LDL-C calculated by Friedewald, Martin-Hopkins, or NIH equation 2 versus beta-quantification: pooled alirocumab trials

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Abstract Accurate assessment of LDL-C levels is important, as they are often used for treatment recommendations. For many years, plasma LDL-C levels were calculated using the Friedewald equation, but there are limitations to this method compared with direct measurement via beta-quantification (BQ). Here, we assessed differences between the Friedewald, Martin-Hopkins, and NIH equation 2 methods of calculating LDL-C and the "gold standard" BQ method using pooled phase 3 data with alirocumab. All randomized patients were included irrespective of the treatment arm (n = 6,122). We compared pairs of LDL-C values (n = 17,077) determined by each equation and BO. We found that BO-derived LDL-C values ranged from 1 to 397 mg/dl (mean 90.68 mg/ There were strong correlations between dl). Friedewald-calculated, Martin-Hopkins-calculated, and NIH equation 2-calculated LDL-C with BQdetermined LDL-C values (Pearson's correlation coefficient = 0.985, 0.981, and 0.985, respectively). Importantly, for BQ-derived LDL-C values \geq 70 mg/ dl, only 3.2%, 1.4%, and 1.8% of Friedewald-calculated, Martin-Hopkins-calculated, and NIH equation 2-calculated values were <70 mg/dl, respectively. When triglyceride (TG) levels were <150 mg/dl, differences between calculated and BQ-derived LDL-C values were minimal, regardless of the LDL-C level (<40, <55, or <70 mg/dl). However, when TG levels were >150 mg/dl, NIH equation 2 provided greater accuracy than Friedewald or Martin-Hopkins. When TGs were >250 mg/dl, inaccuracies were seen with all three methods, although NIH equation 2 remained the most accurate. III In conclusion, LDL-C calculated by any of the three methods can guide treatment decisions for most patients, including those treated with proprotein convertase subtilisin/kexin type 9 inhibitors.

Supplementary key words alirocumab • beta-quantification • calculated LDL-C • cholesterol • drug therapy/hypolipidemic drugs • Friedewald • LDL • Martin-Hopkins • NIH equation 2 • PCSK9

The accurate assessment of LDL-C levels is important, as treatment decisions are often based on achievement of a specific LDL-C goal, a risk-based treatment goal, or reduction of LDL-C levels by at least 50% (1, 2). The gold standard reference method for measurement of LDL-C is beta-quantification (BQ) (3); however, the technique is labor-intensive and timeconsuming, requiring the use of ultracentrifugation (which is not routinely available in all laboratories) to separate lipoprotein classes.

In clinical practice, plasma LDL-C levels were, for many years, indirectly determined by calculation from total cholesterol, HDL-C, and plasma triglyceride (TG) measurements following the method of Friedewald et al. (4), where LDL-C = total cholesterol – (HDL-C + TG/5) (using values in mg/dl). However, there are several limitations with the Friedewald equation. Most importantly, the equation assumes a fixed TG:cholesterol ratio for estimating the concentration of VLDL-C (represented by TG/5 in the formula). In patients with high TGs, this relationship may no longer hold true, and the Friedewald equation is not reliable when TGs are >400 mg/dl (4). Friedewald-calculated LDL-C values have also been reported to lose accuracy in patients with TGs \geq 150 mg/dl or LDL-C <70 mg/dl (5, 6). Other researchers have reported significant loss of accuracy at very-low LDL-C levels (<30 mg/dl) (7).

With the known limitations of the Friedewald equation, more recent methods for calculating LDL-C have been developed. The method developed by Martin *et al.* (hereafter referred to as the Martin-Hopkins method) uses patient-specific ratios of TGs to VLDL-C derived from direct measurement of the latter by vertical spin

J. Lipid Res. (2022) 63(1) 100148 1

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density gradient ultracentrifugation (8) to allow estimation of VLDL-C in conjunction with directly measured TGs, total cholesterol, and HDL-C concentrations. The estimated VLDL-C is subtracted from non-HDL-C to obtain the LDL-C level (LDL-C = total cholesterol - HDL-C - TG/novel factor, where the novel factor is an adjustable factor based on a patient's non-HDL-C and TG levels and is derived from a 174cell 2D table) (9). A more recent method by Sampson et al. (hereafter referred to as the NIH equation 2 method) used TGs and non-HDL-C as independent variables in a least-squares regression to generate an equation for estimation of VLDL-C, which was used in a second equation to calculate LDL-C at both low LDL-C and/or high TG levels (using least-squares regressions, LDL-C = total cholesterol/0.948 - HDL-C/ $0.971 - (TG/8.56 + [TG \times non-HDL-C]/2,140 - TG^2/$ 16100) - 9.44 (6).

Discrepancies between calculated and BQ-derived LDL-C have become more important in the era of proprotein convertase subtilisin/kexin type 9 (PCSK9) inhibitors that can reduce LDL-C to very low levels when added to a statin (10–13). In some phase 3 trials of the PCSK9 inhibitor alirocumab, LDL-C was determined by both Friedewald calculation and BQ methods (14–22). This provided an opportunity to assess potential differences between those two methods for determining LDL-C and to compare the Friedewald calculation to the Martin-Hopkins and NIH equation 2 methods (>5,000 patients).

MATERIALS AND METHODS

We compared pairs of LDL-C values determined by the Friedewald, Martin-Hopkins, or NIH equation 2 calculations, and BQ, using data from 10 ODYSSEY trials. Trial designs have been reported previously (14–22) and are summarized in supplemental Fig. Sl. All studies were conducted in accordance with the Declaration of Helsinki and the International Conference on Harmonization Guidelines for Good Clinical Practice. All study protocols were approved by the appropriate institutional review board/ethics review committees, and patients provided written informed consent.

Patients were eligible for participation if their LDL-C levels were \geq 70 mg/dl or \geq 100 mg/dl at screening (depending on cardiovascular risk), except in LONG TERM, where all patients enrolled had LDL-C \geq 70 mg/dl. Patients with TGs >400 mg/dl at screening were excluded. The primary endpoint in each trial was the percentage reduction in LDL-C from baseline to week 24.

For the present analysis, data were pooled from patients randomized to alirocumab, placebo, ezetimibe, or statin in the 10 trials (total 6,122 randomized patients; supplemental Fig. S1). Patients from the statin control arms in the ODYSSEY OP-TIONS I, OPTIONS II, and ALTERNATIVE studies, and the 75 mg alirocumab every 2 weeks calibration arm in the OD-YSSEY CHOICE I and II studies which investigated the alirocumab every 4 weeks dosing regimen, were also included.

Analyses were performed on the pooled intention-to-treat population (all randomized patients, regardless of treatment adherence). BQ and calculated LDL-C values were assessed at weeks 0 and 24 in all trials, and at weeks 12, 52, and 78 in ODYSSEY LONG TERM. If TG values were >400 mg/dl at one of these postrandomization visits, Friedewald calculation of LDL-C was not performed and LDL-C was derived by BQ instead. All available BQ-derived LDL-C values with a corresponding Friedewald-calculated LDL-C value at that timepoint were included in this analysis.

For the LONG TERM trial, central analysis of lipid parameters and BQ were performed by Covance Central Laboratory Services (Indianapolis, IN, and Geneva, Switzerland); BQ analyses were performed as follows. Serum was centrifuged for 20 h and 15 min using a Beckman Ultracentrifuge (model L7-35, L8-70M, or L-80XP). After centrifugation, VLDL-C and chylomicrons were in the upper layer, and HDL-C and LDL-C were in the bottom layer of the tube. The cholesterol in the bottom fraction was removed using a Beckman CentriTube Slicer and assayed using Roche Modular/Cobas analyzers. LDL-C concentration was calculated as the cholesterol level in the bottom fraction minus the HDL-C concentration. HDL-C was determined enzymatically following the chemical precipitation of LDL with a polyanion dextran sulfate and divalent Mg cation solution. For the enzymatic determination of cholesterol, all cholesteryl esters are split into free cholesterol and fatty acids by cholesterol esterase (Roche Diagnostics, Indianapolis, IN). The total free cholesterol is then oxidized by cholesterol oxidase to cholest-4-en-3-one and hydrogen peroxide. The hydrogen peroxide reacts with phenol in the presence of peroxidase and 4-aminophenozone to form an o-quinoneimine dye; the intensity of the color formed is proportional to the cholesterol concentration and can be measured photometrically. The interassay coefficient of variation (reproducibility) of the BQ assay ranged from 1.77% to 2.12% depending on the cholesterol concentration (supplemental Table S1).

For all other trials, lipid analyses were performed by Medpace Reference Laboratories, with BQ analyses performed using preparative ultracentrifugation as outlined in the Lipid Research Clinics method manual (23), described as follows. Serum or plasma was overlaid with normal saline (density 1.006 g/ml) and centrifuged (Beckman Ultracentrifugation model # L-90K and rotor type 50.4) at 40,000 rpm for 18-22 h at 10 °C to separate VLDL in the supernatant from LDL and HDL in the infranatant. The cholesterol concentration of the infranatant was then measured; all apolipoprotein B-containing lipoproteins (VLDL, LDL, and lipoprotein[a]) were precipitated from the serum using 50 kDa dextran sulfate with magnesium ions (MgCl₂), and the cholesterol in the remaining HDL fraction was measured by enzymatic methods. The HDL-C concentration was subtracted from the infranatant cholesterol concentration to provide the BQ LDL-C value. The interassay coefficient of variation (reproducibility) ranged from 4.3% to 5.1% (supplemental Table S1).

The accuracy of serum total cholesterol, HDL-C, and TG level determinations (used to calculate LDL-C by the Friedewald, Martin-Hopkins, and NIH Equation 2 methods) was tested by participating in the Centers for Disease Control and Prevention–National Heart, Lung, and Blood Institute Lipid Standardization Program assays. Total cholesterol and TGs were determined by enzymatic methods using a Roche Modular analyzer (Covance Central Laboratory Services) or a Beckman Coulter AU series automatic analyzer (Medpace Reference Laboratories). HDL-C was isolated by chemical precipitation and centrifugation (a polyanion dextran sulfate and divalent Mg cation solution interacts with the LDL and



VLDL to form an insoluble complex that is removed by centrifugation) and then analyzed as for total cholesterol. Assays conducted by Covance Central Laboratory Services and Medpace Reference Laboratories conformed to Centers for Disease Control standards; the precision and reproducibility of all lipid parameters for both central laboratories are summarized in supplemental Table SI. Spearman's correlation coefficients were derived to assess the relationship between LDL-C calculated by each of the three methods and BQ-derived LDL-C. The difference between BQ-derived and calculated LDL-C values was determined by calculated minus BQ-derived LDL-C values (both in mg/dl).

Differences between Friedewald-calculated, Martin-Hopkins-calculated, and NIH equation 2-calculated and BQ-derived LDL-C values were further assessed via Bland-Altman-adapted plots (difference vs. BQ LDL-C). Analyses according to quintiles of BQ LDL-C values or according to tertiles of TG values are also presented. In addition, a sensitivity analysis excluding the results from LONG TERM was also performed.

RESULTS

The initial dataset comprised 17,086 pairs of BQderived and Friedewald-calculated LDL-C values; nine LDL-C pairs with BQ-derived LDL-C values between 400 and 584 mg/dl were excluded as there was an insufficient number of values to allow for correct estimation of the correlation between LDL-C pairs in this range. Therefore, 17,077 pairs were included in the current analysis (data from 6,007 patients). BQ-derived LDL-C values ranged from 1 to 397 mg/dl (mean 90.68 mg/dl, median 87.00 mg/dl, Q1:Q3 49.03:120.85 mg/dl; Fig. 1).

Overall, there was a strong correlation between Friedewald-calculated and BQ-derived LDL-C (Pearson's correlation coefficient = 0.985); the mean difference was 0.13 mg/dl (median, -0.8 mg/dl; Ql:Q3, -5.4:5.0 mg/dl). A strong correlation between



Fig. 1. Distribution of BQ-derived LDL-C values. The BQ-derived LDL-C values (n = 17,077) ranged from 1 to 397 mg/dl with the following parameters: mean, 90.68 mg/dl; median, 87.00 mg/dl; and Ql:Q3 49.03:120.85 mg/dl. BQ, beta-quantification.

both Martin-Hopkins– and NIH equation 2–calculated LDL-C values with BQ-derived LDL-C values was also observed (Pearson's correlation coefficient = 0.981 and 0.985, respectively); the mean differences were 3.5 mg/dl (median, 1.8 mg/dl; Ql:Q3, -3.0:8.2 mg/dl) for Martin-Hopkins and 3.0 mg/dl (median, 1.7 mg/dl; Ql:Q3, -3.1:7.5, mg/dl) for NIH equation 2. The overall correlation between all four methods is summarized in supplemental Table S2.

To assess the impact of lower LDL-C levels on the relationship between calculated and measured LDL-C, we examined the correlation by quintile of BQ LDL-C values. The boundaries for the BQ quintiles were 41.7, 73.4, 99.2, and 129.0 mg/dl; however, values rounded to the nearest whole number (40, 70, 100, and 130 mg/dl) were used for the analyses. Although correlations were shown to vary when data were analyzed by quintile of BQ values, there was no observable pattern across quintiles of BQ values with Friedewaldcalculated (Fig. 2A), Martin-Hopkins–calculated (Fig. 2B), or NIH equation 2-calculated LDL-C values (Fig. 2C). In addition, the mean differences between Friedewald-calculated and BQ-derived LDL-C values by quintile of BQ LDL-C values for the entire study sample were -3.0, -1.7, 1.0, 1.2, and 2.8 mg/dl for the first to the fifth quintiles, respectively (median differences: -3.0, -2.0, 1.0, 1.0 and 2.0 mg/dl); the interguartile range increased from the first to the fifth quintile (6.9, 8.3, 10.0, 11.0, and 13.6 mg/dl, respectively). The corresponding mean differences between Martin-Hopkins-calculated and BQ-derived LDL-C values by quintile of BQ LDL-C values were 0.4, 2.8, 5.3, 4.3, and 4.3 mg/dl for the first to fifth quintiles, respectively (median differences: -0.9, 1.3, 4.1, 3.0, and 2.4 mg/dl); the interquartile range from the first to the fifth quintile was 6.9, 9.9, 11.0, 12.5, and 15.3 mg/dl, respectively. Similarly, the corresponding mean differences between NIH equation 2-calculated and BQ-derived LDL-C values by quintile of BQ LDL-C values were -0.8, 1.3, 4.4, 4.3, and 5.2 mg/dl for the first to fifth quintiles, respectively (median differences: -1.7, 0.1, 3.5,3.9, and 4.7 mg/dl); the interquartile range from the first to fifth quintile was 6.3, 8.7, 10.3, 11.3, and 13.6 mg/dl, respectively.

The differences between calculated and BO-derived LDL-C values were also assessed for subgroups of BQderived LDL-C values <40 mg/dl, <55 mg/dl, and <70 mg/dl with TGs >150 mg/dl. The results for all three methods in these subgroups are summarized in Table 1. In the subgroup with LDL-C <40 mg/dl/TGs >150 mg/dl, median differences between calculated and BQ-derived LDL-C values were -9 mg/dl, 3 mg/dl, and -1 mg/dl for the Friedewald, Martin-Hopkins, and NIH equation 2 methods, respectively. For the subgroup with LDL-C <55 mg/dl/TGs >150 mg/dl, the median differences were -8 mg/dl (Friedewald), 4 mg/dl (Martin-Hopkins), and -1 mg/dl (NIH for the subgroup equation 2); with LDL-C



BQ value (mg/dl)

Fig. 2. Scatter plot of calculated versus BQ-derived LDL-C values according to quintiles of BQ-derived LDL-C values. A: Friedewald-calculated versus BQ-derived LDL-C. B: Martin-Hopkins–calculated versus BQ-derived LDL-C. C: NIH equation 2–calculated versus BQ-derived LDL-C. A: Regression equations (Pearson's correlation) for each quintile of BQ-derived LDL-C values are as follows: QI (n = 3,276): y = -2.948 + 0.997x (0.73); Q2 (n = 3,180): y = -7.652 + 1.108x (0.76); Q3 (n = 3,911): y = -1.114 + 1.025x (0.69); Q4 (n = 3,382): y = -1.642 + 1.025x (0.66); and Q5 (n = 3,328): y = -2.592 + 1.032x (0.96). Note that the x = y line is shown as a red dotted line. B: Regression equations (Pearson's correlation) for each quintile of BQ-derived LDL-C values are as follows: Q1 (n = 3,276): y = -0.5315 + 1.147x (0.74); Q3 (n = 3,911): y = 4.784 + 1.006x (0.65); Q4 (n = 3,382): y = 4.059 + 1.002x (0.62); and Q5 (n = 3,328): y = -1.477 + 1.016x (0.95). Note that the x = y line is shown as a red dotted line. C: Regression equations (Pearson's correlation) for each quintile of BQ-derived LDL-C values are as follows: Q1 (n = 3,276): y = -0.586 + 0.991x (0.75); Q2 (n = 3,180): y = -6.466 + 1.140x (0.77); Q3 (n = 3,911): y = 1.674 + 1.032x (0.70); Q4 (n = 3,382): y = -0.586 + 0.991x (0.75); Q2 (n = 3,180): y = -6.466 + 1.140x (0.77); Q3 (n = 3,911): y = 1.674 + 1.032x (0.70); Q4 (n = 3,382): y = 1.746 + 1.023x (0.66); and Q5 (n = 3,328): y = 3.567 + 1.010x (0.96). Note that the x = y line is shown as a red dotted line. The boundaries for the quintiles of BQ LDL-C values are 40, 70, 100, and 130 mg/dl (rounded values of the real boundaries were used, which were 41.7, 73.4, 99.2, and 129.0 mg/dl, respectively), with minimum = 1 mg/dl and maximum = 397 mg/dl. BQ, beta-quantification; Q, quintile; x, BQ-derived LDL-C value; y, calculated LDL-C value.

TABLE 1.	Analysis of difference between	calculated LDL-C values and BO-der	rived LDL-C values for several subg	roups of r	oatients
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		Difference (mg/dl)				
Subgroup	n of LDL-C Pairs	Mean	Median	Ql	Q2	IQR
BO-derived LDL-C <40 mg/dl and TGs >150 mg/dl						
Friedewald versus BQ	626	-7.6	-8.9	-13.0	-4.2	8.8
Martin-Hopkins versus BO	626	5.5	3.1	-0.8	8.9	9.7
NIH equation 2 versus \widetilde{BO}	626	0.9	-0.9	-4.4	3.5	7.9
BO-derived LDL-C <40 mg/dl and TGs <150 mg/dl						
Friedewald versus BO	2,595	-1.9	-2.3	-5.0	1.0	6.0
Martin-Hopkins versus BO	2,595	-0.9	-1.5	-4.3	1.7	6.0
NIH equation 2 versus $B\widetilde{O}$	2,595	-1.2	-1.8	-4.7	1.3	6.0
BO-derived LDL-C <40 mg/dl and TGs >250 mg/dl	<i>,</i>					
Friedewald versus BO	100	-10.0	-14.3	-19.0	-4.8	14.1
Martin-Hopkins versus BO	100	15.6	11.5	5.9	21.3	15.4
NIH equation 2 versus \widetilde{BO}	100	5.4	1.7	-3.4	9.4	12.9
BO-derived LDL-C <55 mg/dl and TGs <150 mg/dl						
Friedewald versus BO	3.810	-1.8	-2.0	-5.0	1.0	6.0
Martin-Hopkins versus BO	3.810	-0.8	-1.4	-4.4	2.0	6.4
NIH equation 2 versus \widetilde{BO}	3.810	-1.1	-1.6	-4.7	1.7	6.4
BO-derived LDL-C $<55 \text{ mg/dl}$ and TGs $>150 \text{ mg/dl}$	- ,					
Friedewald versus BO	1.020	-7.0	-8.0	-13.0	-3.0	10.0
Martin-Hopkins versus BO	1,020	6.4	4.0	-0.1	10.5	10.7
NIH equation 2 versus \widetilde{BO}	1.020	1.4	-0.6	-4.1	4.9	9.0
BO-derived LDL-C $<55 \text{ mg/dl}$ and TGs $>250 \text{ mg/dl}$						
Friedewald versus BO	175	-10.0	-13.5	-20.0	-4.6	15.4
Martin-Hopkins versus BO	175	15.5	12.7	6.2	21.2	15.1
NIH equation 2 versus \widetilde{BO}	175	4.9	1.9	-3.7	9.6	13.3
BO-derived LDL-C $<70 \text{ mg/dl}$ and TGs $<150 \text{ mg/dl}$						
Friedewald versus BO	4.874	-1.5	-2.0	-5.0	1.9	6.9
Martin-Hopkins versus BO	4.874	-0.4	-1.1	-4.3	2.6	6.9
NIH equation 2 versus \widetilde{BO}	4.874	-0.6	-1.3	-4.5	2.4	6.9
BO-derived LDL-C $<70 \text{ mg/dl}$ and TGs $>150 \text{ mg/dl}$	y					
Friedewald versus BO	1.493	-5.5	-7.0	-12.0	-1.0	11.0
Martin-Hopkins versus BO	1.493	7.7	5.4	0.6	12.3	11.7
NIH equation 2 versus \widetilde{BO}	1.493	2.7	0.6	-3.7	6.6	10.3
BO-derived LDL-C $<70 \text{ mg/dl}$ and TGs $>250 \text{ mg/dl}$	· · ·					
Friedewald versus BO	274	-7.2	-10.0	-17.0	-0.8	16.2
Martin-Hopkins versus BO	274	16.8	13.5	7.2	22.8	15.6
NIH equation 2 versus \widetilde{BQ}	274	6.6	4.0	-2.4	12.9	15.3

BQ, beta-quantification; IQR, interquartile range; TG, triglyceride.

<70 mg/dl/TGs >150 mg/dl, the median differences were -7 mg/dl (Friedewald), 5 mg/dl (Martin-Hopkins), and 1 mg/dl (NIH equation 2). Greater median differences were observed for the Friedewald and Martin-Hopkins methods compared with NIH equation 2 for the subgroup with BQ-derived LDL-C <40 mg/dl and TGs >250 mg/dl (Friedewald, -14 mg/dl; Martin-Hopkins, 12 mg/dl; NIH equation 2, 2 mg/dl), the subgroup with BQ-derived LDL-C <55 mg/dl and TGs >250 mg/dl (Friedewald, -14 mg/dl; Martin-Hopkins, 13 mg/dl; NIH equation 2, 2 mg/dl), and the subgroup with BQ-derived LDL-C <70 mg/dl and TGs >250 mg/dl (Friedewald, -10 mg/dl; Martin-Hopkins, 14 mg/dl; NIH equation 2, 4.0 mg/dl; Table 1).

However, when subgroups with BQ-derived LDL-C <40 mg/dl, <55 mg/dl, <70 mg/dl, and TGs <150 mg/dl were examined, the median differences between the three methods were minimal (Friedewald, -2.3, 2.0, and -2.0 mg/dl; Martin-Hopkins, -1.5, -1.4, and -1.1 mg/dl; NIH equation 2, -1.8, -1.6, and -1.3 mg/dl for LDL-C <40 <55, <70 mg/dl, respectively (Table 1).

Analyses of the differences between calculated and BQ-derived LDL-C values versus BQ-derived LDL-C

values were conducted for the total dataset. Overall, small differences were observed across the range of BQ LDL-C values for all three methods (Fig. 3). Comparison of the differences between Friedewald-calculated and BQ-derived LDL-C and BQ LDL-C values implied that the Friedewald method generally overestimates when BQ values are >87 mg/dl and underestimates when BQ values are <87 mg/dl (Fig. 3A). Similarly, the Martin-Hopkins method was found to overestimate across the entire range of BQ LDL-C values (Fig. 3B), and the NIH equation 2 method was found to overestimate when BQ LDL-C values are >13 mg/dl (Fig. 3C).

Analyses of the differences between calculated and BQ-derived LDL-C values versus BQ-derived LDL-C values were also analyzed in more detail according to quintiles of BQ-derived LDL-C values (Table 2; supplemental Fig. S2). When comparing the differences between calculated and BQ-derived LDL-C, the smallest difference was observed with the first quintile of BQ values for all three methods (58.1%, 66.5%, and 66.1% of values differed by 0–5 mg/dl; 28.9%, 25.2%, and 27.6% differed by 5–10 mg/dl; and 13.0%, 8.2%, and 6.3% differed by >10 mg/dl for the Friedewald,



Fig. 3. Bland-Altmann–adapted plots for the difference between calculated and BQ-derived LDL-C values. A: Friedewald-calculated versus BQ-derived LDL-C. B: Martin-Hopkins–calculated versus BQ-derived LDL-C. C: NIH equation 2–calculated versus BQ-derived LDL-C. A: Regression equation (shown as a red line): y = -3.495 + 0.040x. B: Regression equation (shown as a red line): y = 1.338 + 0.0239x. C: Regression equation (shown as a red line): y = -0.497 + 0.0382x. BQ, beta-quantification; TG, triglyceride.

Martin-Hopkins, and NIH equation 2 methods, respectively; Table 2). By comparison, for the overall dataset, 50.3%, 47.9%, and 49.6% of LDL-C pairs differed by 0–5 mg/dl; 29.4%, 28.1%, and 29.4% differed by 5–10 mg/dl; and 20.3%, 24.1%, and 21.3% differed by >10 mg/dl for the Friedewald, Martin-Hopkins, and NIH equation 2 methods, respectively.

We examined concordance between calculated and BQ-derived LDL-C values above and below the LDL-C threshold of <70 mg/dl, used as a clinical target for patients at very-high cardiovascular risk (Table 3). Overall, the discordance was 3.6%, 4.0%, and 3.6% for the Friedewald, Martin-Hopkins, and NIH equation 2

methods. For BQ-derived LDL-C values <70 mg/dl, the concordance was 94.7%, 97.4%, and 96.9% for the Friedewald, Martin-Hopkins, and NIH equation 2 methods, respectively. Furthermore, for BQ-derived LDL-C values \geq 70 mg/dl, 3.2% had a corresponding Friedewald-calculated LDL-C value <70 mg/dl, 1.4% had a corresponding Martin-Hopkins–calculated LDL-C value <70 mg/dl, and 1.8% had a corresponding NIH equation 2–calculated LDL-C value <70 mg/dl.

The number of outliers (absolute difference between calculated and BQ-derived LDL-C values >50 mg/dl) was similar between the Friedewald (45 pairs; 39 distinct patients [6,007 total patients in the analysis]), Martin-

TABLE 2. Summar	y of the differences betw	veen calculated and BQ	2-derived LDL-C va	lues according to	quintile of BQ	2-derived LDL-C values
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	Absolute Difference Between Calculated and BQ-Derived LDL-C Values, n (%) of Values				
Quintile (mg/dl)	0-5 mg/dL	5–10 mg/dL	>10 mg/dL	Total, n	
Friedewald-calculated					
Q1: ≤40	1,902 (58.06)	947 (28.91)	427 (13.03)	3,276	
Q2: >40 to ≤ 70	1,754 (55.16)	908 (28.55)	518 (16.29)	3,180	
Q3: >70 to ≤ 100	2,001 (51.16)	1,202 (30.73)	708 (18.10)	3,911	
Q4: >100 to ≤130	1,603 (47.40)	1,044 (30.87)	735 (21.73)	3,382	
Q5: >130 to ≤ 400	1,329 (39.93)	924 (27.76)	1,075 (32.30)	3,328	
Total, n	8,589	5,025	3,463	17,077	
Martin-Hopkins-calculated					
Q1: ≤40	2,180 (66.54)	827 (25.24)	269 (8.21)	3,276	
Q2: >40 to ≤ 70	1,669 (52.48)	921 (28.96)	590 (18.55)	3,180	
Q3: >70 to ≤ 100	1,702 (43.52)	1,171 (29.94)	1,038 (26.54)	3,911	
Q4: >100 to ≤130	1,402 (41.45)	985 (29.12)	995 (29.42)	3,382	
Q5: >130 to ≤ 400	1,221 (36.69)	890 (26.74)	1,217 (36.57)	3,328	
Total, n	8,174	4,794	4,109	17,077	
NIH equation 2–calculated					
Q1: ≤40	2,166 (66.12)	905 (27.63)	205 (6.26)	3,276	
Q2: >40 to ≤ 70	1,803 (56.70)	927 (29.15)	450 (14.15)	3,180	
Q3: >70 to ≤ 100	1,805 (46.15)	1,231 (31.48)	875 (22.37)	3,911	
Q4: >100 to ≤130	1,438 (42.52)	1,037 (30.66)	907 (26.82)	3,382	
Q5: >130 to ≤ 400	1,228 (36.90)	903 (27.13)	1,197 (35.97)	3,328	
Total, n	8,440	5,003	3,634	17,077	

BQ, beta-quantification; Q, quintile.

Hopkins (63 pairs; 53 distinct patients), and NIH equation 2 (51 pairs; 44 distinct patients) methods. In total, 70 pairs resulted in an absolute difference in LDL-C values >50 mg/dl, of which 49 (70%) pairs were outliers with at least two calculated-LDL-C methods, and 40 (57%) pairs, which were outliers with all three calculated-LDL-C methods. There were 21 LDL-C pairs that resulted in outliers with only one method of LDL-C calculation (Friedewald, two pairs; Martin-Hopkins, 16 pairs; NIH equation 2, three pairs). When the LDL-C outliers were compared for these patients, no obvious patterns were observed over time (supplemental Fig. S3), nor in the lipid parameters used to calculate LDL-C (supplemental Fig. S4).

We further analyzed the data according to TG tertiles to assess whether TG levels affected the results. TGs ranged from 21 to 400 mg/dl, with boundaries of 97 and 150 mg/dl for the tertiles. Overall, no impact on the correlation between calculated versus BQ-derived LDL-C was observed when analyzed by TG tertiles

TABLE 3. Analysis of concordance between calculated and BQderived LDL-C values for the LDL-C threshold of 70 mg/dL

n (%) of LDL-C Pairs	BQ <70 mg/dL	BQ ≥70 mg/dL	Total
Friedewald <70 mg/dL	6.095 (94.7)	341 (5.3)	6.436
Friedewald $\geq 70 \text{ mg/dL}$	282 (2.7)	10,359 (97.3)	10,641
Total	6,377	10,700	17,077
Martin-Hopkins <70 mg/dL	5,853 (97.4)	154 (2.6)	6,007
Martin-Hopkins \geq 70 mg/dL	524 (4.7)	10,546 (95.3)	11,070
Total	6,377	10,700	17,077
NIH equation $2 < 70 \text{ mg/dL}$	5,960 (96.9)	191 (3.1)	6,151
NIH equation $2 \ge 70 \text{ mg/dL}$	417 (3.8)	10,509 (96.2)	10,926
Total	6,377	10,700	17,077

Discordance is 3.6%, 4.0%, and 3.6% for the Friedewald, Martin-Hopkins, and NIH equation 2 methods, respectively.

^BQ, beta-quantification.

(supplemental Fig. S5). However, there was a trend for Friedewald-calculated LDL-C to be less than BQderived LDL-C for the third tertile of TGs (intercept of approximately -7 compared with -1 and -3 for the first and second TG tertiles, respectively; supplemental Fig. S5A). In contrast, there was a trend for Martin-Hopkins-calculated LDL-C to be greater than BQderived LDL-C for the third tertile of TGs (intercept of approximately 8 compared with -2 and +2 for the first and second TG tertiles, respectively; supplemental Fig. S5B). A tendency for calculated values to be higher than BQ-derived LDL-C values for the third tertile of TGs was also observed for NIH equation 2-calculated LDL-C values (intercept of approximately 3 for the third tertile vs. -3 and 0 for the first and second TG tertiles, respectively; supplemental Fig. S5C).

Comparison of the difference between calculated and BQ-derived LDL-C values versus BQ LDL-C according to tertiles of TGs is shown in supplemental Fig. S6. The difference between Friedewald-calculated and BQ-derived LDL-C tended to be greater at higher BQ-LDL-C values, with the difference being greater in the third tertile of TGs versus the first and second tertiles of TGs (supplemental Fig. S6A). For the difference between Martin-Hopkins-calculated LDL-C and BQ-derived LDL-C, a larger difference between values was observed in the third tertile versus the first and second tertiles of TGs (supplemental Fig. S6B). There was no obvious trend in the difference between NIH equation 2-calculated LDL-C and BO-derived LDL-C values when analyzed by TG tertiles (supplemental Fig. S6C).

A sensitivity analysis excluding the results of LONG TERM was also conducted. The BQ-derived LDL-C values (n = 6,667) ranged from 1 to 397 mg/dl (mean

103.2 mg/dl, median 95.0 mg/dl, Q1:Q3 67.0:132.0 mg/dl). As we observed for the total dataset, there was an overall strong correlation between calculated and BQ-derived LDL-C values for all 3 methods (supplemental Tables S3 and S4). When the correlations between calculated and BQ-derived LDL-Clevels were assessed by quintile of BQ LDL-C value, the results were overall consistent with that of the total dataset (supplemental Fig. S7). Similarly, when LONG TERM data were excluded, the differences between calculated and BQderived LDL-C values were similar to the overall dataset when evaluated by quintiles of BQ-derived LDL-C values (supplemental Table S5). When concordance between calculated and BQ-derived LDL-C values was assessed above and below an LDL-C threshold of 70 mg/dl, the overall discordance after removal of LONG TERM data was 4.2%, 4.2%, and 3.8% for the Friedewald, Martin-Hopkins, and NIH equation 2 methods (supplemental Table S6), respectively, again consistent with the total dataset. Finally, when the LDL-C outliers were compared (absolute difference between calculated and BQ-derived LDL-C values >50 mg/dl) after exclusion of the LONG TERM data, no obvious patterns were observed over time (supplemental Fig. S8).

DISCUSSION

This analysis of the available data in the ODYSSEY trials shows that the discrepancy between Friedewaldcalculated, Martin-Hopkins-calculated, and NIH equation 2-calculated LDL-C with BQ-derived LDL-C values is small and clinically insignificant, with approximately 50% or greater of LDL-C values differing by less than 5 mg/dl. This conclusion includes levels in the lowest quintile of BQ-derived LDL-C values (<40 mg/dl) where the majority of LDL-C values also differed by less than 5 mg/dl (58.1%, 66.5%, and 66.1% for the Friedewald, Martin-Hopkins, and NIH equation 2 methods, respectively). Furthermore, even in the lowest quintile of BQ-derived LDL-C values ($\leq 40 \text{ mg/dl}$), the mean difference between calculated and BQ-derived LDL-C values was relatively small compared with the mean difference for the overall dataset for all three methods. In addition, no impact on the overall correlation between calculated and BQ-derived LDL-C values was observed when analyzed by TG tertiles. Although the correlation between Friedewald-calculated and BQ-derived LDL-C diminished as TGs increased, even in the highest tertile of TGs (>150 mg/dl), the correlation remained high (0.98).

The results are in agreement with those of previous analyses (24–28) as well as current guidelines (1, 2), indicating that Friedewald-calculated LDL-C provides a reasonable estimation of BQ-derived LDL-C in most cases. A previous analysis of samples from patients treated to LDL-C levels <40 mg/dl with the PCSK9

inhibitor evolocumab reported an overall small median absolute difference (4 mg/dl) between Friedewaldcalculated and BQ-derived LDL-C values; greater variation was observed for patients with LDL-C <40 mg/dl and TG levels ≥150 mg/dl, with median absolute difference of 10 mg/dl between Friedewald-calculated and BQ-derived LDL-C values (29). Furthermore, in the previous analysis, for the subgroup of patients with LDL-C <40 mg/dl and TG levels \geq 150 mg/dl, a smaller median difference between calculated and BQ-derived LDL-C was observed for the Martin-Hopkins versus the Friedewald method (2 vs. -10 mg/dl, respectively) (29). In the present analysis, for the subgroup of BQ-derived LDL-C values <40 mg/dl with TGs >150 mg/dl, a greater median difference between calculated LDL-C values and BQ-derived LDL-C was observed for Friedewald (-9 mg/dl) compared with the Martin-Hopkins (3 mg/dl) and NIH equation 2 (-1 mg/dl) methods. Differences across the three methods were also observed for LDL-C <40, <55, or <70 mg/dl and TGs >150 mg/dl and LDL-C <40, <55, or <70 mg/dl and TGs >250 mg/dl (Table 1). For each set of lipid cut points, Friedewald underestimated, Martin-Hopkins overestimated, and NIH equation 2 was essentially the same as the BQ value. Importantly, when TGs were <150 mg/dl, which was the case in the largest group of subjects by far, differences between the three methods for estimation of LDL-C <40, <55, and <70 mg/dl were minimal to nil.

In the present analysis, we found good concordance between calculated and BQ-derived values of LDL-C <70 or \geq 70 mg/dl, with a small proportion (3.6%, 4.0%, and 3.6% for the Friedewald, Martin-Hopkins, and NIH equation 2 methods, respectively) of samples that had a BQ-derived value <70 mg/dl having a corresponding calculated value of \geq 70 mg/dl, or vice versa. Importantly, in the lowest quintile of BQ-derived LDL-C values (<40 mg/dl), there were only very small differences between BQ-derived and calculated LDL-C concentrations, with the majority of LDL-C values only differing by 0–5 mg/dl, as noted above.

We identified a number of outliers where the difference between calculated and BQ-derived LDL-C values was >50 mg/dl; the number of outliers was similar between the Friedewald (45 LDL-C pairs), Martin-Hopkins (63 LDL-C pairs), and NIH equation 2 (51 LDL-C pairs) methods. In total, there were 70 LDL-C pairs that were outliers; the majority (70%; 40 LDL-C pairs) were outliers with all three calculated-LDL-C methods. For most patients with an outlier, there was only one outlying value. We cannot exclude the possibility that there were errors in the original lipid panel and/or BQ measurements at certain time points that led to the observed outlying values.

The 2018 US cholesterol management guidelines (2) state that the Friedewald equation loses accuracy at LDL-C levels <70 mg/dl and suggest that alternative calculation methods may be used, such as those of

Martin *et al.* (5, 9) or measurement of direct LDL-C. The 2019 European guidelines note that alternative modes of calculating LDL-C or direct LDL-C measurement may be considered, but the former have not been shown to be superior to the Friedewald equation in estimating cardiovascular risk, and the latter have limitations of precision and accuracy, especially in patients with high TG levels (1). The recent NIH equation 2 method (6) for calculating LDL-C has been shown to have greater accuracy for patients with hypertriglyceridemia (TGs 400-800 mg/dl) compared with both the Friedewald and Martin equations. In the present analysis, NIH equation 2 was found to be more accurate than both other methods with TGs >150 mg/dl. Results to date have not clearly indicated a benefit for using direct LDL-C over Friedewald in routine use (24, 26, 27, 30, 31). As an alternative to LDL-C, guidelines also recommend that analysis of non-HDL-C and apolipoprotein B may be used to estimate cardiovascular risk (1, 2).

The Martin-Hopkins method is now recommended in the 2018 US cholesterol management guidelines (2) when measurement of LDL-C levels is needed at very low LDL-C. In a previous analysis of clinical data from the FOURIER trial with evolocumab, Martin-Hopkins-calculated LDL-C values were found to have a significantly stronger correlation with BQderived LDL-C values compared with Friedewaldcalculated LDL-C values when Friedewald-calculated LDL-C values were <40 mg/dl (29). In contrast, in the present analysis, an overall similar correlation was observed between Friedewald-calculated, Martin-Hopkins-calculated, and NIH equation 2-calculated LDL-C with BQ-derived LDL-C values (Pearson's correlation coefficient of 0.985 and 0.981, respectively).

As expected, NIH equation 2-calculated and BQderived LDL-C values were also shown to be strongly correlated within the current dataset (Pearson's correlation coefficient = 0.985). In a previous comparison with BQ, NIH equation 2-calculated LDL-C values were found to be more accurate than either the Friedewald or Martin-Hopkins methods for calculating LDL-C for patients particularly with levels, hypertriglyceridemia (6). As the present dataset was limited to TG values <400 mg/dl, this may have resulted in the greater similarity in correlation between NIH equation 2-calculated and Friedewald-calculated LDL-C values observed in the present analyses.

Study limitations

The present analysis utilized pooled data from 10 trials in which patients were randomized to alirocumab, placebo, ezetimibe, or statin; we cannot be certain that the various background therapies did not introduce heterogeneity into the dataset. The standard lipid measurements and those derived by the BQ method were carried out in two different laboratories, with a

40/60% split of samples between the two. In addition, all lipid measurements were taken in the fasted state; therefore, conclusions cannot be extended to lipid assessments of nonfasted samples. Furthermore, we did not compare Friedewald-calculated LDL-C with methods that directly precipitate LDL from serum.

Summary

The results of this analysis suggest that the correlations between Friedewald-calculated, Martin-Hopkins–calculated, and NIH equation 2–calculated LDL-C values with BQ-derived LDL-C values are high, with all three methods providing a suitable alternative to BQderived LDL-C. This remains true even with LDL-C levels <40 mg/dl, levels that can be achieved in many individuals when PCSK9 inhibitors are added to standard statin therapy. When TG levels are >150 mg/dl, which occurs in the top 25%–30% of the population, NIH equation 2 provides the greatest accuracy.

Data availability

Qualified researchers may request access to study documents (including the clinical study report, study protocol with any amendments, blank case report form, statistical analysis plan) that support the methods and findings reported in this article. Individual anonymized participant data will be considered for sharing once the product and indication have been approved by major health authorities (e.g., FDA, EMA, PMDA, etc), if there is legal authority to share the data and there is not a reasonable likelihood of participant re-identification. Submit requests to https://vivli.org/.

Supplemental data

This article contains supplemental data.

Acknowledgments

The authors thank the patients, their families, and all investigators involved in the ODYSSEY studies included in this analysis. Medical writing support under the direction of the authors was provided by Rachel Dunn, PhD, Susanne Ulm, PhD, and Rob Campbell, PhD, of Prime Global (Knutsford, UK), funded by Regeneron Pharmaceuticals, Inc., according to Good Publication Practice guidelines (http://annals.org/aim/article/2424869/good-publication-practice-communi cating-company-sponsored-medical-research-gpp3).

This study was funded by Regeneron Pharmaceuticals, Inc. The sponsors were involved in the study design and collection, analysis, and interpretation of data, as well as data checking of information provided in the article. The authors were involved in the study design and collection, analysis, and interpretation of data. All authors had full access to all the data in this study and take complete responsibility for the integrity of the data and accuracy of the data analysis.

Author contributions

H. N. G. and C. P. C. conceptualization; H. N. G., R. S. R., G. K. H., A. L., R. S., Y. P., and C. P. C. methodology; H. N. G., G. K. H., and C. P. C. investigation; H. N. G., R. S. R., G. K. H.,

A. L., R. S., Y. P., and C. P. C. writing-review and editing; A. L. and Y. P. formal analysis.

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Conflict of interest

H. N. G. has received research grants from Sanofi, Regeneron Pharmaceuticals, Inc., Pfizer, AstraZeneca, and Amgen and modest consultant/advisory board fees from Amarin, Amgen, AstraZeneca, Ionis, Janssen, Kowa, Merck, Novartis, Sanofi, Regeneron Pharmaceuticals, Inc., and Pfizer Inc. R. S. R. has received research grants through his institution from Amgen, Arrowhead, Novartis, and Regeneron Pharmaceuticals, Inc. G. K. H. is a holder of a Vidi Grant (016.156.445) from the Netherlands Organisation for Scientific Research (NWO) and Klinkerpad, has received research support through his institution from Aegerion, Amgen, and Sanofi and speaker fees from Amgen, Aegerion, Sanofi, Regeneron Pharmaceuticals, Inc., and Pfizer. C. P. C. has received research grants from Amgen, Boehringer Ingelheim, Bristol-Myers Squibb, Daiichi Sankyo, Janssen, Merck, and Pfizer and consulting fees from Aegerion, Alnylam, Amarin, Amgen, Applied Therapeutics, Ascendia, Boehringer Ingelheim, Bristol-Myers Squibb, Corvidia, HLS Therapeutics, Innovent, Janssen, Kowa, Merck, Pfizer, and Sanofi. R. S. R. serves on advisory boards from Amgen, Amyrt, C5, CVS Caremark, and Regeneron Pharmaceuticals, Inc., receives honoraria from Amgen, Kowa, and Regeneron Pharmaceuticals, Inc., receives royalties from UpToDate, and has stock ownership in MediMergent. G. K. H. has served as a consultant for Amgen, Aegerion, Sanofi, Regeneron Pharmaceuticals, Inc., and Pfizer and has been employed part-time by Novo Nordisk AS and Copenhagen, Denmark, since April 2019. A. L. is an employee of and stockholder in Sanofi. R. S. is an employee of and stockholder in Regeneron Pharmaceuticals, Inc. Y. P. is employed by a company that is contracted to Sanofi. All other authors declare that they have no conflicts of interest with the contents of this article.

Abbreviations

BQ, beta-quantification; PCSK9, proprotein convertase subtilisin/kexin type 9.

Manuscript received June 16, 2021, and in revised from November 3, 2021. Published, JLR Papers in Press, November 11, 2021, https://doi.org/10.1016/j.jlr.2021.100148

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