

Chronic Pulmonary Disease Due to *Mycobacterium monacense* Infection: The First Case from Iran

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We herein report a case in which the recently characterized species *Mycobacterium monacense* was isolated from the sputum of an Iranian patient. This case represents the first isolation of *M. monacense* from Iran. The isolate was identified by conventional and molecular techniques. Our findings show that *M. monacense* infection is not restricted to developed countries.

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INTRODUCTION

Mycobacterium monacense is a rapidly growing *Mycobacterium* (RGM) that was first isolated from independent clinical specimens in 2006 [1]. Subsequently, 3 additional studies from the United States, Germany, and India have reported the isolation of *M. monacense* from human clinical samples [2-4]. Here, we report the first case of human isolate of *M. monacense* from a chronic respiratory infection in Iran identified by a combination of phenotypic and molecular tests.

CASE REPORT

A 57-yr-old female was admitted to a hospital in the suburb of Isfahan, Iran, for respiratory impairment with chronic productive cough and chest pain. She had no history of mycobacterial infection. A purified protein derivative (PPD) skin test was positive

to 5 TU (>12 mm). A chest X radiograph revealed clusters of small (>5 mm) nodules. Based on radiological findings, a positive tuberculin test, the presence of acid-fast bacilli (AFB) in a direct smear of sputum, and isolation of the same strain on 3 repeated examinations, the patient was prescribed anti-tuberculosis drugs and was entered into the tuberculosis register. However, an examination of the cultured infecting organism revealed a yellow-pigmented, rapidly growing scotochromogenic *Mycobacterium*. Therefore, following the primary identification of the causative agent, the patient's treatment was changed to combination therapy with amikacin and ciprofloxacin. After the initial 45 days of the anti-tuberculosis therapy and the following 45 days of antimicrobial therapy of non-tuberculous pulmonary infection, the patient's general condition improved. Her sputum smears and culture became negative and remained negative at each follow-up examination every 3 months. After discharge from hospital, the patient received a 12-month course of contin-

ued treatment at a peripheral center affiliated with the national health care system. During the 24-month follow-up period, the patient did not relapse.

A battery of conventional phenotypic and molecular tests was used to conclusively identify the M11 strain. The phenotypic tests were carried out using standard culture and biochemical methods, as previously described [5]. The susceptibility of the isolate “M11” to major anti-mycobacterial agents was performed using the microdilution method for rapidly growing mycobacteria [6]. For molecular testing a panel of previously defined molecular markers for mycobacteria were used including, PCR amplification of a genus-specific region of the 65-kDa heat shock protein (*hsp*) gene [7], PCR restriction fragment length polymorphism analysis (PRA) of a 644-bp fragment of the *hsp65* gene [8], amplification and direct sequence analysis of the near-full length 16S rDNA and 16S-23S rDNA internal transcribed spacer (ITS) genes, and amplification and direct sequence analysis of partial *hsp65* and *rpoB* genes [9-13]. The GenBank accession numbers of the M11 strain genetic markers in the present study are GU142931, HM229791, HM229792, and HM229793.

Acid-fast bacilli were observed by microscopic evaluation of the sputum sample; the organism was then cultured on Löwenstein-Jensen (LJ) medium. The same isolate was isolated from 3 consecutive sputum samples. The M11 isolate was a rapidly growing (<7 days), scotochromogenic *Mycobacterium* capable of growth at 25°C, 37°C, and 45°C. Subcultures of the primary isolate had both smooth and rough colony types. Strain M11 was positive for semi-quantitative (100-mm foam) and heat-stable (68°C) catalases, nitrate reduction, Tween® hydrolysis, growth on LJ medium with 5% NaCl, and tellurite reduction. Strain M11 was negative for urease activity, growth on MacConkey agar without crystal violet, arylsulfatase activity (14 days), niacin production, and iron uptake. The M11 isolate was susceptible to amikacin (≤ 1 µg/mL), ceftioxin (2 µg/mL), ciprofloxacin (≤ 0.12 µg/mL), clarithromycin (≤ 0.12 µg/mL), doxycycline (≤ 1 µg/mL), ethambutol (≤ 0.5 µg/mL), imipenem (1 µg/mL), rifampicin (≤ 0.06 µg/mL), streptomycin (2 µg/mL), and sulfamethoxazole (1 µg/mL).

Genus-specific PCR amplification yielded a characteristic 228-bp fragment of the *hsp65* gene, confirming that strain M11 belonged to the genus *Mycobacterium*. The M11 isolate had a short helix 18 in the 16S rDNA gene, which is a typical molecular signature of rapidly growing mycobacteria.

In the PRA method targeting the 644-bp fragment, the M11 isolate’s restriction profile consisted of 456/180-bp, 270/161/117-bp and 320/207-bp fragments from digestion with *Avall*, *HphI*, and *HpaII*, respectively. This restriction profile is distinct from

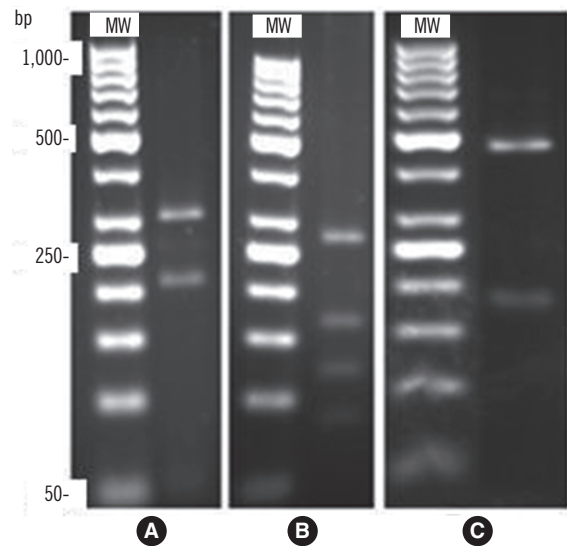


Fig. 1. PCR restriction fragment length polymorphism patterns obtained from digestion of the amplified *hsp65* gene of the isolate “M11” with *HphI* (A), *HpaII* (B) and *Avall* (C), respectively. MW: the 50 bp molecular weight marker.

those of other previously characterized atypical mycobacteria (Fig. 1).

The near-full length (1,438-bp) 16S rDNA gene sequence of the M11 isolate shared 99.93% identity with that of *M. monacense* DSM44395^T, 99.23% identity with that of *M. doricum* DSM44339^T, and 97.89% identity with that of *M. vaccae* ATCC 15483^T. Accordingly, there were 1, 13, and 29 nucleotide differences, respectively. The first and second hypervariable signature sequences of the M11 isolate (positions 128-270 and 408-503; *Escherichia coli* numbering) shared 100% identity with those of the *M. monacense* type strain (Table 1).

In a phylogeny generated using the 16S rDNA gene, the M11 isolate was classified as an RGM species, and was closely related to *M. monacense* (Fig. 2).

The M11 isolate’s *hsp65* gene sequence shared 99.09% identity with that of *M. monacense*. Its ITS gene sequence also shared 97.83% identity with that of *M. monacense* clone F1-05352. The M11 isolate’s 745-bp fragment of the *rpoB* gene had a unique sequence compared to those of other atypical mycobacteria; however a comparison was not made between the M11 isolate and of the *M. monacense* type strain because its relevant data was not available in the molecular databases.

DISCUSSION

In countries with limited resources, such as Iran, the identification of non-tuberculous mycobacteria (NTM) species by only

Table 1. Alignment of selected stretches of 16S rDNA gene of Iranian strain of M11 compared with those of closely related mycobacteria

Strain	Genbank accession number	16S rDNA positions according to <i>E. coli</i> numbering system																						
		95	96	97	181	183	184	186	192	193	193	194	258	262	264	268	269	307	407	441	464	465	466	467
<i>M. monacense</i> DSM44395 ^T	AF 107039	T	A	C	A	C	G	C	G	C	G	G	A	T	C	T	C	C	G	A	G	C	A	A
<i>M. doricum</i> DSM44339 ^T	AF 264700	.	G	T	A	T	.	.	.	T	G	.	.
<i>M. flavescens</i> ATCC14474 ^T	X52932	.	.	.	T	A	.	G	G	T	C	T	.	A	G
M 11	GU142931	.	G	.	Y

“.”Indicates that the base pair was identical to that of type strain of *M. monacense*.

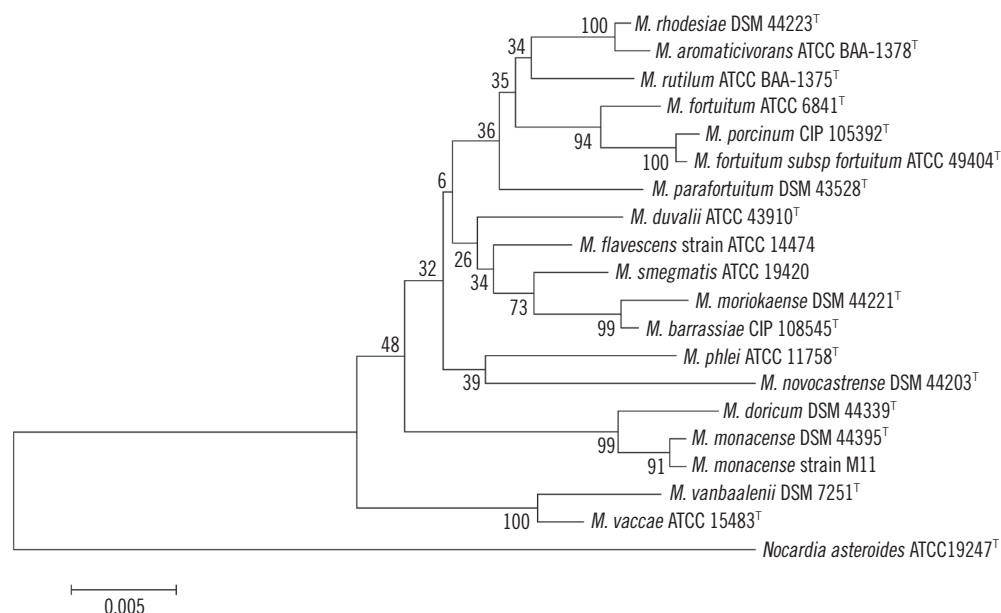


Fig. 2. Alignment of selected stretches of 16S rDNA gene of the Iranian isolate “M11” compared with those of closely related mycobacteria.

routine conventional tests (e.g., direct microscopy and culture analysis) may result in erroneous or incomplete diagnosis. As a result, infections caused by these organisms may be underdiagnosed or misdiagnosed and, accordingly, mistreated [14, 15]. Fortunately, the use of molecular tests in conjunction with conventional phenotypic tests has shown significant promise in accurately identifying clinically relevant NTM [14-16].

In 2006, *M. monacense* was first characterized as a new species following its isolation from the bronchial lavage of an elderly patient and from a biopsy of an injury-acquired fistula in a healthy child [1]. Since then, 3 additional cases of *M. monacense* isolation from clinical samples have been published. Taieb et al. [2] reported the isolation of *M. monacense* from a hand infection of an American diabetic patient. Hogardt et al. [3] reported a pulmonary tuberculosis-like infection in a Chinese patient who had traveled to Europe, suggesting that it might have originated in China. In the most recent report, Therese et al. reported the first

case of *M. monacense* isolation from the sputum of a female patient in India [4].

In all but one of the reported cases, the clinical significance of isolation of *M. monacense* remains uncertain. Specifically, Hogardt et al. [3] presented the only case of a pulmonary disease associated with *M. monacense* which was isolated from a pulmonary tumor of a Chinese patient. The current report described a pulmonary disease attributable to infection with *M. monacense*; this could support a clinically relevant role for this organism. In addition to its phenotypic features, the molecular tests used in the current study suggested that the Iranian isolate belonged to the *M. monacense* species. Specifically, it shared the highest identity with the 16S rDNA and *hsp65* gene sequences of *M. monacense*, as opposed to those of other published mycobacteria.

The M11 isolate was considered the causative agent of disease in this case, because the acid-fast bacilli were microscopi-

cally observed in more than one diseased specimen and were subsequently recovered from a pure culture. Furthermore, no such organism had been isolated in our laboratory previously or during the same time. Our case also met the minimum evaluation criteria for patients suspected of non-tuberculous mycobacterial lung disease set by the American Thoracic Society [17]. These criteria include pulmonary symptoms, a positive chest radiograph, and 3 sputum specimens positive for AFB.

This report follows a case in the United States, which confirms Hogardt's suggestion that *M. monacense* infection is not restricted to Europe [12]. Consistent with previous reports on most non-tuberculous mycobacteria, our findings indicate that molecular markers are important for the identification of rare clinical isolates of atypical mycobacteria. Consequently, we recommend that developing countries centralize the laboratory diagnosis of rare NTM to a regional referral laboratory in order to avoid misidentification.

Authors' Disclosures of Potential Conflicts of Interest

No potential conflict of interest relevant to this article was reported.

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REFERENCES

1. Reischl U, Melzl H, Kroppenstedt RM, Miethke T, Naumann L, Mariottini A, et al. *Mycobacterium monacense* sp. nov. Int J Syst Evol Microbiol 2006;56:2575-8.
2. Taieb A, Ikeguchi R, Yu VL, Rihs JD, Sharma M, Wolfe J, et al. *Mycobacterium monacense*: a mycobacterial pathogen that causes infection of the hand. J Hand Surg Am 2008;33:94-6.
3. Hogardt M, Schreff AM, Naumann L, Reischl U, Sing A. *Mycobacterium monacense* in a patient with a pulmonary tumor. Jpn J Infect Dis 2008;61:77-8.
4. Therese KL, Gayathri R, Thirupathi K, Madhavan HN. First report on isolation of *Mycobacterium monacense* from sputum specimen in India. Lung India 2011;28:124-6.
5. Kent PT and Kubica GP. Public health mycobacteriology: a guide for the level III laboratory. Centers for Disease Control, U.S. Department of Health and Human Services, Atlanta, Ga.; 1985.
6. National Committee for Clinical Laboratory Standards. Susceptibility testing of mycobacteria, nocardiae, and other aerobic actinomycetes; Approved Standard. Wayne, PA: NCCLS; 2003. Document No. M24-A.
7. Khan IU and Yadav JS. Development of a single-tube, cell lysis-based, genus-specific PCR method for rapid identification of mycobacteria: optimization of cell lysis, PCR primers and conditions, and restriction pattern analysis. J Clin Microbiol 2004;42:453-7.
8. Kim H, Kim SH, Shim TS, Kim MN, Bai GH, Park YG, et al. PCR restriction fragment length polymorphism analysis (PRA)-algorithm targeting 644 bp Heat Shock Protein 65 (*hsp65*) gene for differentiation of *Mycobacterium* spp. J Microbiol Methods 2005;62:199-09.
9. Shojaei H, Magee JG, Freeman R, Yates M, Horadagoda NU, Goodfellow M. *Mycobacterium elephantis* sp. nov., a rapidly growing non-chromogenic *Mycobacterium* isolated from an elephant. Int J Syst Evol Microbiol 2000;50 Pt 5:1817-20.
10. Roth A, Fischer M, Hamid ME, Michalke S, Ludwig W, Mauch H. Differentiation of phylogenetically related slowly growing mycobacteria based on 16S-23S rRNA gene internal transcribed spacer sequences. J Clin Microbiol 1998;36:139-47.
11. Kim H, Kim SH, Shim TS, Kim MN, Bai GH, Park YG, et al. Differentiation of *Mycobacterium* species by analysis of the heat-shock protein 65 gene (*hsp65*). Int J Syst Evol Microbiol 2005;55:1649-56.
12. Adekambi T, Colson P, Drancourt M. *rpoB*-based identification of non-pigmented and late-pigmenting rapidly growing mycobacteria. J Clin Microbiol 2003;41:5699-708.
13. Jeon YS, Chung H, Park S, Hur I, Lee JH, Chun J. jPHYDIT: a JAVA-based integrated environment for molecular phylogeny of ribosomal RNA sequences. Bioinformatics 2005;21:3171-3.
14. Shojaei H, Heidarieh P, Hashemi A, Feizabadi MM, Daei Naser A. Species identification of neglected nontuberculous mycobacteria in a developing country. Jpn J Infect Dis 2011;64:265-71.
15. Tortoli E, Rogasi PG, Fantoni E, Beltrami C, De Francisci A, Mariottini A. Infection due to a novel *Mycobacterium*, mimicking multidrug-resistant *Mycobacterium tuberculosis*. Clin Microbiol Infect 2010;16:1130-4.
16. Tortoli E, Bartoloni A, Böttger EC, Emler S, Garzelli C, Magliano E, et al. Burden of unidentifiable mycobacteria in a reference laboratory. J Clin Microbiol 2001;39:4058-65.
17. Glassroth J. Pulmonary disease due to nontuberculous mycobacteria. Chest 2008;133:243-51.