



Complete Genome Sequence of *Labrenzia* sp. Strain CP4, Isolated from a Self-Regenerating Biocathode Biofilm

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Here, we present the complete genome sequence of *Labrenzia* sp. strain CP4, isolated from an electricity-consuming marine biocathode biofilm. *Labrenzia* sp. strain CP4 consists of a circular 5.2 Mbp chromosome and an 88 Kbp plasmid.

Received 16 March 2016 Accepted 17 March 2016 Published 12 May 2016

Citation Wang Z, Eddie BJ, Malanoski AP, Hervey WJ, IV, Lin B, Strycharz-Glaven SM. 2016. Complete genome sequence of *Labrenzia* sp. strain CP4, isolated from a self-regenerating biocathode biofilm. Genome Announc 4(3):e00354-16. doi:10.1128/genomeA.00354-16.

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L abrenzia spp. are aerobic, motile, Gram-negative bacteria recently assigned to the family *Rhodobacterales* (1). Currently, the genus *Labrenzia* consists of five recognized species: *L. alexandrii*, *L. aggregata*, *L. alba*, *L. marina*, and *L. suaedae* (1, 2), all isolated from hypersaline habitats. However, the only complete genome currently available is that of *L. alexandrii* type strain (DFL-11^T) (3). The general characteristics of this genus include polar flagella, ubiquinone 10 (Q10) as the major respiratory lipoquinone, nitrate reduction to nitrite or nitrogen, and chemoheterotrophic and nonfermentative growth under aerobic and anaerobic conditions (2). Notably, *Labrenzia* spp. have been isolated from microbial communities enriched from the Deepwater Horizon (DWH) site, as well as other oil-contaminated coastal marine sediments (4–6), and found to degrade polycyclic aromatic hydrocarbons (PAHs).

Labrenzia sp. strain CP4 was isolated from a biocathode microbial community, known as Biocathode-MCL (*Marinobacter*, *Chromatiaceae*, *Labrenzia*), originally enriched from seawater collected at Rutgers Marine Field Station (RUMFS) in Tuckerton, New Jersey, USA (7). Biocathode-MCL uses electrons supplied by a cathode to drive CO_2 fixation and O_2 reduction and *Labrenzia* has been shown to be one of the most abundant active constituents (8, 9). Genome analysis of CP4 supports ongoing multiomics studies of Biocathode-MCL to determine the mechanism of extracellular electron transfer (EET) at the biocathode for microbial electrosynthesis (8, 9).

The genome of CP4 was sequenced by DNA Link USA, Inc. (San Diego, CA, USA) using the PacBio RS II sequencing platform (Pacific Biosciences, Menlo Park, CA, USA). Genomic DNA was extracted using the Wizard genomic DNA purification kit (Promega, Madison, WI, USA) and used to prepare a 10-kb insert library that was sequenced using two single-molecule real-time (SMRT) sequencing cells and P4-C2 chemistry. This resulted in 17,249 filtered and preassembled sequence reads with a mean length of 7,314 bp and 111× genome coverage. Assembly (via SMRTpipe HGAP.2 and SMRTpipe Celera Assembler) and consensus polishing (SMRTpipe Quiver) yielded two circular contigs with sizes of 5,249,082 and 87,984 bp (58.97% G+C content), which represent a chromosome and a plasmid, respectively. The

genome is predicted to contain 5,746 protein-coding sequences (CDS), 3 rRNA operons, and 60 tRNAs using RAST (Rapid Annotation using Subsystem Technology). Labrenzia spp. are known to oxidize CO to CO2 by proteins encoded by the carbon monoxide dehydrogenase (CODH) operon (coxLMS) (10, 11). Identification of two forms of CODH (form I and II) in CP4 suggests that this bacterium has this capability. It is noted that CP4 contains homologous genes to those for the photorespiratory glycerate pathway in cyanobacteria (12). However, in contrast to L. alexandrii, CP4 does not contain the photosynthetic reaction center genes *pufLM* or bacteriochlorophyll synthase genes. The plasmid shares many features with plasmid LADFL 5 in L. alexandrii, such as a P-type ATPase translocating heavy-metal ions and mercury and cobalt-zinc-cadmium resistance systems (3). In addition, it also harbors multiple genes encoding cytochrome c and cytochrome oxidase biogenesis proteins.

Nucleotide sequence accession numbers. The complete genome sequences of *Labrenzia* sp. CP4 have been deposited in GenBank under accession numbers CP011927 (chromosome) and CP011928 (plasmid).

ACKNOWLEDGMENTS

We acknowledge support listed below from the Office of Naval Research (ONR) and the Naval Research Laboratory (NRL).

The opinions and assertions contained herein are those of the authors and are not to be construed as those of U.S. Navy, military service at large, or the U.S. government.

FUNDING INFORMATION

This work was funded by the ONR via U.S. NRL core funds, as well as under ONR award numbers N0001413WX20995, N0001414WX20485, N0001414WX20518, and N0001415WX00195 to S.S.-G.

REFERENCES

- 1. Bibi F, Jeong JH, Chung EJ, Jeon CO, Chung YR. 2014. *Labrenzia suaedae* sp. nov., a marine bacterium isolated from a halophyte, and emended description of the genus *Labrenzia*. Int J Syst Evol Microbiol 64:1116–1122. http://dx.doi.org/10.1099/ijs.0.052860-0.
- 2. Biebl H, Pukall R, Lunsdorf H, Schulz S, Allgaier M, Tindall BJ, Wagner-Dobler I. 2007. Description of *Labrenzia alexandrii* gen. nov., sp. nov., a novel alphaproteobacterium containing bacteriochlorophyll *a*, and

a proposal for reclassification of *Stappia aggregata* as *Labrenzia aggregata* comb. nov., of *Stappia marina* as *Labrenzia marina* comb. Nov. and of *Stappia alba* as *Labrenzia alba* comb. nov., and emended descriptions of the genera *Pannonibacter*, *Stappia* and *Roseibium*, and of the species *Roseibium denhamense* and *Roseibium hamelinense*. Int J Syst Evol Microbiol 57:1095–1107.

- Fiebig A, Pradella S, Petersen J, Päuker O, Michael V, Lünsdorf H, Göker M, Klenk HP, Wagner-Döbler I. 2013. Genome of the R-body producing marine alphaproteobacterium *Labrenzia alexandrii* type strain (DFL-11(T)). Stand Genomic Sci 7:413–426. http://dx.doi.org/10.4056/ sigs.3456959.
- Bacosa HP, Liu Z, Erdner DL. 2015. Natural sunlight shapes crude oil-degrading bacterial communities in Northern Gulf of Mexico surface waters. Front Microbiol 6:1325. http://dx.doi.org/10.3389/ fmicb.2015.01325.
- Overholt WA, Green SJ, Marks KP, Venkatraman R, Prakash O, Kostka JE. 2013. Draft genome sequences for oil-degrading bacterial strains from beach sands impacted by the Deepwater Horizon oil spill. Genome Announc 1(6):e01015-13. http://dx.doi.org/10.1128/genomeA.01015-13.
- Militon C, Jézéquel R, Gilbert F, Corsellis Y, Sylvi L, Cravo-Laureau C, Duran R, Cuny P. 2015. Dynamics of bacterial assemblages and removal of polycyclic aromatic hydrocarbons in oil-contaminated coastal marine sediments subjected to contrasted oxygen regimes. Environ Sci Pollut Res Int 22:15260–15272. http://dx.doi.org/10.1007/s11356-015-4510-y.
- Strycharz-Glaven SM, Glaven RH, Wang Z, Zhou J, Vora GJ, Tender LM. 2013. Electrochemical investigation of a microbial solar cell reveals a

nonphotosynthetic biocathode catalyst. Appl Environ Microbiol **79**: 3933–3942. http://dx.doi.org/10.1128/AEM.00431-13.

- Wang Z, Leary DH, Malanoski AP, Li RW, Hervey WJt, Eddie BJ, Tender GS, Yanosky SG, Vora GJ, Tender LM, Lin B, Strycharz-Glaven SM. 2015. A previously uncharacterized, nonphotosynthetic member of the *Chromatiaceae* is the primary CO2-fixing constituent in a selfregenerating biocathode. Appl Environ Microbiol 81:699–712. http:// dx.doi.org/10.1128/AEM.02947-14.
- Leary DH, Hervey WJ, IV, Malanoski AP, Wang Z, Eddie BJ, Tender GS, Vora GJ, Tender LM, Lin B, Strycharz-Glaven SM. 2015. Metaproteomic evidence of changes in protein expression following a change in electrode potential in a robust biocathode microbiome. Proteomics 15: 3486–3496. http://dx.doi.org/10.1002/pmic.201400585.
- Weber CF, King GM. 2007. Physiological, ecological, and phylogenetic characterization of *Stappia*, a marine CO-oxidizing bacterial genus. Appl Environ Microbiol 73:1266–1276. http://dx.doi.org/10.1128/ AEM.01724-06.
- King GM, Weber CF. 2007. Distribution, diversity and ecology of aerobic CO-oxidizing bacteria. Nat Rev Microbiol 5:107–118. http://dx.doi.org/ 10.1038/nrmicro1595.
- 12. Eisenhut M, Kahlon S, Hasse D, Ewald R, Lieman-Hurwitz J, Ogawa T, Ruth W, Bauwe H, Kaplan A, Hagemann M. 2006. The plant-like C2 glycolate cycle and the bacterial-like glycerate pathway cooperate in phosphoglycolate metabolism in cyanobacteria. Plant Physiol 142:333–342. http://dx.doi.org/10.1104/pp.106.082982.