Molecular Therapy Methods & Clinical Development

Correction



Expression of Human ACE2 in Lactobacillus and Beneficial Effects in Diabetic Retinopathy in Mice

Amrisha Verma, Kang Xu, Tao Du, Ping Zhu, Zhibing Liang, Shengquan Liao, Juantao Zhang, Mohan K. Raizada, Maria B. Grant, and Qiuhong Li Correspondence: Qiuhong Li. E-mail: qli@ufl.edu https://doi.org/10.1016/j.omtm.2020.02.002

(Molecular Therapy: Methods & Clinical Development 14, 161-170; September 13, 2019)

Following publication, the authors realized that the design of CTB fusion constructs used in the study had not been sufficiently explained in the Materials and Methods. This information is now provided below and has been incorporated into the original online file.

Expression Vector Construction

The concept of using CTB fusion for systemic protein delivery by the oral route was originally reported by Limaye et al.⁸⁹ and has been used for oral delivery of a number of therapeutic proteins, including ACE2 and Ang-(1-7).^{35,90} A mutant form of CTB, which retains the binding to GM1-ganglioside for cellular uptake but lacks immunomodulatory and toxic activity,^{91,92} was used in this study. The codons of the CTB sequence were also extensively changed for optimal expression in *Lactobacillus*. The FC site was based on our previously published sequence,³⁶ which is an artificial FC site designed after careful comparison of available natural protein substrates of furin protease, with enhanced affinity for FC. The codons were also optimized for expression in *Lactobacillus* species. The DNA sequence for codon-optimized CTB for expression in *Lactobacillus* species is as follows: 5'-ATGATTAAGTTAAGTTAAGTTTGGTGTTTTTTTTTATCAGTTTTATCAGCTTACGCTCACGGTTACCACAAAACATTACTGATTTATGTGCTGAAATACCACAACACTCAAAATTCACACTTTAAAGATTAAGTTTAAGTTAAGTTAAGATTAAGTTTAACGATAAGATTTTTTCAAAGTTGAAGATTAAGTTAACGAAAACAGTTAAGGTGAAATACCACAACACTCAAAATTCACACTTTAAAGTTGAAGGTTGAAATACCACAACACTCAAAATTCAACGTTAAAGTTTAACGTTAACGTAAGGTTGAAAGGTTCAAGGTTGAAAAGAGGTGCTACTTTTAAGTTTAACGTTAACGTAAGGTTGAAAAGAGGCTAATTGAAGGTAAAGGATACTTTAACGTAACGATAACGATAAGGTTGAAAAGAGGCTAATTGAAGGTAAGGATACTTTAACGTAATGGCTAACTTTAACGTAAGGTTGAAAGGTTGAAAAGGTTGAAAGGTTGAAAGGTTAAGGTAAGGATACTTTAACGTAATGGCTAACTTAACGTAAGGTTGAAAAGAGGCTAATTGAAGGTAAGGATACTTTAACGTAATGGCTAAC-3'.

REFERENCES

- Limaye, A., Koya, V., Samsam, M., and Daniell, H. (2006). Receptor-mediated oral delivery of a bioencapsulated green fluorescent protein expressed in transgenic chloroplasts into the mouse circulatory system. FASEB J. 20, 959–961.
- 90. Shenoy, V., Kwon, K.C., Rathinasabapathy, A., Lin, S., Jin, G., Song, C., Shil, P., Nair, A., Qi, Y., Li, Q., et al. (2014). Oral delivery of Angiotensin-converting enzyme 2 and Angiotensin-(1-7) bioencapsulated in plant cells attenuates pulmonary hypertension. Hypertension 64, 1248–1259.
- 91. Rodighiero, C., Fujinaga, Y., Hirst, T.R., and Lencer, W.I. (2001). A cholera toxin B-subunit variant that binds ganglioside G(M1) but fails to induce toxicity. J. Biol. Chem. 276, 36939–36945.
- 92. Aman, A.T., Fraser, S., Merritt, E.A., Rodigherio, C., Kenny, M., Ahn, M., Hol, W.G., Williams, N.A., Lencer, W.I., and Hirst, T.R. (2001). A mutant cholera toxin B subunit that binds GM1- ganglioside but lacks immunomodulatory or toxic activity. Proc. Natl. Acad. Sci. USA 98, 8536–8541.