Soo In Kim<sup>1</sup>, Yeon Seok Jang<sup>1</sup>, Seung Hee Han<sup>1</sup>, Myeong Jin Choi<sup>1</sup>, Eun Hye Go<sup>1</sup>, Yong-Pil Cheon<sup>2</sup>, Jung Sick Lee<sup>3</sup> and Sung-Ho Lee<sup>1,†</sup>

<sup>1</sup>Dept. of Green Life Science, Sangmyung University, Seoul 110-743, Korea <sup>2</sup>Dept. of Biology, Sungshin Women's University, Seoul 142-732, Korea <sup>3</sup>Dept. of Aqualife Medicine, Chonnam National University, Yeosu 550-749, Korea

**ABSTRACT** : Manganese  $(Mn^{2+})$  is a trace element that is essential for normal physiology, and is predominantly obtained from food. Several lines of evidence, however, demonstrated that overexposure to MnCl<sub>2</sub> exerts serious neurotoxicity, immunotoxicity and developmental toxicity, particularly in male. The present study aimed to evaluate the effect of 0, 1.0, 3.3, and 10 mg/kg/day doses of MnCl<sub>2</sub> on the reproductive organs in the immature female rats. Rats (PND 22; S.D. strain) were exposed to MnCl<sub>2</sub> (MnCl<sub>2</sub> · 4H<sub>2</sub>O) dissolved in drinking water for 2 weeks. The animals were sacrificed on PND 35, then the tissues were immediately removed and weighed. Histological studies were performed using the uteri tissue samples. Serum LH and FSH levels were measured with the specific ELISA kits. Body weights of the experimental group animals were not significantly different from those of control group animals. However, ovarian tissue weights in 1 mg and 3.3 mg MnCl<sub>2</sub> dose groups were significantly lower than those of control animals (p < 0.05 and p < 0.01, respectively). Uterine tissue weights of 3.3 mg dose MnCl<sub>2</sub> groups were significantly lower than those of control animals (p < 0.01), while the 1 mg MnCl<sub>2</sub> dose and 10 mg MnCl<sub>2</sub> dose failed to induce any change in uterine weight. Similarly, only 3.3 mg MnCl<sub>2</sub> dose could induce the significant decrease in the oviduct weight compared to the control group (p < 0.05). Non-reproductive tissues such as adrenal and kidney failed to respond to all doses of MnCl<sub>2</sub> exposure. The uterine histology revealed that the MnCl<sub>2</sub> exposure could affect the myometrial cell proliferation particularly in 3.3 mg dose and 10mg dose group. Serum FSH levels were significantly decreased in 1mg MnCl<sub>2</sub> dose and 10 MnCl<sub>2</sub> mg groups (p < 0.05 and p < 0.01, respectively). In contrast, treatment with 1 mg MnCl<sub>2</sub> dose induced a significant increment of serum LH level (p < 0.05). The present study demonstrated that MnCl<sub>2</sub> exposure is capable of inducing abnormal development of reproductive tissues, at least to some extent, and altered gonadotropin secretions in immature female rats. Combined with the well-defined actions of this metal on GnRH and prolactin secretion, one can suggest the Mn<sup>2+</sup> might be a potential environmental mediator which is involved in the female pubertal process.

Key words : Manganese (Mn2+), Immature female rats, Ovary, Uterus, Gonadotropins, Pubertal process

## INTRODUCTION

Manganese  $(Mn^{2+})$  is a trace metal and is essential element that is required for normal mammalian physiology. Several enzyme systems have been reported to interact with or depend on  $Mn^{2+}$  for their optimal catalytic or regulatory function (Gunter et al., 2006). On the other hand, excessive exposure to  $Mn^{2+}$  seems to cause serious neurotoxicity, immunotoxicity and developmental toxicity, particularly in male (Michalke et al., 2007).

Concerning the reproductive toxicity of  $Mn^{2+}$ , studies have been focused on the mammalian testicular dysfunction. Chronic exposure to  $Mn_3O_4$  (1,050 ppm) retarded the sexual development shown significantly smaller testis, seminal vesicle, and preputial gland weights in mice (Gray & Laskey, 1980; Webster & Valois, 1987). In human,

<sup>&</sup>lt;sup>\*</sup> Corresponding author: Sung-Ho Lee, Dept. of Green Life Science, Sangmyung University, Seoul 110-743, Korea. Phone: +82-2-2287-5139, Fax: +82-2-2287-0098, E-mail: shlee@smu.ac.kr

occupational exposure to  $Mn^{2+}$  decreased libido and impotency (Emara et al., 1971; Mena et al., 1967), and may result in lowered sperm count and semen quality (Hjollund et al., 1998).

So far, however, research on the effect of  $Mn^{2+}$  exposure on female reproductive physiology has been conducted poorly. The present study aimed to evaluate the effect of 0, 1.0, 3.3, and 10 mg/kg/day doses of  $MnCl_2$  on the reproductive organs in the immature female rats.

## **MATERIALS & METHODS**

## 1. Animals

Timely pregnant Sprague-Dawley rats were obtained from DBL (Chungcheongbuk-do, Korea) and reared in sangmyung university animal facility under conditions of 12-h light/dark cycle (lights on at 07:00 h) and constant temperature of  $22\pm1$  °C. During pregnancy and lactation, the mothers had free access to normal chow and tap water. All procedures used were approved by the Animal Care and Use Committee at Sangmyung University.

## 2. Experimental design

The day after weaning (postnatal day 22, PND 22), female dams were randomly assigned to the following exposure groups (n=8 dams/group) from PND 22 until PND 35: (1) oral administration of 0.9% Saline (300  $\mu l$ /100 g BW/day, Control group), (2) oral administration of 1.0 mg/kg BW/day MnCl<sub>2</sub> · 4H<sub>2</sub>O (Sigma-Aldrich), (3) oral administration of 3.3 mg/kg BW/day MnCl<sub>2</sub> · 4H<sub>2</sub>O, and (4) oral administration of 10 mg/kg BW/day MnCl<sub>2</sub> · 4H<sub>2</sub>O. Both saline and Mn<sup>2+</sup> solution were supplied daily for 2 weeks. At PND 35, animals were sacrificed and the tissues(ovary, uterus, oviduct, adrenal, and kidney) were immediately removed and weighed at 1,800 hour.

## 3. Histology

Uterine tissue specimens were fixed 4% paraformaldehyde then were serially dehydrated in graded ethanol and xylene. Specimens were paraffin embedded and sectioned at 5  $\mu$ m thickness. Sections were stained with Hematoxylin-Eosin (H & E) stain and examined under light microscope.

### 4. Measurement of serum gonadotropin levels

The trunk blood samples were collected and centrifuged at 3,000×g for 15 min. The measurements of the serum FSH and LH were carried out according to the commercial instructions for the specific ELISA kits (USCN, China). The sensitivities of the FSH and LH assay were 0.55 ng/ml and 122.5 pg/ml, respectively. The intra-assay and inter-assay coefficients of variation were <10% and <12% for both hormones, respectively.

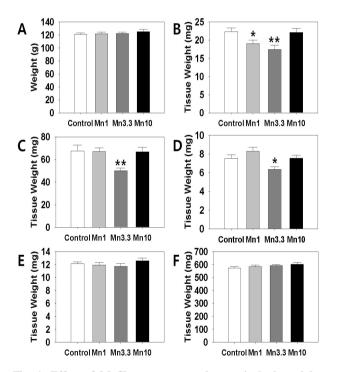
#### 5. Statistical analysis

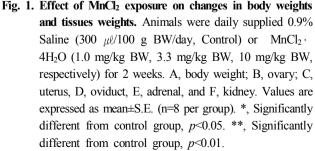
All values are expressed as the means ( $\pm$ S.E.). Differences between control and treatment groups were analysed by Student's t-test. *P* values less than 0.05 were considered significant. The IBM PC programs INSTAT and PRISM 3.0 (GraphPad, San Diego, CA, USA) were used to calculate and plot the results.

# RESULTS

#### 1. Tissue weights

In order to evaluate a potential effect of  $Mn^{2+}$  on female reproductive organs, we measured the tissues weights of ovary, uterus and oviduct. Kidney and adrenal weight served as non-reproductive tissues. After 2 weeks of administration, body weights of the all  $Mn^{2+}$  exposure group animals were not significantly different from those of control group animals (Fig. 1, A). However, ovarian tissue weights (Fig. 1, B). in 1 mg MnCl<sub>2</sub> dose group (19.06±0.98 mg, *p*<0.05) and 3.3 mg MnCl<sub>2</sub> dose group (17.47±1.16 mg, *p*<0.01) were significantly lower than those of control animals (22.39±0.97 mg). Uterine tissue weights (Fig. 1, C). of 3.3 mg dose MnCl<sub>2</sub> groups were significantly lower than those of control animals (Control : 3.3 mg dose MnCl<sub>2</sub> =  $67.52\pm5.15$  :  $50.04\pm2.41$  mg, p<0.01), while the 1 mg MnCl<sub>2</sub> dose (Control : 1 mg dose MnCl<sub>2</sub>= $67.52\pm5.15$  :  $67.11\pm3.28$  mg) and 10 mg MnCl<sub>2</sub> dose (Control : 10 mg dose MnCl<sub>2</sub> =  $67.52\pm5.15$  :  $66.70\pm4.03$  mg) failed to induce any change in uterine weight. Similarly, only 3.3 mg MnCl<sub>2</sub> dose could induce the significant decrease in the oviduct weight (Fig. 1, D). compared to the control group (Control : 3.3 mg dose MnCl<sub>2</sub> =  $7.52\pm0.40$  :  $6.35\pm0.29$  mg, p<0.05). Non-reproductive tissues such as adrenal (Control : 1 mg : 3.3 mg : 10 mg dose MnCl<sub>2</sub>





=  $12.13\pm0.28$  :  $11.93\pm0.38$  :  $11.75\pm0.42$  :  $12.59\pm0.43$ mg) and kidney (Control : 1 mg : 3.3 mg : 10 mg dose MnCl<sub>2</sub> =  $574.74\pm8.99$  :  $584.56\pm12.38$  :  $592.42\pm7.82$  :  $603.67\pm14.06$  mg) failed to respond to all doses of MnCl<sub>2</sub> exposure (Fig. 1, E & F, respectively).

# 2. Uterine histology

To access the histological changes in MnCl<sub>2</sub> exposured uteri, standard paraffin section and hematoxylin-eosin staining method were employed. 3.3 mg and 10 mg MnCl<sub>2</sub> dose groups shown the thickend myometrial layer when compared to control (Fig. 2, Mn3.3 & Mn10). In contrast, 1 mg MnCl<sub>2</sub> exposure reduced the thickness of myometrium (Fig. 2, Mn1).

# 3. ELISA

Serum LH and FSH levels were measured using specific ELISA kits. Fig. 3 (A) shows that the secretion of FSH was significantly decreased by treatment with 1 mg or 3.3 mg MnCl<sub>2</sub> (Control:1 mg dose MnCl<sub>2</sub> =  $3.54\pm0.18$  :  $3.07\pm0.14$  ng/ml, p<0.05; Control : 10 mg dose MnCl<sub>2</sub> =  $3.54\pm0.18:2.75\pm0.10$  ng/ml, p<0.01). Serum LH level was significantly elevated by 1 mg MnCl<sub>2</sub> exposure (Control : 1 mg dose MnCl<sub>2</sub> =  $39.85\pm8.93:76.71\pm12.36$  ng/ml, p<0.05, Fig. 3, B). Higher doses of MnCl<sub>2</sub> exposure failed to change the serum LH levels.

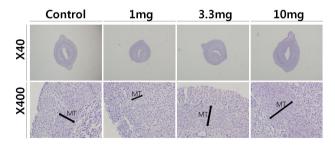


Fig. 2. Effect of MnCl<sub>2</sub> exposure on histological changes in uteri from immature rats. Standard paraffin section and hematoxylin-eosin staining method were employed. Mn 1, 1.0 mg/kg BW/ day of MnCl<sub>2</sub> · 4H<sub>2</sub>O exposure; Mn 3.3, 3.3 mg/kg BW/day of MnCl<sub>2</sub> · 4H<sub>2</sub>O exposure; Mn 10, 10 mg/kg BW/day of MnCl<sub>2</sub> · 4H<sub>2</sub>O exposure, respectively.

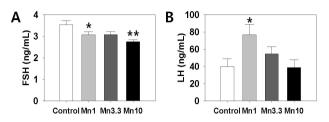


Fig. 3. Measurement of serum FSH and LH levels in response to  $Mn^{2+}$  exposure. The trunk blood samples were collected and centrifuged at 3,000×g for 15 min. The measurements of the serum FSH and LH were carried out according to the commercial instructions for the specific ELISA kits (USCN, China). Values are expressed as mean±S.E. (n=6 per group). \*, Significantly different from control group, *p*<0.05. \*\*, Significantly different from control group, *p*<0.01.

#### DISCUSSION

In the present study we demonstrated that the  $Mn^{2+}$  exposure could change the weights of reproductive tissues in immature female rats. Although we failed to find the advance or delay of puberty onset in the  $Mn^{2+}$  exposured animals (data not shown), the potential reproductive toxicity of this metal in immature female rats cannot be ruled out.  $Mn^{2+}$  exposure, Indeed, not only affected the proliferative activity in myometrial layer, but induced significant changes in the serum FSH and LH levels.

More defined doses and exposure periods will be helpful to verify the physiological relevance of Mn<sup>2+</sup> exposure in female reproduction.

Occupational exposure to  $Mn^{2+}$  could be occurred often at the workplaces such are mines and dried battery factories (Emara et al., 1971; Mena et al., 1967). In this respect, most studies on the toxicological effects of  $Mn^{2+}$  have been focused on the men. Furthermore, a relatively small portion of the studies dealt with reproductive toxicity of  $Mn^{2+}$  exposure using male animal models.  $Mn^{2+}$  exposure for 2 and 4 h inhibited rat primary Leydig cell steroidogenesis by decreasing StAR protein expression while 24 and 48 h exposure of MnCl<sub>2</sub> caused adverse effects on both StAR protein and P450scc and 3b-HSD enzyme activity to reduce steroid-ogenesis (Cheng et al., 2003).

Pine et al. (2005) reported that  $Mn^{2+}$  administered acutely into the third ventricle shown dose-dependently to stimulate LH release in prepubertal female rat, and this effect was due to a Mn<sup>2+</sup>-induced stimulation of GnRH. The authors demonstrated that Mn<sup>2+</sup> can stimulate specific puberty-related hormones and suggested that it may facilitate the normal onset of puberty. According to them, Mn<sup>2+</sup> may contribute to precocious puberty if an individual is exposed to elevated levels of Mn<sup>2+</sup> too early in developmental process. This hypothesis was verified by same research group; Lee et al. (2006) demonstrated that Mn<sup>2+</sup> is a direct stimulator of prepubertal GnRH/LH secretion and may facilitate the normal onset of male puberty. Their data that Mn<sup>2+</sup> can cause GnRH release in adult males, suggested this action should be considered in relation to age, gender, as well as mechanistic and functional differences between adult and immature animals.

In the present study we failed to confirm the advanced puberty onset in  $Mn^{2+}$  exposed female rats (up to 10 mg/kg BW, data not shown). Pine et al. (2005) shown that same dose of Mn<sup>2+</sup> exposure initiated a moderate but significant advancement in age at vaginal opening (VO) in terms of days (1.5 days). The only difference between the two studies was timing and duration of Mn<sup>2+</sup> exposure: Our treatment regimen was PND 21-35 (14 days), and theirs was PND 12-29 (18 days). Though Mn<sup>2+</sup> can acutely induce GnRH secretion in adult males, one should consider that additional action of Mn<sup>2+</sup> to release GABA, a GnRH inhibitor, may ultimately contribute to suppressed reproductive function observed in adult animals following exposure to high chromic levels of Mn<sup>2+</sup> (Prestifilippo et al., 2008). Because neurotransmitter secretory circuits in female during peripubertal period are remarkably unstable, the extent of GABA-

GnRH regulation, and probably more neuronal regulation, may vary responding to the minor difference. Studies indicate the more specific action mechanism of Mn<sup>2+</sup> within the hypothalamus; Mn<sup>2+</sup> activates soluble guanylate cyclase (sGC) directly and/or as a cofactor with available nitric oxide (NO), hence generating cGMP and resulting in prepubertal GnRH release (Lee et al., 2007). More recently, Mn<sup>2+</sup>, through the upregulation of IGF-1 and COX-2, may promote maturational events and glialneuronal communications facilitating the increased neurosecretory activity, including that of GnRH, resulting in precocious pubertal development (Hiney et al., 2011).

Special emphasis should be made on the action of Mn<sup>2+</sup> in brain. Inhalation of the mixture of MnCl<sub>2</sub> and Mn(OAc)<sub>3</sub> for 5 months developed movement abnormalities, significant loss of substantia nigra compacta (SNc) dopaminergic neurons; these symptoms similar to those observed in Parkinson disease (PD) (Ordoñez-Librado et al., 2010). Dopamine (DA) depletion is closely related to pituitary prolactin biosynthesis. Male rats exposed to  $Mn^{2+}$  for 4 or 13 weeks showed a progressive and significant decrease in hypothalamic DA, whereas prolactin and Pit-1 mRNA levels increased in response to Mn<sup>2+</sup> exposure (Kim et al., 2009). These results suggest that exposure to Mn<sup>2+</sup> decreases hypothalamic DA and promotes the production of prolactin in the pituitary and that Pit-1 might be a regulator of DA and prolactin. Furthermore, such Mn<sup>2+</sup> exposure induced significant increase in serum prolactin levels seemed to be highly correlated with the testis toxicity (Lee. 2009). In this context, relationship between Mn<sup>2+</sup> exposure and DA/ prolactin secretions in immature female rats will be helpful to understand the function of the metal during pubertal development.

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