elsevier: San Diego, CA, USA, 2014; pp. 795–802. [[CrossRef](#)]
191. Kulbe, H.; Chakravarty, P.; Leinster, D.A.; Charles, K.A.; Kwong, J.; Thompson, R.G.; Coward, J.I.; Schioppa, T.; Robinson, S.C.; Gallagher, W.M.; et al. A Dynamic Inflammatory Cytokine Network in the Human Ovarian Cancer Microenvironment. *Cancer Res.* **2012**, *72*, 66–75. [[CrossRef](#)]
192. Larkin, J.M.G.; Ferguson, T.R.; Pickering, L.M.; Edmonds, K.; James, M.G.; Thomas, K.; Banerji, U.; Berns, B.; de Boer, C.; Gore, M.E. A Phase I/II Trial of Sorafenib and Infliximab in Advanced Renal Cell Carcinoma. *Br. J. Cancer* **2010**, *103*, 1149–1153. [[CrossRef](#)] [[PubMed](#)]
193. Hwang, H.S.; Park, Y.Y.; Shin, S.J.; Go, H.; Park, J.M.; Yoon, S.Y.; Lee, J.L.; Cho, Y.M. Involvement of the TNF- α Pathway in TKI Resistance and Suggestion of TNFR1 as a Predictive Biomarker for TKI Responsiveness in Clear Cell Renal Cell Carcinoma. *J. Korean Med. Sci.* **2020**, *35*, e31. [[CrossRef](#)] [[PubMed](#)]
194. Harrison, M.L.; Obermueller, E.; Maisey, N.R.; Hoare, S.; Edmonds, K.; Li, N.F.; Chao, D.; Hall, K.; Lee, C.; Timotheadou, E.; et al. Tumor Necrosis Factor Alpha as a New Target for Renal Cell Carcinoma: Two Sequential Phase II Trials of Infliximab at Standard and High Dose. *J. Clin. Oncol.* **2007**, *25*, 4542–4549. [[CrossRef](#)]
195. Maisey, N. Antitumor Necrosis Factor (TNF- α) Antibodies in the Treatment of Renal Cell Cancer. *Cancer Invest.* **2007**, *25*, 589–593. [[CrossRef](#)]
196. Navarro-Sarabia, F.; Ariza-Ariza, R.; Hernández-Cruz, B.; Villanueva, I. Adalimumab for Treating Rheumatoid Arthritis. *J. Rheumatol.* **2006**, *33*, 1075–1081. [[CrossRef](#)]
197. Kobelt, D.; Zhang, C.; Clayton-Lucey, I.A.; Glauen, R.; Voss, C.; Siegmund, B.; Stein, U. Pro-Inflammatory TNF- α and IFN- γ Promote Tumor Growth and Metastasis via Induction of MACC1. *Front. Immunol.* **2020**, *11*, 980. [[CrossRef](#)]
198. Prattichizzo, F.; Giuliani, A.; Recchioni, R.; Bonafè, M.; Marcheselli, F.; De Carolis, S.; Campanati, A.; Giuliadori, K.; Rippo, M.R.; Brugè, F.; et al. Anti-TNF- α Treatment Modulates SASP and SASP-Related MicroRNAs in Endothelial Cells and in Circulating Angiogenic Cells. *Oncotarget* **2016**, *7*, 11945–11958. [[CrossRef](#)]
199. Steed, P.M.; Tansey, M.G.; Zalevsky, J.; Zhukovsky, E.A.; Desjarlais, J.R.; Szymkowski, D.E.; Abbott, C.; Carmichael, D.; Chan, C.; Cherry, L.; et al. Inactivation of TNF Signaling by Rationally Designed Dominant-Negative TNF Variants. *Science* **2003**, *301*, 1895–1898. [[CrossRef](#)]
200. Xu, J.; Chakrabarti, A.K.; Tan, J.L.; Ge, L.; Gambotto, A.; Vujanovic, N.L. Essential Role of the TNF-TNFR2 Cognate Interaction in Mouse Dendritic Cell-Natural Killer Cell Crosstalk. *Blood* **2007**, *109*, 3333–3341. [[CrossRef](#)]
201. Vujanovic, L.; Szymkowski, D.E.; Alber, S.; Watkins, S.C.; Vujanovic, N.L.; Butterfield, L.H. Virally Infected and Matured Human Dendritic Cells Activate Natural Killer Cells via Cooperative Activity of Plasma Membrane-Bound TNF and IL-15. *Blood* **2010**, *116*, 575–583. [[CrossRef](#)]
202. Vujanovic, N.L. Role of TNF Superfamily Ligands in Innate Immunity. *Immunol. Res.* **2011**, *50*, 159–174. [[CrossRef](#)] [[PubMed](#)]
203. Andrea, S.V.; Lazar, V.; Albert, B.D.L.; Fernando, C.B.; Robert, L.F.; Yan, L.; Nikola, L.V. Inhibition of Soluble Tumor Necrosis Factor Prevents Chemically Induced Carcinogenesis in Mice. *Cancer Immunol. Res.* **2016**, *4*, 441–451. [[CrossRef](#)]
204. Slamon, D.J.; Godolphin, W.; Jones, L.A.; Holt, J.A.; Wong, S.G.; Keith, D.E.; Levin, W.J.; Stuart, S.G.; Udove, J.; Ullrich, A. Studies of the HER-2/Neu Proto-Oncogene in Human Breast and Ovarian Cancer. *Science* **1989**, *244*, 707–712. [[CrossRef](#)] [[PubMed](#)]

205. Braun, S.; Schlimok, G.; Heumos, I.; Schaller, G.; Riethdorf, L.; Riethmüller, G.; Pantel, K. ErbB2 Overexpression on Occult Metastatic Cells in Bone Marrow Predicts Poor Clinical Outcome of Stage I-III Breast Cancer Patients. *Cancer Res.* **2001**, *61*, 1890–1895. [[PubMed](#)]
206. Potti, A.; Willardson, J.; Forseen, C.; Kishor Ganti, A.; Koch, M.; Hebert, B.; Levitt, R.; Mehdi, S.A. Predictive Role of HER-2/Neu Overexpression and Clinical Features at Initial Presentation in Patients with Extensive Stage Small Cell Lung Carcinoma. *Lung Cancer* **2002**, *36*, 257–261. [[CrossRef](#)]
207. Canoz, O.; Ozkan, M.; Arsav, V.; Er, O.; Coskun, H.S.; Soyuer, S.; Altinbas, M. The Role of C-ErbB-2 Expression on the Survival of Patients with Small-Cell Lung Cancer. *Lung* **2006**, *184*, 267–272. [[CrossRef](#)] [[PubMed](#)]
208. Minami, T.; Kijima, T.; Kohmo, S.; Arase, H.; Otani, Y.; Nagatomo, I.; Takahashi, R.; Miyake, K.; Higashiguchi, M.; Morimura, O.; et al. Overcoming Chemoresistance of Small-Cell Lung Cancer through Stepwise HER2-Targeted Antibody-Dependent Cell-Mediated Cytotoxicity and VEGF-Targeted Antiangiogenesis. *Sci. Rep.* **2013**, *3*, 2669. [[CrossRef](#)] [[PubMed](#)]
209. Krop, I.E.; Kim, S.-B.; González-Martín, A.; LoRusso, P.M.; Ferrero, J.-M.; Smitt, M.; Yu, R.; Leung, A.C.F.; Wildiers, H. Trastuzumab Emtansine versus Treatment of Physician’s Choice for Pretreated HER2-Positive Advanced Breast Cancer (TH3RESA): A Randomised, Open-Label, Phase 3 Trial. *Lancet Oncol.* **2014**, *15*, 689–699. [[CrossRef](#)]
210. Singh, R.; Lillard, J.W.J.; Singh, S. Chemokines: Key Players in Cancer Progression and Metastasis. *Front. Biosci.* **2011**, *3*, 1569–1582. [[CrossRef](#)]
211. Balkwill, F.R. The Chemokine System and Cancer. *J. Pathol.* **2012**, *226*, 148–157. [[CrossRef](#)]
212. Hobbs, S.S.; Goettel, J.A.; Liang, D.; Yan, F.; Edelblum, K.L.; Frey, M.R.; Mullane, M.T.; Polk, D.B. TNF Transactivation of EGFR Stimulates Cytoprotective COX-2 Expression in Gastrointestinal Epithelial Cells. *Am. J. Physiol. Gastrointest. Liver Physiol.* **2011**, *301*, G220–G229. [[CrossRef](#)] [[PubMed](#)]
213. Argast, G.M.; Campbell, J.S.; Brooling, J.T.; Fausto, N. Epidermal Growth Factor Receptor Transactivation Mediates Tumor Necrosis Factor-Induced Hepatocyte Replication. *J. Biol. Chem.* **2004**, *279*, 34530–34536. [[CrossRef](#)] [[PubMed](#)]
214. Ueno, Y.; Sakurai, H.; Matsuo, M.; Choo, M.K.; Koizumi, K.; Saiki, I. Selective Inhibition of TNF-Alpha-Induced Activation of Mitogen-Activated Protein Kinases and Metastatic Activities by Gefitinib. *Br. J. Cancer* **2005**, *92*, 1690–1695. [[CrossRef](#)] [[PubMed](#)]
215. Tsatsanis, C.; Androulidaki, A.; Venihaki, M.; Margioris, A.N. Signalling Networks Regulating Cyclooxygenase-2. *Int. J. Biochem. Cell Biol.* **2006**, *38*, 1654–1661. [[CrossRef](#)]
216. Chun, K.-S.; Surh, Y.-J. Signal Transduction Pathways Regulating Cyclooxygenase-2 Expression: Potential Molecular Targets for Chemoprevention. *Biochem. Pharmacol.* **2004**, *68*, 1089–1100. [[CrossRef](#)]
217. Giovannucci, E.; Rimm, E.B.; Stampfer, M.J.; Colditz, G.A.; Ascherio, A.; Willett, W.C. Aspirin Use and the Risk for Colorectal Cancer and Adenoma in Male Health Professionals. *Ann. Intern. Med.* **1994**, *121*, 241–246. [[CrossRef](#)]
218. Greenberg, E.R.; Baron, J.A.; Freeman, D.H.J.; Mandel, J.S.; Haile, R. Reduced Risk of Large-Bowel Adenomas among Aspirin Users. The Polyp Prevention Study Group. *J. Natl. Cancer Inst.* **1993**, *85*, 912–916. [[CrossRef](#)]
219. Son, D.-S.; Kabir, S.M.; Dong, Y.; Lee, E.; Adunyah, S.E. Characteristics of Chemokine Signatures Elicited by EGF and TNF in Ovarian Cancer Cells. *J. Inflamm.* **2013**, *10*, 25. [[CrossRef](#)]
220. Freedman, M.H.; Cohen, A.; Grunberger, T.; Bunin, N.; Luddy, R.E.; Saunders, E.F.; Shahidi, N.; Lau, A.; Estrov, Z. Central Role of Tumour Necrosis Factor, GM-CSF, and Interleukin 1 in the Pathogenesis of Juvenile Chronic Myelogenous Leukaemia. *Br. J. Haematol.* **1992**, *80*, 40–48. [[CrossRef](#)]
221. Shan, D.; Ledbetter, J.A.; Press, O.W. Signaling Events Involved in Anti-CD20-Induced Apoptosis of Malignant Human B Cells. *Cancer Immunol. Immunother.* **2000**, *48*, 673–683. [[CrossRef](#)]
222. Adami, F.; Guarini, A.; Pini, M.; Siviero, F.; Sancetta, R.; Massaia, M.; Trentin, L.; Foà, R.; Semenzato, G. Serum Levels of Tumour Necrosis Factor-Alpha in Patients with B-Cell Chronic Lymphocytic Leukaemia. *Eur. J. Cancer* **1994**, *30A*, 1259–1263. [[CrossRef](#)]
223. Saulep-Easton, D.; Vincent, F.B.; Le Page, M.; Wei, A.; Ting, S.B.; Croce, C.M.; Tam, C.; Mackay, F. Cytokine-Driven Loss of Plasmacytoid Dendritic Cell Function in Chronic Lymphocytic Leukemia. *Leukemia* **2014**, *28*, 2005–2015. [[CrossRef](#)] [[PubMed](#)]
224. Francisco, L.M.; Sage, P.T.; Sharpe, A.H. The PD-1 Pathway in Tolerance and Autoimmunity. *Immunol. Rev.* **2010**, *236*, 219–242. [[CrossRef](#)] [[PubMed](#)]
225. Guleria, I.; Khosroshahi, A.; Ansari, M.J.; Habicht, A.; Azuma, M.; Yagita, H.; Noelle, R.J.; Coyle, A.; Mellor, A.L.; Khoury, S.J.; et al. A Critical Role for the Programmed Death Ligand 1 in Fetomaternal Tolerance. *J. Exp. Med.* **2005**, *202*, 231–237. [[CrossRef](#)] [[PubMed](#)]
226. Butte, M.J.; Keir, M.E.; Phamduy, T.B.; Sharpe, A.H.; Freeman, G.J. Programmed Death-1 Ligand 1 Interacts Specifically with the B7-1 Costimulatory Molecule to Inhibit T Cell Responses. *Immunity* **2007**, *27*, 111–122. [[CrossRef](#)]
227. Curiel, T.J.; Wei, S.; Dong, H.; Alvarez, X.; Cheng, P.; Mottram, P.; Krzysiek, R.; Knutson, K.L.; Daniel, B.; Zimmermann, M.C.; et al. Blockade of B7-H1 Improves Myeloid Dendritic Cell-Mediated Antitumor Immunity. *Nat. Med.* **2003**, *9*, 562–567. [[CrossRef](#)]
228. Latchman, Y.E.; Liang, S.C.; Wu, Y.; Chernova, T.; Sobel, R.A.; Klemm, M.; Kuchroo, V.K.; Freeman, G.J.; Sharpe, A.H. PD-L1-Deficient Mice Show That PD-L1 on T Cells, Antigen-Presenting Cells, and Host Tissues Negatively Regulates T Cells. *Proc. Natl. Acad. Sci. USA* **2004**, *101*, 10691–10696. [[CrossRef](#)]
229. Freeman, G.J.; Long, A.J.; Iwai, Y.; Bourque, K.; Chernova, T.; Nishimura, H.; Fitz, L.J.; Malenkovich, N.; Okazaki, T.; Byrne, M.C.; et al. Engagement of the PD-1 Immunoinhibitory Receptor by a Novel B7 Family Member Leads to Negative Regulation of Lymphocyte Activation. *J. Exp. Med.* **2000**, *192*, 1027–1034. [[CrossRef](#)]
230. Leach, D.R.; Krummel, M.F.; Allison, J.P. Enhancement of Antitumor Immunity by CTLA-4 Blockade. *Science* **1996**, *271*, 1734–1736. [[CrossRef](#)]

231. Hodi, F.S.; O'Day, S.J.; McDermott, D.F.; Weber, R.W.; Sosman, J.A.; Haanen, J.B.; Gonzalez, R.; Robert, C.; Schadendorf, D.; Hassel, J.C.; et al. Improved Survival with Ipilimumab in Patients with Metastatic Melanoma. *N. Engl. J. Med.* **2010**, *363*, 711–723. [[CrossRef](#)]
232. Wang, X.; Yang, L.; Huang, F.; Zhang, Q.; Liu, S.; Ma, L.; You, Z. Inflammatory Cytokines IL-17 and TNF- α up-Regulate PD-L1 Expression in Human Prostate and Colon Cancer Cells. *Immunol. Lett.* **2017**, *184*, 7–14. [[CrossRef](#)] [[PubMed](#)]
233. Qu, Q.-X.; Xie, F.; Huang, Q.; Zhang, X.-G. Membranous and Cytoplasmic Expression of PD-L1 in Ovarian Cancer Cells. *Cell. Physiol. Biochem. Int. J. Exp. Cell. Physiol. Biochem. Pharm.* **2017**, *43*, 1893–1906. [[CrossRef](#)] [[PubMed](#)]
234. Hartley, G.; Regan, D.; Guth, A.; Dow, S. Regulation of PD-L1 Expression on Murine Tumor-Associated Monocytes and Macrophages by Locally Produced TNF- α . *Cancer Immunol. Immunother.* **2017**, *66*, 523–535. [[CrossRef](#)] [[PubMed](#)]
235. Li, Z.; Zhang, C.; Du, J.-X.; Zhao, J.; Shi, M.-T.; Jin, M.-W.; Liu, H. Adipocytes Promote Tumor Progression and Induce PD-L1 Expression via TNF- α /IL-6 Signaling. *Cancer Cell Int.* **2020**, *20*, 179. [[CrossRef](#)]
236. Lv, Y.; Zhao, Y.; Wang, X.; Chen, N.; Mao, F.; Teng, Y.; Wang, T.; Peng, L.; Zhang, J.; Cheng, P.; et al. Increased Intratumoral Mast Cells Foster Immune Suppression and Gastric Cancer Progression through TNF- α -PD-L1 Pathway. *J. Immunother. Cancer* **2019**, *7*, 54. [[CrossRef](#)]
237. Ju, X.; Zhang, H.; Zhou, Z.; Chen, M.; Wang, Q. Tumor-Associated Macrophages Induce PD-L1 Expression in Gastric Cancer Cells through IL-6 and TNF- α Signaling. *Exp. Cell Res.* **2020**, *396*, 112315. [[CrossRef](#)]
238. Lim, S.-O.; Li, C.-W.; Xia, W.; Cha, J.-H.; Chan, L.-C.; Wu, Y.; Chang, S.-S.; Lin, W.-C.; Hsu, J.-M.; Hsu, Y.-H.; et al. Deubiquitination and Stabilization of PD-L1 by CSN5. *Cancer Cell* **2016**, *30*, 925–939. [[CrossRef](#)]
239. Rotte, A.; D'Orazi, G.; Bhandaru, M. Nobel Committee Honors Tumor Immunologists. *J. Exp. Clin. Cancer Res.* **2018**, *37*, 262. [[CrossRef](#)]
240. Zingg, D.; Arenas-Ramirez, N.; Sahin, D.; Rosalia, R.A.; Antunes, A.T.; Haeusel, J.; Sommer, L.; Boyman, O. The Histone Methyltransferase Ezh2 Controls Mechanisms of Adaptive Resistance to Tumor Immunotherapy. *Cell Rep.* **2017**, *20*, 854–867. [[CrossRef](#)]
241. Jeong, H.; Kim, S.; Hong, B.-J.; Lee, C.-J.; Kim, Y.-E.; Bok, S.; Oh, J.-M.; Gwak, S.-H.; Yoo, M.Y.; Lee, M.S.; et al. Tumor-Associated Macrophages Enhance Tumor Hypoxia and Aerobic Glycolysis. *Cancer Res.* **2019**, *79*, 795–806. [[CrossRef](#)]
242. Boutsikou, E.; Domvri, K.; Hardavella, G.; Tsiouda, D.; Zarogoulidis, K.; Kontakiotis, T. Tumour Necrosis Factor, Interferon-Gamma and Interleukins as Predictive Markers of Antiprogrammed Cell-Death Protein-1 Treatment in Advanced Non-Small Cell Lung Cancer: A Pragmatic Approach in Clinical Practice. *Ther. Adv. Med. Oncol.* **2018**, *10*. [[CrossRef](#)] [[PubMed](#)]
243. Blidner, A.G.; Choi, J.; Cooksley, T.; Dougan, M.; Glezerman, I.; Ginex, P.; Girotra, M.; Gupta, D.; Johnson, D.; Shannon, V.R.; et al. Cancer Immunotherapy-Related Adverse Events: Causes and Challenges. *Support. Care Cancer Off. J. Multinat. Assoc. Support. Care Cancer* **2020**, *28*, 6111–6117. [[CrossRef](#)] [[PubMed](#)]
244. Flaherty, K.T.; Infante, J.R.; Daud, A.; Gonzalez, R.; Kefford, R.F.; Sosman, J.; Hamid, O.; Schuchter, L.; Cebon, J.; Ibrahim, N.; et al. Combined BRAF and MEK Inhibition in Melanoma with BRAF V600 Mutations. *N. Engl. J. Med.* **2012**, *367*, 1694–1703. [[CrossRef](#)] [[PubMed](#)]
245. Wang, D.Y.; Salem, J.-E.; Cohen, J.V.; Chandra, S.; Menzer, C.; Ye, F.; Zhao, S.; Das, S.; Beckermann, K.E.; Ha, L.; et al. Fatal Toxic Effects Associated With Immune Checkpoint Inhibitors: A Systematic Review and Meta-Analysis. *JAMA Oncol.* **2018**, *4*, 1721–1728. [[CrossRef](#)]
246. Boutros, C.; Tarhini, A.; Routier, E.; Lambotte, O.; Ladurie, F.L.; Carbonnel, F.; Izzeddine, H.; Marabelle, A.; Champiat, S.; Berdelou, A.; et al. Safety Profiles of Anti-CTLA-4 and Anti-PD-1 Antibodies Alone and in Combination. *Nat. Rev. Clin. Oncol.* **2016**, *13*, 473–486. [[CrossRef](#)]
247. Larkin, J.; Chiarion-Sileni, V.; Gonzalez, R.; Grob, J.J.; Cowey, C.L.; Lao, C.D.; Schadendorf, D.; Dummer, R.; Smylie, M.; Rutkowski, P.; et al. Combined Nivolumab and Ipilimumab or Monotherapy in Untreated Melanoma. *N. Engl. J. Med.* **2015**, *373*, 23–34. [[CrossRef](#)]
248. Naidoo, J.; Page, D.B.; Li, B.T.; Connell, L.C.; Schindler, K.; Lacouture, M.E.; Postow, M.A.; Wolchok, J.D. Toxicities of the Anti-PD-1 and Anti-PD-L1 Immune Checkpoint Antibodies. *Ann. Oncol.* **2015**, *26*, 2375–2391. [[CrossRef](#)]
249. Ribas, A.; Wolchok, J.D. Cancer Immunotherapy Using Checkpoint Blockade. *Science* **2018**, *359*, 1350–1355. [[CrossRef](#)]
250. Jacobberger-Foissac, C.; Blake, S.J.; Liu, J.; McDonald, E.; Triscott, H.; Nakamura, K.; Smyth, M.J.; Teng, M.W. Concomitant or Delayed Anti-TNF Differentially Impact on Immune-Related Adverse Events and Antitumor Efficacy after Anti-CD40 Therapy. *J. Immunother. Cancer* **2020**, *8*. [[CrossRef](#)]
251. Verheijden, R.J.; May, A.M.; Blank, C.U.; Aarts, M.J.B.; van den Berkortel, F.W.P.J.; van den Eertwegh, A.J.M.; de Groot, J.W.B.; Boers-Sonderen, M.J.; van der Hoeven, J.J.M.; Hospers, G.A.; et al. Association of Anti-TNF with Decreased Survival in Steroid Refractory Ipilimumab and Anti-PD1-Treated Patients in the Dutch Melanoma Treatment Registry. *Clin. Cancer Res.* **2020**, *26*, 2268–2274. [[CrossRef](#)]
252. Montfort, A.; Dufau, C.; Colacios, C.; Andrieu-Abadie, N.; Levade, T.; Filleron, T.; Delord, J.-P.; Ayyoub, M.; Meyer, N.; Ségui, B. Anti-TNF, a Magic Bullet in Cancer Immunotherapy? *J. Immunother. Cancer* **2019**, *7*, 303. [[CrossRef](#)] [[PubMed](#)]
253. Upadhaya, S.; Hubbard-Lucey, V.M.; Yu, J.X. Immuno-Oncology Drug Development Forges on despite COVID-19. *Nat. Rev. Drug Discov.* **2020**, 751–752. [[CrossRef](#)] [[PubMed](#)]
254. Phelan, K.W.; Advani, A.S. Novel Therapies in Acute Lymphoblastic Leukemia. *Curr. Hematol. Malig. Rep.* **2018**, *13*, 289–299. [[CrossRef](#)] [[PubMed](#)]
255. Dai, H.; Wang, Y.; Lu, X.; Han, W. Chimeric Antigen Receptors Modified T-Cells for Cancer Therapy. *J. Natl. Cancer Inst.* **2016**, *108*. [[CrossRef](#)] [[PubMed](#)]
256. Frey, N. The What, When and How of CAR T Cell Therapy for ALL. *Best Pr. Res. Clin. Haematol.* **2017**, *30*, 275–281. [[CrossRef](#)]
257. Grupp, S.A.; Kalos, M.; Barrett, D.; Aplenc, R.; Porter, D.L.; Rheingold, S.R.; Teachey, D.T.; Chew, A.; Hauck, B.; Wright, J.F.; et al. Chimeric Antigen Receptor-Modified T Cells for Acute Lymphoid Leukemia. *N. Engl. J. Med.* **2013**, *368*, 1509–1518. [[CrossRef](#)]

258. Raponi, S.; De Propriis, M.S.; Intoppa, S.; Milani, M.L.; Vitale, A.; Elia, L.; Perbellini, O.; Pizzolo, G.; Foà, R.; Guarini, A. Flow Cytometric Study of Potential Target Antigens (CD19, CD20, CD22, CD33) for Antibody-Based Immunotherapy in Acute Lymphoblastic Leukemia: Analysis of 552 Cases. *Leuk. Lymphoma* **2011**, *52*, 1098–1107. [[CrossRef](#)]
259. Scheuermann, R.H.; Racila, E. CD19 Antigen in Leukemia and Lymphoma Diagnosis and Immunotherapy. *Leuk. Lymphoma* **1995**, *18*, 385–397. [[CrossRef](#)]
260. Maude, S.L.; Barrett, D.; Teachey, D.T.; Grupp, S.A. Managing Cytokine Release Syndrome Associated with Novel T Cell-Engaging Therapies. *Cancer J.* **2014**, *20*, 119–122. [[CrossRef](#)]
261. Maus, M.V.; Haas, A.R.; Beatty, G.L.; Albelda, S.M.; Levine, B.L.; Liu, X.; Zhao, Y.; Kalos, M.; June, C.H. T Cells Expressing Chimeric Antigen Receptors Can Cause Anaphylaxis in Humans. *Cancer Immunol. Res.* **2013**, *1*, 26–31. [[CrossRef](#)]
262. Brentjens, R.J.; Davila, M.L.; Riviere, I.; Park, J.; Wang, X.; Cowell, L.G.; Bartido, S.; Stefanski, J.; Taylor, C.; Olszewska, M.; et al. CD19-Targeted T Cells Rapidly Induce Molecular Remissions in Adults with Chemotherapy-Refractory Acute Lymphoblastic Leukemia. *Sci. Transl. Med.* **2013**, *5*, 177ra38. [[CrossRef](#)] [[PubMed](#)]
263. Elia, A.R.; Grioni, M.; Basso, V.; Curnis, F.; Freschi, M.; Corti, A.; Mondino, A.; Bellone, M. Targeting Tumor Vasculature with TNF Leads Effector T Cells to the Tumor and Enhances Therapeutic Efficacy of Immune Checkpoint Blockers in Combination with Adoptive Cell Therapy. *Clin. Cancer Res.* **2018**, *24*, 2171–2181. [[CrossRef](#)] [[PubMed](#)]
264. Piali, L.; Fichtel, A.; Terpe, H.J.; Imhof, B.A.; Gisler, R.H. Endothelial Vascular Cell Adhesion Molecule 1 Expression Is Suppressed by Melanoma and Carcinoma. *J. Exp. Med.* **1995**, *181*, 811–816. [[CrossRef](#)] [[PubMed](#)]
265. Bellone, M.; Calcinotto, A. Ways to Enhance Lymphocyte Trafficking into Tumors and Fitness of Tumor Infiltrating Lymphocytes. *Front. Oncol.* **2013**, *3*, 231. [[CrossRef](#)] [[PubMed](#)]
266. Sharma, P.; Hu-Lieskovan, S.; Wargo, J.A.; Ribas, A. Primary, Adaptive, and Acquired Resistance to Cancer Immunotherapy. *Cell* **2017**, *168*, 707–723. [[CrossRef](#)]
267. Bellone, M.; Elia, A.R. Constitutive and Acquired Mechanisms of Resistance to Immune Checkpoint Blockade in Human Cancer. *Cytokine Growth Factor Rev.* **2017**, *36*, 17–24. [[CrossRef](#)]
268. Calcinotto, A.; Grioni, M.; Jachetti, E.; Curnis, F.; Mondino, A.; Parmiani, G.; Corti, A.; Bellone, M. Targeting TNF- α to Neoangiogenic Vessels Enhances Lymphocyte Infiltration in Tumors and Increases the Therapeutic Potential of Immunotherapy. *J. Immunol.* **2012**, *188*, 2687–2694. [[CrossRef](#)]
269. Manzo, T.; Sturmheit, T.; Basso, V.; Petrozziello, E.; Hess Michelini, R.; Riba, M.; Freschi, M.; Elia, A.R.; Grioni, M.; Curnis, F.; et al. T Cells Redirected to a Minor Histocompatibility Antigen Instruct Intratumoral TNF α Expression and Empower Adoptive Cell Therapy for Solid Tumors. *Cancer Res.* **2017**, *77*, 658–671. [[CrossRef](#)]
270. Plautz, G.E.; Touhalisky, J.E.; Shu, S. Treatment of Murine Gliomas by Adoptive Transfer of Ex Vivo Activated Tumor-Draining Lymph Node Cells. *Cell. Immunol.* **1997**, *178*, 101–107. [[CrossRef](#)]
271. Peng, L.; Shu, S.; Krauss, J.C. Treatment of Subcutaneous Tumor with Adoptively Transferred T Cells. *Cell. Immunol.* **1997**, *178*, 24–32. [[CrossRef](#)]
272. Tanaka, H.; Yoshizawa, H.; Yamaguchi, Y.; Ito, K.; Kagamu, H.; Suzuki, E.; Gejyo, F.; Hamada, H.; Arakawa, M. Successful Adoptive Immunotherapy of Murine Poorly Immunogenic Tumor with Specific Effector Cells Generated from Gene-Modified Tumor-Primed Lymph Node Cells. *J. Immunol.* **1999**, *162*, 3574–3582. [[PubMed](#)]
273. Ye, Z.; Shi, M.; Chan, T.; Sas, S.; Xu, S.; Xiang, J. Engineered CD8+ Cytotoxic T Cells with Fiber-Modified Adenovirus-Mediated TNF-Alpha Gene Transfection Counteract Immunosuppressive Interleukin-10-Secreting Lung Metastasis and Solid Tumors. *Cancer Gene* **2007**, *14*, 661–675. [[CrossRef](#)] [[PubMed](#)]
274. Pedersen, A.E.; Thorn, M.; Gad, M.; Walter, M.R.; Johnsen, H.E.; Gaarsdal, E.; Nikolajsen, K.; Buus, S.; Claesson, M.H.; Svane, I.M. Phenotypic and Functional Characterization of Clinical Grade Dendritic Cells Generated from Patients with Advanced Breast Cancer for Therapeutic Vaccination. *Scand. J. Immunol.* **2005**, 147–156. [[CrossRef](#)] [[PubMed](#)]
275. Brunner, C.; Seiderer, J.; Schlamp, A.; Bidlingmaier, M.; Eigler, A.; Haimerl, W.; Lehr, H.A.; Krieg, A.M.; Hartmann, G.; Endres, S. Enhanced Dendritic Cell Maturation by TNF-Alpha or Cytidine-Phosphate-Guanosine DNA Drives T Cell Activation in Vitro and Therapeutic Anti-Tumor Immune Responses in Vivo. *J. Immunol.* **2000**, *165*, 6278–6286. [[CrossRef](#)] [[PubMed](#)]
276. Cumberbatch, M.; Kimber, I. Tumour Necrosis Factor-Alpha Is Required for Accumulation of Dendritic Cells in Draining Lymph Nodes and for Optimal Contact Sensitization. *Immunology* **1995**, *84*, 31–35.
277. Hogquist, K.A.; Jameson, S.C.; Heath, W.R.; Howard, J.L.; Bevan, M.J.; Carbone, F.R. T Cell Receptor Antagonist Peptides Induce Positive Selection. *Cell* **1994**, *76*, 17–27. [[CrossRef](#)]
278. Liu, Y.; Saxena, A.; Zheng, C.; Carlsen, S.; Xiang, J. Combined Alpha Tumor Necrosis Factor Gene Therapy and Engineered Dendritic Cell Vaccine in Combating Well-Established Tumors. *J. Gene Med.* **2004**, *6*, 857–868. [[CrossRef](#)]
279. Klebanoff, C.A.; Khong, H.T.; Antony, P.A.; Palmer, D.C.; Restifo, N.P. Sinks, Suppressors and Antigen Presenters: How Lymphodepletion Enhances T Cell-Mediated Tumor Immunotherapy. *Trends Immunol.* **2005**, *26*, 111–117. [[CrossRef](#)]
280. Dudley, M.E.; Wunderlich, J.R.; Yang, J.C.; Sherry, R.M.; Topalian, S.L.; Restifo, N.P.; Royal, R.E.; Kammula, U.; White, D.E.; Mavroukakis, S.A.; et al. Adoptive Cell Transfer Therapy Following Non-Myeloablative but Lymphodepleting Chemotherapy for the Treatment of Patients with Refractory Metastatic Melanoma. *J. Clin. Oncol.* **2005**, *23*, 2346–2357. [[CrossRef](#)]
281. Dudley, M.E.; Gross, C.A.; Langan, M.M.; Garcia, M.R.; Sherry, R.M.; Yang, J.C.; Phan, G.Q.; Kammula, U.S.; Hughes, M.S.; Citrin, D.E.; et al. CD8+ Enriched “Young” Tumor Infiltrating Lymphocytes Can Mediate Regression of Metastatic Melanoma. *Clin. Cancer Res.* **2010**, *16*, 6122–6131. [[CrossRef](#)]

282. Santos, J.M.; Cervera-carrascon, V.; Havunen, R.; Zafar, S.; Siurala, M.; Sorsa, S.; Anttila, M.; Kanerva, A.; Hemminki, A. Adenovirus Coding for Interleukin-2 and Tumor Necrosis Factor Alpha Replaces Lymphodepleting Chemotherapy in Adoptive T Cell Therapy. *Mol. Ther.* **2018**, *26*, 2243–2254. [[CrossRef](#)] [[PubMed](#)]
283. Siurala, M.; Havunen, R.; Saha, D.; Lumen, D.; Airaksinen, A.J.; Tähtinen, S.; Cervera-Carrascon, V.; Bramante, S.; Parviainen, S.; Vähä-Koskela, M.; et al. Adenoviral Delivery of Tumor Necrosis Factor- α and Interleukin-2 Enables Successful Adoptive Cell Therapy of Immunosuppressive Melanoma. *Mol. Ther.* **2016**, *24*, 1435–1443. [[CrossRef](#)] [[PubMed](#)]
284. Restifo, N.P.; Dudley, M.E.; Rosenberg, S.A. Adoptive Immunotherapy for Cancer: Harnessing the T Cell Response. *Nat. Rev. Immunol.* **2012**, *12*, 269–281. [[CrossRef](#)] [[PubMed](#)]
285. Rosenberg, S.A.; Yang, J.C.; Sherry, R.M.; Kammula, U.S.; Hughes, M.S.; Phan, G.Q.; Citrin, D.E.; Restifo, N.P.; Robbins, P.F.; Wunderlich, J.R.; et al. Durable Complete Responses in Heavily Pretreated Patients with Metastatic Melanoma Using T-Cell Transfer Immunotherapy. *Clin. Cancer Res.* **2011**, *17*, 4550–4557. [[CrossRef](#)]
286. Landsberg, J.; Kohlmeyer, J.; Renn, M.; Bald, T.; Rogava, M.; Cron, M.; Fatho, M.; Lennerz, V.; Wölfel, T.; Hölzel, M.; et al. Melanomas Resist T-Cell Therapy through Inflammation-Induced Reversible Dedifferentiation. *Nature* **2012**, *490*, 412–416. [[CrossRef](#)] [[PubMed](#)]
287. Mehta, A.; Kim, Y.J.; Robert, L.; Tsoi, J.; Comin-Anduix, B.; Berent-Maoz, B.; Cochran, A.J.; Economou, J.S.; Tumei, P.C.; Puig-Saus, C.; et al. Immunotherapy Resistance by Inflammation-Induced Dedifferentiation. *Cancer Discov.* **2018**, *8*, 935–943. [[CrossRef](#)]
288. Hugo, W.; Zaretsky, J.M.; Sun, L.; Song, C.; Moreno, B.H.; Hu-Lieskovan, S.; Berent-Maoz, B.; Pang, J.; Chmielowski, B.; Cherry, G.; et al. Genomic and Transcriptomic Features of Response to Anti-PD-1 Therapy in Metastatic Melanoma. *Cell* **2017**, *168*, 542. [[CrossRef](#)]
289. Willrich, M.A.V.; Murray, D.L.; Snyder, M.R. Tumor Necrosis Factor Inhibitors: Clinical Utility in Autoimmune Diseases. *Transl. Res.* **2015**, *165*, 270–282. [[CrossRef](#)]
290. Roda, G.; Jharap, B.; Neeraj, N.; Colombel, J.-F. Loss of Response to Anti-TNFs: Definition, Epidemiology, and Management. *Clin. Transl. Gastroenterol.* **2016**, *7*, e135. [[CrossRef](#)]
291. Rubbert-Roth, A.; Atzeni, F.; Masala, I.F.; Caporali, R.; Montecucco, C.; Sarzi-Puttini, P. TNF Inhibitors in Rheumatoid Arthritis and Spondyloarthritis: Are They the Same? *Autoimmun. Rev.* **2018**, *17*, 24–28. [[CrossRef](#)]
292. Liu, J.; Dong, Z.; Zhu, Q.; He, D.; Ma, Y.; Du, A.; He, F.; Zhao, D.; Xu, X.; Zhang, H.; et al. TNF- α Promoter Polymorphisms Predict the Response to Etanercept More Powerfully than That to Infliximab/Adalimumab in Spondyloarthritis. *Sci. Rep.* **2016**, *6*, 1–9. [[CrossRef](#)] [[PubMed](#)]
293. Murdaca, G.; Gulli, R.; Spanò, F.; Lantieri, F.; Burlando, M.; Parodi, A.; Mandich, P.; Puppo, F. TNF- α Gene Polymorphisms: Association with Disease Susceptibility and Response to Anti-TNF- α Treatment in Psoriatic Arthritis. *J. Invest. Derm.* **2014**, *134*, 2503–2509. [[CrossRef](#)] [[PubMed](#)]
294. Hochberg, M.C.; Lebowitz, M.G.; Plevy, S.E.; Hobbs, K.F.; Yocum, D.E. The Benefit/Risk Profile of TNF-Blocking Agents: Findings of a Consensus Panel. *Semin. Arthritis Rheum.* **2005**, *34*, 819–836. [[CrossRef](#)] [[PubMed](#)]
295. Taylor, P.C. Antibody Therapy for Rheumatoid Arthritis. *Curr. Opin. Pharm.* **2003**, *3*, 323–328. [[CrossRef](#)]
296. Chang, J.; Girgis, L. Clinical Use of Anti-TNF-Alpha Biological Agents—A Guide for GPs. *Aust. Fam. Physician* **2007**, *36*, 1035–1038.
297. Roach, D.R.; Bean, A.G.D.; Demangel, C.; France, M.P.; Briscoe, H.; Britton, W.J. TNF Regulates Chemokine Induction Essential for Cell Recruitment, Granuloma Formation, and Clearance of Mycobacterial Infection. *J. Immunol.* **2002**, *168*, 4620–4627. [[CrossRef](#)]
298. Antoni, C.; Braun, J. Side Effects of Anti-TNF Therapy: Current Knowledge. *Clin. Exp. Rheumatol.* **2002**, *20* (Suppl. S28), S152–S157.
299. Bean, A.G.; Roach, D.R.; Briscoe, H.; France, M.P.; Korner, H.; Sedgwick, J.D.; Britton, W.J. Structural Deficiencies in Granuloma Formation in TNF Gene-Targeted Mice Underlie the Heightened Susceptibility to Aerosol Mycobacterium Tuberculosis Infection, Which Is Not Compensated for by Lymphotoxin. *J. Immunol.* **1999**, *162*, 3504–3511.
300. Her, M.; Kavanaugh, A. Alterations in Immune Function with Biologic Therapies for Autoimmune Disease. *J. Allergy Clin. Immunol.* **2016**, *137*, 19–27. [[CrossRef](#)]
301. Leombruno, J.P.; Einarson, T.R.; Keystone, E.C. The Safety of Anti-Tumour Necrosis Factor Treatments in Rheumatoid Arthritis: Meta and Exposure-Adjusted Pooled Analyses of Serious Adverse Events. *Ann. Rheum. Dis.* **2009**, *68*, 1136–1145. [[CrossRef](#)]
302. Thompson, A.E.; Rieder, S.W.; Pope, J.E. Tumor Necrosis Factor Therapy and the Risk of Serious Infection and Malignancy in Patients with Early Rheumatoid Arthritis: A Meta-Analysis of Randomized Controlled Trials. *Arthritis Rheum.* **2011**, *63*, 1479–1485. [[CrossRef](#)] [[PubMed](#)]
303. Grijalva, C.G.; Chen, L.; Delzell, E.; Baddley, J.W.; Beukelman, T.; Winthrop, K.L.; Griffin, M.R.; Herrinton, L.J.; Liu, L.; Ouellet-Hellstrom, R.; et al. Initiation of Tumor Necrosis Factor- α Antagonists and the Risk of Hospitalization for Infection in Patients with Autoimmune Diseases. *JAMA* **2011**, *306*, 2331–2339. [[CrossRef](#)] [[PubMed](#)]
304. Murdaca, G.; Negrini, S.; Pellicchio, M.; Greco, M.; Schiavi, C.; Giusti, F.; Puppo, F. Update upon the Infection Risk in Patients Receiving TNF Alpha Inhibitors. *Expert Opin. Drug Saf.* **2019**, *18*, 219–229. [[CrossRef](#)] [[PubMed](#)]
305. Zhang, Z.; Fan, W.; Yang, G.; Xu, Z.; Wang, J.; Cheng, Q.; Yu, M. Risk of Tuberculosis in Patients Treated with TNF- α Antagonists: A Systematic Review and Meta-Analysis of Randomised Controlled Trials. *BMJ Open* **2017**, *7*, 1–8. [[CrossRef](#)]
306. Ramiro, S.; Sepriano, A.; Chatzidionysiou, K.; Nam, J.L.; Smolen, J.S.; van der Heijde, D.; Dougados, M.; van Vollenhoven, R.; Bijlsma, J.W.; Burmester, G.R.; et al. Safety of Synthetic and Biological DMARDs: A Systematic Literature Review Informing the 2016 Update of the EULAR Recommendations for Management of Rheumatoid Arthritis. *Ann. Rheum. Dis.* **2017**, *76*, 1101–1136. [[CrossRef](#)]

307. Cao, B.L.; Qasem, A.; Sharp, R.C.; Abdelli, L.S.; Naser, S.A. Systematic Review and Meta-Analysis on the Association of Tuberculosis in Crohn's Disease Patients Treated with Tumor Necrosis Factor- α Inhibitors (Anti-TNF α). *World J. Gastroenterol.* **2018**, *24*, 2764–2775. [[CrossRef](#)]
308. McDonald, J.R.; Zeringue, A.L.; Caplan, L.; Ranganathan, P.; Xian, H.; Burroughs, T.E.; Fraser, V.J.; Cunningham, F.; Eisen, S.A. Herpes Zoster Risk Factors in a National Cohort of Veterans with Rheumatoid Arthritis. *Clin. Infect. Dis.* **2009**, *48*, 1364–1371. [[CrossRef](#)]
309. Che, H.; Lukas, C.; Morel, J.; Combe, B. Risk of Herpes/Herpes Zoster during Anti-Tumor Necrosis Factor Therapy in Patients with Rheumatoid Arthritis. Systematic Review and Meta-Analysis. *Jt. Bone Spine* **2014**, *81*, 215–221. [[CrossRef](#)]
310. Winthrop, K.L.; Baddley, J.W.; Chen, L.; Liu, L.; Grijalva, C.G.; Delzell, E.; Beukelman, T.; Patkar, N.M.; Xie, F.; Saag, K.G.; et al. Association between the Initiation of Anti-Tumor Necrosis Factor Therapy and the Risk of Herpes Zoster. *JAMA* **2013**, *309*, 887–895. [[CrossRef](#)]
311. Deepak, P.; Stobaugh, D.J.; Sherid, M.; Sifuentes, H.; Ehrenpreis, E.D. Neurological Events with Tumour Necrosis Factor Alpha Inhibitors Reported to the Food and Drug Administration Adverse Event Reporting System. *Aliment. Pharm.* **2013**, *38*, 388–396. [[CrossRef](#)]
312. Kaltsonoudis, E.; Zikou, A.K.; Voulgari, P.V.; Konitsiotis, S.; Argyropoulou, M.I.; Drosos, A.A. Neurological Adverse Events in Patients Receiving Anti-TNF Therapy: A Prospective Imaging and Electrophysiological Study. *Arthritis Res.* **2014**, *16*, R125. [[CrossRef](#)] [[PubMed](#)]
313. Robinson, W.H.; Genovese, M.C.; Moreland, L.W. Demyelinating and Neurologic Events Reported in Association with Tumor Necrosis Factor Alpha Antagonism: By What Mechanisms Could Tumor Necrosis Factor Alpha Antagonists Improve Rheumatoid Arthritis but Exacerbate Multiple Sclerosis? *Arthritis Rheum.* **2001**, *44*, 1977–1983. [[CrossRef](#)]
314. Solomon, A.J.; Spain, R.I.; Kruer, M.C.; Bourdette, D. Inflammatory Neurological Disease in Patients Treated with Tumor Necrosis Factor Alpha Inhibitors. *Mult. Scler.* **2011**, *17*, 1472–1487. [[CrossRef](#)] [[PubMed](#)]
315. Saffra, N.; Astafurov, K. Visual Loss Induced by Adalimumab Used for Plaque Psoriasis. *Case Rep. Dermatol.* **2017**, 60–64. [[CrossRef](#)] [[PubMed](#)]
316. Tanno, M.; Nakamura, I.; Kobayashi, S.; Kurihara, K.; Ito, K. New-Onset Demyelination Induced by Infliximab Therapy in Two Rheumatoid Arthritis Patients. *Clin. Rheumatol.* **2006**, *25*, 929–933. [[CrossRef](#)] [[PubMed](#)]
317. Balakumar, P.; Singh, M. Anti-Tumour Necrosis Factor-Alpha Therapy in Heart Failure: Future Directions. *Basic Clin. Pharm. Toxicol.* **2006**, *99*, 391–397. [[CrossRef](#)]
318. Bucalo, A.; Rega, F.; Zangrilli, A.; Silvestri, V.; Valentini, V.; Scafetta, G.; Marraffa, F.; Grassi, S.; Rogante, E.; Piccolo, A.; et al. Paradoxical Psoriasis Induced by Anti-TNF α Treatment: Evaluation of Disease-Specific Clinical and Genetic Markers. *Int. J. Mol. Sci.* **2020**, *21*, 7873. [[CrossRef](#)]
319. Bonovas, S.; Minozzi, S.; Lytras, T.; González-Lorenzo, M.; Pecoraro, V.; Colombo, S.; Polloni, I.; Moja, L.; Cinquini, M.; Marino, V.; et al. Risk of Malignancies Using Anti-TNF Agents in Rheumatoid Arthritis, Psoriatic Arthritis, and Ankylosing Spondylitis: A Systematic Review and Meta-Analysis. *Expert Opin. Drug Saf.* **2016**, *15*, 35–54. [[CrossRef](#)]
320. Chen, Y.; Sun, J.; Yang, Y.; Huang, Y.; Liu, G. Malignancy Risk of Anti-Tumor Necrosis Factor Alpha Blockers: An Overview of Systematic Reviews and Meta-Analyses. *Clin. Rheumatol.* **2016**, *35*, 1–18. [[CrossRef](#)]
321. Shelton, E.; Laharie, D.; Scott, F.I.; Mamtani, R.; Lewis, J.D.; Colombel, J.-F.F.; Ananthakrishnan, A.N. Cancer Recurrence Following Immune-Suppressive Therapies in Patients With Immune-Mediated Diseases: A Systematic Review and Meta-Analysis. *Gastroenterology* **2016**, *151*, 97–109.e4. [[CrossRef](#)]
322. Bongartz, T.; Sutton, A.J.; Sweeting, M.J.; Buchan, I.; Matteson, E.L.; Montori, V. Anti-TNF Antibody Therapy in Rheumatoid Arthritis and the Risk of Serious Infections and Malignancies: Systematic Review and Meta-Analysis of Rare Harmful Effects in Randomized Controlled Trials. *JAMA* **2006**, *295*, 2275–2285. [[CrossRef](#)] [[PubMed](#)]
323. Askling, J.; Fored, C.M.; Baecklund, E.; Brandt, L.; Backlin, C.; Ekbom, A.; Sundström, C.; Bertilsson, L.; Cöster, L.; Geborek, P.; et al. Haematopoietic Malignancies in Rheumatoid Arthritis: Lymphoma Risk and Characteristics after Exposure to Tumour Necrosis Factor Antagonists. *Ann. Rheum. Dis.* **2005**, *64*, 1414–1420. [[CrossRef](#)] [[PubMed](#)]
324. Minozzi, S.; Bonovas, S.; Lytras, T.; Pecoraro, V.; González-Lorenzo, M.; Bastiampillai, A.J.; Gabrielli, E.M.; Lonati, A.C.; Moja, L.; Cinquini, M.; et al. Risk of Infections Using Anti-TNF Agents in Rheumatoid Arthritis, Psoriatic Arthritis, and Ankylosing Spondylitis: A Systematic Review and Meta-Analysis. *Expert Opin. Drug Saf.* **2016**, *15* (Suppl. S1), 11–34. [[CrossRef](#)] [[PubMed](#)]
325. Dignass, A.; Van Assche, G.; Lindsay, J.O.; Lémann, M.; Söderholm, J.; Colombel, J.F.; Danese, S.; D'Hoore, A.; Gassull, M.; Gomollón, F.; et al. The Second European Evidence-Based Consensus on the Diagnosis and Management of Crohn's Disease: Current Management. *J. Crohns. Colitis* **2010**, *4*, 28–62. [[CrossRef](#)] [[PubMed](#)]
326. Li, H.H.; Zhu, H.; Liu, L.S.; Huang, Y.; Guo, J.; Li, J.; Sun, X.P.; Chang, C.X.; Wang, Z.H.; Zhai, K. Tumour Necrosis Factor- α Gene Polymorphism Is Associated with Metastasis in Patients with Triple Negative Breast Cancer. *Sci. Rep.* **2015**, *5*, 10244. [[CrossRef](#)]
327. Azmy, I.A.F.; Balasubramanian, S.P.; Wilson, A.G.; Stephenson, T.J.; Cox, A.; Brown, N.J.; Reed, M.W.R. Role of Tumour Necrosis Factor Gene Polymorphisms (-308 and -238) in Breast Cancer Susceptibility and Severity. *Breast Cancer Res.* **2004**, *6*, 395–400. [[CrossRef](#)]
328. Yi, F.; Shi, X.; Pei, X.; Wu, X. Tumor Necrosis Factor-Alpha-308 Gene Promoter Polymorphism Associates with Survival of Cancer Patients: A Meta-Analysis. *Medicine (Baltimore)* **2018**, *97*, e13160. [[CrossRef](#)]
329. Balkwill, F.; Mantovani, A. Cancer and Inflammation: Implications for Pharmacology and Therapeutics. *Clin. Pharm.* **2010**, *87*, 401–406. [[CrossRef](#)]
330. Lentz, M.; Kumar, K. Reduction of Plasma Levels of Soluble Tumor Necrosis Factor and Interleukin-2 Receptors by Means of a Novel Immunoabsorption Column. *Ther. Apher. Dial.* **2008**, *12*, 491–499. [[CrossRef](#)]

331. Krippner-Heidenreich, A.; Grunwald, I.; Zimmermann, G.; Kühnle, M.; Gerspach, J.; Sterns, T.; Shnyder, S.D.; Gill, J.H.; Männel, D.N.; Pfizenmaier, K.; et al. Single-Chain TNF, a TNF Derivative with Enhanced Stability and Antitumoral Activity. *J. Immunol.* **2008**, *180*, 8176–8183. [[CrossRef](#)]
332. Josephs, S.F.; Ichim, T.E.; Prince, S.M.; Kesari, S.; Marincola, F.M.; Escobedo, A.R.; Jafri, A. Unleashing Endogenous TNF-Alpha as a Cancer Immunotherapeutic. *J. Transl. Med.* **2018**, *16*, 1–8. [[CrossRef](#)] [[PubMed](#)]
333. Chen, X.; Subleski, J.J.; Kopf, H.; Howard, O.M.Z.; Männel, D.N.; Oppenheim, J.J. Cutting Edge: Expression of TNFR2 Defines a Maximally Suppressive Subset of Mouse CD4+CD25+FoxP3+ T Regulatory Cells: Applicability to Tumor-Infiltrating T Regulatory Cells. *J. Immunol.* **2008**, *180*, 6467–6471. [[CrossRef](#)] [[PubMed](#)]
334. Facciabene, A.; Motz, G.T.; Coukos, G. T-Regulatory Cells: Key Players in Tumor Immune Escape and Angiogenesis. *Cancer Res.* **2012**, *72*, 2162–2171. [[CrossRef](#)]
335. Smyth, M.J.; Ngiew, S.F.; Teng, M.W.L. Targeting Regulatory T Cells in Tumor Immunotherapy. *Immunol. Cell Biol.* **2014**, *92*, 473–474. [[CrossRef](#)]
336. Okubo, Y.; Mera, T.; Wang, L.; Faustman, D.L. Homogeneous Expansion of Human T-Regulatory Cells via Tumor Necrosis Factor Receptor 2. *Sci. Rep.* **2013**, *3*, 3153. [[CrossRef](#)]
337. Ham, B.; Wang, N.; D’Costa, Z.; Fernandez, M.C.; Bourdeau, F.; Auguste, P.; Illemann, M.; Eefsen, R.L.; Høyer-Hansen, G.; Vainer, B.; et al. TNF Receptor-2 Facilitates an Immunosuppressive Microenvironment in the Liver to Promote the Colonization and Growth of Hepatic Metastases. *Cancer Res.* **2015**, *75*, 5235–5247. [[CrossRef](#)] [[PubMed](#)]
338. Wang, J.; Al-Lamki, R.S. Tumor Necrosis Factor Receptor 2: Its Contribution to Acute Cellular Rejection and Clear Cell Renal Carcinoma. *Biomed Res. Int.* **2013**, *2013*, 821310. [[CrossRef](#)] [[PubMed](#)]
339. Hamilton, K.E.; Simmons, J.G.; Ding, S.; Van Landeghem, L.; Lund, P.K. Cytokine Induction of Tumor Necrosis Factor Receptor 2 Is Mediated by STAT3 in Colon Cancer Cells. *Mol. Cancer Res.* **2011**, *9*, 1718–1731. [[CrossRef](#)]
340. Nakayama, S.; Yokote, T.; Tsuji, M.; Akioka, T.; Miyoshi, T.; Hirata, Y.; Hiraoka, N.; Iwaki, K.; Takayama, A.; Nishiwaki, U.; et al. Expression of Tumour Necrosis Factor- α and Its Receptors in Hodgkin Lymphoma. *Br. J. Haematol.* **2014**, 574–577. [[CrossRef](#)]
341. Rauert, H.; Stühmer, T.; Bargou, R.; Wajant, H.; Siegmund, D. TNFR1 and TNFR2 Regulate the Extrinsic Apoptotic Pathway in Myeloma Cells by Multiple Mechanisms. *Cell Death Dis.* **2011**, *2*, e194. [[CrossRef](#)]
342. Uhlén, M.; Björling, E.; Agaton, C.; Szigartyo, C.A.-K.; Amini, B.; Andersen, E.; Andersson, A.-C.; Angelidou, P.; Asplund, A.; Asplund, C.; et al. A Human Protein Atlas for Normal and Cancer Tissues Based on Antibody Proteomics. *Mol. Cell. Proteom.* **2005**, *4*, 1920–1932. [[CrossRef](#)] [[PubMed](#)]
343. Williams, G.S.; Mistry, B.; Guillard, S.; Ulrichsen, J.C.; Sandercock, A.M.; Wang, J.; González-Muñoz, A.; Parmentier, J.; Black, C.; Soden, J.; et al. Phenotypic Screening Reveals TNFR2 as a Promising Target for Cancer Immunotherapy. *Oncotarget* **2016**, *7*, 68278–68291. [[CrossRef](#)] [[PubMed](#)]
344. Goukassian, D.A.; Qin, G.; Dolan, C.; Murayama, T.; Silver, M.; Curry, C.; Eaton, E.; Luedemann, C.; Ma, H.; Asahara, T.; et al. Tumor Necrosis Factor-Alpha Receptor P75 Is Required in Ischemia-Induced Neovascularization. *Circulation* **2007**, *115*, 752–762. [[CrossRef](#)] [[PubMed](#)]
345. Naserian, S.; Abdelgawad, M.E.; Afshar Bakshloo, M.; Ha, G.; Arouche, N.; Cohen, J.L.; Salomon, B.L.; Uzan, G. The TNF/TNFR2 Signaling Pathway Is a Key Regulatory Factor in Endothelial Progenitor Cell Immunosuppressive Effect. *Cell Commun. Signal.* **2020**, *18*, 94. [[CrossRef](#)] [[PubMed](#)]
346. Vanamee, É.S.; Faustman, D.L. TNFR2: A Novel Target for Cancer Immunotherapy. *Trends Mol. Med.* **2017**, *23*, 1037–1046. [[CrossRef](#)]
347. Torrey, H.; Butterworth, J.; Mera, T.; Okubo, Y.; Wang, L.; Baum, D.; Defusco, A.; Plager, S.; Warden, S.; Huang, D.; et al. Targeting TNFR2 with Antagonistic Antibodies Inhibits Proliferation of Ovarian Cancer Cells and Tumor-Associated Tregs. *Sci. Signal.* **2017**, *10*, eaaf8608. [[CrossRef](#)] [[PubMed](#)]
348. Yang, M.; Tran, L.; Torrey, H.; Song, Y.; Perkins, H.; Case, K.; Zheng, H.; Takahashi, H.; Kuhlreiber, W.M.; Faustman, D.L. Optimizing TNFR2 Antagonism for Immunotherapy with Tumor Microenvironment Specificity. *J. Leukoc. Biol.* **2020**, *107*, 971–980. [[CrossRef](#)]