

Ankyrin Repeat-Rich Membrane Spanning (ARMS)/Kidins220 Scaffold Protein Regulates Neuroblastoma Cell Proliferation through p21

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Cell proliferation is tightly controlled by the cell-cycle regulatory proteins, primarily by cyclins and cyclin-dependent kinases (CDKs) in the G₁ phase. The ankyrin repeat-rich membrane spanning (ARMS) scaffold protein, also known as kinase D-interacting substrate of 220 kDa (Kidins 220), has been previously identified as a prominent downstream target of neurotrophin and ephrin receptors. Many studies have reported that ARMS/Kidins220 acts as a major signaling platform in organizing the signaling complex to regulate various cellular responses in the nervous and vascular systems. However, the role of ARMS/Kidins220 in cell proliferation and cell-cycle progression has never been investigated. Here we report that knockdown of ARMS/Kidins220 inhibits mouse neuroblastoma cell proliferation by inducing slowdown of cell cycle in the G₁ phase. This effect is mediated by the upregulation of a CDK inhibitor p21, which causes the decrease in cyclin D1 and CDK4 protein levels and subsequent reduction of pRb hyperphosphorylation. Our results suggest a new role of ARMS/Kidins220 as a signaling platform to regulate tumor cell proliferation in response to the extracellular stimuli.

INTRODUCTION

The ankyrin repeat-rich membrane spanning (ARMS) protein (Kong et al., 2001), also known as a kinase D-interacting substrate of 220 kDa (Kidins 220) (Iglesias et al., 2000), has been previously identified as a downstream interactor of neurotrophin and ephrin receptors (reviewed in Neubrand et al., 2012). As a transmembrane scaffold protein, ARMS/Kidins220 is highly expressed in the nervous system, such as hippocampus, olfactory bulb, motor neurons in the spinal cord (Iglesias et al., 2000; Kong et al., 2001). ARMS/Kidins220 is tyrosine phosphorylated specifically by Trk and Eph receptors (Kong et al., 2001). It also

directly interacts with all three Trk receptors and with p75NTR to form a ternary complex (Arevalo et al., 2004; Chang et al., 2004). Accordingly, ARMS has many important functions in the nervous system, such as neuronal survival (Duffy et al., 2011; Lopez-Menendez et al., 2009; Sniderhan et al., 2008), cytoskeletal remodeling and neuronal differentiation (Bracale et al., 2007; Higuero et al., 2010; Neubrand et al., 2010; Park et al., 2010), dendrite and synapse development (Chen et al., 2012; Cortes et al., 2007; Wu et al., 2009) and neuronal activity and synaptic plasticity (Arevalo et al., 2010; Cesca et al., 2012; Sutachan et al., 2010; Wu et al., 2010).

Interestingly, ARMS/Kidins220 acts as a signaling platform for sustained activation of the mitogen-activated protein kinase (MAPK) signaling through the recruitment of CrkL-C3G complex and Rap1 (Arevalo et al., 2004; 2006). Since MAPK signaling is a major survival pathway and Trk receptors were initially identified as oncogenes (Martin-Zanca et al., 1986), ARMS/Kidins220 has also been suggested as a potential oncogene (Neubrand et al., 2012). ARMS/Kidins220 is highly expressed in melanoma, and it contributes to melanoma progression by sustained MAPK signaling and inhibiting stress-induced apoptosis (Liao et al., 2007). Recently, it is also reported that ARMS/Kidins220 is expressed in human neuroblastoma cells and stabilizes NGF-induced survival signaling (Rogers and Schor, 2013).

Although ARMS/Kidins220 is involved in tumor cell survival and contributes to tumor progression, its role in cell proliferation and cell-cycle progression has never been investigated. In this study, we report that knockdown of ARMS/Kidins220 in mouse neuroblastoma cells induces a slowdown of cell cycle in the G₁ phase, which results in a decrease in cell proliferation. The G₁ phase arrest is associated with the decreased expressions of cyclin D1 and cyclin-dependent kinase 4 (CDK4) and hyperphosphorylated pRb, which is mediated by p21 upregulation. Thus, the p21-cyclinD1/CDK4 signaling pathway plays a critical role in ARMS/Kidins220-dependent cell-cycle progression. Taken together, our study reveals a novel mechanism by which ARMS/Kidins220 scaffold protein regulates tumor cell proliferation.

MATERIALS AND METHODS

Cell culture

Neuro2a mouse neuroblastoma cells were grown in Dulbecco's modified Eagle's medium (DMEM; Hyclone) supplemented with 10% fetal bovine serum (FBS; Gibco), 10 U/ml penicillin, and 10 g/ml streptomycin at 37°C with 5% CO₂.

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Antibodies

The following antibodies were used: rabbit polyclonal (clone 892) against the C-terminus of ARMS/Kidins220 (Kong et al., 2001); cyclin D1, cyclin A, cyclin E, CDK4, ERK, pERK and phospho-pRb (Ser780) (Cell Signaling); CDK2, p21, p16 and α -tubulin (Sigma).

Generation of ARMS/Kidins220 short hairpin RNA (shRNA) retroviral vectors and ARMS/Kidins220-knockdown Neuro2a cell lines

Retroviral vectors producing shRNA against ARMS/Kidins220 were constructed by inserting oligonucleotide sequence 5'-CACCGCCAGAGAGCAGATCAGTATTCGAAAATACTGATCTGCTCTCTGGC-3' (forward) and 5'-AAAAGCCAGAGAGCAGATCAGTATTTTCGAATACTGATCTGCTCTCTGGC-3' (reverse) (Li et al., 2008) into pSuper.retro vector (OligoEngine) as previously described (Brummelkamp et al., 2002). Briefly, the pSUPER.retro vector with shARMS sequences was co-transfected with VM- γ P and VSV-G plasmids into 293T cells using Lipofectamine (Invitrogen) according to the manufacturer's protocol. After 48 h, the culture media were filtered through a 0.45 μ m filter (Satorius) to obtain retroviral supernatants. To obtain highly concentrated virus stock, the supernatant was ultra-centrifuged at 250,000 \times g for 1.5 h at 4°C using a Beckman SW26 rotor. Virus pellets were re-suspended in a small amount (1% of original volume) of serum-free DMEM. Titration of virus was performed using NIH3T3 cells infected with 10-fold serial dilutions and virus titer was determined at day 5 post-inoculation by counting the number of puromycin (2 μ g/ml)-resistant colonies. To construct the control and ARMS/Kidins220-knockdown Neuro2a stable cell lines, Neuro2a cells were infected with retroviral supernatants containing 4 μ g/ml polybrene for 5 h. After adsorption, cells were allowed to recover for 48 h in DMEM containing 10% FBS and cultured in selection media containing puromycin (2 μ g/ml) for next 7 days.

Cell proliferation assay

For the cellular growth assay, cells were plated in 24-well plates (5×10^3 cells/well) and counted at the indicated time points after being trypsinized and incubated with trypan blue solution (Gibco). The cell proliferation was also measured using EZ-CyTox (Daeil Science, Korea) according to the manufacturer's protocol. Cells were plated in 96-well plates (3×10^3 cells/well), and 10 μ l of assay solution were added to each well. Cells were then incubated for 1.5 h at 37°C, and the absorbance was measured at 450 nm using a microplate reader (Bio-Rad).

Fluorescence-activated cell sorting analysis

The cell-cycle profile was measured by flow cytometry using propidium iodide (PI; Sigma) at indicated times (0, 24, and 48 h). Briefly, trypsinized cells were collected by centrifugation, washed twice with cold PBS, fixed with cold 70% methanol drop-wise while vortexing gently, and then left on ice at least 1 h. After washing twice with PBS, DNA was stained with 50 μ g/ml PI and 0.1 mg/ml RNase A (Sigma) for 30 min in the dark (Jadhav et al., 2007). The DNA content of the nuclei was analyzed using a FACS Calibur system (Becton Dickinson). Cell cycle distribution was analyzed with ModFit LT™ software (Becton Dickinson).

Immunoblotting

To synchronize cells in the Go phase, cells were cultured in serum-free DMEM for 24 h and then stimulated to enter the cell cycle in DMEM containing 10% FBS. For immunoblotting,

cell lysates were prepared at the indicated times (0, 4, 8 and 24 h). Briefly, cells were rinsed with ice-cold PBS twice, and were harvested in 10 mM Tris-HCl (pH 8.0), 150 mM NaCl, 1mM EDTA, and 1% NP-40 (TNE buffer) containing protease and phosphatase inhibitors. Equal amount of proteins (100 μ g/lane) were resolved by 6-20% gradient SDS-PAGE, and western blotting was performed with antibodies against different proteins.

Statistical analysis

Data were analyzed using Student's *t*-test and expressed as mean \pm standard error of mean (SEM). Probability (*P*) values < 0.05 were considered statistically significant.

RESULTS

Downregulation of ARMS/Kidins220 results in decreased neuroblastoma cell proliferation

The expression of ARMS/Kidins220 in melanoma cells has been shown to confer resistance to UVB-induced apoptosis by activating ERK signaling pathway (Liao et al., 2007). Despite the fact that ARMS is a potential oncogene and is involved in tumor cell survival signaling (Liao et al., 2007; 2011), little is known about its effects on tumor cell proliferation. Thus, in this study, we sought to examine the role of ARMS/Kidins220 in tumor cell proliferation.

First, we established ARMS/Kidins220-knockdown Neuro2a stable cell lines (ARMS-KD) using ARMS-specific shRNA (shARMS)-containing retroviral vectors. Control Neuro2a cells were infected with a virus containing vector alone. We confirmed that ARMS/Kidins220 expression was reduced to $35.9 \pm 2\%$ in ARMS-KD cells compared with the control cells (Fig. 1A). Next, we performed cell proliferation assay using trypan blue staining to investigate whether ARMS/Kidins220 can regulate proliferation in these cells. The control and ARMS-KD cells were synchronized in the Go phase by serum starvation for 24 h. They passed through the cell cycle after the addition of 10% FBS. Cell proliferation was inferred by the number of cells at the indicated times. As shown in Fig. 1B, there was no statistical difference in cell growth between the two cell lines on the 1st day. However, on the 2nd day, the number of cells in the control was significantly higher than the number of ARMS-KD cells. On the 4th day, there were 1.82 times more control cells than ARMS-KD cells (Control, $24 \times 10^3 \pm 0.24$ vs ARMS-KD, $13.8 \times 10^3 \pm 0.11$). The doubling times of control and ARMS-KD cells are approximately 1.88 days and 4.26 days, respectively.

To confirm the previous result, we also examined cell proliferation using EZ-CyTox assay. As expected from the results of the cell counting assay, ARMS-KD cells showed decreased proliferation compared to the control cells on the 2nd day, with maximal apparent differences on the 4th day (Control, 2.01 ± 0.08 vs ARMS-KD, 1.03 ± 0.07) (Fig. 1C). Taken together, these data suggest that the expression level of ARMS/Kidins220 influences on the proliferation of Neuro2a cells.

Downregulation of ARMS/Kidins220 results in a prolonged G1 phase of the cell cycle

To investigate whether the effect of ARMS/Kidins220 on neuroblastoma cell growth is due to alteration of the cell cycle, we analyzed cell-cycle phase progression using fluorescence-activated cell sorting (FACS) on the control and ARMS-KD cells (Fig. 2A). Cells were synchronized in the Go phase of cell cycle by serum starvation for 24 h. There were no significant differences in the proportion of both cells in the all phases of cell

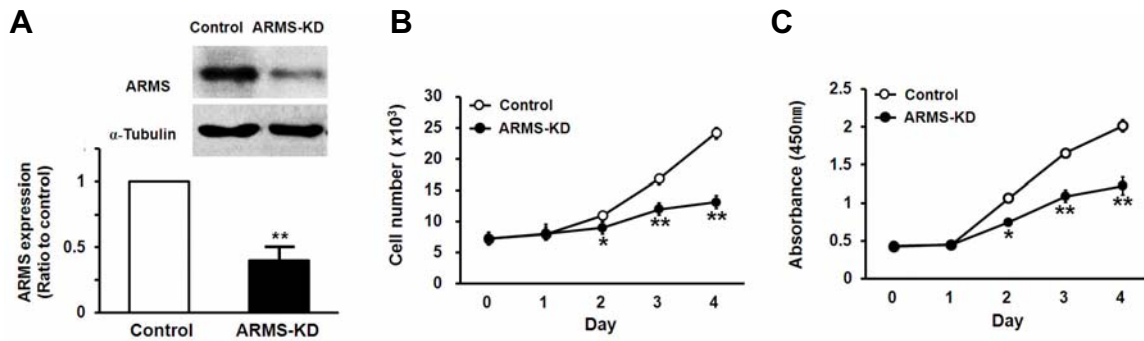


Fig. 1. Downregulation of ARMS/Kidins220 inhibits Neuro2a cell proliferation. (A) Western blot analysis of Neuro2a cell lysates after shControl and shARMS retroviral infection. To verify the reduction of ARMS/Kidins220 protein expression, Western blotting was performed with ARMS/Kidins220 (892) antibody and α -tubulin antibody. (B) Cell counting assay for cell proliferation. Cell proliferation was determined by counting the cell number with trypan blue staining at the indicated times. (C) Cell proliferation assay. On each day, the cells were incubated with EZ-CyTox solution for 1.5 h and the absorbance was measured using an ELISA reader at 450 nm. The values are represented as the mean \pm SEM of three independent experiments, and statistical significance is evaluated by Student's *t*-test (* p < 0.05, ** p < 0.01).

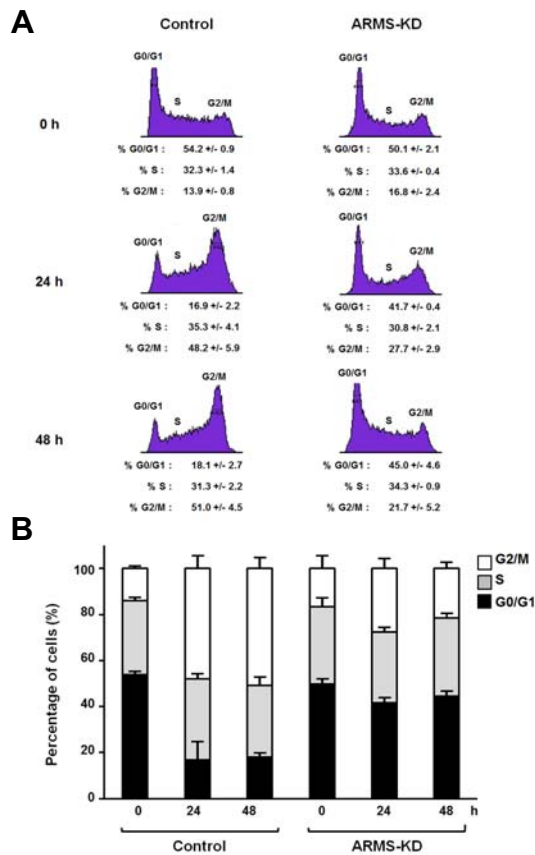


Fig. 2. Downregulation of ARMS/Kidins220 inhibits cell cycle progression in the G₀/G₁ phase. After Go synchronization in serum-free DMEM for 24 h, the control and ARMS-KD neuroblastoma cells were cultured with DMEM containing 10% FBS to stimulate re-entry into the cell cycle. (A) FACS analysis in the control and ARMS-KD cells. At each time point, cells were fixed, stained with propidium iodide (PI), and analyzed for DNA content by the flow cytometry. The distribution of cells in G₀/G₁, S, and G₂/M phase is indicated. (B) The bar graph shows the percentage of the cells in G₀/G₁, S, and G₂/M phase in the control and ARMS-KD cells. The values are represented as the mean \pm SEM of three independent experiments.

cycle at 0 h. However, at 24 h after the addition of 10% FBS, the proportion of the control cells in the G₀/G₁ phase significantly decreased from 54.2 \pm 0.9% to 16.9 \pm 2.2%, whereas the proportion of ARMS-KD cells in the G₀/G₁ phase decreased from 50.0 \pm 2.1% to 41.7 \pm 0.4% (Fig. 2B). At the same time, the proportion of control cells in the G₂/M phase dramatically increased from 13.9 \pm 0.8% to 48.2 \pm 5.9%, whereas the proportion of ARMS-KD cells in the G₂/M phase increased from 16.8 \pm 2.4% to 27.7 \pm 2.9% (Fig. 2B). These results suggest that downregulation of ARMS/Kidins220 induced a slowdown of cell cycle in the G₁ phase, leading to retarded cell growth.

Downregulation of ARMS/Kidins220 is associated with the decreased levels of cyclin D1 and CDK4

Cell growth is controlled by the expression and activity of cell-cycle regulatory proteins, such as cyclins and CDKs. D-type cyclins are important for cell-cycle progression through the G₁ phase (Sherr, 1995). Decreased cyclin D1 expression prevents cells from entering the S phase, leading to cell-cycle arrest in the G₁ phase (Baldin et al., 1993). Since downregulation of ARMS/Kidins220 induced G₁ phase arrest, we performed Western blotting to examine the expression of the G₁ phase regulatory protein, cyclin D1, at the different time points of cell cycle progression. Downregulation of ARMS/Kidins220 resulted in significant decreases in cyclin D1 protein levels at 8, 24 and 48 h, when compared with control cells (p < 0.01) (Figs. 3A and 3B). Although there was some decrease in cyclin D1 protein levels at 0 h in ARMS-KD cells, this decrease did not reach statistical significance.

To examine the effects of ARMS/Kidins220-knockdown on the other types of cyclins, we also analyzed the expression of cyclin E and cyclin A in both cells. Cyclin E is involved in the S phase, and cyclin A mainly functions in G₂/M phase (Furuno et al., 1999). In contrast to cyclin D1, there were no significant differences in cyclin E and cyclin A expressions between the control and ARMS-KD cells (Figs. 3A, 3C, and 3D).

We next examined the kinase partner of each cyclin at different time points. Downregulation of ARMS/Kidins220 resulted in significant decreases in the protein levels of CDK4, the kinase partner of cyclin D1, at the all-time points (p < 0.01) (Figs. 3A and 3E). Interestingly, even at 0 h, without cell-cycle progression, CDK4 protein level was significantly decreased in ARMS-KD cells. In contrast, the expression of CDK2, the kinase part-

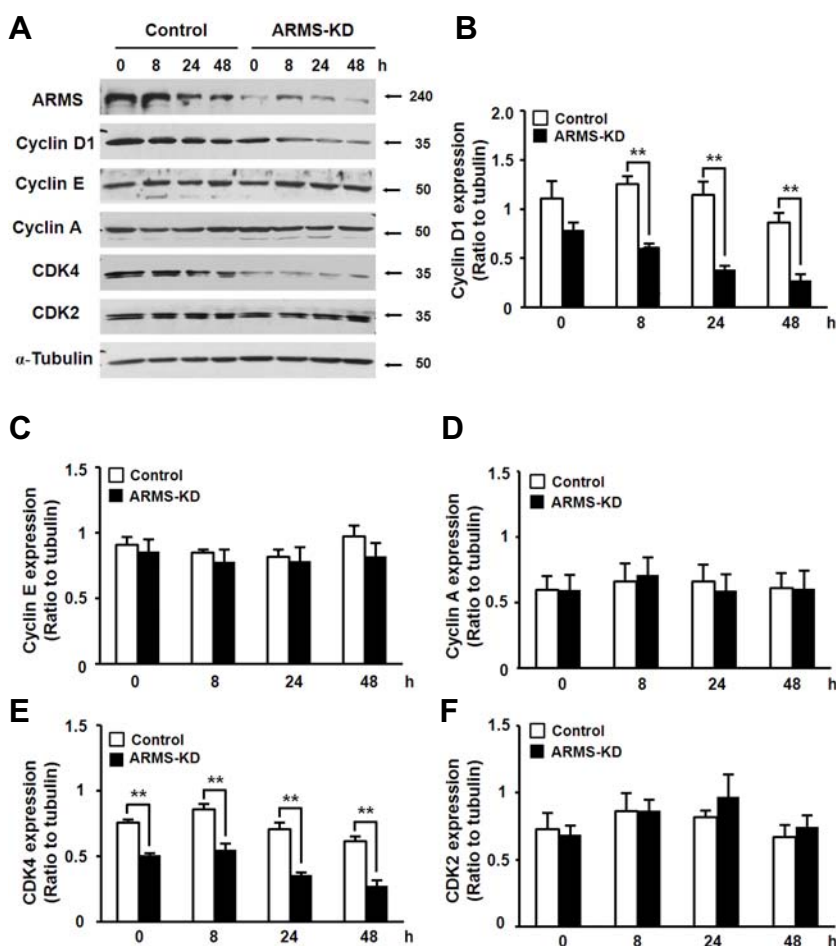


Fig. 3. Downregulation of ARMS/Kidins220 is associated with the decreased levels of cyclin D1 and CDK4. After Go synchronization in serum-free DMEM for 24 h, the control and ARMS-KD neuroblastoma cells were cultured with DMEM containing 10% FBS to stimulate re-entry into the cell cycle. (A) Western blot analysis of cyclins and CDKs in the control and ARMS-KD neuroblastoma cells at 0, 8, 24, 48 h after re-entry into the cell cycle. The loading control is α -tubulin. (B-F) Band intensities from three independent experiments were analyzed and normalized to α -tubulin. The values are mean \pm SEM and statistical significance was evaluated by Student's *t*-test (**p* < 0.05, ***p* < 0.01).

ner of cyclin E and cyclin A, was similar in both cells (Fig. 3F). These results indicate that knockdown of ARMS/Kidins220 is associated with the decreased expressions of cyclin D1 and CDK4 in the G₁ phase but has no effect on other cell cycle regulatory proteins.

Downregulation of ARMS/Kidins220 inhibits the hyperphosphorylation of pRb via reduction of cyclin D1 and CDK4

To confirm whether the reduction of cyclin D1 and CDK4 causes an effect on the downstream pRb pathway, we performed Western blotting with a phospho-pRb (Ser780) antibody, which specifically binds to CDK4-phosphorylated pRb. pRb hyperphosphorylation occurs at each G₁/S transition in both normal and cancer cells, and Ser780 of pRb is a site phosphorylated specifically by CDK4 (Chen et al., 1989; Jiao et al., 2006). As expected, the level of hyperphosphorylated pRb (ppRb) was lower in ARMS-KD cells than the control cells at 24 h although there is no difference between the two cells at earlier time points (0, 4, 8 h) (Fig. 4D). This result suggests that reduction of cyclin D1 and CDK4 decreases pRb phosphorylation, which in turn leads to a slowdown of cell cycle in the G₁ phase (Figs. 4A-4D).

To further confirm the role of ARMS/Kidins220 in regulation of cyclin D1/CDK4, we analyzed the expression of cyclin-dependent kinase inhibitors (CKIs) in G₁/S transition, p21 and p16. p21 is involved in regulating the function of cyclin D1/CDK4 and cyclin E/CDK2 complexes (El-Deiry et al., 1993;

Harper et al., 1993; Xiong et al., 1993a). Another CKI protein p16 specifically inhibits the function of cyclin D1/CDK4 in the G₁ phase (Bates et al., 1994; Xiong et al., 1993b). At all-time points in our experiment, the levels of p21 protein in ARMS-KD cells were significantly higher than the control cells (Fig. 4E). In contrast, there was no difference in the levels of p16 protein between the two cells (Fig. 4F). These results indicate that knockdown of ARMS/Kidins220 upregulates p21 protein expression, thus inhibiting cyclin D1/CDK4 function in the G₁ phase of cell cycle.

Next, we examined the p42- and p44-kDa extracellular signal-regulated kinases (ERKs) activation in ARMS-KD cells since ARMS/Kidins220 was proven to act as a platform for sustained Erk activation in neurons through the recruitment of CrkL (Arevalo et al., 2004; 2006) and ARMS/Kidins220-knockdown downregulates ERK activity in melanoma cells (Liao et al., 2007; 2011). Consistent with the previous studies, the level of pERK protein in ARMS-KD cells was decreased at early time points compared with the control cells, especially at 60 min (*p* < 0.05) (Supplementary Fig. 1). We also observed the level of pERK protein in both cells at 4, 8, 24 h. The level of pERK protein in ARMS-KD cells was significantly decreased at 24 h when compared with the control cells (*p* < 0.05) (Figs. 4A and 4G).

Taken together, our results suggest that downregulation of ARMS/Kidins220 inhibits cell-cycle progression at G₁/S check point through the upregulation of p21 protein in neuroblastoma cells (Fig. 5).

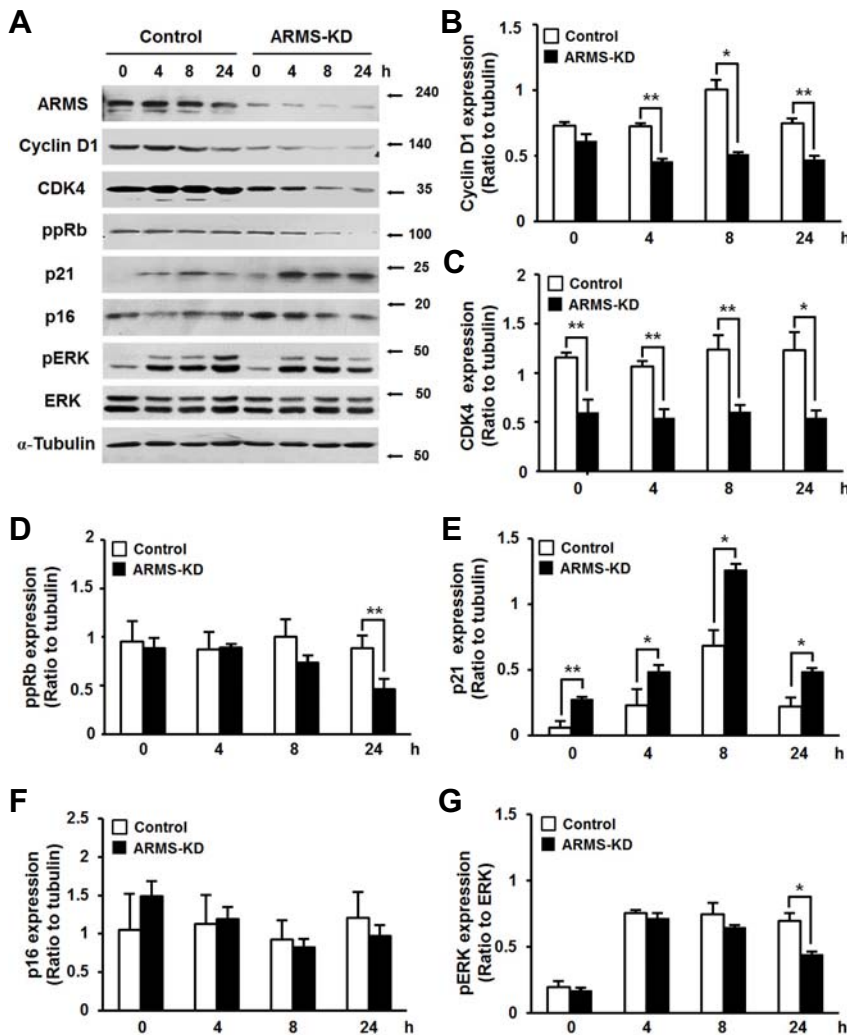


Fig. 4. Downregulation of ARMS/Kidins220 is associated with the decreased levels of ppRb and pERK, and the increased levels of p21. After G0 synchronization in serum-free DMEM for 24 h, the control and ARMS-KD neuroblastoma cells were cultured with DMEM containing 10% FBS to stimulate re-entry into the cell cycle. (A) Western blot analysis of cyclin D1, CDK4, ppRb, p21, p16, pERK and ERK in the control and ARMS-KD neuroblastoma cells at 0, 4, 8, 24 h after re-entry into the cell cycle. The loading control is α -tubulin. (B-G) The bar graphs show the changes in cyclin D1, CDK4, ppRb, p21, p16, pERK and ERK during cell cycle progression in the control and ARMS-KD cells. Band intensities from three independent experiments were analyzed and normalized to α -tubulin. The values are mean \pm SEM and statistical significance was evaluated by Student's *t*-test (**p* < 0.05, ***p* < 0.01).

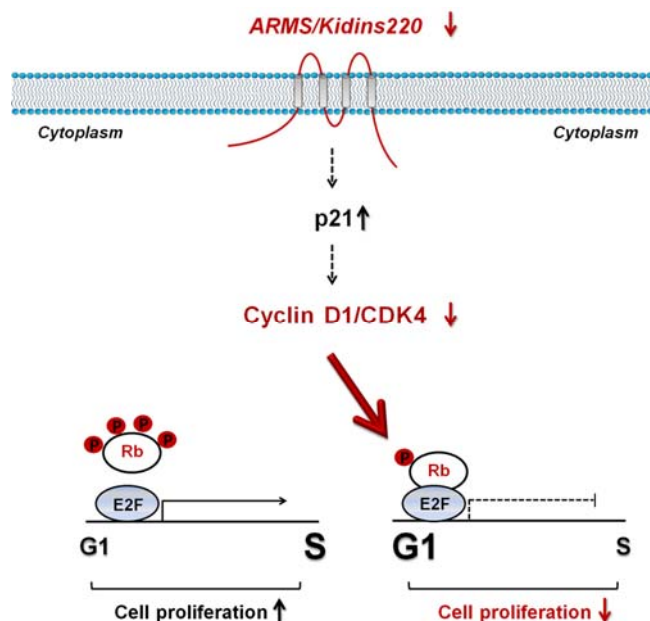


Fig. 5. Diagram depicting how ARMS/Kidins220 regulates cell-cycle progression. ARMS/Kidins220-knockdown upregulates p21 protein expression, which in turn reduces the levels of cyclin D1 and CDK4 in G₁ phase. Inhibition of cyclin D1 and CDK4 results in the increased level of hypophosphorylated pRb and subsequent sequestration of E2F transcription factors. The transcriptional activation of E2F-responsive genes is essential for DNA synthesis and cell cycle progression. Taken together, reduction of ARMS/Kidins220 expression upregulates p21 expression, leading to inhibition of cyclin D1 and CDK4, and reducing pRb hyperphosphorylation.

DISCUSSION

ARMS/Kidins220 is a transmembrane protein highly expressed in the nervous system, and has critical roles in neuronal differentiation, survival and development. As a multifunctional scaffold protein, it has various domains for protein-protein interactions (Iglesias et al., 2000; Kong et al., 2001). Thus, ARMS/Kidins220 has been suggested to play important roles in organizing the assembly of protein complexes of receptors and effectors which transduce the extracellular stimuli to signaling pathways inside the cell (Chao, 2003; Neubrand et al., 2012). Accordingly, there is an increasing body of evidence that ARMS/Kidins220 has various physiological functions not only in the nervous system but also in the vascular system (Cesca et al., 2012).

However, the pathological functions of ARMS/Kidins220 have not been well elucidated. Specifically, ARMS/Kidins220 acts as a major signaling platform for prolonged MAP kinase signaling by neurotrophins (Arevalo et al., 2004; 2006). ARMS/Kidins220-knockdown also decreases the levels of pMAPK and increases neuronal cell death mediated by excitotoxicity in cerebral ischemia (Lopez-Menendez et al., 2009).

ARMS/Kidins220 is colocalized with TrkA on the cell surface of sympathetic neurons in the developing nervous system (Kong et al., 2001). Interestingly, ARMS/Kidins220 expression is highly increased in human melanoma compared to benign skin lesions, and prevents stress-induced apoptosis in melanoma cells by activating ERK pathway (Liao, 2007). Its expression is also increased in human neuroblastoma, another neural crest tumor, and stabilizes NGF-induced survival signaling (Rogers and Schor, 2013). Therefore, similar to Trk receptors which have certain roles as oncogenes (Martin-Zanca et al., 1986), ARMS/Kidins220 may also have the potential to act as an oncogene (Neubrand et al., 2012).

One of the important functions of oncogenes is to increase cell proliferation by stimulating cell-cycle progression. As a tumor suppressor, pRb inhibits cell-cycle progression at G₁/S by blocking the activity of the E2F family of transcription factors, which activate the genes required for induction of S phase. Rb function is regulated through cell cycle by phosphorylation (Chen et al., 1989). Hyperphosphorylation of pRb by cyclin-CDK complexes in the G₁ phase inactivates its growth suppression function; consequently, the G₁ restriction point in cell cycle is bypassed (reviewed in Malumbres and Barbacid, 2009). p21, one of the CKIs in G₁/S transition, was originally identified as a downstream mediator of tumor suppressor p53 (El-Deiry et al., 1993; Harper et al., 1993; Xiong et al., 1993a). Upregulation of p21 inhibits the kinase activity of G₁ cyclin/CDK complexes, thus preventing the hyperphosphorylation of pRb and blocking cell-cycle progression (Dulic et al., 1994).

In this study, ARMS/Kidins220-knockdown induced a slowdown of cell cycle in the G₁ phase, leading to decreased neuroblastoma cell proliferation. Downregulation of ARMS/Kidins220 is associated with the decreased expressions of cyclin D1 and CDK4 in the G₁ phase but has no effect on other cell-cycle regulatory proteins, such as cyclin A, cyclin E and CDK2. Cyclin D is expressed at G₁/S transition where diverse mitogenic signaling cascades are converged to mediate engagement of the cell cycle machinery. Thus, cyclin D1 is often overexpressed and associated with tumorigenesis in many cancers (Masamha and Benbrook, 2009). In some cells, the downstream cyclin E can replace cyclin D to facilitate G₁/S transition. But, loss of cyclin D is sufficient to cause G₁ arrest in other cellular contexts (Masamha and Benbrook, 2009).

Interestingly, even without cell-cycle progression, CDK4 protein level was significantly decreased in ARMS-KD cells. Thus, it is likely that CDK4 activity is consistently inhibited in these cells. We further confirmed that the decreased cyclin D1/CDK4 levels were tightly associated with p21 upregulation at all-time points in our experiment. In contrast, the level of p16 protein, another CKI in G₁/S transition, was unchanged in ARMS-KD cells. Indeed, the level of ppRb was also lower in these cells than the controls at 24 h after the cell-cycle entry.

Consistent with the previous studies, we also observed that ARMS/Kidins220-knockdown decreases pERK level (Arevalo et al., 2004; Liao et al., 2007; 2011). Since ARMS/Kidins220 mediates prolonged MAPK signaling by interacting with CrkL (Arevalo et al., 2004; 2006), it will be interesting to examine which adaptor(s) or effector(s) is (are) involved in ARMS/Kidins220-p21 pathway for cell-cycle regulation. Identifying the specific domain of ARMS/Kidins220 involved in the regulation of p21 expression using the serial deletion constructs of ARMS/Kidins220 will be useful to further delineate this pathway. The other interesting question to address will be whether ARMS/Kidins220 is also involved in other tumor cells and normal cell proliferation by the same mechanism demonstrated in our study. Although the tumor suppressor role of p21 is still controversial (reviewed in Besson et al., 2008), it is clear that p21 inhibits cyclinD1/CDK4, which results in retarded cell proliferation in this study.

In summary, we demonstrate that ARMS/Kidins220 regulates the neuroblastoma cell proliferation through p21 protein. Our results suggest that ARMS/Kidins220 acts as an important platform in organizing the signaling complex to regulate cell-cycle progression in response to the extracellular stimuli. In addition, as a potential oncogene, ARMS/Kidins220 may serve as an important prognostic factor of tumors in the nervous system.

Note: Supplementary information is available on the Molecules and Cells website (www.molcells.org).

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