

## Whole-Genome Sequence of *Ralstonia solanacearum* P673, a Strain Capable of Infecting Tomato Plants at Low Temperatures

## Ana M. Bocsanczy,<sup>a</sup> Jose C. Huguet-Tapia,<sup>b</sup> David J. Norman<sup>a</sup>

Department of Plant Pathology, University of Florida, IFAS, Mid-Florida Research and Education Center, Apopka, Florida, USA<sup>a</sup>; Department of Plant Pathology, University of Florida, Gainesville, Florida, USA<sup>b</sup>

*Ralstonia solanacearum* is the causal agent of bacterial wilt, one of the most destructive bacterial plant diseases. We present the whole-genome sequence of the strain P673 (phylotype IIB, sequevar 4). This strain is capable of producing disease in tomato plants at low temperatures. P673 has 311 unique genes.

Received 30 January 2014 Accepted 3 February 2014 Published 20 February 2014

Citation Bocsanczy AM, Huguet-Tapia JC, Norman DJ. 2014. Whole-genome sequence of *Ralstonia solanacearum* P673, a strain capable of infecting tomato plants at low temperatures. Genome Announc. 2(1):e00106-14. doi:10.1128/genomeA.00106-14.

Copyright © 2014 Bocsanczy et al. This is an open-access article distributed under the terms of the Creative Commons Attribution 3.0 Unported license.

Address correspondence to Ana M. Bocsanczy, anamariab@ufl.edu, or David J. Norman, djn@ufl.edu.

**R**alstonia solanacearum (Smith 1896) Yabuuchi et al. 1996 is a Gram-negative soil-borne bacterium that causes bacterial wilt, one of the most destructive bacterial diseases in tropical, subtropical, and recently some temperate areas of the world (1, 2). *R. solanacearum* exhibits such phenotypic and genotypic diversity that it is considered a species complex (3). All established populations of *R. solanacearum* found in the southern United States have been classified as race 1 and biovars 1 and 3 (4). P673 is a race 1 biovar 1 (R1B1) strain with special characteristics. It was isolated from an *Epipremnum aureum* (pothos) ornamental plant (5) and is able to infect tomato and, to a lesser degree, potato plants at 18°C in chamber tests, when inoculated naturally through the soil (6). It was classified as phylotype IIb, sequevar 4, which places it in the same group as the exotic strains in the Caribbean that have been detected in pond and irrigation water in Florida (7).

Genomic DNA was extracted using the UltraClean microbial DNA isolation kit (Mo Bio Laboratories, Inc.), according to the manufacturer's instructions. A combination of two Illumina single-read and two 454 shotgun libraries produced >18 million reads and an additional 152,375 reads, respectively. The reads were assembled into 251 contigs using the hybrid strategy MIRA software version 3.05 (8). Contigs were annotated by the Prokaryotic Genomes Annotation Pipeline (PGAP) at NCBI (9). There are 4,690 coding sequences (CDSs), 2 rRNAs, 3 noncoding RNAs (ncRNAs), and 45 tRNAs predicted. The annotated contigs were deposited in the NCBI database. Additionally, contigs were assembled in two replicons or gapped chromosomes by the Geneious 6.0 software (Biomatters Ltd.) using the genomic sequence of the closely related strain R. solanacearum Po82 (10) as a reference. The chromosome is represented by 3.48 Mbp and the megaplasmid by 1.91 Mbp. The overall length of approximately 5.39 Mbp is shorter than the sequence of Po82 (5.43 Mbp) (10). The overall G+Ccontent of the P673 genome is 66.9%.

Biological functions were assigned to 63% of the predicted proteins. The most represented categories were amino acid, protein, carbohydrate, and lipid metabolism (30%) and stress response, virulence, and cell wall biogenesis (3% each). Hypothetical protein and no-match categories were assigned to 37% of proteins. A comparison of P673 replicons with those of 7 complete genome sequences (10, 11) revealed that 311 genes are unique to P673. Elucidating the genomic differences between P673 and similar strains that do not infect their host at low temperatures will facilitate the discovery of key cool virulence proteins.

**Nucleotide sequence accession numbers.** This whole-genome shotgun project has been deposited at DDBJ/EMBL/GenBank under the accession number JALO00000000. The version described in this paper is version JALO01000000.

## ACKNOWLEDGMENTS

This research was funded by the U.S. Department of Agriculture Floriculture and Nursery Research Initiative and the University of Florida Institute of Food and Agricultural Sciences.

We thank the International Center for Biotechnology Research (ICBR) genomics core of the University of Florida, Gainesville, FL, for the preparation of the libraries and sequencing of the *R. solanacearum* P673 genome.

## REFERENCES

- 1. Elphinstone JG. 2005. The current bacterial wilt situation: a global overview, p 9–28. *In* Allen C, Prior P, Hayward AC (ed), Bacterial wilt disease and the *Ralstonia solanacearum* species complex. APS Publishing, St. Paul, MN.
- Janse JD, van den Beld HE, Elphinstone J, Simpkins S, Tjou-Tam-Sin NNA, van Vaerenbergh J. 2004. Introduction to Europe of *Ralstonia* solanacearum biovar. 2, race 3 in *Pelargonium zonale* cuttings. J. Plant Pathol. 86:147–155.
- 3. Fegan M, Prior P. 2005. How complex is the "*Ralstonia solanacearum* species complex," p 449–462. *In* Allen C, Prior P, Hayward AC (ed), Bacterial wilt disease and the *Ralstonia solanacearum* species complex. APS Publishing, St. Paul, MN.
- Ji PS, Allen C, Sanchez-Perez A, Yao J, Elphinstone JG, Jones JB, Momol AT. 2007. New diversity of *Ralstonia solanacearum* strains associated with vegetable and ornamental crops in Florida. Plant Dis. 91: 195–203. http://dx.doi.org/10.1094/PDIS-91-2-0195.
- Norman DJ, Zapata M, Gabriel DW, Duan YP, Yuen JM, Mangravita-Novo A, Donahoo RS. 2009. Genetic diversity and host range variation of *Ralstonia solanacearum* strains entering North America. Phytopathology 99:1070–1077. http://dx.doi.org/10.1094/PHYTO-99-9-1070.

- Bocsanczy AM, Achenbach UC, Mangravita-Novo A, Yuen JM, Norman DJ. 2012. Comparative effect of low temperature on virulence and twitching motility of *Ralstonia solanacearum* strains present in Florida. Phytopathology 102:185–194. http://dx.doi.org/10.1094/PHYTO-05-11-0145.
- 7. Hong JC, Momol MT, Jones JB, Ji P, Olson SM, Allen C, Perez A, Pradhanang P, Guven K. 2008. Detection of *Ralstonia solanacearum* in irrigation ponds and aquatic weeds associated with the ponds in North Florida. Plant Dis. 92:1674–1682. http://dx.doi.org/10.1094/PDIS-92-12-1674.
- Chevreux B, Wetter T, Suhai S. 1999. Genome sequence assembly using trace signals and additional sequence information, p 45–56. *In* Computer science and biology: proceedings of the German Conference on Bioinformatics (GCB '99). German Conference on Bioinformatics, Hannover, Germany. http://www.bioinfo.de/isb/gcb99/talks/chevreux/.
- Angiuoli SV, Gussman A, Klimke W, Cochrane G, Field D, Garrity G, Kodira CD, Kyrpides N, Madupu R, Markowitz V, Tatusova T, Thomson N, White O. 2008. Toward an online repository of Standard Operating Procedures (SOPs) for (meta)genomic annotation. Omics 12: 137–141. http://dx.doi.org/10.1089/omi.2008.0017.
- Xu J, Zheng HJ, Liu L, Pan ZC, Prior P, Tang B, Xu JS, Zhang H, Tian Q, Zhang LQ, Feng J. 2011. Complete genome sequence of the plant pathogen *Ralstonia solanacearum* strain Po82. J. Bacteriol. 193: 4261–4262. http://dx.doi.org/10.1128/JB.05384-11.
- Remenant B, Coupat-Goutaland B, Guidot A, Cellier G, Wicker E, Allen C, Fegan M, Pruvost O, Elbaz M, Calteau A, Salvignol G, Mornico D, Mangenot S, Barbe V, Medigue C, Prior P. 2010. Genomes of three tomato pathogens within the *Ralstonia solanacearum* species complex reveal significant evolutionary divergence. BMC Genomics 11:379. http://dx .doi.org/10.1186/1471-2164-11-379.