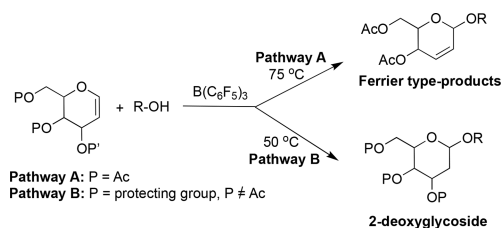




### Scheme 1. BCF-Catalysed Synthesis of 2,3-Unsaturated Glycosides from “Disarmed” Glycals (Pathway A) and 2- $\alpha$ -Deoxyglycosides from “armed” Glycals (Pathway B)



temperatures; changing the solvent to  $\text{CH}_2\text{Cl}_2$  or  $\text{CH}_3\text{CN}$  was also detrimental to the reaction (see Table S1 in the Supporting Information (SI) for solvent and temperature screen details). Having established the optimal reaction conditions, our attention then turned to exploring the substrate scope of the reaction between **1a** and a range of OH nucleophiles **2b–2k** (Table 1). In all cases, reactions proceeded smoothly within 1.5–4 h and in good to excellent yields and a clear preference for the  $\alpha$ -products, demonstrating the reaction is tolerant of primary, secondary, and phenolic OH nucleophiles, as well as common alcohol protecting groups (e.g., acetals, ethers, and esters).

Glycosylations with primary alcohols **2b–2e** afforded the corresponding 2,3-unsaturated glycosides in 72–86% yield within 2 h and with a 30:1  $\alpha$ : $\beta$  ratio (entries 2–5). Reactions with secondary alcohols such as 4-methoxyphenol **2i**, cholesterol **2j**, or *N*-hydroxysuccinimide **2k** (entries 9–11) proceeded smoothly giving the desired products in similar high  $\alpha$ -selectivity (>30:1 to >20:1,  $\alpha$ / $\beta$  ratio) and yields of 77–82%. Reactions with glycosides **2g**, **2h**, and propargyl alcohol **2f** (entries 6–8) prove to be more challenging and afforded the products in lower yields and stereoselectivity, albeit in favor of the  $\alpha$ -products (67–86%, 6:1–7:1  $\alpha$ : $\beta$  ratio). The ability to effectively activate galactals is noteworthy, as generally glycal substrates that favor a bigger shift toward  $^5\text{H}_4$  conformations (e.g., glucals) undergo rearrangement more readily than their C-4 epimer galactals where the equilibria is shifted toward the  $^4\text{H}_5$  form and as a result galactals often give mixtures of products, as well as lower overall yields.<sup>10</sup>

The scope of the reaction was further investigated with regards to glycal donor. To this end, a series of peracetylated glycals: D-glucal **1b**, L-fucal **1c**, D-xylal **1d** and L-rhamnal **1e** were subjected to the reaction conditions with **2e** as the model OH nucleophile (Scheme 2). In general, moderate to good yields (65–86%) and  $\alpha$ -selectivities were obtained in all cases leading to the formation of 2-deoxy and 2,6-dideoxy Ferrier-type products. Best diastereoselectivities were observed for 2,6-dideoxyglycals (15:1–30:1  $\alpha$ : $\beta$ , **1c–1e**), while glucal **1b** yielded **4b** in 86% yield and a 3:1  $\alpha$ : $\beta$  ratio.

The synthetic utility of our strategy was further exemplified on the preparation of rare glycoside analogues **6–8**, which are often difficult to access by traditional methods<sup>11</sup> bearing a Boc-protected amino propyl linker that could be used for array conjugation (Scheme 3). Glycosylation of 3-(Boc-amino)-1-propanol with **1c**, followed by ester deprotection gave 2,3-unsaturated fucoside **5** in 83% yield (2 steps) and a 10:1  $\alpha$ / $\beta$  ratio. Alkene reduction of **5** with  $\text{Rh}-\text{Al}_2\text{O}_3$  afforded  $\alpha$ -L-Rhodinose **6** in 76% yield. Alternatively, subjecting **5** to reduction followed by treatment with Dess-Martin periodinane gave  $\alpha$ -L-cinerulose **7** (55%), while direct alcohol oxidation of **5** yields  $\alpha$ -L-aculose **8** (79%).

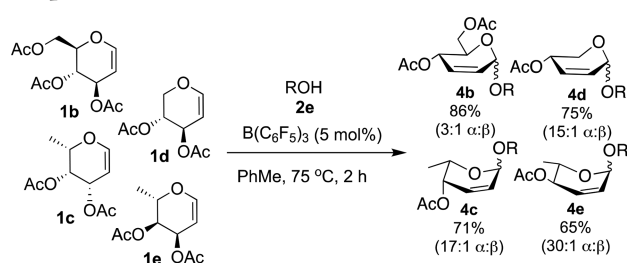
### Table 1. Glycosylation Reactions with Galactal **1a**<sup>c</sup>

Entry	ROH	Time (h)	Yield (%) <sup>[a]</sup>	$\alpha$ : $\beta$ <sup>[b]</sup>
1	<b>2a</b>	2	88 <sup>[c,d]</sup>	30:1
2	BnOH <b>2b</b>	1.5	84	20:1
3	<b>2c</b>	1.5	72	30:1
4	<b>2d</b>	2	72	30:1
5	<b>2e</b>	2	75	30:1
6	<b>2f</b>	3	86	7:1
7	<b>2g</b>	3	68	6:1
8	<b>2h</b>	2	67	6:1
9	<b>2i</b>	1.5	82	30:1
10	<b>2j</b>	3	79	20:1
11	<b>2k</b>	4	91	30:1

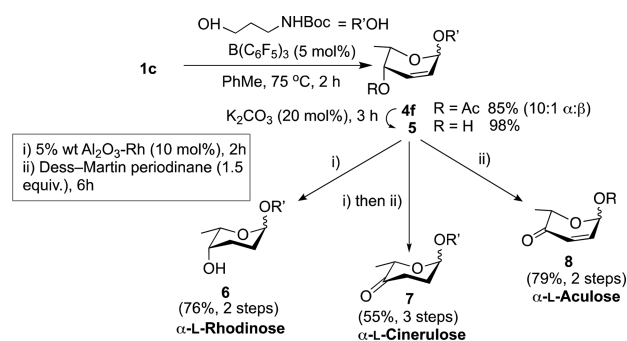
<sup>a</sup>Isolated yield. <sup>b</sup>Determined by  $^1\text{H-NMR}$ . <sup>c</sup>Reaction did not proceed in the absence of catalyst. <sup>d</sup>Activation with  $\text{BF}_3\cdot\text{OEt}_2$  afforded **3a** in 19% as a 6:1 mixture of anomers.

Next, we explored whether “armed” glycosides lacking a leaving group at C-3 could undergo BCF-activation and give substitution products selectively. To probe this, reactions between perbenzylated galactal **9a**, acceptor **2a** and BCF were screened at different catalyst loadings, solvents and temperatures as before. Best results were found when 5 mol %  $\text{B}(\text{C}_6\text{F}_5)_3$  was used in toluene at 50 °C to give 2-deoxyglycoside **10a** after 2 h (88%,  $\alpha$ / $\beta$  > 30:1, entry 1, Table 2). As before, reactions were less efficient at lower catalyst loadings or temperatures below 50 °C and changing

Scheme 2. Reaction of Glycals 1a–1e, with Glycoside Acceptors 2e



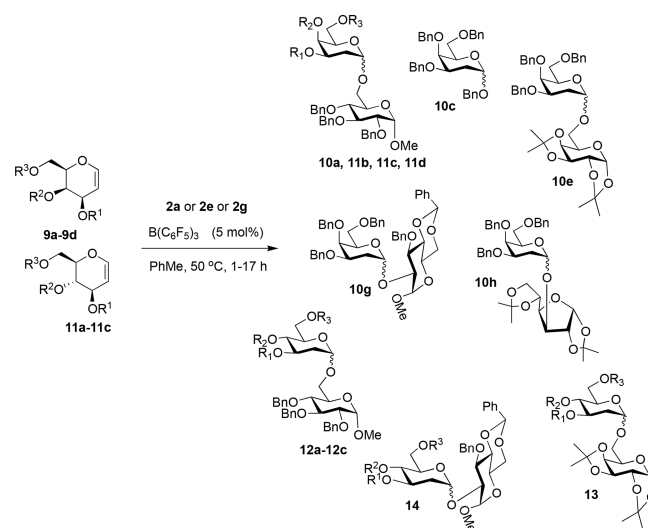
Scheme 3. Synthesis of Rare Glycosides 6–8



the solvent to  $\text{CH}_2\text{Cl}_2$ ,  $\text{CH}_3\text{CN}$  or  $\text{CF}_3\text{Ph}$  was also detrimental (SI Table S2 for full details).

To explore the substrate scope of the glycosylation, galactals **9b–d** and glucals **11a–c** were reacted with a range of primary and secondary OH nucleophiles **2a**, **2e**, or **2g** under the optimized reaction conditions. In all cases, reactions proceeded smoothly in yields of 62–94% and high  $\alpha$ -selectivity (20:1–30:1  $\alpha/\beta$ ), with secondary OHs requiring longer reaction times (entries 4 and 5 vs 1–3). Subsequently, a series of differentially protected galactals **9b–d** and glucals **11a–c** bearing benzyl, methoxymethyl acetal, silyl ethers and acetals, and siloxane protecting groups were prepared and subjected to the reaction conditions to investigate the effect of glycal donor on the reaction. Pleasingly, reactions involving all galactals were complete within 1–7 h, in good yields (71–82%) and high  $\alpha$ -selectivities (20:1 to 30:1  $\alpha/\beta$ ) (entries 6–8). The reaction was also amenable to glycosylations with glucal substrates, albeit required longer reaction times (17 h) and afforded the glycoside products in moderate to good yields (54–86%) with similarly high  $\alpha$ -stereocontrol. Siloxane protected donors **11b** and **11c** gave better yields than perbenzylated glucal **11a** (entries 9–13) as expected.<sup>4c</sup> These results further highlight that the catalytic system works well across a range of reactivity profiles in both the glycal moiety and nucleophile acceptor.

Table 2. Reaction of Glycals 9a–d and 11a–c with Model Glycoside Acceptors 2a, 2e, or 2g

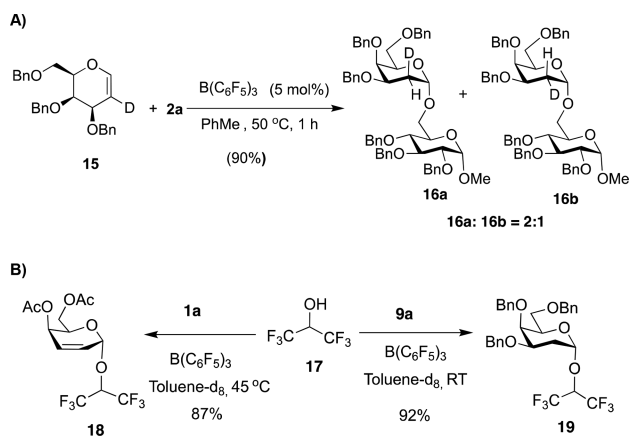


entry		R <sup>1</sup>	R <sup>2</sup>	R <sup>3</sup>	product	time (h)	yield (%) <sup>a</sup>	$\alpha/\beta$ <sup>b</sup>
1	9a	Bn	Bn	Bn	10a	1	88 <sup>d,e</sup>	21:1
2	9a	Bn	Bn	Bn	10c	1	94	26:1
3	9a	Bn	Bn	Bn	10e	2	84	30:1
4	9a	Bn	Bn	Bn	10g	7	62	25:1
5	9a	Bn	Bn	Bn	10h	7	68	20:1
6	9b	TBS	TBS	TBS	11b	1	92	30:1
7	9c	MOM	MOM	MOM	11c	1	71	20:1
8	9d	Si( <i>i</i> -Pr) <sub>2</sub>	MOM	MOM	11d	1	82	20:1
9	11a	Bn	Bn	Bn	12a <sup>c</sup>	17	54	30:1
10	11b	O[Si( <i>i</i> -Pr) <sub>2</sub> ] <sub>2</sub>		Bn	12b	17	78	30:1
11	11c	O[Si( <i>i</i> -Pr) <sub>2</sub> ] <sub>2</sub>		TIPS	12c	17	75	30:1
12	11c	O[Si( <i>i</i> -Pr) <sub>2</sub> ] <sub>2</sub>		TIPS	13	17	86	30:1
13	11c	O[Si( <i>i</i> -Pr) <sub>2</sub> ] <sub>2</sub>		TIPS	14	17	64	20:1

<sup>a</sup>Isolated yield. <sup>b</sup>Determined by <sup>1</sup>H NMR. <sup>c</sup>10% Ferrier product **12a'** also isolated. <sup>d</sup>Reaction did not proceed in the absence of catalyst. <sup>e</sup>Reaction with  $\text{BF}_3\cdot\text{OEt}_2$  afforded **10a** (<35%) and a mixture of products and starting material.

To probe the mechanism of this versatile reaction, deuterated perbenzylated galactal **15** was reacted with **2a** to yield  $\alpha$ -glycoside **16a** and **16b** (90% yield) as a 2:1 mixture of cis/trans products, with a preference for syn addition of both H and O-nucleophile across the double bond (Scheme 4A).

**Scheme 4. Model Glycosylations of 2a or 17 with Glycal Donors 15, 1a, or 9a**

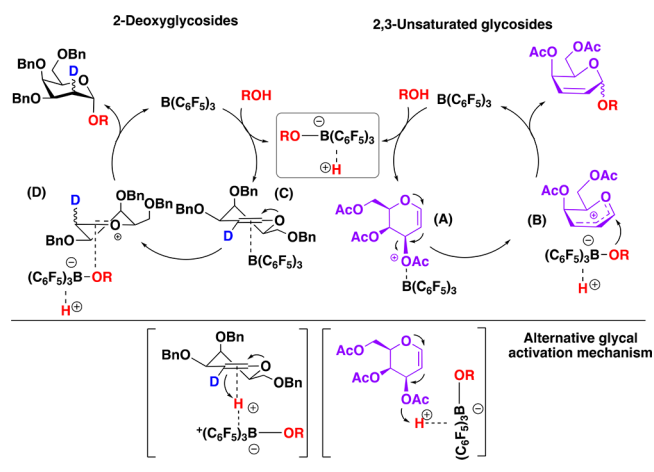


Addition of  $K_2CO_3$  to the reaction between either **1a** or **9a** with **2a** inhibited the reaction, which supports an acid catalyzed process. Monitoring the reaction between **1a** or **9a** and hexafluoroisopropanol **17** (Scheme 4B) by  $^1H$  NMR over 60 min at 45 °C or 90 min at RT, respectively, only showed anomeric signals corresponding to the starting material and product, without any observable changes in the anomeric ratio of the product throughout the time scales of the reaction (SI Figures S1 and S3).<sup>13</sup> Moreover, subjecting a 4:1  $\alpha/\beta$ -anomeric mixture of **10a** to the reaction conditions in the presence of acceptor **2a** gave no change in the anomeric ratio (see SI for details). These results suggest the reaction proceeds via short-lived intermediates and that the high  $\alpha$ -selectivity is not likely the result of anomerization.  $^{19}F$ -NMR of the reactions (SI Figures S2 and S4) showed the appearance of fluorinated signals assigned to products **18** and **19**, respectively, and also shifts associated with the formation of other BCF species, suggesting the presence of BCF-adducts. Moreover,  $^1H$  NMR spectroscopy studies in Toluene- $d_8$  of a 1:1 mixture of BCF with galactal donor **1a** or **9a**, also showed H-shifts associated with the enol ether alkene protons, in each case (SI Figures S5 and S7).  $^{19}F$ -NMR of the same mixtures showed additional signals associated with several distinct BCF-species (SI Figures S6 and S8), suggesting activation of the glycal enol ethers by BCF can take place and formation of adducts. Interestingly,  $^1H$  NMR equimolar mixtures of  $B(C_6F_5)_3$  and OH nucleophile **2a** at room temperature showed proton shifts associated with **2a** (Figure S9). This effect was more evident in the  $^{19}F$ -NMR spectra of the same mixtures (SI Figure S10) which showed the shift of the fluorine signals from the catalysts and appearance of different fluorinated species, further supporting the formation of an adduct between the catalyst and the OH nucleophile. This is in agreement to previous reports of glycosyl acceptor activation with boron-based catalysts such as BCF and  $PhBF_2$  in the acid–base activation of trichloroacetimidate glycosyl donors.<sup>8b,12</sup>

As our preliminary findings suggest, BCF could act as a Lewis acid to promote the effective allylic rearrangement<sup>13</sup> (A)

of deactivated glycols such as **1a** to form transient oxocarbenium ion (B) that can undergo nucleophilic substitution by the BCF-activated nucleophile adduct (H—BCF—OR) in a stereoselective manner to give 2,3-unsaturated glycosides. In the presence of more reactive glycols, which lack a leaving group at C-3 (e.g., **9a**), enol ether direct activation to form oxocarbenium ion (D) might take place, which after nucleophilic substitution by the BCF-activated nucleophile and concomitant protonolysis leads to deoxyglycoside products. In both instances, there is a clear preference for an  $\alpha$ -face nucleophilic approach, likely due to sterics and a favorable anomeric effect<sup>14</sup> (Scheme 5, top). However, in the presence

**Scheme 5. Proposed Mechanism**



of Lewis basic oxygen atoms, BCF coordination to the pyran oxygen in the donor is also possible. Therefore, an acid–base catalyzed mechanism whereby the boron ate adduct promotes both oxocarbenium ion formation and nucleophile activation can not be discarded and it is likely to occur in parallel (Scheme 5, bottom). Further investigations are ongoing to better understand the mechanism of this reaction.

## CONCLUSIONS

In summary, we have described the unprecedented BCF-catalyzed substrate-controlled stereoselective synthesis of  $\alpha$ -deoxyglycosides directly from glycols. We show that 2,3-unsaturated  $\alpha$ -O-glycoside products are obtained with deactivated glycols at 75 °C, while 2- $\alpha$ -deoxyglycosides are formed with activated glycols lacking a leaving group at C-3 at slightly lower temperatures. This metal-free and versatile reaction is applicable to a range of glycal donors and nucleophile acceptors, and is tolerant of most common protecting groups. The reaction proceeds with good to excellent yields and high selectivity for the  $\alpha$ -anomer. We exemplify the robustness and utility of the approach in the stereoselective synthesis of a series of oligosaccharides, glycosyl-amino acids, and other glyco-conjugates including rare glycosides analogues  $\alpha$ -L-Rhodinose  $\alpha$ -L-cinerulose and  $\alpha$ -L-aculose. Work from our lab is currently underway to exploit this chemistry for the stereoselective synthesis of other important glycosides.

## EXPERIMENTAL SECTION

**General Experimental Procedures.** Chemicals were purchased and used without further purification. Glycal donors **1a–1e**, **9a**, and **11a** were purchased from Carbosynth and OH acceptors **2b**, **2e**, **2f**,



2h, 2i, 2j, and 2k were obtained from Sigma-Aldrich. Galactal donors 9b and 9c and glycosyl acceptors 2a, 2c, and 2d were prepared following literature procedures,<sup>4d</sup> while glucal 11b and 11c and glycosyl acceptor 2g were synthesized by Balmond et al. reported methods.<sup>4c</sup> Dry solvents were obtained by distillation using standard procedures or by passage through a column of anhydrous alumina using equipment from Anhydrous Engineering (University of Bristol) based on the Grubbs' design. Reactions requiring anhydrous conditions were performed under nitrogen; glassware and needles were either flame-dried immediately prior to use or placed in an oven (150 °C) for at least 2 h and allowed to cool either in a desiccator or under reduced pressure; liquid reagents, solutions, or solvents were added via syringe through rubber septa; solid reagents were added via Schlenk type adapters. Teflon rings were used between the joints of the condensers and round-bottom flasks. Reactions were monitored by TLC on Kieselgel 60 F254 (Merck). Detection was by examination under UV light (254 nm) and by charring with 10% sulfuric acid in ethanol. Flash column chromatography was performed using silica gel [Merck, 230–400 mesh (40–63 μm)]. Extracts were concentrated in vacuo using both a Büchi rotary evaporator (bath temperatures up to 40 °C) at a pressure of either 15 mmHg (diaphragm pump) or 0.1 mmHg (oil pump), as appropriate, and a high vacuum line at room temperature. <sup>1</sup>H NMR and <sup>13</sup>C NMR spectra were measured in the solvent stated at 400 or 500 MHz. Chemical shifts are quoted in parts per million from residual solvent peak (CDCl<sub>3</sub>: <sup>1</sup>H–7.26 ppm and 13C–77.16 ppm) and coupling constants (J) given in Hertz. Multiplicities are abbreviated as b (broad), s (singlet), d (doublet), t (triplet), q (quartet), m (multiplet), or combinations thereof. Mass spectra were determined by the University of Bristol mass spectrometry service by electrospray ionization (SI) modes. The units of the specific rotation, (deg·mL)/(g·dm), are implicit and are not included with the reported value. Concentration *c* is given in g/100 mL.

**General Glycosylation Procedure.** Glycal donor (1.0 equiv), OH nucleophile acceptor (0.75 equiv), and B(C<sub>6</sub>F<sub>5</sub>)<sub>3</sub> (5 mol %) were weighed into an oven-dried microwave vial, sealed and placed under vacuum for 1 h. Then the vial was filled with N<sub>2</sub> followed by the addition of ~1.0 mL of anhydrous toluene. The solutions were stirred and heated at 75 °C for Ferrier glycosylation and 50 °C for 2-deoxy glycosylation until the reaction was determined to be complete by either TLC or NMR analysis of the crude material (times are given in Tables S1 and S2 and Tables 1 and 2 of the main manuscript). The reaction mixture was concentrated in vacuo and the dried residue was purified by silica gel column chromatography.

**Methyl-6-O-(4,6-di-O-acetyl-2,3-dideoxy-α-D-threo-hex-2-enopyranosyl)-2,3,4-tri-O-benzyl-α-D-glucopyranoside (3a).** Following the general glycosylation procedure, donor 1a (50 mg, 0.18 mmol), B(C<sub>6</sub>F<sub>5</sub>)<sub>3</sub> (5 mg 0.01 mmol), and acceptor 2a (64 mg, 0.14 mmol) to afford following purification by silica gel column chromatography (Hexane:EtOAc, 6:1 to 3:1) 3a as a colorless oil (82 mg, 88%). Spectroscopic data in agreement with literature.<sup>15</sup>

**Benzyl-4,6-di-O-acetyl-2,3-dideoxy-α-D-threo-hex-2-enopyranoside (3b).** Following the general glycosylation procedure, donor 1a (50 mg, 0.18 mmol), B(C<sub>6</sub>F<sub>5</sub>)<sub>3</sub> (5 mg 0.01 mmol) and acceptor 2b (15 mg, 0.14 mmol) to afford following purification by column chromatography (Hexane:EtOAc, 12:1 to 8:1) 3b as a colorless oil (37 mg, 84%). Spectroscopic data in agreement with literature.<sup>15</sup>

**Methyl-6-O-(4,6-di-O-acetyl-2,3-dideoxy-α-D-threo-hex-2-enopyranosyl)-2,3,4-tri-O-benzyl-α-D-glucopyranoside (3c).** Following the general glycosylation procedure, donor 1a (50 mg, 0.18 mmol), B(C<sub>6</sub>F<sub>5</sub>)<sub>3</sub> (5 mg 0.01 mmol) and acceptor 2c (70 mg, 0.14 mmol) to afford following purification by silica gel column chromatography (Hexane: EtOAc, 5:1 to 2:1) 3c as a colorless oil (72 mg, 72%). <sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>) δ 7.98 (dd, *J* = 18.7, 7.7 Hz, SH, Ar–H), 7.87 (d, *J* = 7.8 Hz, 2H, Ar–H), 7.55–7.52 (m, 2H, Ar–H), 7.39 (t, *J* = 7.7 Hz, 4H, Ar–H), 7.32–7.28 (m, 2H, Ar–H), 6.20–6.09 (m, 2H, H-2, H-2'), 6.04 (dd, *J* = 10.0, 3.0 Hz, 1H, H-3'), 5.72 (t, *J* = 9.8 Hz, 1H, H-4), 5.30–5.26 (m, 2H, H-1, H-3), 5.13 (d, *J* = 2.9 Hz, 1H, H-1'), 5.04 (dd, *J* = 5.5, 2.5 Hz, 1H, H-4'), 4.39 (ddd, *J* = 7.8, 5.4, 2.4 Hz, 1H, H-5'), 4.26 (dt, *J* = 10.4, 3.7 Hz, 1H, H-5), 4.16–3.99 (m,

3H, H-6a, H-6a', H-6b'), 3.74 (dd, *J* = 11.1, 3.0 Hz, 1H, H-6b), 3.50 (s, 3H, OCH<sub>3</sub>), 2.07 (s, 3H, COCH<sub>3</sub>), 1.86 (s, 3H, COCH<sub>3</sub>). <sup>13</sup>C NMR (126 MHz, CDCl<sub>3</sub>) δ 170.5 (COCH<sub>3</sub>), 170.3 (COCH<sub>3</sub>), 165.8 (COPh), 165.8 (COPh), 165.2 (COPh), 133.5 (Ar–C), 133.4 (Ar–C), 133.1 (Ar–C), 130.1 (Ar–C), 129.9 (Ar–C), 129.8 (Ar–C), 129.2 (C-2'), 129.1 (Ar–C), 128.5 (Ar–C), 128.4 (Ar–C), 128.3 (Ar–C), 125.3, (C-3') 97.1 (C-1), 93.9 (C-1'), 72.1 (C-3), 70.6 (C-2), 69.4 (C-4), 68.3 (C-5), 66.6 (C-5'), 66.1 (C-6), 62.6 (2C-4', 6'), 55.7 (OCH<sub>3</sub>), 20.8 (COCH<sub>3</sub>), 20.5 (COCH<sub>3</sub>); ESI-HRMS for C<sub>38</sub>H<sub>38</sub>O<sub>14</sub>Na<sup>+</sup> (MNa<sup>+</sup>) calculated: 741.2159; found: 741.2161

**Thiophenyl-6-O-(4,6-di-O-acetyl-2,3-dideoxy-α-D-threo-hex-2-enopyranosyl)-2,3,4-tri-O-benzyl-α-D-glucopyranoside (3d).** Following the general glycosylation procedure, donor 1a (50 mg, 0.18 mmol), B(C<sub>6</sub>F<sub>5</sub>)<sub>3</sub> (5 mg 0.01 mmol) and acceptor 2d (80 mg, 0.14 mmol) to afford following purification by column chromatography (Hexane:EtOAc, 5:1 to 1:1) 3d as a colorless oil (85 mg, 72%). <sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>) δ 8.00–7.96 (m, 2H, Ar–H), 7.96–7.91 (m, 2H, Ar–H), 7.82–7.78 (m, 2H, Ar–H), 7.57–7.50 (m, 4H, Ar–H), 7.41 (m, 5H, Ar–H), 7.35–7.25 (m, 5H, Ar–H), 6.10 (ddd, *J* = 10.1, 5.5, 1.0 Hz, 1H, H-2'), 5.96 (dd, *J* = 10.0, 3.1 Hz, 1H, H-3'), 5.91 (t, *J* = 9.5 Hz, 1H, H-3), 5.63 (t, *J* = 9.7 Hz, 1H, H-4), 5.49 (t, *J* = 9.7 Hz, 1H, H-2), 5.12 (d, *J* = 3.1 Hz, 1H, H-1'), 5.08 (d, *J* = 10.0 Hz, 1H, H-1), 4.97 (dd, *J* = 5.5, 2.5 Hz, 1H, H-4'), 4.32 (ddd, *J* = 7.8, 5.4, 2.5 Hz, 1H, H-5'), 4.16–4.08 (m, 2H, H-6a', H-6b'), 4.07–4.00 (m, 2H, H-5, H-6a), 3.83–3.77 (m, 1H, H-6b), 2.08 (s, 3H, COCH<sub>3</sub>), 1.93 (s, 3H, COCH<sub>3</sub>). <sup>13</sup>C NMR (126 MHz, CDCl<sub>3</sub>) δ 170.5 (COCH<sub>3</sub>), 170.3 (COCH<sub>3</sub>), 165.8 (COPh), 165.1 (COPh), 165.0 (COPh), 133.5 (Ar–C), 133.2 (Ar–C), 132.6 (Ar–C), 132.1 (Ar–C), 130.1 (C-3'), 129.9 (Ar–C), 129.8 (Ar–C), 129.7 (Ar–C), 129.2 (Ar–C), 128.9 (Ar–C), 128.9 (Ar–C), 128.8 (Ar–C), 128.5 (Ar–C), 128.4 (Ar–C), 128.3 (Ar–C), 128.2 (Ar–C), 125.3 (C-2'), 93.8 (C-1'), 85.9 (C-1), 77.2 (C-5), 74.3 (C-3), 70.5 (C-2), 69.5 (C-4) 66.7 (C-5') 66.5 (C-6), 62.7 (C-6'), 62.6 (C-4'), 20.8 (COCH<sub>3</sub>), 20.6 (COCH<sub>3</sub>). ESI-HRMS for C<sub>43</sub>H<sub>40</sub>O<sub>13</sub>SNa<sup>+</sup> (MNa<sup>+</sup>) calculated: 819.2087; found: 819.2103.

**6-O-(4,6-di-O-acetyl-2,3-dideoxy-α-D-threo-hex-2-enopyranosyl)-1,2,4,5-di-O-isopropylidene-α-D-galactopyranoside (3e).** Following the general glycosylation procedure, donor 1a (50 mg, 0.18 mmol), B(C<sub>6</sub>F<sub>5</sub>)<sub>3</sub> (5 mg 0.01 mmol), and acceptor 2e (36 mg, 0.14 mmol) to afford following purification by silica gel column chromatography (Hexane:EtOAc, 7:1 to 4:1) 3e as a colorless oil (49 mg, 75%). Spectroscopic data in agreement with literature.<sup>15</sup>

**2'-Propynyl-4,6-di-O-acetyl-2,3-dideoxy-α/β-D-threo-hex-2-enopyranoside (3f).** Following the general glycosylation procedure, donor 1a (50 mg, 0.18 mmol), B(C<sub>6</sub>F<sub>5</sub>)<sub>3</sub> (5 mg 0.01 mmol) and acceptor 2f (7.73 mg, 0.14 mmol) to afford following purification by silica gel column chromatography (Hexane:EtOAc, 8:1 to 4:1) 3f as a colorless oil (32 mg, 86%). Spectroscopic data in agreement with literature.<sup>15</sup>

**Methyl 3-O-(4,6-di-O-acetyl-2,3-dideoxy-α-D-threo-hex-2-enopyranosyl)-4,6-O-benzylidene-2-O-benzyl-α-D-glucopyranoside (3g).** Following the general glycosylation procedure, donor 1a (50 mg, 0.18 mmol), B(C<sub>6</sub>F<sub>5</sub>)<sub>3</sub> (5 mg 0.01 mmol) and acceptor 2g (51 mg, 0.14 mmol) to afford following purification by silica gel column chromatography (Hexane:EtOAc, 5:1 to 3:1) 3g as a colorless oil (55 mg, 68%). <sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>) δ 7.42 (dd, *J* = 6.7, 2.8 Hz, 2H, Ar–H), 7.37–7.32 (m, 7H, Ar–H), 7.32–7.29 (m, 1H, Ar–H), 6.07 (dd, *J* = 3.2, 1.8 Hz, 2H, H-2', H-3'), 5.54 (d, *J* = 2.0 Hz, 1H, H-1), 5.51 (s, 1H, H-PhCH), 5.00 (dd, *J* = 4.3, 2.7 Hz, 1H, H-4'), 4.81–4.69 (m, 1H, PhCHH), 4.57 (d, *J* = 12.2 Hz, 1H, PhCHH), 4.54 (d, *J* = 3.7 Hz, 1H, H-1), 4.51–4.46 (m, 1H, H-5'), 4.41 (t, *J* = 9.4 Hz, 1H, H-3), 4.25 (ddt, *J* = 13.2, 7.3, 3.3 Hz, 2H, H-6a', H-6a), 4.05 (dd, *J* = 11.2, 7.2 Hz, 1H, H-6b'), 3.82 (td, *J* = 9.9, 4.8 Hz, 1H, H-5), 3.69 (t, *J* = 10.3 Hz, 1H, H-6b), 3.56 (t, *J* = 9.5 Hz, 1H, H-4), 3.46 (ddd, *J* = 13.1, 9.4, 3.6 Hz, 1H, H-2), 3.35 (s, 3H, OCH<sub>3</sub>), 2.06 (d, *J* = 6.1 Hz, 6H, 2 COCH<sub>3</sub>). <sup>13</sup>C NMR (126 MHz, CDCl<sub>3</sub>) δ 170.8 (COCH<sub>3</sub>), 170.4 (COCH<sub>3</sub>), 138.2 (Ar–C), 137.2 (Ar–C), 130.8 (C-3'), 129.0 (Ar–C), 128.4 (Ar–C), 128.3 (Ar–C), 128.1 (Ar–C), 127.9 (Ar–C), 126.0 (Ar–C), 124.9 (C-2'), 101.4 (C-PhCH), 99.1 (C-1), 93.6 (C-1'), 82.8 (C-4), 78.1 (C-2), 73.4 (C-PhCH<sub>2</sub>), 73.1 (C-3), 69.1

(C-6), 66.4 (C-5'), 62.7 (C-4'), 62.2 (C-6'), 62.0 (C-5), 55.2 (OCH<sub>3</sub>), 20.8 (COCH<sub>3</sub>), 20.70 (COCH<sub>3</sub>). ESI-HRMS for C<sub>31</sub>H<sub>36</sub>O<sub>11</sub>Na<sup>+</sup> (MNa<sup>+</sup>) calculated: 607.2155; found: 607.2155.

**3-O-(4,6-di-O-acetyl-2,3-dideoxy- $\alpha$ -D-threo-hex-2-enopyranosyl)-1,2:5,6-di-O-isopropylidene- $\alpha$ -D-glucofuranose (3h).** Following the general glycosylation procedure, donor **1a** (50 mg, 0.18 mmol), B(C<sub>6</sub>F<sub>5</sub>)<sub>3</sub> (5 mg 0.01 mmol), and acceptor **2h** (36 mg, 0.14 mmol) to afford following purification by silica gel column chromatography (Hexane:EtOAc, 7:1 to 3:1) **3h** as a colorless oil (44 mg, 67%). <sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>)  $\delta$  6.16 (ddd, *J* = 10.0, 5.5, 1.1 Hz, 1H, H-2'), 6.09–5.99 (m, 1H, H-3'), 5.90 (d, *J* = 3.6 Hz, 1H, H-1), 5.39–5.34 (m, 1H, H-1'), 5.04 (dt, *J* = 5.5, 2.8 Hz, 1H, H-4'), 4.63 (d, *J* = 3.6 Hz, 1H, H-2), 4.41–4.28 (m, 3H, H-5', H-3, H-6a'), 4.25–4.18 (m, 2H, H-4, H-6b'), 4.16–4.08 (m, 2H, H-5, H-6a), 3.99 (dd, *J* = 8.6, 5.1 Hz, 1H, H-6b), 2.13 (s, 3H), 2.10 (s, 3H), 1.52 (s, 3H), 1.42 (s, 3H), 1.33 (d, *J* = 4.1 Hz, 6H). <sup>13</sup>C NMR (126 MHz, CDCl<sub>3</sub>)  $\delta$  170.8 (COCH<sub>3</sub>), 170.3 (COCH<sub>3</sub>), 130.0 (C-3'), 125.4 (C-2'), 112.0 (4°C), 109.17(4°C), 105.4 (C-1), 94.9 (C-1'), 84.3 (C-2), 81.3 (C-4), 80.77 (C-3), 72.7 (C-5), 67.8 (C-6), 67.2 (C-5'), 63.1 (C-6'), 62.8 (C-4'), 27.0 (CCH<sub>3</sub>), 26.9 (CCH<sub>3</sub>), 26.5 (CCH<sub>3</sub>), 25.4 (CCH<sub>3</sub>), 20.8 (COCH<sub>3</sub>), 20.7 (COCH<sub>3</sub>). ESI-HRMS for C<sub>22</sub>H<sub>32</sub>O<sub>11</sub>Na<sup>+</sup> (MNa<sup>+</sup>) calculated: 495.1842; found: 495.1832.

**4-Methoxyphenyl-4,6-di-O-acetyl-2,3-dideoxy- $\alpha$ -D-threo-hex-2-enopyranoside (3i).** Following the general glycosylation procedure, donor **1a** (50 mg, 0.18 mmol), B(C<sub>6</sub>F<sub>5</sub>)<sub>3</sub> (5 mg 0.01 mmol) and acceptor **2i** (17 mg, 0.14 mmol) to afford following purification by silica gel column chromatography (Hexane:EtOAc, 9:1 to 3:1) **3i** as a colorless oil (36 mg, 82%) <sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>)  $\delta$  7.10–7.05 (m, 4H, Ar–H), 6.88–6.83 (m, 2H, Ar–H), 6.27 (ddd, *J* = 9.9, 5.4, 1.0 Hz, 1H, H-2), 6.21 (dd, *J* = 10.0, 3.0 Hz, 1H, H-3), 5.64 (dt, *J* = 3.0, 0.6 Hz, 1H, H-1), 5.13 (dd, *J* = 5.4, 2.5 Hz, 1H, H-4), 4.54 (ddd, *J* = 7.7, 5.3, 2.5 Hz, 1H, H-5), 4.29–4.23 (m, 2H, H-6a, H-6b), 3.80 (s, 3H, OCH<sub>3</sub>), 2.12 (s, COCH<sub>3</sub>), 1.98 (s, 3H, COCH<sub>3</sub>). <sup>13</sup>C NMR (126 MHz, CDCl<sub>3</sub>)  $\delta$  170.6 (COCH<sub>3</sub>), 170.4 (COCH<sub>3</sub>), 155.2 (Ar–C), 151.0 (Ar–C), 129.9 (C-3), 125.9 (C-2), 118.8 (Ar–C), 114.5 (Ar–C), 93.7 (C-1), 67.5 (C-5), 62.6 (C-4), 62.5 (C-6), 55.7 (OCH<sub>3</sub>), 20.8 (COCH<sub>3</sub>), 20.7 (COCH<sub>3</sub>). ESI-HRMS for C<sub>17</sub>H<sub>20</sub>O<sub>7</sub>Na<sup>+</sup> (MNa<sup>+</sup>) calculated: 359.1107; found: 359.1117.

**Cholesteryl-4,6-diacetyl-2,3-dideoxy- $\alpha$ -D-threo-2-enopyranoside (3j).** Following the general glycosylation procedure, donor **1a** (50 mg, 0.18 mmol), B(C<sub>6</sub>F<sub>5</sub>)<sub>3</sub> (5 mg 0.01 mmol) and acceptor **2j** (53 mg, 0.14 mmol) to afford following purification by silica gel column chromatography (Hexane:EtOAc, 4:1 to 2:1) **3j** as a colorless oil (65 mg, 79%) <sup>1</sup>H NMR (500 MHz, Chloroform-*d*)  $\delta$  6.12 (dd, *J* = 10.0, 5.3 Hz, 1H, H-3), 6.03 (dd, *J* = 10.0, 3.0 Hz, 1H, H-2), 5.38 (dd, *J* = 4.9, 2.4 Hz, 1H, C = CH), 5.23 (d, *J* = 3.1 Hz, 1H, H-1), 5.04 (dd, *J* = 5.4, 2.5 Hz, 1H, H-4), 4.43 (ddd, *J* = 7.7, 5.6, 2.5 Hz, 1H, H-5), 4.27–4.20 (m, 2H, H-6a, H-6b), 3.59 (m, 1H), 2.44 (ddd, *J* = 13.4, 5.2, 2.1 Hz, 1H), 2.39–2.24 (m, 1H), 2.09 (d, *J* = 5.2 Hz, 6H, 2 COCH<sub>3</sub>), 2.05–1.95 (m, 3H), 1.93–1.79 (m, 3H), 1.63–1.05 (m, 16H), 1.02 (s, 5H), 0.93 (d, *J* = 6.5 Hz, 4H), 0.88 (dd, *J* = 6.6, 2.2 Hz, 8H), 0.69 (s, 3H). <sup>13</sup>C NMR (126 MHz, CDCl<sub>3</sub>)  $\delta$  170.7 (COCH<sub>3</sub>), 170.4 (COCH<sub>3</sub>), 140.8 (C = CH), 131.2 (C-2), 125.0 (C-3), 121.8 (C = CH), 92.4 (C-1), 78.0, 66.7 (C-5), 63.0 (2C-6, 4), 56.8 (CH), 56.2 (CH), 50.2 (CH), 42.3 (4 °C), 40.4 (CH<sub>2</sub>), 39.8 (CH<sub>2</sub>), 39.5 (CH<sub>2</sub>), 37.2 (4 °C), 36.7 (CH<sub>2</sub>), 36.2 (CH), 35.8, 31.9, 31.9, 28.2, 28.0, 24.3, 23.8, 22.8, 22.6, 21.1 (COCH<sub>3</sub>), 20.8 (COCH<sub>3</sub>), 19.3 (CH<sub>3</sub>), 18.7 (CH<sub>3</sub>), 11.9 (CH<sub>3</sub>). ESI-HRMS for C<sub>37</sub>H<sub>58</sub>O<sub>6</sub>Na<sup>+</sup> (MNa<sup>+</sup>) calculated: 621.4131; found: 621.4126.

**N-Succinimido-4,6-diacetyl-2,3-dideoxy- $\alpha$ -D-threo-2-enopyranoside (3k).** Following the general glycosylation procedure, donor **1a** (50 mg, 0.18 mmol), B(C<sub>6</sub>F<sub>5</sub>)<sub>3</sub> (5 mg 0.01 mmol) and acceptor **2k** (16 mg, 0.14 mmol) to afford following purification by silica gel column chromatography (Hexane:EtOAc, 3:1 to 1:1) **3k** as a colorless oil (41 mg, 91%). <sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>)  $\delta$  6.36 (ddd, *J* = 10.0, 5.6, 1.2 Hz, 1H, H-3), 6.20 (ddd, *J* = 10.0, 3.1, 0.6 Hz, 1H, H-2), 5.64 (ddd, *J* = 3.2, 1.2, 0.6 Hz, 1H, H-1), 5.15 (ddd, *J* = 5.6, 2.7, 0.6 Hz, 1H, H-4), 4.81 (td, *J* = 6.3, 2.7 Hz, 1H, H-5), 4.33 (dd, *J* = 11.3, 6.2 Hz, 1H, H-6a), 4.07 (dd, *J* = 11.3, 6.5 Hz, 1H, H-6b), 2.75 (s, 4H, 2CH<sub>2</sub>), 2.08 (s, 3H, COCH<sub>3</sub>), 2.07 (s, 3H, COCH<sub>3</sub>). <sup>13</sup>C NMR (126

MHz, CDCl<sub>3</sub>)  $\delta$  171.0 (2 NHS CO), 170.6 (COCH<sub>3</sub>), 170.1 (COCH<sub>3</sub>), 128.8 (C-3), 125.9 (C-2), 97.5 (C-1), 68.3 (C-5), 61.9 (C-4), 61.8 (C-6), 25.5 (2 CH<sub>2</sub>), 20.8 (COCH<sub>3</sub>), 20.7 (COCH<sub>3</sub>). ESI-HRMS for C<sub>14</sub>H<sub>17</sub>NO<sub>8</sub>Na<sup>+</sup> (MNa<sup>+</sup>) calculated: 350.0852; found: 350.0862.

**6-O-(4,6-di-O-acetyl-2,3-dideoxy- $\alpha$ -D-erythro-hex-2-enopyranosyl)-1,2:4,5-di-O-isopropylidene- $\alpha$ -D-galactopyranoside (4b).** Following the general glycosylation procedure, donor **1b** (50 mg, 0.18 mmol), B(C<sub>6</sub>F<sub>5</sub>)<sub>3</sub> (5 mg 0.01 mmol) and acceptor **2e** (36 mg, 0.14 mmol) to afford following purification by column chromatography (Hexane:EtOAc, 8:1 to 4:1) **4b** as a yellow oil (56 mg, 86%). Spectroscopic data in agreement with literature.<sup>9b</sup>

**4-O-(acetyl)-2,3,6-trideoxy- $\alpha$ -L-hex-2-enopyranosyl-(1  $\rightarrow$  6)-1,2:3,4-di-O-isopropylidene- $\alpha$ -D-galactopyranoside (4c).** Following the general glycosylation procedure, donor **1c** (50 mg, 0.12 mmol), B(C<sub>6</sub>F<sub>5</sub>)<sub>3</sub> (6 mg 0.011 mmol), and acceptor **2e** (46 mg, 0.10 mmol) to afford following purification by silica gel column chromatography (Hexane:EtOAc, 8:1 to 5:1) **4c** as a yellow oil (52 mg, 71%). <sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>)  $\delta$  6.14–5.95 (m, 2H, H-2', H-3'), 5.53 (d, *J* = 5.0 Hz, 1H, H-1), 5.10 (d, *J* = 1.8 Hz, 1H, H-1'), 4.92 (dd, *J* = 4.4, 2.5 Hz, 1H, H-4'), 4.60 (dd, *J* = 7.9, 2.4 Hz, 1H, H-3), 4.31 (dd, *J* = 5.1, 2.4 Hz, 1H, H-2), 4.28–4.21 (m, 2H, H-5', H-4), 3.99–3.91 (m, 2H, H-5, H-6a), 3.72–3.64 (m, 1H, H-6b), 2.10 (s, 3H, COCH<sub>3</sub>), 1.53 (s, 3H, CCH<sub>3</sub>), 1.45 (s, 3H, CCH<sub>3</sub>), 1.33 (s, 5H, CCH<sub>3</sub>), 1.22 (d, *J* = 6.6 Hz, 3H, CCH<sub>3</sub>). <sup>13</sup>C NMR (126 MHz, CDCl<sub>3</sub>)  $\delta$  170.7 (COCH<sub>3</sub>), 130.5 (C-3'), 125.8 (C-2'), 109.2 (CCH<sub>3</sub>), 108.5 (CCH<sub>3</sub>), 96.3 (C-1), 94.0 (C-1'), 71.1 (C-5'), 70.6 (C-3), 70.5 (C-2), 67.0 (C-5), 66.1 (C-6), 65.1 (C-4'), 64.6 (C-4), 26.1 (CCH<sub>3</sub>), 26.0 (CCH<sub>3</sub>), 24.9 (CCH<sub>3</sub>), 24.5 (CCH<sub>3</sub>), 20.9 (COCH<sub>3</sub>), 15.9 (CCH<sub>3</sub>). ESI-HRMS for C<sub>20</sub>H<sub>30</sub>O<sub>9</sub>Na<sup>+</sup> (MNa<sup>+</sup>) calculated: 437.1788; found: 437.1788.

**6-O-(R-2,3-dihydro-2H-pyran-4-yl acetate)-1,2:4,5-di-O-isopropylidene- $\alpha$ -D-galactopyranoside (4d).** Following the general glycosylation procedure, donor **1d** (50 mg, 0.18 mmol), B(C<sub>6</sub>F<sub>5</sub>)<sub>3</sub> (6 mg 0.011 mmol) and acceptor **2e** (36 mg, 0.14 mmol) to afford following purification by silica gel column chromatography (Hexane:EtOAc, 10:1 to 6:1) **4d** as a colorless oil (56 mg, 75%). <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  6.09–5.99 (m, 2H, H-2', H-3'), 5.54 (d, *J* = 5.1 Hz, 1H, H-1), 5.09 (d, *J* = 2.3 Hz, 1H, H-1'), 4.95–4.91 (m, 1H, H-4'), 4.59 (dd, *J* = 7.9, 2.4 Hz, 1H, H-3), 4.30 (dd, *J* = 5.1, 2.4 Hz, 1H, H-2), 4.23–4.15 (m, 2H, H-4, H-6a), 3.99 (ddd, *J* = 7.1, 4.8, 1.9 Hz, 1H, H-5), 3.90–3.72 (m, 3H, H-6b, H-5a', H-5b'), 2.08 (s, 3H, COCH<sub>3</sub>), 1.52 (s, 3H, CCH<sub>3</sub>), 1.43 (s, 3H, CCH<sub>3</sub>), 1.32 (s, 6H, 2 CCH<sub>3</sub>). <sup>13</sup>C NMR (101 MHz, CDCl<sub>3</sub>)  $\delta$  170.6 (COCH<sub>3</sub>), 130.9 (C-3'), 124.7 (C-2'), 109.3 (CCH<sub>3</sub>), 108.5 (CCH<sub>3</sub>), 96.3 (C-1), 93.5 (C-1'), 71.2 (C-4'), 70.7 (C-3), 70.4 (C-2), 67.4 (C-5), 67.3 (C-5'), 63.4 (C-4), 61.4 (C-6), 26.0 (CCH<sub>3</sub>), 26.0 (CCH<sub>3</sub>), 24.9 (CCH<sub>3</sub>), 24.5 (CCH<sub>3</sub>), 21.1 (COCH<sub>3</sub>). ESI-HRMS for C<sub>19</sub>H<sub>28</sub>O<sub>9</sub>Na<sup>+</sup> (MNa<sup>+</sup>) calculated: 423.1631; found: 423.1623.

**4-O-(acetyl)-2,3,6-trideoxy- $\alpha$ -L-hex-2-enopyranosyl-(1  $\rightarrow$  6)-1,2:3,4-di-O-isopropylidene- $\alpha$ -D-galactopyranoside (4e).** Following the general glycosylation procedure, donor **1e** (50 mg, 0.12 mmol), B(C<sub>6</sub>F<sub>5</sub>)<sub>3</sub> (6 mg 0.011 mmol) and acceptor **2e** (46 mg, 0.10 mmol) to afford following purification by silica gel column chromatography (Hexane:EtOAc, 15:1 to 8:1) **4e** as a yellow oil (48 mg, 65%). Spectroscopic data in agreement with literature.<sup>9b</sup>

**3-N-Boc-propyl acetyl-2,3,6-trideoxy- $\alpha$ -L-hex-2-enopyranosyl (4f).** Following the general glycosylation procedure, donor **1e** (50 mg, 0.12 mmol), B(C<sub>6</sub>F<sub>5</sub>)<sub>3</sub> (6 mg 0.011 mmol), and acceptor **2l** (31 mg, 0.10 mmol) to afford following purification by silica gel column chromatography (Hexane:EtOAc, 5:1 to 3:1) **4f** as a yellow oil (50 mg, 85%). <sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>)  $\delta$  1.24 (d, *J* = 6.6 Hz, 3H, CCH<sub>3</sub>), 1.45 (s, 9H, OC(CH<sub>3</sub>)<sub>3</sub>), 1.80 (dt, *J* = 11.6, 5.9 Hz, 2H, CH<sub>2</sub>), 2.12 (s, 3H, COCH<sub>3</sub>), 3.18–3.29 (m, 2H, NHCH<sub>2</sub>), 3.56 (dt, *J* = 9.9, 6.0 Hz, 1H, OCHH), 3.82–3.87 (m, 1H, OCHH), 4.23 (qd, *J* = 6.7, 2.3 Hz, 1H, H-5), 4.70 (s, 1H, NH), 4.93 (dd, *J* = 5.4, 2.5 Hz, 1H, H-4), 5.02 (d, *J* = 2.9 Hz, 1H, H-1), 6.02 (ddt, *J* = 9.9, 3.1, 0.7 Hz, 1H, H-2), 6.09 (ddd, *J* = 10.0, 5.4, 1.0 Hz, 1H, H-3). <sup>13</sup>C NMR (126 MHz, CDCl<sub>3</sub>)  $\delta$  170.7 (COCH<sub>3</sub>), 155.9 (NHCO), 130.3 (C-2), 125.9 (C-3), 94.3 (C-1), 69.5 (OC(CH<sub>3</sub>)<sub>3</sub>), 66.3 (OCH<sub>2</sub>), 65.0 (C-4),



64.7 (C-5), 38.2 (NHCH<sub>2</sub>), 29.9 (CH<sub>2</sub>), 28.4 (OC(CH<sub>3</sub>)<sub>3</sub>), 20.9 (COCH<sub>3</sub>), 16.1. ESI-HRMS for C<sub>16</sub>H<sub>27</sub>NO<sub>6</sub>Na<sup>+</sup> (MNa<sup>+</sup>) calculated: 352.1736; found: 352.1748

(3-*N*-Boc-propyl)-acetyl-2,3,6-trideoxy- $\alpha$ -L-hex-2-enopyranosyl (5). To a stirring solution of glycoside 4f (200 mg, 0.18 mmol) in 5 mL of methanol, 20 mol % K<sub>2</sub>CO<sub>3</sub> (36 mg, 0.14 mmol) was added. The solution was stirred at RT until the reaction was determined to be complete by TLC. The reaction mixture was concentrated in vacuo and the dried residue diluted in CHCl<sub>3</sub> (20 mL) and washed with water (20 mL), brine (20 mL), and dried over anhydrous MgSO<sub>4</sub>. The organic phase was concentrated in vacuo and purified by silica gel column chromatography (Hexane:EtOAc, 3:1 to 1:1) to afford 5 as a colorless oil (170 mg, 75%). <sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>)  $\delta$  1.30 (dd, *J* = 6.7, 2.3 Hz, 3H, CCH<sub>3</sub>), 1.44 (s, 9H, OC(CH<sub>3</sub>)<sub>3</sub>), 1.79 (p, *J* = 6.9 Hz, 2H, CH<sub>2</sub>), 3.24 (s, 2H, NHCH<sub>2</sub>), 3.51–3.63 (m, 2H, OCHH, H-4), 3.84 (dddd, *J* = 10.4, 9.3, 5.6, 2.8 Hz, 1H, OCHH), 4.11 (qd, *J* = 6.7, 3.2 Hz, 1H, H-5), 4.72 (s, 1H, NH), 4.95 (s, 1H, H-1), 5.84–5.92 (m, 1H, H-2), 6.15–6.22 (m, 1H, H-3). <sup>13</sup>C NMR (126 MHz, CDCl<sub>3</sub>)  $\delta$  155.9 (NHCO), 130.3 (C-3), 128.1 (C-2), 94.6 (C-1), 79.1 (OC(CH<sub>3</sub>)<sub>3</sub>), 66.4 (C-5), 66.2 (OCH<sub>2</sub>), 63.9 (C-4), 38.2 (NHCH<sub>2</sub>), 29.9 (CH<sub>2</sub>), 28.4 (OC(CH<sub>3</sub>)<sub>3</sub>), 16.1 (CCH<sub>3</sub>). ESI-HRMS for C<sub>14</sub>H<sub>23</sub>NO<sub>5</sub>Na<sup>+</sup> (MNa<sup>+</sup>) calculated: 310.1630; found: 310.1629.

(3-*N*-Boc-propyl)- $\alpha$ -L-Rhodinoside (6). To a stirring solution of glycoside 5 (100 mg 0.348 mmol) in 2 mL of a 1:6 mixture of ethyl acetate;toluene at RT, 5 mol % of Rh–Al<sub>2</sub>O<sub>3</sub> (25 mg) was added. The reaction mixture was placed under a H<sub>2</sub> atmosphere (balloon) and was stirred at RT for 5h. The reaction mixture was filtered through diatomaceous earth and the filtrate was concentrated under vacuum. The dry residue was purified by silica gel column chromatography (Hexane:EtOAc, 5:1 to 2:1) 6 as a colorless oil (77 mg, 76%). <sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>)  $\delta$  4.87 (s, 1H, NH), 4.73–4.77 (m, 1H, H-1), 3.90 (q, *J* = 6.7, 6.0 Hz, 1H, H-5), 3.70 (ddd, *J* = 9.9, 7.2, 5.1 Hz, 1H, OCHH), 3.55 (s, 1H, H-4), 3.39–3.48 (m, 1H, OCHH), 3.19 (dd, *J* = 26.9, 5.8 Hz, 2H, NHCH<sub>2</sub>), 2.08 (s, 1H, OH), 1.89–2.02 (m, 2H, H-2a, H-3a), 1.67–1.81 (m, 3H, H-3b, CH<sub>2</sub>), 1.48–1.55 (m, 1H, H-2b), 1.41 (s, 9H, OC(CH<sub>3</sub>)<sub>3</sub>), 1.15 (d, *J* = 6.6 Hz, 3H, CCH<sub>3</sub>). <sup>13</sup>C NMR (126 MHz, CDCl<sub>3</sub>)  $\delta$  155.9 (NHCO), 97.2 (C-1), 73.9 (OC(CH<sub>3</sub>)<sub>3</sub>), 67.2 (C-5), 66.2 (C-4), 65.4 (OCH<sub>2</sub>), 38.7 (NHCH<sub>2</sub>), 29.5 (CH<sub>2</sub>), 28.4 (OC(CH<sub>3</sub>)<sub>3</sub>), 25.7 (C-3), 23.4 (C-2), 17.1 (CCH<sub>3</sub>). ESI-HRMS for C<sub>14</sub>H<sub>27</sub>NO<sub>3</sub>Na<sup>+</sup> (MNa<sup>+</sup>) calculated: 312.1787; found: 312.1781.

(3-*N*-Boc-propyl)- $\alpha$ -L-Cineruloside (7). To a stirring solution of 5 (100 mg 0.348 mmol) in 2 mL of a 1:6 mixture of ethyl acetate;toluene at RT, 5% Rh–Al<sub>2</sub>O<sub>3</sub> (25 mg) was added. The reaction mixture was placed under a H<sub>2</sub> atmosphere (balloon) and was stirred at room temperature for 5 h. The reaction mixture was filtered through diatomaceous earth, and the filtrate was concentrated under vacuum. The dry residue was purified by silica gel column chromatography (Hexane:EtOAc, 4:1 to 2:1) 7 as a colorless oil (77 mg, 76%). <sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>)  $\delta$  4.97 (t, *J* = 4.7 Hz, 1H, H-1), 4.73 (s, 1H, NH), 4.24 (q, *J* = 6.7 Hz, 1H, H-5), 3.83 (ddd, *J* = 10.0, 6.8, 5.5 Hz, 1H, OCHH), 3.55 (ddd, *J* = 10.0, 6.5, 5.4 Hz, 1H, OCHH), 3.25 (q, *J* = 6.0 Hz, 2H, NHCH<sub>2</sub>), 2.53 (ddd, *J* = 16.1, 8.0, 5.7 Hz, 1H, H-3b), 2.44 (ddd, *J* = 16.1, 8.7, 5.8 Hz, 1H, H-3a), 2.25–2.32 (m, 1H, H-2a), 2.01 (dddd, *J* = 14.1, 8.3, 5.7, 4.3 Hz, 1H, H-2b), 1.75–1.85 (m, 2H, CH<sub>2</sub>), 1.44 (s, 9H, OC(CH<sub>3</sub>)<sub>3</sub>), 1.29 (d, *J* = 6.7 Hz, 3H, CCH<sub>3</sub>). <sup>13</sup>C NMR (126 MHz, CDCl<sub>3</sub>)  $\delta$  210.3 (C-4, C<sub>2</sub>CO), 155.9 (NHCO), 96.8 (C-1), 79.2 (OC(CH<sub>3</sub>)<sub>3</sub>), 71.0 (C-5), 65.8 (OCH<sub>2</sub>), 38.3 (NHCH<sub>2</sub>), 33.6 (C-3), 29.8 (CH<sub>2</sub>), 29.0 (CH<sub>2</sub>), 28.4 (OC(CH<sub>3</sub>)<sub>3</sub>), 14.9 (CCH<sub>3</sub>). ESI-HRMS for C<sub>14</sub>H<sub>25</sub>NO<sub>5</sub>Na<sup>+</sup> (MNa<sup>+</sup>) calculated: 310.1630; found: 310.1630.

(3-*N*-Boc-propyl)- $\alpha$ -L-Aculoside (8). To a flask loaded with 5 (100 mg 0.18 mmol), Dess-Martin periodinane (221 mg 0.52 mmol) in 5 mL CH<sub>2</sub>Cl<sub>2</sub> solvent was added and the reaction left to stir for 6 h at RT. The reaction mixture was then filtered through diatomaceous earth, and the filtrate was concentrated under vacuum. The dry

residue was purified by silica gel column chromatography (Hexane:EtOAc, 5:1 to 3:1) 8 as a colorless oil (84 mg 86% yield). <sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>)  $\delta$  6.84 (dd, *J* = 10.2, 3.5 Hz, 1H, H-2), 6.09 (d, *J* = 10.4 Hz, 1H, H-3), 5.18 (d, *J* = 3.5 Hz, 1H, H-1), 4.67 (s, 1H, NH), 4.55 (q, *J* = 6.8 Hz, 1H, H-5), 3.92 (dt, *J* = 9.9, 6.0 Hz, 1H, OCHH), 3.64 (dt, *J* = 9.9, 6.0 Hz, 1H, OCHH), 3.21–3.31 (m, 2H, NHCH<sub>2</sub>), 1.83 (p, *J* = 6.6 Hz, 2H, CH<sub>2</sub>), 1.45 (s, 9H, OC(CH<sub>3</sub>)<sub>3</sub>), 1.40 (d, *J* = 6.8 Hz, 3H, CCH<sub>3</sub>). <sup>13</sup>C NMR (126 MHz, CDCl<sub>3</sub>)  $\delta$  196.9 (C-4 CO), 155.9 (NHCO), 143.3 (C-2), 127.3 (C-3), 93.3 (C-1), 70.4 (OC(CH<sub>3</sub>)<sub>3</sub>), 67.2 (C-5), 38.0 (CH<sub>2</sub>), 30.0 (CH<sub>2</sub>), 29.7 (CH<sub>2</sub>), 28.0 (OC(CH<sub>3</sub>)<sub>3</sub>), 15.3 (CCH<sub>3</sub>). ESI-HRMS for C<sub>14</sub>H<sub>23</sub>NO<sub>5</sub>Na<sup>+</sup> (MNa<sup>+</sup>) calculated: 308.1474; found: 308.1461.

Methyl-2,3,4-tri-*O*-benzyl-6-*O*-(2-deoxy-3,4,6-tri-*O*-benzyl- $\alpha$ -D-lyxo-hexapyranosyl)- $\alpha$ -D-glucopyranoside (10a). Following the general glycosylation procedure. Donor 9a (50 mg, 0.10 mmol), B(C<sub>6</sub>F<sub>5</sub>)<sub>3</sub> (3 mg 0.006 mmol), and acceptor 2a (42 mg, 0.09 mmol) to afford following purification by column chromatography (Hexane:EtOAc, 6:1 to 3:1) 10a as a colorless oil (82 mg, 88%). Spectroscopic data in agreement with literature.<sup>9a</sup>

Benzyl 2-deoxy-3,4,6-tri-*O*-benzyl- $\alpha$ -D-lyxo-hexapyranoside (10b). Following the general glycosylation procedure. Donor 9a (50 mg, 0.10 mmol), B(C<sub>6</sub>F<sub>5</sub>)<sub>3</sub> (3 mg 0.006 mmol) and acceptor 2b (10 mg, 0.09 mmol) to afford following purification by column chromatography (Hexane:EtOAc, 20:1 to 10:1) 10b as a colorless oil (46 mg, 94%). Spectroscopic data in agreement with literature.<sup>9a</sup>

6-*O*-(3,4,6-tri-*O*-benzyl- $\alpha$ -D-lyxo-hexapyranosyl)-1,2,3,4-di-*O*-isopropylidene- $\alpha$ -D-galactopyranoside (10e). Following the general glycosylation procedure. Donor 9a (50 mg, 0.10 mmol), B(C<sub>6</sub>F<sub>5</sub>)<sub>3</sub> (3 mg 0.006 mmol), and acceptor 2e (23 mg, 0.09 mmol) to afford following purification by column chromatography (Hexane:EtOAc, 6:1 to 4:1) 10e as a colorless oil (50 mg, 84%). Spectroscopic data in agreement with literature.<sup>4</sup>

Methyl 3-*O*-benzyl-2-*O*-(2-deoxy-3,4,6-tri-*O*-benzyl- $\alpha$ -D-lyxo-hexapyranosyl)-4,6-*O*-benzylidene- $\alpha$ -D-glucopyranoside (10g). Following the general glycosylation procedure. Donor 9a (50 mg, 0.10 mmol), B(C<sub>6</sub>F<sub>5</sub>)<sub>3</sub> (3 mg 0.006 mmol), and acceptor 2e (34 mg, 0.09 mmol) to afford following purification by column chromatography (Hexane:EtOAc, 6:1 to 4:1) 10g as a colorless oil (45 mg, 62%). Spectroscopic data in agreement with literature.<sup>9a</sup>

3-*O*-(2-deoxy-3,4,6-tri-*O*-benzyl- $\alpha$ -D-lyxo-hexapyranoside)-1,2,5,6-di-*O*-isopropylidene- $\alpha$ -D-glucofuranoside (10h). Following the general glycosylation procedure. Donor 9a (50 mg, 0.10 mmol), B(C<sub>6</sub>F<sub>5</sub>)<sub>3</sub> (3 mg 0.006 mmol) and acceptor 2h (23 mg, 0.09 mmol) to afford following purification by column chromatography (Hexane:EtOAc, 6:1 to 3:1) 10h as a colorless oil (41 mg, 68%). Spectroscopic data in agreement with literature.<sup>9d</sup>

Methyl-2,3,4-tri-*O*-benzyl-6-*O*-(2-deoxy-3,4,6-tri-*O*-tert-butylidimethylsilyl- $\alpha$ -D-lyxo-hexapyranosyl)- $\alpha$ -D-glucopyranoside (11b). Following the general glycosylation procedure. Donor 9b (50 mg, 0.10 mmol), B(C<sub>6</sub>F<sub>5</sub>)<sub>3</sub> (3 mg 0.006 mmol) and acceptor 2a (36 mg, 0.08 mmol) to afford following purification by column chromatography (Hexane:EtOAc, 10:1 to 8:1) 11b as a colorless oil (68 mg, 92%). Spectroscopic data in agreement with literature.<sup>9a</sup>

Methyl-2,3,4-tri-*O*-benzyl-6-*O*-(2-deoxy-3,4,6-tri-*O*-methoxymethyl ether- $\alpha$ -D-lyxo-hexapyranosyl)- $\alpha$ -D-glucopyranoside (11c). Following the general glycosylation procedure, donor 9c (50 mg, 0.10 mmol), B(C<sub>6</sub>F<sub>5</sub>)<sub>3</sub> (5 mg 0.009 mmol) and acceptor 2a (63 mg, 0.11 mmol) to afford following purification by silica gel column chromatography (Hexane:EtOAc, 7:1 to 5:1) 11c as a colorless oil (71 mg, 71%). Spectroscopic data in agreement with literature.<sup>9a</sup>

Methyl-2,3,4-tri-*O*-benzyl-6-*O*-(2-deoxy-4,6-*O*-[bis(tert-butyl)silylene]-3-*O*-methoxymethyl ether- $\alpha$ -D-lyxo-hexapyranosyl)- $\alpha$ -D-glucopyranoside (11d). Following the general glycosylation procedure, donor 9d (50 mg, 0.10 mmol), B(C<sub>6</sub>F<sub>5</sub>)<sub>3</sub> (4 mg 0.008 mmol) and acceptor 2a (53 mg, 0.11 mmol) to afford following purification by silica gel column chromatography (Hexane:EtOAc, 10:1 to 7:1) 11d as a colorless oil (74 mg, 82%). <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  7.31 (dddd, *J* = 20.0, 16.0, 8.5, 5.8 Hz, 15H, Ar–H), 5.02–4.93 (m, 3H, H-1', PhCHH), 4.83–4.77 (m, 3H, PhCHH, OCH–H), 4.74–4.65 (m, 2H, PhCHH, OCH–H), 4.61 (d, *J* = 3.5 Hz, 1H, H-1), 4.55 (d, *J* = 11.4 Hz, 1H, PhCHH), 4.38 (d, *J* = 2.0 Hz, 1H,

H-4'), 4.05–3.96 (m, 3H, H-3, H-6a', H-6b'), 3.88 (ddd,  $J = 12.2$ , 4.4, 2.7 Hz, 1H, H-3'), 3.78 (m, 2H, H-5, H-6a), 3.63 (d,  $J = 9.6$  Hz, 1H, H-6b), 3.54 (dd,  $J = 9.7$ , 3.6 Hz, 1H, H-2), 3.51–3.45 (m, 1H, H-4), 3.42 (d,  $J = 3.1$  Hz, 4H, H-5', CH<sub>2</sub>OCH<sub>3</sub>), 3.38 (s, 3H, OCH<sub>3</sub>), 2.18 (td,  $J = 12.4$ , 3.6 Hz, 1H, H-2a'), 1.82 (dd,  $J = 12.5$ , 4.7 Hz, 1H, H-2b'), 1.04 (s, 18H, 2C(CH<sub>3</sub>)<sub>3</sub>). <sup>13</sup>C NMR (101 MHz, CDCl<sub>3</sub>)  $\delta$  138.6 (Ar–C), 138.3 (Ar–C), 138.1 (Ar–C), 128.5 (Ar–C), 128.4 (Ar–C), 127.9 (Ar–C), 127.7 (Ar–C), 127.2 (Ar–C), 98.2 (C-1'), 97.9 (C-1), 95.0 (OCH<sub>2</sub>–CH<sub>3</sub>), 82.2 (C-3), 80.1 (C-2), 78.1 (C-4), 75.8 (PhCH<sub>2</sub>), 74.8 (PhCH<sub>2</sub>), 73.3 (PhCH<sub>2</sub>), 72.1 (C-3'), 70.4 (C-4'), 69.7 (C-5), 67.56 (2C, C-6', C-5'), 65.9 (C-6), 55.7 (OCH<sub>2</sub>CH<sub>3</sub>), 55.1 (OCH<sub>3</sub>), 30.1 (C-2'), 27.6 (C(CH<sub>3</sub>)<sub>3</sub>), 27.4 (C(CH<sub>3</sub>)<sub>3</sub>), 23.4 (C(CH<sub>3</sub>)<sub>3</sub>), 20.8 (C(CH<sub>3</sub>)<sub>3</sub>). ESI-HRMS for C<sub>44</sub>H<sub>62</sub>O<sub>11</sub>SiNa<sup>+</sup> (MNa<sup>+</sup>) calculated: 817.3959; found: 817.3965.

**Methyl-2,3,4-tri-O-benzyl-6-O-(2-deoxy-3,4,6-tri-O-benzyl- $\alpha$ -D-lyxo-hexapyranosyl)- $\alpha$ -D-glucopyranoside (12a) and Methyl-6-O-(4,6-di-O-acetyl-2,3-dideoxy- $\alpha$ -D-erythro-hex-2-enopyranosyl)-2,3,4-tri-O-benzyl- $\alpha$ -D-glucopyranoside (12a')**. Following the general glycosylation procedure. Donor 11a (50 mg, 0.10 mmol), B(C<sub>6</sub>F<sub>5</sub>)<sub>3</sub> (3 mg 0.006 mmol) and acceptor 2a (42 mg, 0.09 mmol) to afford following purification by column chromatography (Hexane:EtOAc, 6:1 to 3:1) 12a as a colorless oil (44 mg, 54%) and 12a' as a colorless oil (7 mg, 10%). Spectroscopic data in agreement with literature.<sup>4c</sup>

**Methyl-2,3,4-tri-O-benzyl-6-O-(2-deoxy-3,4-O-(1,1,3,3-tetraisopropylidisiloxane-1,3-diyl)-6-O-benzyl- $\alpha$ -D-erythro-hexapyranosyl)- $\alpha$ -D-glucopyranoside (12b)**. Following the general glycosylation procedure, donor 11b (50 mg, 0.10 mmol), B(C<sub>6</sub>F<sub>5</sub>)<sub>3</sub> (3 mg 0.006 mmol), and acceptor 2a (36 mg, 0.08 mmol) to afford following purification by silica gel column chromatography (Hexane:EtOAc, 12:1 to 8:1) 12b as a colorless oil (56 mg, 78%). Spectroscopic data in agreement with literature.<sup>9a</sup>

**Methyl-2,3,4-tri-O-benzyl-6-O-(2-deoxy-3,4-O-(1,1,3,3-tetraisopropylidisiloxane-1,3-diyl)-6-O-triisopropylsilyl- $\alpha$ -D-erythro-hexapyranosyl)- $\alpha$ -D-glucopyranoside (12c)**. Following the general glycosylation procedure, donor 11c (50 mg, 0.10 mmol), B(C<sub>6</sub>F<sub>5</sub>)<sub>3</sub> (2 mg 0.005 mmol) and acceptor 2a (32 mg, 0.07 mmol) to afford following purification by silica gel column chromatography (Hexane:EtOAc, 12:1 to 8:1) 12c as a colorless oil (51 mg, 73%). Spectroscopic data in agreement with literature.<sup>9a</sup>

**6-O-(2-deoxy-3,4-O-(1,1,3,3-tetraisopropylidisiloxane-1,3-diyl)-6-O-triisopropylsilyl- $\alpha$ -D-erythro-hexapyranosyl)-1,2,3,4-di-O-isopropylidene- $\alpha$ -D-galactopyranoside (13)**. Following the general glycosylation procedure, donor 11c (50 mg, 0.10 mmol), B(C<sub>6</sub>F<sub>5</sub>)<sub>3</sub> (2 mg 0.005 mmol), and acceptor 2e (18 mg, 0.07 mmol) to afford following purification by column chromatography (Hexane:EtOAc, 16:1 to 10:1) 13 as a colorless oil (48 mg, 86%). <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  0.83–1.18 (m, 49H, 7 $\times$  SiCH(CH<sub>3</sub>)<sub>2</sub>), 1.32 (s, 6H, 2 O<sub>2</sub>CCH<sub>3</sub>), 1.43 (s, 3H, O<sub>2</sub>CCH<sub>3</sub>), 1.51 (s, 3H, O<sub>2</sub>CCH<sub>3</sub>), 1.66 (ddd,  $J = 13.3$ , 11.4, 3.7 Hz, 1H, H-2a'), 2.10 (dd,  $J = 12.9$ , 5.3 Hz, 1H, H-2b'), 3.49 (dd,  $J = 9.4$ , 8.3 Hz, 1H, H-4'), 3.54–3.61 (m, 1H, H-5'), 3.65 (dd,  $J = 10.7$ , 6.2 Hz, 1H, H-6b'), 3.79 (ddd,  $J = 17.8$ , 10.7, 6.4 Hz, 2H, H-6b, H-6a'), 3.97 (td,  $J = 6.5$ , 1.6 Hz, 1H, H-5), 4.00–4.08 (m, 2H, H-6a, H-3'), 4.20 (dd,  $J = 7.9$ , 1.8 Hz, 1H, H-4), 4.30 (dd,  $J = 5.0$ , 2.4 Hz, 1H, H-2), 4.60 (dd,  $J = 7.9$ , 2.3 Hz, 1H, H-3), 4.95 (d,  $J = 3.1$  Hz, 1H, H-1'), 5.51 (d,  $J = 5.0$  Hz, 1H, H-1). <sup>13</sup>C NMR (101 MHz, CDCl<sub>3</sub>)  $\delta$  109.2 (O<sub>2</sub>CCH<sub>3</sub>), 108.4 (O<sub>2</sub>CCH<sub>3</sub>), 96.3 (C-1), 96.0 (C-1'), 74.6 (C-4'), 73.4 (C-5'), 71.6 (C-3'), 71.1 (C-4), 70.7 (2C, C-2, C-3), 65.5 (C-5), 64.5 (C-6'), 63.3 (C-6), 38.0 (C-2'), 26.0 (O<sub>2</sub>CCH<sub>3</sub>), 25.9 (O<sub>2</sub>CCH<sub>3</sub>), 24.9 (O<sub>2</sub>CCH<sub>3</sub>), 24.4 (O<sub>2</sub>CCH<sub>3</sub>), 18.0, 17.9, 17.6, 17.4, 17.37, 17.3, 17.2 (Si(CH(CH<sub>3</sub>)<sub>2</sub>)), 13.0, 12.8, 12.4, 12.3, 12.0 (Si(CH(CH<sub>3</sub>)<sub>2</sub>)). ESI-HRMS for C<sub>39</sub>H<sub>76</sub>O<sub>11</sub>Si<sub>3</sub>Na<sup>+</sup> (MNa<sup>+</sup>) calculated: 827.4593; found: 827.4586

**Methyl-3-O-benzyl-2-O-(2-deoxy-3,4-O-(1,1,3,3-tetraisopropylidisiloxane-1,3-diyl)-6-O-triisopropylsilyl- $\alpha$ -D-erythro-hexapyranosyl)-4,6-O-benzylidene- $\alpha$ -D-glucopyranoside (14)**. Following the general glycosylation procedure, donor 11c (50 mg, 0.10 mmol), B(C<sub>6</sub>F<sub>5</sub>)<sub>3</sub> (2 mg 0.005 mmol), and acceptor 2g (26 mg, 0.07 mmol) to afford following purification by column chromatography (Hexane:EtOAc, 10:1 to 5:1) 14 as a colorless oil (41 mg, 64%). Spectroscopic data in agreement with literature.<sup>9a</sup>

**Methyl-2,3,4-tri-O-benzyl-6-O-(2-deoxy-3,4,6-tri-O-benzyl-(axial/equatorial)-2-<sup>2</sup>H- $\alpha$ -D-lyxo-hexapyranosyl)- $\alpha$ -D-glucopyranoside (16a/16b)**. Following the General Glycosylation Procedure, galactal 9 (50 mg, 0.120 mmol), B(C<sub>6</sub>F<sub>5</sub>)<sub>3</sub> (3 mg 0.006 mmol), and acceptor 2a (42 mg, 0.090 mmol). Following purification by silica gel column chromatography (8:1 to 4:1, Hexane:EtOAc) to afford glycoside 16a/16b as an oil (70 mg, 88%).<sup>9a</sup>

**Hexafluoroisopropyl-4,6-di-O-acetyl-2,3-dideoxy- $\alpha$ -D-threo-hex-2-enopyranoside (18)**. Following the General Glycosylation Procedure, galactal 1a (50 mg, 0.18 mmol), B(C<sub>6</sub>F<sub>5</sub>)<sub>3</sub> (5 mg 0.009 mmol) acceptor 17 (42 mg, 0.090 mmol) at 45°C. Following purification by silica gel column chromatography (6:1 to 4:1, Hexane:EtOAc) product 18 was obtained as an oil (45 mg, 87%). <sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>)  $\delta$  6.29 (ddd,  $J = 10.0$ , 5.7, 1.2 Hz, 1H, H-3), 6.09 (dd,  $J = 10.0$ , 3.0 Hz, 1H, H-2), 5.34 (d,  $J = 2.6$  Hz, 1H, H-1), 5.08 (dd,  $J = 5.7$ , 2.5 Hz, 1H, H-4), 4.63 (hept,  $J = 5.4$  Hz, 1H, HC(CF<sub>3</sub>)<sub>2</sub>), 4.35 (ddd,  $J = 7.3$ , 4.6, 2.4 Hz, 1H, H-5), 4.30 (dd,  $J = 11.6$ , 4.6 Hz, 1H, H-6a), 4.18 (dd,  $J = 11.6$ , 7.5 Hz, 1H, H-6b), 2.09 (s, 3H, COCH<sub>3</sub>), 2.07 (s, 3H, COCH<sub>3</sub>). <sup>13</sup>C NMR (126 MHz, CDCl<sub>3</sub>)  $\delta$  170.5 (COCH<sub>3</sub>), 170.1 (COCH<sub>3</sub>), 127.9(C-2), 126.9 (C-3), 122.9 (CF<sub>3</sub>), 122.2 (CF<sub>3</sub>), 94.8 (C-1), 71.6–71.1 (C(CF<sub>3</sub>)<sub>2</sub>), 68.1 (C-5), 62.4 (C-6), 62.1 (C-4), 20.7 (COCH<sub>3</sub>), 20.5 (COCH<sub>3</sub>). <sup>19</sup>F NMR (470 MHz, CDCl<sub>3</sub>)  $\delta$  –73.52 (m, CF<sub>3</sub>), –73.61 (m, CF<sub>3</sub>). ESI-HRMS for C<sub>13</sub>H<sub>14</sub>F<sub>6</sub>O<sub>6</sub>Na<sup>+</sup> (MNa<sup>+</sup>) calculated: 403.0592; found: 403.0600.

**Hexafluoroisopropyl-2-deoxy-3,4,6-tri-O-benzyl- $\alpha$ -D-lyxo-hexapyranoside (19)**. Following the General Glycosylation Procedure, galactal 9a (50 mg, 0.120 mmol), B(C<sub>6</sub>F<sub>5</sub>)<sub>3</sub> (3 mg 0.006 mmol) acceptor 17 (42 mg, 0.090 mmol) at RT. Following purification by silica gel column chromatography (12:1 to 8:1, Hexane:EtOAc) product 19 was obtained as an oil (48 mg, 92%). <sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>)  $\delta$  7.46–7.23 (m, 15H, Ar–H), 5.28 (d,  $J = 3.6$  Hz, 1H, H-1), 4.97 (d,  $J = 11.5$  Hz, 1H, PhCHH), 4.67–4.61 (m, 3H, PhCHH, PhCHH), 4.50 (q,  $J = 11.9$  Hz, 3H, PhCHH, HC(CF<sub>3</sub>)<sub>2</sub>), 4.06–3.99 (m, 2H, H-4, H-5), 3.95 (ddd,  $J = 12.1$ , 4.6, 2.3 Hz, 1H, H-3), 3.63 (dd,  $J = 9.3$ , 7.0 Hz, 1H, H-6a), 3.58 (dd,  $J = 9.3$ , 5.9 Hz, 1H, H-6b), 2.37 (td,  $J = 12.7$ , 3.9 Hz, 1H, H-2a), 2.19 (dd,  $J = 13.2$ , 4.6 Hz, 1H, H-2b). <sup>13</sup>C NMR (126 MHz, CDCl<sub>3</sub>)  $\delta$  138.6 (Ar–C), 138.1 (Ar–C), 138.0 (Ar–C), 128.5 (Ar–C), 128.4 (Ar–C), 128.3 (Ar–C), 128.2 (Ar–C), 127.7 (Ar–C), 127.7 (Ar–C), 127.6 (Ar–C), 127.6 (Ar–C), 127.4 (Ar–C), 122.4 (CF<sub>3</sub>), 120.2 (CF<sub>3</sub>), 100.0 (C-1), 74.4 (PhCH<sub>2</sub>), 73.8 (C-3), 73.4 (PhCH<sub>2</sub>), 72.5 (C-4), 71.5 (C(CF<sub>3</sub>)<sub>2</sub>), 71.4 (C-5), 70.7 (PhCH<sub>2</sub>), 68.8 (C-6), 30.2 (C-2). <sup>19</sup>F NMR (470 MHz, CDCl<sub>3</sub>)  $\delta$  –73.24 (m, CF<sub>3</sub>), –73.40 (m, CF<sub>3</sub>). ESI-HRMS for C<sub>30</sub>H<sub>30</sub>F<sub>6</sub>O<sub>5</sub>Na<sup>+</sup> (MNa<sup>+</sup>) calculated: 607.1895; found: 607.1903.

**Synthesis of Methyl-2,3,4-tri-O-benzyl-6-O-(2-deoxy-3,4,6-tri-O-benzyl- $\alpha$ -D-lyxohexapyranosyl)- $\alpha$ -D-glucopyranoside (10a)**. The glycosyl donor 9a (1 equiv) and acceptor 2a (0.83 equiv) were weighed into a microwave vial and placed under vacuum for 1 h, after which time the microwave vial was filled with N<sub>2</sub>. A solution mixture containing (R)-3,3'-Bis[3,5-bis(trifluoromethyl)phenyl]-1,1'-binaphthyl-2,2'-diyl hydrogenphosphate (0.1 equiv) and thiourea (0.1 equiv) in anhydrous CH<sub>3</sub>CN (1 mL) was stirred for 30 min, before adding it to the microwave vial containing 1a and 2a. The reaction mixture was stirred at RT for 4 h and then was purified by silica gel column chromatography (Hexane:EtOAc, 7:1 to 4:1) affording disaccharide 3a as a colorless oil (66 mg 70%, 4:1  $\alpha$ : $\beta$ ). The spectroscopic data was in agreement with previously reported data.<sup>15</sup>

**In Situ Anomerization Test of 10a in the Presence of 2a**. Disaccharide 10a (4:1  $\alpha$ : $\beta$ , 1 equiv), acceptor monosaccharide 2a (1 equiv), and B(C<sub>6</sub>F<sub>5</sub>)<sub>3</sub> (5 mol %) were weighed into an oven-dried microwave vial, sealed and placed under vacuum for 1 h. Then the vial was filled with N<sub>2</sub> followed by the addition of ~1.0 mL of anhydrous toluene. The solutions were stirred and heated at 50 °C for 2 h without observing any change in the anomeric ratio (4:1  $\alpha$ / $\beta$ ) as monitored by NMR of the crude mixture.



## ■ ASSOCIATED CONTENT

### Supporting Information

The Supporting Information is available free of charge on the ACS Publications website at DOI: 10.1021/acs.joc.8b02613.

Reaction optimization tables and NMR reaction studies and NMR spectra for all compounds (PDF)

## ■ AUTHOR INFORMATION

### Corresponding Author

\*E-mail: m.c.galan@bris.ac.uk

### ORCID

M. Carmen Galan: 0000-0001-7307-2871

### Notes

The authors declare no competing financial interest.

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