# Review Article Roles of Identified Long Noncoding RNA in Diabetic Nephropathy

# Yan Li,<sup>1</sup> Keyang Xu,<sup>2</sup> Kechen Xu,<sup>3</sup> Sixiang Chen,<sup>2</sup> Yifang Cao,<sup>4</sup> and Huakui Zhan <sup>1</sup>

<sup>1</sup>The First Clinical Medical College, Chengdu University of Traditional Chinese Medicine, Chengdu, 610075 Sichuan, China <sup>2</sup>Zhejiang Chinese Medical University, Hangzhou, 310053 Zhejiang, China

<sup>3</sup>The Fourth Affiliated Hospital, Zhejiang University School of Medicine, Yiwu, 322000 Zhejiang, China

<sup>4</sup>The First Hospital of Jiaxing, Jiaxing, 314001 Zhejiang, China

Correspondence should be addressed to Huakui Zhan; zhanhuak@qq.com

Received 11 December 2018; Accepted 28 January 2019; Published 12 February 2019

Academic Editor: Akira Sugawara

Copyright © 2019 Yan Li et al. This is an open access article distributed under the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Diabetes mellitus is the leading chronic disease in the world, and diabetic nephropathy (DN) as one of its complications could increase the mortality. The development of DN is associated to abnormal hemodynamic factors like cytokine networks and the intervention of metabolic risk factors like blood pressure, blood glucose, and blood lipid. However, the pathogenesis of DN is still poorly understood. Although glucose-lowering drugs and insulins have significant effects on blood glucose, the fluctuation of blood glucose or other risk factors could continuously damage the kidney. Recent studies reported that the progression of DN is closely related to the expression of long noncoding RNA (lncRNA), which is important for the early diagnosis and targeted intervention of DN. In this review, we briefly summarize the published studies on the functions and potential mechanism of reported lncRNA in the regulation of DN.

# 1. Diabetic Nephropathy

Diabetic nephropathy (DN) is a progressive kidney disease that develops consequently to diabetes and is the important cause of chronic renal disease worldwide [1]. And DN accounts for approximately 40% of diagnosed end-stage kidney failure [2]. The early features of DN include glomerular mesangial expansion, hypertrophy, and increased renal accumulation of extracellular matrix (ECM) proteins such as collagens and fibronectin, as well as podocyte effacement [3, 4]. Albuminuria is used to stage DN and is regarded as a biomarker for diagnosis [5, 6]. But the typical pathological characteristics of DN can also be characterized by excessive proliferation of ECM and diffuse glomerular basement thickening of mesangial cells (MCs), which can eventually lead to glomerular sclerosis and renal interstitial fibrosis when exposed to high glucose [7, 8], because MCs can secrete various cytokines, such as transforming growth factor  $\beta$ 1 (TGF- $\beta$ 1), collagen (COL), and fibronectin (FN) [9]. In addition, genetic factors are also important for disease risk [10, 11]; recent evidence has shown the involvement of epigenetic factors, such as DNA methylation, histone

posttranslational modifications, microRNAs (miRNAs), and lncRNAs which are involved in the development of renal diseases, including DN [11]. It has been reported that many factors which are crucially linked with the progress of DN, such as inflammation, oxidative stress, activated hexose, renal ECM, and the polyol pathway, are related to epigenetic factors [5].

# 2. Long Noncoding RNA (IncRNA)

lncRNA was first described in the large-scale sequencing of full-length eDNA library in mice in 2002. lncRNA is a group of transcription materials with >200 nucleotides, which lacks a specific complete open reading frame and has no protein-coding function [12]. lncRNA accounts for 80% in the whole mammalian genome transcripts [12]. The majority of lncRNAs are produced by the transcribing of RNA polymerase [13]. Similar to mRNA, lncRNA is commonly expressed in eukaryotic genomes with 5' cap structures and a poly adenosine tail [14]. According to the location of lncRNA and proximal protein-coding genes in the genome, lncRNA can be classified into six categories: exon sense overlapping,

intron sense overlapping, intronic antisense, natural antisense, and bidirectional and intergenic lncRNA [15]. Recent studies have shown that lncRNA regulates gene expression on a variety of levels, mainly including epigenetic transcription and posttranscriptional modification, and the regulation modes include chromosomal modification, transcriptional interference, or transcriptional activation [16]. Expression disorders of lncRNA are found in many types of tumors and neurological and cardiovascular diseases [17, 18]. Accumulating evidence has indicated the significant roles of lncRNAs in the pathophysiology of DN, and the crosstalk between lncRNA and DN was widely reported in recent years [19-21]. lncRNAs are involved in the procession of DN through regulating many important factors, such as pathologic process in MCs, reactive oxidative products (ROS), mechanisms involving ECM accumulation, and actions of miRNAs [1, 22, 23]. For example, ENSMUST00000147869 is downregulated in diabetic renal tissue, and mesangial cell proliferation and fibrosis are significantly enhanced through silencing its expression [24]. ROS-induced expression of ASncmtRNA-2 may contribute to DN fibrosis through regulating TGF- $\beta$ 1 [25]. CYP4B1-PS1-001 is significantly downregulated in early DN, and it can significantly inhibit the proliferation and fibrosis of MCs through inducing its overexpression [26]. NR\_033515 promoted cell proliferation, fibrogenesis, and the EMT (epithelial-mesenchymal transition) process by miR-743b-5p [1]. Therefore, abnormal expression of lncRNA plays key roles in the occurrence and development of DN.

#### 3. IncRNA Upregulated in DN

3.1. LncPVT1. PVT1 (plasmacytoma variant translocation 1) is a lncRNA (1.9 kb) that encodes a number of alternative transcripts. When it is amplified and overexpressed, it will increase cell proliferation and inhibit apoptosis [27]. PVTI is the first ncRNA reported to associate with kidney disease [28], which is highly expressed in human renal MCs under a high-glucose condition and significantly promotes the expression of fiber connection protein (FN1), type IV collagen, TGF- $\beta$ 1, and type 1 plasminogen activator inhibitor (PAI-1). More importantly, the deletion of PVT1 gene in MCs significantly reduced the expression of major ECM proteins and their regulatory factors, including FN1, COL4A1, TGF- $\beta$ 1, and PAI-1 [1, 29], while PAI-1 is the main inhibitor of glomerular ECM degradation [30]. It indicates that PVT1 may participate in the genesis and development of DN by regulating the accumulation of ECM. Some studies also suggest that some of the effects of PVT1 on ECM factors may be mediated through the actions of miRNAs, such as miR-1207-5p and miR-1207-3p [1].

3.2. *lncRNA MALAT1*. Metastasis-associated lung adenocarcinoma transcript 1 (MALAT1) is broadly expressed in mammalian tissues including the kidney and in tumors [31]. And MALAT1 is also aberrantly upregulated in early DN [32].  $\beta$ -Catenin is a key mediator in the WNT signaling pathway, which can contribute to podocyte malfunction and albuminuria as well as kidney fibrosis [33]. MALAT1 could promote the translocation of  $\beta$ -catenin into the nuclei via enhancing serine/arginine splicing factor 1, and nuclear accumulation of  $\beta$ -catenin can cause podocyte damage and eventually lead to DN [20, 34, 35]. In addition, MALAT1 is a real culprit as an initiator of inflammation and oxidative stress, which can regulate glucose-induced upregulation of inflammatory mediators IL-6 and TNF- $\alpha$  through activation of serum amyloid antigen 3 (SAA3) [36]. Such changes may influence endothelial stability which is essential for all organs and for macro- and microvessels, which in the end leads to DN [37–39]. Furthermore, MALAT1 regulates renal tubular epithelial pyroptosis by modulated miR-23c targeting of ELAVL1 in DN [40]. Therefore, MALAT1 may be a potential therapeutic target for DN.

LincRNA *Gm4419*. Gm4419 (Ensembl ID 3.3. ENSMUST00000180671) is a LincRNA, which is located in chromosome 12 (Chr12:21417911-21419803, 1730 bp) [41], and it is a regulator of the transcription factor nuclear factor kappa light-chain enhancer of activated B cells (NF-κB), which is a crucial inflammatory stimulus for DN [42]. Gm4419 can directly interact with p50 to regulate the NF- $\kappa$ B/NLRP3 inflammasome signaling pathway and mediate inflammatory molecular expressions in MCs, and it is associated with the development of inflammation, fibrosis, and proliferation of MCs with high glucose [23]. The silencing of Gm4419 expression leads to significant inhibition of cell inflammation, fibrosis, and proliferation in MCs with high-glucose conditions [23]. Thus, Gm4419 may have a functional role in DN inflammation through NF-κB/NLRP3 inflammasome signaling and may act as a novel and specific therapeutic target for DN.

3.4. IncRNA GM5524. Cellular autophagy and apoptosis are essential for the maintenance of normal tissue homeostasis under physiological conditions [43]. The disorder of cellular autophagy and apoptosis has been reported in diabetes and its complications [44]. Gm5524 is significantly upregulated in DN tissues and podocytes under high-glucose conditions [45]. Gm5524 may be involved in DN by affecting these two processes: the first process is Gm5524 having effects on apoptosis and autophagy-associated factors through inhibiting antiapoptotic Bcl2 protein expression. And proapoptotic protein Bax expression is increased in Gm5524 knockdown podocytes [46, 47]. The second process is Gm5524 which promotes the development of DN by activating the LC3/ATG signaling pathway, which is a well-established biochemical assay to determine the activation of autophagy [45]. Thus, Gm5524 may further the understanding of the involvement of lncRNAs in DN.

3.5. IncRNA NR\_033515. NR\_033515 is significantly upregulated in serum of DN patients, and the expression level is related to the different stages of DN and positively associated with diagnostic markers of DN (KIM-1 and NGAL). Overexpression of NR\_033515 promotes MC proliferation and inhibits MC apoptosis. And it also increases the expression of proliferation-related genes (PCNA and cyclin D1), fibrogenesis-related gene proteins (P38, ASK1, fibronectin, and  $\alpha$ -SMA), and EMT biomarkers (E-cadherin and vimentin) by regulating miR-743b-5p expression [19]. Thus, NR\_033515 may be a pivotal target for the early diagnosis and treatment of DN.

3.6. IncRNA Erbb4-IR. Erbb4-IR is located within the intron region between the first and second exons of ErBb4 gene on chromosome 1 of the mouse genome. A pathogenic role of Erbb4-IR is revealed in human T2DN tissues, and its molecular mechanism is also elucidated in a novel T2DN mouse stain in S3KO db/db mice [48]. The functional role of Erbb4-IR in T2DN is revealed by kidney-specific silencing of Erbb4-IR to protect against the development of T2DN such as elevated microalbuminuria, serum creatinine, and progressive renal fibrosis in db/db mice. In addition, Erbb4-IR can directly inhibit the transcription of renoprotective miR-29b, and then TGF- $\beta$ /Smad3 signaling is activated. Therefore, renal fibrosis and renal dysfunction are largely promoted by Erbb4-IR during the progression of T2DN [49]. Thus, Erbb4-IR may represent a precise therapeutic target for DN.

3.7. IncRNA ASncmtRNA-2. Antisense mitochondrial noncoding RNA-2 (ASncmtRNA-2) is a mitochondrial lncRNA that is expressed in the mitochondria and exported to the nucleus [50, 51]. Previous studies have revealed that it is involved in the tumorigenesis and mitochondrial retrograde signaling pathways [51]. Notably, it has been demonstrated that ASncmtRNA-2 is overexpressed during aging and replicative senescence in human endothelial cells [52]. And ASncmtRNA-2 potentially serves a role in physiological oxidative stress and overproduction of oxidative products such as ROS, which can induce injury to the human kidneys through the following mechanisms: (i) inducing lipid peroxidation, protein crosslinking, and the formation of DNA adducts, leading to tissue damage; (ii) inducing direct damage to cellular DNA; and (iii) activating multiple cellular signaling pathways, including NF- $\kappa$ B and TGF- $\beta$ 1. These mechanisms induce further generation of ROS, synthesis, secretion of cytokines, and deposition of ECM components, which induce more severe damage to the kidneys. Therefore, ASncmtRNA-2 can be a novel method to regulate ROS generation to reduce renal damage in DN in a clinical setting [25, 53–55].

3.8. lncRNA Lnc-MGC. Lnc-MGC can serve as a scaffold for a cluster of 40 miRNAs and appears to induce features of early DN [56]. Lnc-MGC can regulate megacluster (MGC), and its 3' region overlaps with Mirg, and the middle region with Gm2922, other ncRNAs. Lnc-MGC is regulated by an endoplasmic reticulum (ER) stress-related transcription factor, CHOP (C/EBP homologous protein), via TGF- $\beta$ 1-dependent and TGF- $\beta$ 1-independent pathways [57]. ER stress has been observed to be increased in patients with progressive DN, and expression of renal CHOP and albuminuria is significantly increased in aged diabetic mice by promoting ER stress [58, 59]. Furthermore, a chemically modified oligonucleotide (gapmer) targeting Lnc-MGC can inhibit cluster miRNAs and then decrease protein synthesis, ER stress,

glomerular ECM, and hypertrophy in diabetic mice and human DN tissues [57]. These results demonstrate the translational implications of targeting Lnc-MGC for controlling DN progression.

#### 4. IncRNAs Downregulated in DN

4.1. lncRNA TUG1. lncRNA taurine-upregulated gene 1 (TUG1) was firstly identified as a part of photoreceptors and retinal development in mouse retinal cells, a lncRNA located at chromosome 22q12 [60]. TUG1 was considered to be involving in regulating carcinogenesis in several malignant tumors, and it also has been reported to play a key role in the progression of DN [61]. TUG1 is significantly repressed in the podocytes of diabetic mice by rescuing PPARy coactivator  $\alpha$  (PGC-1 $\alpha$ ) expression, which is an important member of the nuclear receptor superfamily and well known to have an important role in mitochondrial bioenergetics and respiration [62, 63], and ameliorating the courses of DN, for regulating glomerular MCs proliferation, cell cycle, and diabetic glomerular ECM synthesis [64–66]. TUG1 acts as an endogenous sponge of miR-377 and downregulates miR-377 expression levels and thereby can relieve the inhibition of its target gene PGC-1 $\alpha$  and alleviates ECM accumulation and cytokine secretion in MCs, including PAI-1, TGF- $\beta$ 1, FN, and collagen IV (Col IV) under high glucose [67]. Overall, TUG1 provides a novel insight of DN pathogenesis.

4.2. IncRNA MIAT. Myocardial infarction-associated transcript (MIAT), also known as retinal noncoding RNA 2 (RNCR2), is identified in myocardial infarction [68]. Dysfunction of kidney tubules in the tubular system of a diabetic kidney is proposed as the initial event in the development of DN [69]. In diabetic rats, MIAT shows the lower level and its expression is negatively correlated with serum creatinine and BUN [70]. MIAT can regulate proximal convoluted tubule cell viability via stabilizing nuclear factor erythroid 2-related factor 2 (Nrf2) expression, which is the key molecule of cellular defense against high blood glucose-induced oxidative stress and genotoxicity of cells. And Nrf2 can pathologically and functionally protect the kidney against diabetic damage [71, 72]. Interestingly, expression of Nrf2 can be enhanced by MIAT overexpression in 45 mM glucoseincubated renal tubular epithelial cell line (HK-2 cells) [70]. In summary, the data suggest that MIAT/Nrf2 served as an important signaling pathway for DN and it might be the potential therapeutic to reduce the burden of this disease.

4.3. *lncRNA CASC2*. Cancer susceptibility candidate 2 (CASC2) has been showed to have critical functions in tumorigenesis [73]. Recently, it was reported that the low expression of CASC2 has diagnostic values in serum and renal tissues for diabetes complicated with chronic renal failure [74]. A follow-up showed that patients with low serum level of CASC2 had significantly higher incidence of chronic renal failure by inhibiting the JNK pathway, which is common in patients with DN. Overexpression of CASC2 significantly inhibited the apoptosis of podocytes. In addition,

treatment with a JNK activator significantly reduced the inhibitory effects of CASC2 overexpression on apoptosis of podocytes [75, 76]. Furthermore, the ROC curve analysis showed that the CASC2 level in renal tissues and serum is effective in diagnosing type 2 diabetes complicated with chronic renal failure [74]. Thus, CASC2 may serve as a predictive factor and target for the treatment and prevention of DN with chronic renal failure.

4.4. IncRNA ENSMUST00000147869. ENSMUST00000147869 has been found to be downregulated in DN, proliferation and fibrosis indexes are reversed in MCs with the action of Cyp4a12a, a neighboring gene locus to ENSMUST00000147869, and it is the predominant 20hydroxyeicosatetraenoic acid synthase involved in determining sex- and strain-specific differences in susceptibility to hypertension and other cardiovascular diseases [77]. Cyp4a12a is a member of Cyp4a isoforms. CYP4 proteins metabolize fatty acids, eicosanoids, and vitamin D and are important for chemical defense, and the production of kidney CYP4 arachidonic acid metabolites can contribute to the abnormalities in renal function [78]. Downregulated Cyp4a12a is defined as a target gene, which can be recruited during ENSMUST00000147869 overexpression [77]. ENSMUST00000147869 can affect the synthesis of ECM and dramatically decreased the levels of fibronectin and Col IV in MCs under a high-glucose condition [24]. So the overexpression of ENSMUST00000147869 can significantly reduce the expression of the proliferation index (PCNA and cyclin D1) and fibrosis index (collagen I and FN) in MCs, as well as the growth rate. Thus, intergenic IncRNA ENSMUST00000147869 with nearby Cyp4a12a can be regarded as the potential therapeutic target and molecular biomarker for DN.

4.5. LincRNA 1700020I14Rik. 1700020I14Rik is located in chromosome 2 (Chr2: 119594296-119600744) [79], which has been found to be downregulated and acts as an endogenous RNA to regulate the miRNAs in DN. The induced overexpression of 1700020I14Rik can reduce the expression of miR-34a-5p via the silent information regulator T1/hypoxia-inducible factor-1 $\alpha$  (Sirt1/HIF-1 $\alpha$ ) signal pathway and eventually promotes proliferation and fibrosis in MCs [22]. Intriguingly, it has been reported that Sirt1 is a direct target of miR-34a-5p [22, 42], so the Sirt1/HIF-1 $\alpha$  signaling pathway plays a significant role in the proliferation and fibrosis of DN [80]. However, knockdown of 1700020I14Rik will reverse the upper processes. Furthermore, the expressions of renal fibrosis genes including TGF- $\beta$ 1, FN, and Col IV also decreased by induced overexpression of 1700020I14Rik [22]. These results provide new insights into the regulation between 1700020I14Rik and miR-34a-5p/Sirt1/HIF-1α signaling pathway during the progression of DN.

4.6. *lncRNA CYP4B1-PS1-001*. CYP4B1-PS1-001 is located within a cluster of genes on chromosome 4 related to cytochrome P450 (CYP450) and is important in many reactions involving drug metabolism and synthesis of cholesterol, steroids, and other lipids [81]. CYP4B1-PS1-001 is significantly downregulated in response to early DN. While overexpression of CYP4B1-PS1-001 can inhibit proliferation and fibrosis of MCs due to an interaction with nucleolin (NCL). Furthermore, degradation of CYP4B1-PS1-001-associated NCL is mediated by a ubiquitin proteasome-dependent pathway [26]. The results show that overexpression of CYP4B1-PS1-001 decreases the levels of FN and collagen I as the major components of ECM in MCs under a highglucose condition [81]. Overall, CYP4B1-PS1-001 could provide a potential therapeutic target and molecular biomarker in DN pathogenesis.

4.7. *lncRNA Gm15645*. Gm15645 is significantly downregulated in DN tissue podocytes in a high-glucose condition. The mechanism of Gm15645 is opposite with that of Gm5524, which may affect podocyte apoptosis and autophagy via regulation of the Bcl2/Bax and LC3/ATG pathways in DN [45].

4.8. IncRNA LINC01619. LINC01619 can regulate miR-27a/-FoxO1 (forkhead box protein O1) and endoplasmic reticulum (ER) stress-mediated podocyte injury in DN by serving as a "sponge" for miR-27a. FOXO1 is the earliest discovered transcription factor of the FOXO subfamily and plays an important physiological function in proliferation, apoptosis, differentiation, oxidative stress, and other biological processes involved in cell metabolic diseases such as diabetes [82]. FOXO1 abolishment not only upregulates CHOP and GRP78 expression in podocytes but also increases podocyte foot process effacement [83]. Thus, the recovery of LINC01619 can alleviate oxidative stress and podocyte injury, and the silence of LINC01619 can induce oxidative stress and podocyte injury, diffuse podocyte foot process effacement, and decrease renal function [83]. Downregulation of LINC01619 contributes to proteinuria and declines renal function in DN patients; therefore, targeting LINC01619 may be a therapeutic approach for preventing DN.

#### 5. Conclusion

lncRNAs play a crucial role in the pathogenesis and progression of DN. The upregulated lncRNAs have a common function: they can promote the excessive proliferation of ECM, glomerular sclerosis and renal interstitial fibrosis, and inflammation and thickening of MCs. On the contrary, the downregulated lncRNAs appear to function as the protective factor against DN (Figure 1). In the previous studies, most studies of DN were focused on the functions of miRNAs. The regulation of lncRNA in DN is complex, involving interactions among multiple molecules and signaling pathways. Detailed functional verification in multiple models of DN is essential for the identification of lncRNA with clinical application potentials. Furthermore, recent studies suggested that N6-methyladenosine (m<sup>6</sup>A) modification of lncRNAs may play key roles in many diseases, such as cancer, leukaemia, Parkinson's disease, obesity, and diabetes [84, 85]. Regulatory roles between lncRNAs and m<sup>6</sup>A methylation in DN need to be further clarified.

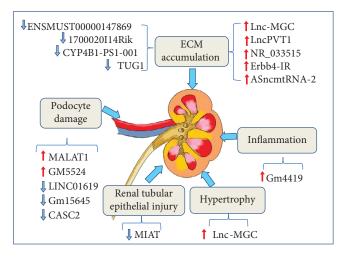


FIGURE 1: The roles of lncRNAs in diabetic nephropathy. Dysregulation of upregulated lncRNA (LncPVT1, MALAT1, Gm4419, GM5524, NR\_033515, Erbb4-IR, ASncmtRNA-2, and Lnc-MGC) and downregulated lncRNA (TUG1, MIAT, CASC2, ENSMUST00000147869, 1700020114Rik, CYP4B1-PS1-001, Gm15645, and LINC01619) involved in ECM maintenance and regulation of inflammation and podocyte damage, as described in the text. Red arrows indicate the damaging function of lncRNA upregulated in DN, and blue arrows indicate the protective functions of lncRNA downregulated in DN.

#### **Conflicts of Interest**

The authors declare that they have no conflicts of interest.

#### **Authors' Contributions**

Yan Li and Keyang Xu contributed equally to this article.

### Acknowledgments

This work was supported by the Sichuan Science and Technology Department (grant number 2016Jy0156, to Huakui Zhan) and the Jiaxing Science and Technology Project (grant number 2018AD32091, to Yifang Cao).

#### References

- M. L. Alvarez, M. Khosroheidari, E. Eddy, and J. Kiefer, "Role of microRNA 1207-5P and its host gene, the long non-coding RNA Pvt1, as mediators of extracellular matrix accumulation in the kidney: implications for diabetic nephropathy," *PLoS One*, vol. 8, no. 10, article e77468, 2013.
- [2] D. Choudhury, M. Tuncel, and M. Levi, "Diabetic nephropathy—a multifaceted target of new therapies," *Discovery Medicine*, vol. 10, no. 54, pp. 406–415, 2010.
- [3] Y. S. Kanwar, L. Sun, P. Xie, F. Y. Liu, and S. Chen, "A glimpse of various pathogenetic mechanisms of diabetic nephropathy," *Annual Review of Pathology*, vol. 6, no. 1, pp. 395–423, 2011.
- [4] M. Kato and R. Natarajan, "MicroRNAs in diabetic nephropathy: functions, biomarkers, and therapeutic targets," *Annals of the New York Academy of Sciences*, vol. 1353, no. 1, pp. 72–88, 2015.

- [5] M. K. Arora and U. K. Singh, "Molecular mechanisms in the pathogenesis of diabetic nephropathy: an update," *Vascular Pharmacology*, vol. 58, no. 4, pp. 259–271, 2013.
- [6] M. Kitada, K. Kanasaki, and D. Koya, "Clinical therapeutic strategies for early stage of diabetic kidney disease," *World Journal of Diabetes*, vol. 5, no. 3, pp. 342–356, 2014.
- [7] R. A. Risdon, J. C. Sloper, and H. E. De Wardener, "Relationship between renal function and histological changes found in renal-biopsy specimens from patients with persistent glomerular nephritis," *The Lancet*, vol. 2, no. 7564, pp. 363–366, 1968.
- [8] M. Mourtada-Maarabouni and G. T. Williams, "Role of GAS5 noncoding RNA in mediating the effects of rapamycin and its analogues on mantle cell lymphoma cells," *Clinical Lymphoma*, *Myeloma & Leukemia*, vol. 14, no. 6, pp. 468–473, 2014.
- [9] X. Chen, D. D. Wang, T. Wei, S. M. He, G. Y. Zhang, and Q. L. Wei, "Effects of astragalosides from Radix Astragali on high glucose-induced proliferation and extracellular matrix accumulation in glomerular mesangial cells," *Experimental and Therapeutic Medicine*, vol. 11, no. 6, pp. 2561–2566, 2016.
- [10] M. C. Thomas, P. H. Groop, and K. Tryggvason, "Towards understanding the inherited susceptibility for nephropathy in diabetes," *Current Opinion in Nephrology and Hypertension*, vol. 21, no. 2, pp. 195–202, 2012.
- [11] M. C. Thomas, "Epigenetic mechanisms in diabetic kidney disease," *Current Diabetes Reports*, vol. 16, no. 3, p. 31, 2016.
- [12] P. Kapranov, J. Cheng, S. Dike et al., "RNA maps reveal new RNA classes and a possible function for pervasive transcription," *Science*, vol. 316, no. 5830, pp. 1484–1488, 2007.
- [13] C. A. Brosnan and O. Voinnet, "The long and the short of noncoding RNAs," *Current Opinion in Cell Biology*, vol. 21, no. 3, pp. 416–425, 2009.
- [14] Y. Okazaki, M. Furuno, T. Kasukawa et al., "Analysis of the mouse transcriptome based on functional annotation of 60, 770 full-length cDNAs," *Nature*, vol. 420, no. 6915, pp. 563–573, 2002.
- [15] C. P. Ponting, P. L. Oliver, and W. Reik, "Evolution and functions of long noncoding RNAs," *Cell*, vol. 136, no. 4, pp. 629–641, 2009.
- [16] B. Jayaram, I. Syed, C. N. Kyathanahalli, C. J. Rhodes, and A. Kowluru, "Arf nucleotide binding site opener [ARNO] promotes sequential activation of Arf6, Cdc42 and Rac1 and insulin secretion in INS 832/13 β-cells and rat islets," *Biochemical Pharmacology*, vol. 81, no. 8, pp. 1016–1027, 2011.
- [17] T. Saito, J. Kurashige, S. Nambara et al., "A long non-coding RNA activated by transforming growth factor- $\beta$  is an independent prognostic marker of gastric cancer," *Annals of Surgical Oncology*, vol. 22, Supplement 3, pp. 915–922, 2015.
- [18] A. Roth and S. Diederichs, "Long noncoding RNAs in lung cancer," *Current Topics in Microbiology and Immunology*, vol. 394, pp. 57–110, 2016.
- [19] J. Gao, W. Wang, F. Wang, and C. Guo, "LncRNA-NR\_033515 promotes proliferation, fibrogenesis and epithelial-tomesenchymal transition by targeting miR-743b-5p in diabetic nephropathy," *Biomedicine & Pharmacotherapy*, vol. 106, pp. 543–552, 2018.
- [20] M. Hu, R. Wang, X. Li et al., "LncRNA MALAT1 is dysregulated in diabetic nephropathy and involved in high glucose-induced podocyte injury via its interplay with β-catenin," *Journal of Cellular and Molecular Medicine*, vol. 21, no. 11, pp. 2732– 2747, 2017.

- [21] J. Long and F. R. Danesh, "Values and limitations of targeting lncRNAs in diabetic nephropathy," *Diabetes*, vol. 67, no. 4, pp. 552-553, 2018.
- [22] A. Li, R. Peng, Y. Sun, H. Liu, H. Peng, and Z. Zhang, "LincRNA 1700020I14Rik alleviates cell proliferation and fibrosis in diabetic nephropathy via miR-34a-5p/Sirt1/HIF-1α signaling," *Cell Death & Disease*, vol. 9, no. 5, p. 461, 2018.
- [23] H. Yi, R. Peng, L. Y. Zhang et al., "LincRNA-Gm4419 knockdown ameliorates NF-κB/NLRP3 inflammasome-mediated inflammation in diabetic nephropathy," *Cell Death & Disease*, vol. 8, no. 2, article e2583, 2017.
- [24] M. Wang, D. Yao, S. Wang, Q. Yan, and W. Lu, "Long non-coding RNA ENSMUST00000147869 protects mesangial cells from proliferation and fibrosis induced by diabetic nephropathy," *Endocrine*, vol. 54, no. 1, pp. 81–92, 2016.
- [25] Y. Gao, Z. Y. Chen, Y. Wang, Y. Liu, J. X. Ma, and Y. K. Li, "Long non-coding RNA ASncmtRNA-2 is upregulated in diabetic kidneys and high glucose-treated mesangial cells," *Experimental and Therapeutic Medicine*, vol. 13, no. 2, pp. 581–587, 2017.
- [26] S. Wang, X. Chen, M. Wang et al., "Long non-coding RNA CYP4B1-PS1-001 inhibits proliferation and fibrosis in diabetic nephropathy by interacting with nucleolin," *Cellular Physiology and Biochemistry*, vol. 49, no. 6, pp. 2174–2187, 2018.
- [27] Y. Guan, W. L. Kuo, J. L. Stilwell et al., "Amplification of PVT1 contributes to the pathophysiology of ovarian and breast cancer," *Clinical Cancer Research*, vol. 13, no. 19, pp. 5745–5755, 2007.
- [28] R. L. Hanson, D. W. Craig, M. P. Millis et al., "Identification of PVT1 as a candidate gene for end-stage renal disease in type 2 diabetes using a pooling-based genome-wide single nucleotide polymorphism association study," *Diabetes*, vol. 56, no. 4, pp. 975–983, 2007.
- [29] M. L. Alvarez and J. K. DiStefano, "Functional characterization of the plasmacytoma variant translocation 1 gene (PVT1) in diabetic nephropathy," *PLoS One*, vol. 6, no. 4, article e18671, 2011.
- [30] H. B. Lee and H. Ha, "Plasminogen activator inhibitor-1 and diabetic nephropathy," *Nephrology*, vol. 10, Supplement 2, pp. S11–S13, 2005.
- [31] X. Zhou, S. Liu, G. Cai et al., "Long non coding RNA MALAT1 promotes tumor growth and metastasis by inducing epithelial-mesenchymal transition in oral squamous cell carcinoma," *Scientific Reports*, vol. 5, no. 1, 2015.
- [32] B. Yan, Z. F. Tao, X. M. Li, H. Zhang, J. Yao, and Q. Jiang, "Aberrant expression of long noncoding RNAs in early diabetic retinopathy," *Investigative Ophthalmology & Visual Science*, vol. 55, no. 2, pp. 941–951, 2014.
- [33] M. Edeling, G. Ragi, S. Huang, H. Pavenstädt, and K. Susztak, "Developmental signalling pathways in renal fibrosis: the roles of Notch, Wnt and Hedgehog," *Nature Reviews Nephrology*, vol. 12, no. 7, pp. 426–439, 2016.
- [34] K. Reidy, H. M. Kang, T. Hostetter, and K. Susztak, "Molecular mechanisms of diabetic kidney disease," *The Journal of Clinical Investigation*, vol. 124, no. 6, pp. 2333–2340, 2014.
- [35] C. Dai, D. B. Stolz, L. P. Kiss, S. P. Monga, L. B. Holzman, and Y. Liu, "Wnt/β-catenin signaling promotes podocyte dysfunction and albuminuria," *Journal of the American Society of Nephrology*, vol. 20, no. 9, pp. 1997–2008, 2009.
- [36] Y. T. Zhuang, D.-Y. Xu, G.-Y. Wang, J.-L. Sun, Y. Huang, and S.-Z. Wang, "IL-6 induced lncRNA MALAT1 enhances

TNF-*α* expression in LPS-induced septic cardiomyocytes via activation of SAA3," *European Review for Medical and Pharmacological Sciences*, vol. 21, no. 2, pp. 302–309, 2017.

- [37] J. Xu and M. H. Zou, "Molecular insights and therapeutic targets for diabetic endothelial dysfunction," *Circulation*, vol. 120, no. 13, pp. 1266–1286, 2009.
- [38] C. E. Tabit, W. B. Chung, N. M. Hamburg, and J. A. Vita, "Endothelial dysfunction in diabetes mellitus: molecular mechanisms and clinical implications," *Reviews in Endocrine* & *Metabolic Disorders*, vol. 11, no. 1, pp. 61–74, 2010.
- [39] P. Puthanveetil, S. Chen, B. Feng, A. Gautam, and S. Chakrabarti, "Long non-coding RNA MALAT1 regulates hyperglycaemia induced inflammatory process in the endothelial cells," *Journal of Cellular and Molecular Medicine*, vol. 19, no. 6, pp. 1418–1425, 2015.
- [40] X. Li, L. Zeng, C. Cao et al., "Long noncoding RNA MALAT1 regulates renal tubular epithelial pyroptosis by modulated miR-23c targeting of ELAVL1 in diabetic nephropathy," *Experimental Cell Research*, vol. 350, no. 2, pp. 327–335, 2017.
- [41] M. C. Tsai, O. Manor, Y. Wan et al., "Long noncoding RNA as modular scaffold of histone modification complexes," *Science*, vol. 329, no. 5992, pp. 689–693, 2010.
- [42] J. Lee and J. K. Kemper, "Controlling SIRT1 expression by microRNAs in health and metabolic disease," Aging, vol. 2, no. 8, pp. 527–534, 2010.
- [43] P. Ravanan, I. F. Srikumar, and P. Talwar, "Autophagy: the spotlight for cellular stress responses," *Life Sciences*, vol. 188, pp. 53–67, 2017.
- [44] M. Hamzawy, S. A. A. Gouda, L. Rashid, M. Attia Morcos, H. Shoukry, and N. Sharawy, "The cellular selection between apoptosis and autophagy: roles of vitamin D, glucose and immune response in diabetic nephropathy," *Endocrine*, vol. 58, no. 1, pp. 66–80, 2017.
- [45] Y. Feng, S. Chen, J. Xu et al., "Dysregulation of lncRNAs GM5524 and GM15645 involved in high-glucose-induced podocyte apoptosis and autophagy in diabetic nephropathy," *Molecular Medicine Reports*, vol. 18, no. 4, pp. 3657–3664, 2018.
- [46] X. Qian, J. Tan, L. Liu et al., "MicroRNA-134-5p promotes high glucose-induced podocyte apoptosis by targeting bcl-2," *American Journal of Translational Research*, vol. 10, no. 3, pp. 989–997, 2018.
- [47] X. Zhang, S. Song, and H. Luo, "Regulation of podocyte lesions in diabetic nephropathy via miR-34a in the Notch signaling pathway," *Medicine*, vol. 95, no. 44, article e5050, 2016.
- [48] Q. Zhou, A. C. K. Chung, X. R. Huang, Y. Dong, X. Yu, and H. Y. Lan, "Identification of novel long noncoding RNAs associated with TGF-β/Smad3-mediated renal inflammation and fibrosis by RNA sequencing," *The American Journal of Pathol*ogy, vol. 184, no. 2, pp. 409–417, 2014.
- [49] S. F. Sun, P. M. K. Tang, M. Feng et al., "Novel lncRNA Erbb4-IR promotes diabetic kidney injury in *db/db* mice by targeting miR-29b," *Diabetes*, vol. 67, no. 4, pp. 731–744, 2018.
- [50] V. A. Burzio, C. Villota, J. Villegas et al., "Expression of a family of noncoding mitochondrial RNAs distinguishes normal from cancer cells," *Proceedings of the National Academy of Sciences of the United States of America*, vol. 106, no. 23, pp. 9430–9434, 2009.
- [51] O. Rackham, A. M. J. Shearwood, T. R. Mercer, S. M. K. Davies, J. S. Mattick, and A. Filipovska, "Long noncoding RNAs are generated from the mitochondrial genome and

regulated by nuclear-encoded proteins," RNA, vol. 17, no. 12, pp. 2085–2093, 2011.

- [52] V. Bianchessi, I. Badi, M. Bertolotti et al., "The mitochondrial lncRNA ASncmtRNA-2 is induced in aging and replicative senescence in endothelial cells," *Journal of Molecular and Cellular Cardiology*, vol. 81, pp. 62–70, 2015.
- [53] M. Brownlee, "The pathobiology of diabetic complications: a unifying mechanism," *Diabetes*, vol. 54, no. 6, pp. 1615– 1625, 2005.
- [54] F. Giacco and M. Brownlee, "Oxidative stress and diabetic complications," *Circulation Research*, vol. 107, no. 9, pp. 1058–1070, 2010.
- [55] N. S. Chandel and G. R. S. Budinger, "The cellular basis for diverse responses to oxygen," *Free Radical Biology & Medicine*, vol. 42, no. 2, pp. 165–174, 2007.
- [56] F. Leti, E. Morrison, and J. K. DiStefano, "Long noncoding RNAs in the pathogenesis of diabetic kidney disease: implications for novel therapeutic strategies," *Personalized Medicine*, vol. 14, no. 3, pp. 271–278, 2017.
- [57] M. Kato, M. Wang, Z. Chen et al., "An endoplasmic reticulum stress-regulated lncRNA hosting a microRNA megacluster induces early features of diabetic nephropathy," *Nature Communications*, vol. 7, no. 1, p. 12864, 2016.
- [58] M. T. Lindenmeyer, M. P. Rastaldi, M. Ikehata et al., "Proteinuria and hyperglycemia induce endoplasmic reticulum stress," *Journal of the American Society of Nephrology*, vol. 19, no. 11, pp. 2225–2236, 2008.
- [59] J. Wu, R. Zhang, M. Torreggiani et al., "Induction of diabetes in aged C57B6 mice results in severe nephropathy: an association with oxidative stress, endoplasmic reticulum stress, and inflammation," *The American Journal of Pathology*, vol. 176, no. 5, pp. 2163–2176, 2010.
- [60] T. L. Young, T. Matsuda, and C. L. Cepko, "The noncoding RNA taurine upregulated gene 1 is required for differentiation of the murine retina," *Current Biology*, vol. 15, no. 6, pp. 501– 512, 2005.
- [61] S. Y. Li and K. Susztak, "The long noncoding RNA *Tug1* connects metabolic changes with kidney disease in podocytes," *The Journal of Clinical Investigation*, vol. 126, no. 11, pp. 4072– 4075, 2016.
- [62] J. Lin, C. Handschin, and B. M. Spiegelman, "Metabolic control through the PGC-1 family of transcription coactivators," *Cell Metabolism*, vol. 1, no. 6, pp. 361–370, 2005.
- [63] M. B. Hock and A. Kralli, "Transcriptional control of mitochondrial biogenesis and function," *Annual Review of Physiol*ogy, vol. 71, no. 1, pp. 177–203, 2009.
- [64] L. Wang, Z. Teng, S. Cai, D. Wang, X. Zhao, and K. Yu, "The association between the PPARγ2 Pro12Ala polymorphism and nephropathy susceptibility in type 2 diabetes: a meta-analysis based on 9, 176 subjects," *Diagnostic Pathology*, vol. 8, no. 1, p. 118, 2013.
- [65] M. Flaquer, N. Lloberas, M. Franquesa et al., "The combination of sirolimus and rosiglitazone produces a renoprotective effect on diabetic kidney disease in rats," *Life Sciences*, vol. 87, no. 5-6, pp. 147–153, 2010.
- [66] J. Long, S. S. Badal, Z. Ye et al., "Long noncoding RNA *Tug1* regulates mitochondrial bioenergetics in diabetic nephropathy," *Journal of Clinical Investigation*, vol. 126, no. 11, pp. 4205–4218, 2016.
- [67] L. J. Duan, M. Ding, L. J. Hou, Y. T. Cui, C. J. Li, and D. M. Yu, "Long noncoding RNA TUG1 alleviates extracellular matrix"

accumulation via mediating microRNA-377 targeting of PPARy in diabetic nephropathy," *Biochemical and Biophysical Research Communications*, vol. 484, no. 3, pp. 598–604, 2017.

- [68] B. Yu and S. Wang, "Angio-LncRs: LncRNAs that regulate angiogenesis and vascular disease," *Theranostics*, vol. 8, no. 13, pp. 3654–3675, 2018.
- [69] V. Vallon and S. C. Thomson, "Renal function in diabetic disease models: the tubular system in the pathophysiology of the diabetic kidney," *Annual Review of Physiology*, vol. 74, no. 1, pp. 351–375, 2012.
- [70] L. Zhou, D. Y. Xu, W. G. Sha, L. Shen, G. Y. Lu, and X. Yin, "Long non-coding MIAT mediates high glucose-induced renal tubular epithelial injury," *Biochemical and Biophysical Research Communications*, vol. 468, no. 4, pp. 726–732, 2015.
- [71] L. Xiao, X. Xu, F. Zhang et al., "The mitochondria-targeted antioxidant MitoQ ameliorated tubular injury mediated by mitophagy in diabetic kidney disease via Nrf2/PINK1," *Redox Biology*, vol. 11, pp. 297–311, 2017.
- [72] A. Uruno, Y. Furusawa, Y. Yagishita et al., "The Keap1-Nrf2 system prevents onset of diabetes mellitus," *Molecular and Cellular Biology*, vol. 33, no. 15, pp. 2996–3010, 2013.
- [73] Z. Wang, X. Wang, H. Zhou, X. Dan, L. Jiang, and Y. Wu, "Long non-coding RNA CASC2 inhibits tumorigenesis via the miR-181a/PLXNC1 axis in melanoma," *Acta Biochimica et Biophysica Sinica*, vol. 50, no. 3, pp. 263–272, 2018.
- [74] L. Wang, N. Su, Y. Zhang, and G. Wang, "Clinical significance of serum lncRNA cancer susceptibility candidate 2 (CASC2) for chronic renal failure in patients with type 2 diabetes," *Medical Science Monitor*, vol. 24, pp. 6079–6084, 2018.
- [75] H. Lian, Y. Cheng, and X. Wu, "TMEM16A exacerbates renal injury by activating P38/JNK signaling pathway to promote podocyte apoptosis in diabetic nephropathy mice," *Biochemical and Biophysical Research Communications*, vol. 487, no. 2, pp. 201–208, 2017.
- [76] H. Yang, Q. Kan, Y. Su, and H. Man, "Long non-coding RNA CASC2 improves diabetic nephropathy by inhibiting JNK pathway," *Experimental and Clinical Endocrinology & Diabetes*, 2018.
- [77] D. N. Muller, C. Schmidt, E. Barbosa-Sicard et al., "Mouse Cyp4a isoforms: enzymatic properties, gender- and strain-specific expression, and role in renal 20-hydroxyeicosatetraenoic acid formation," *The Biochemical Journal*, vol. 403, no. 1, pp. 109– 118, 2007.
- [78] X. Zhao and J. D. Imig, "Kidney CYP450 enzymes: biological actions beyond drug metabolism," *Current Drug Metabolism*, vol. 4, no. 1, pp. 73–84, 2003.
- [79] G. Wang, J. Yao, Z. Li et al., "miR-34a-5p inhibition alleviates intestinal ischemia/reperfusion-induced reactive oxygen species accumulation and apoptosis via activation of SIRT1 signaling," *Antioxidants & Redox Signaling*, vol. 24, no. 17, pp. 961–973, 2016.
- [80] Y. Shao, C. Lv, C. Wu, Y. Zhou, and Q. Wang, "Mir-217 promotes inflammation and fibrosis in high glucose cultured rat glomerular mesangial cells via Sirt1/HIF-1α signaling pathway," *Diabetes/Metabolism Research and Reviews*, vol. 32, no. 6, pp. 534–543, 2016.
- [81] M. Wang, S. Wang, D. Yao, Q. Yan, and W. Lu, "A novel long non-coding RNA CYP4B1-PS1-001 regulates proliferation and fibrosis in diabetic nephropathy," *Molecular and Cellular Endocrinology*, vol. 426, pp. 136–145, 2016.

- [82] K. Wilhelm, K. Happel, G. Eelen et al., "FOXO1 couples metabolic activity and growth state in the vascular endothelium," *Nature*, vol. 529, no. 7585, pp. 216–220, 2016.
- [83] X. Bai, J. Geng, X. Li et al., "Long noncoding RNA LINC01619 regulates microRNA-27a/forkhead box protein O1 and endoplasmic reticulum stress-mediated podocyte injury in diabetic nephropathy," *Antioxidants & Redox Signaling*, vol. 29, no. 4, pp. 355–376, 2018.
- [84] L. Peng, X. Yuan, B. Jiang, Z. Tang, and G. C. Li, "LncRNAs: key players and novel insights into cervical cancer," *Tumour Biology*, vol. 37, no. 3, pp. 2779–2788, 2016.
- [85] A. Maity and B. Das, "N6-methyladenosine modification in mRNA: machinery, function and implications for health and diseases," *The FEBS Journal*, vol. 283, no. 9, pp. 1607–1630, 2016.