

OPEN

# Diagnostic Accuracy of Procalcitonin in Bacterial Meningitis Versus Nonbacterial Meningitis

## *A Systematic Review and Meta-Analysis*

Ting-Ting Wei, MM, Zhi-De Hu, MM, Bao-Dong Qin, MM, Ning Ma, MM, Qing-Qin Tang, MM, Li-Li Wang, MM, Lin Zhou, MD, PhD, and Ren-Qian Zhong, MD, PhD

**Abstract:** Several studies have investigated the diagnostic accuracy of procalcitonin (PCT) levels in blood or cerebrospinal fluid (CSF) in bacterial meningitis (BM), but the results were heterogeneous.

The aim of the present study was to ascertain the diagnostic accuracy of PCT as a marker for BM detection.

A systematic search of the EMBASE, Scopus, Web of Science, and PubMed databases was performed to identify studies published before December 7, 2015 investigating the diagnostic accuracy of PCT for BM. The quality of the eligible studies was assessed using the revised Quality Assessment for Studies of Diagnostic Accuracy method. The overall diagnostic accuracy of PCT detection in CSF or blood was pooled using the bivariate model.

Twenty-two studies involving 2058 subjects were included in this systematic review and meta-analysis. The overall specificities and sensitivities were 0.86 and 0.80 for CSF PCT, and 0.97 and 0.95 for blood PCT, respectively. Areas under the summary receiver operating characteristic curves were 0.90 and 0.98 for CSF PCT and blood PCT, respectively.

The major limitation of this systematic review and meta-analysis was the small number of studies included and the heterogeneous diagnostic thresholds adopted by eligible studies.

Our meta-analysis shows that PCT is a useful biomarker for BM diagnosis.

(*Medicine* 95(11):e3079)

**Abbreviations:** AMa = acute meningitis, AUC = area under curve, BM = bacterial meningitis, CSF = cerebrospinal fluid, DOR = diagnostic odds ratio, FP = false-positive, PCT = procalcitonin, SROC = summary receiver operating characteristic, TP = true-positive.

Editor: Mihalis Panagiotidis.

Received: November 11, 2015; revised: December 30, 2015; accepted: February 23, 2016.

From the Department of Laboratory Diagnostics, Changzheng Hospital, The Second Military Medical University (T-TW, Z-DH, B-DQ, NM, Q-QT, L-LW, LZ, R-QZ); and Department of Laboratory Medicine, The General Hospital, Ji'nan Military Region of PLA, Ji'nan, Shandong, China (Z-DH). Correspondence: Ren-Qian Zhong, Lin Zhou, Department of Laboratory Diagnostics, Changzheng Hospital, The Second Military Medical University, Shanghai 200003, China

(e-mail: 13901628473@163.com, lynnzhou36@163.com).

T-TW and Z-DH contributed equally to this work.

This study program was supported by grants from the National Natural Science Foundation of China (81302541, 81471608).

The authors have no conflicts of interest to disclose.

Copyright © 2016 Wolters Kluwer Health, Inc. All rights reserved.

This is an open access article distributed under the Creative Commons Attribution-NonCommercial-NoDerivatives License 4.0, where it is permissible to download, share and reproduce the work in any medium, provided it is properly cited. The work cannot be changed in any way or used commercially.

ISSN: 0025-7974

DOI: 10.1097/MD.0000000000003079

## INTRODUCTION

Acute meningitis (AM) is an extremely severe and life-threatening infection, and early diagnosis and prompt treatment are critically important for AM patients due to the high rates of mortality and morbidity associated with the infection.<sup>1</sup> AM is classified into bacterial meningitis (BM) and nonbacterial meningitis (NBM). Differentiation of BM from NBM is critical for early and prompt intervention for BM patients. Furthermore, differentiation of BM from NBM helps avoid unnecessary hospitalization, antibiotic abuse, and increased medical burden. However, differentiating the 2 forms of AM is challenging because they share many similar clinical symptoms, such as fever and headache.<sup>2</sup> Positive cerebrospinal fluid (CSF) bacterial culture, Gram staining, or detection of bacterial antigens in the CSF represent the gold standard of clinical testing in BM diagnosis. However, although they have high specificity, the sensitivity is poor. Furthermore, bacterial culture is time-consuming. The serum and CSF markers currently used as supplementary markers in BM diagnosis, such as C-reactive protein, are also characterized by inadequate sensitivity and specificity.<sup>3,4</sup> Therefore, discovery of more sensitive and specific markers for BM is desirable.

Procalcitonin (PCT) is a 116-amino-acid protein that is produced primarily by the C cells of the thyroid gland and secreted from leukocytes in the peripheral blood.<sup>5</sup> In healthy individuals, PCT is secreted at levels that are below the detectable limit. However, serum PCT levels increase markedly in patients suffering from bacterial infections.<sup>6</sup> Therefore, elevated PCT levels may serve as useful diagnostic markers for BM.<sup>7</sup> During the past decades, many studies have investigated the diagnostic accuracy of serum or CSF PCT in BM. However, the results were not unequivocal. Therefore, we performed a systematic review and meta-analysis to ascertain the diagnostic value of serum and CSF PCT in BM.

## MATERIAL AND METHODS

### Literature Search

Using the search terms “(PCT or procalcitonin) and meningitis”, the authors ZDH and TTW independently searched PubMed, Scopus, Web of Science, and EMBASE to identify eligible studies published before December 7, 2015. Manual searches were also conducted by reviewing the references of the eligible studies. The 2 authors (ZDH and TTW) independently reviewed the titles and abstracts of all studies retrieved by independent searches to identify potentially eligible studies. If necessary, a full-text review was conducted, and any disagreements concerning study selection were resolved by full-text review. Since our work is based on available studies, patient consent was waived.

## Inclusion and Exclusion Criteria

Inclusion criteria were: studies that evaluated the diagnostic accuracy of PCT for BM in CSF or blood; sample size of BM or NBM patients greater than 10, to avoid selection bias; and 2 by 2 tables constructed from the reported sensitivity and specificity values. Exclusion criteria were: animal studies; non-English publications; and conference abstracts.

## Data Extraction and Quality Assessment

Publication years, national origin, BM or NBM patient status, PCT-testing methods, references, area under the receiver operating characteristic curve (area under curve [AUC]), and PCT detection thresholds were independently extracted in duplicate by ZDH and TTW. A 3rd reviewer resolved any discrepancies or disagreements between the independently extracted datasets. The true-positive (TP), false-positive (FP), false-negative (FN), and true-negative (TN) rates were calculated according to the BM and NBM sample size based on the reported sensitivity and specificity of each study as follows: TP = number of BM patients  $\times$  sensitivity; FN = number of BM patients  $\times$  (1 – sensitivity); TN = number of NBM patients  $\times$  specificity; FP = number of NBM patients  $\times$  (1 – specificity).

TTW and ZDH independently assessed the eligible studies using the revised Quality Assessment for Studies of Diagnostic Accuracy tool.<sup>8</sup> The items or domains were labeled as unknown in Quality Assessment for Studies of Diagnostic Accuracy tool if the corresponding design characteristics were not reported. Any disagreement in quality assessment was resolved by consensus.

## Statistical Analysis

This meta-analysis was performed and reported in accordance with the Preferred Reporting Items for Systematic Reviews and Meta-Analyses (PRISMA) guidelines for systematic reviews and meta-analyses (Table S1).<sup>9</sup> Data for the overall diagnostic sensitivity and specificity of PCT for meningitis were pooled using the bivariate model. The bivariate model uses paired sensitivity and specificity as the starting point of analysis and may represent a more reliable indicator of diagnostic accuracy of an index test in meta-analysis when compared with the traditional summary receiver operating characteristic (SROC) approach, which uses the diagnostic odds ratio (DOR) as the main outcome measure.<sup>10</sup> Additionally, since the bivariate model uses a random effects approach for both specificity and sensitivity, the degree of heterogeneity beyond chance may be attributed to clinical and methodological differences between studies. Pooled positive and negative likelihood ratios were calculated according to the summary estimates of sensitivity and specificity. Funnel plots and Deeks test were used to test for potential publication bias.<sup>11</sup> All analyses were performed using Stata 13.0<sup>12</sup> (Stata Corp LP, College Station, TX), and a *P* value less than 0.05 was considered statistically significant.

## RESULTS

### Study Eligibility

Twenty-two studies were included in this systematic review and meta-analysis.<sup>13–34</sup> A flowchart of the eligible studies is shown in Supplementary Figure 1, and the characteristics of the studies included in this report are summarized in Table 1. Nine of the included studies were conducted in Asia<sup>21–23,26–28,32–34</sup> and

11 were conducted in Europe.<sup>13–20,25,29,30</sup> The sample sizes in each study ranged from 30 to 254, with a total combined sample size of 2058. To evaluate the efficacy of PCT measurement in BM diagnosis, 2 of the studies<sup>30,33</sup> investigated the diagnostic performance of CSF PCT detection, 17 of the studies focused on serum or plasma PCT detection,<sup>13–15,17–22,24–29,31,32</sup> and 3 of the studies focused on both serum and CSF PCT detection.<sup>16,23,34</sup> Two of the studies enrolled neurosurgery patients,<sup>27,33</sup> 8 studies enrolled pediatric patients,<sup>13,20–22,24,26,29,32</sup> and 9 studies included adult patients.<sup>14,15,17–19,23,25,31,34</sup> The remaining 3 studies<sup>16,28,30</sup> did not report the demographics of the enrolled patients. The references used for BM diagnosis varied among the included studies. All studies set CSF culture as an item of reference, and a few studies set one or more of the following as additional items of reference: CSF Gram staining, blood culture, CSF antigen test, clinical signs or symptoms, and laboratory findings. Thirteen of the studies used the immunoluminometric assay (ILMA) LUMI test (BRAHMS Diagnostica, Berlin, Germany) to determine PCT,<sup>13–17,20–24,26,30,33</sup> 2 used commercial VIDAS PCT assays,<sup>27,33</sup> 1 used a commercial Elecsys PCT assay,<sup>29</sup> 3 used commercial Kryptor PCT assays,<sup>18,19,25</sup> and 2 used commercial Raybiotech PCT assays.<sup>28,31</sup> Fourteen of the studies<sup>14–18,21,24–28,31,32,34</sup> were prospective and 3 of the studies<sup>20,29,33</sup> were retrospective. The remaining 5 studies<sup>13,19,22,23,30</sup> did not report whether their data collection was prospective or retrospective.

### Quality Assessment of Eligible Studies

Table 2 lists the quality assessment of eligible studies. The patient selection method is unknown in 6 of the studies,<sup>16,19,21,22,27,29,30</sup> because the authors failed to report whether the subjects were enrolled consecutively or randomly. The patient selection domain was labeled “high” in 7 studies because healthy individuals were enrolled in the study,<sup>24,28</sup> the study included appropriate exclusion criteria<sup>18,25,31</sup> or the authors mentioned retrospective design.<sup>17,20</sup> The index test domain was labeled “unknown” in 7 studies due to small sample sizes and the lack of a report by the authors indicating whether or not the thresholds were prespecified.<sup>16,24,26–28,31,32</sup> The index test domain was labeled “high” in 3 studies because the threshold was not prespecified.<sup>14,18,29</sup> The reference standard domain of all eligible studies, except for one,<sup>32</sup> was labeled “low” because the reference standard that was used in each eligible study correctly classified the BM and was interpreted without knowledge of the PCT results. The follow-up and timing domains of 6 studies were labeled “unknown” because it was uncertain whether partial verification bias was avoided in those studies.<sup>14,16,18,19,21–25</sup> The follow-up and timing domains were labeled “high” in 4 studies because not all patients were included in their analysis.<sup>17,20,27,29</sup>

### Diagnostic Accuracy of PCT

Table 3 summarizes the diagnostic accuracy of all eligible studies. The overall diagnostic accuracy was pooled using the bivariate model. A forest plot depicting the diagnostic sensitivity and specificity of blood PCT and CSF PCT detection is illustrated in Figure 1. Overall, the diagnostic sensitivity of CSF PCT detection was 0.80 (95% CI, 0.61–0.91), specificity was 0.86 (95% CI, 0.70–0.95), positive likelihood ratio (PLR) was 5.9 (95% CI, 2.4–14.0), negative likelihood ratios (NLR) was 0.23 (95% CI, 0.12–0.47), and DOR was 25 (95% CI, 8–78).<sup>12</sup> I<sup>2</sup> across all eligible CSF PCT studies was 0.67 (95% CI, 0.26–1.00), and only 9% of the observed heterogeneity was attributed

**TABLE 1.** Summary of Eligible Studies

Author	Year	Country	No	Subjects Characteristics	References*					Assay	Matrix	Data Collection
					C	G	A	B	O			
Gendrel et al <sup>13</sup>	1997	France	59	Pediatric meningitis	●	–	–	–	–	Lumi test	Plasma	Unknown
Viallon et al <sup>14</sup>	1999	France	80	Suspected meningitis	●	●	–	–	–	Lumi test	Serum	Prospective
Schwarz et al <sup>15</sup>	2000	Germany	30	Meningitis	●	●	●	●	●	Lumi test	Serum	Prospective
Jereb et al <sup>16</sup>	2001	Slovenia	45	NR	●	●	–	●	–	Lumi test	CSF and serum	Prospective
Dubos et al <sup>17</sup>	2006	France	152	Suspected meningitis	●	●	●	●	–	Lumi test	Serum	Retrospective
Ray et al <sup>18</sup>	2007	France	151	Meningitis and a negative Gram-stained smear	●	–	●	●	●	Kryptor	Serum	Prospective
Knudsen et al <sup>19</sup>	2007	Denmark	52	Suspected meningitis	●	●	●	–	●	Kryptor	Serum	Unknown
Dubos et al <sup>20</sup>	2008	Europe	190	Pediatric meningitis	●	●	●	●	●	Lumi test	Serum	Retrospective
Onal et al <sup>21</sup>	2008	Turkey	30	Pediatric meningitis	●	●	●	–	●	Lumi test	Plasma	Prospective
Steinberg et al <sup>22</sup>	2010	Russia	232	Pediatric meningitis	●	●	●	–	–	Lumi test	Serum	Unknown
Makoo et al <sup>23</sup>	2010	Iran	50	Meningitis	●	●	–	–	●	Lumi test	CSF and serum	Unknown
Alkholi et al <sup>24</sup>	2011	Egypt	50	Pediatric meningitis	●	●	–	–	●	Lumi test	Serum	Prospective
Viallon et al <sup>25</sup>	2011	France	254	Suspected meningitis and a Negative Gram-stained smear or antigen detection	●	–	–	–	–	Kryptor	Serum	Prospective
Ibrahim et al <sup>26</sup>	2011	Saudi	43	Pediatric meningitis	●	●	–	–	●	Lumi test	Serum	Prospective
Choi and Choi <sup>27</sup>	2013	Korea	44	Patients underwent neurosurgery and had CSF pleocytosis	●	–	–	–	–	VIDAS	Serum	Prospective
Prasad et al <sup>28</sup>	2013	India	70	NR	●	●	–	–	●	Raybiotech	Serum	Prospective
Casado et al <sup>29</sup>	2014	Spain	85	Pediatric meningitis	●	–	–	●	●	Elecsys	Serum	Retrospective
Konstantinidis et al <sup>30</sup>	2014	Greece	58	NR	●	–	–	–	●	Lumi test	CSF	Unknown
Abdelkader et al <sup>31</sup>	2014	Egypt	40	Acute meningitis and a negative Gram-stained smear	●	–	–	–	●	Raybiotech	Serum	Prospective
Umran and Radhi <sup>32</sup>	2014	Iraq	45	Suspected pediatric meningitis	–	–	–	–	●	ELIZA M6	Serum	Prospective
Li et al <sup>33</sup>	2014	China	178	Patients underwent neurosurgery with clinical symptoms of meningitis	●	●	–	–	●	VIDAS	CSF	Retrospective
Shen et al <sup>34</sup>	2015	China	120	Suspected meningitis	●	●	–	–	●	Lumi test	CSF and serum	Prospective

A black dot (●) indicates inclusion of the corresponding items in the reference standard, while “–” indicates exclusion of the corresponding item from the reference standard. CSF = cerebrospinal fluid, NR = not reported.

\*Reference composition: A = CSF antigen test, B = blood culture, C = CSF bacterial culture, G = Gram stain, O = others.

to the threshold effect. The overall diagnostic sensitivity of blood PCT detection was 0.95 (95% CI, 0.89–0.97), specificity was 0.97 (95% CI, 0.89–0.99), PLR was 31.7 (95% CI, 8.0–124.8), NLR was 0.06 (95% CI, 0.03–0.11), DOR was 568 (95% CI, 103–3141). I<sup>2</sup> across all eligible studies was 0.96 (95% CI, 0.92–0.99). It is likely that only 27% of the observed heterogeneity was due to the threshold effect.

The SROC curves for CSF PCT and blood PCT are shown in Figure 2. The AUCs for CSF PCT and blood PCT were 0.90 (95% CI, 0.87–0.92) and 0.98 (95% CI, 0.97–0.99), respectively. The 95% CIs for the AUCs of CSF PCT and blood PCT did not overlap, indicating that the overall diagnostic accuracy of blood PCT detection was superior to CSF PCT.

The diagnostic sensitivity, specificity, PLR, NLR, DOR, AUCs for SROCs, I<sup>2</sup>, and proportion of heterogeneity attributed to the threshold effect for blood PCT and CSF PCT are listed in Table 4.

We next analyzed the CSF PCT or blood PCT posttest probability of BM. As shown in Figure 3, the pretest probabilities of BM for blood PCT and CSF PCT were 0.34 and 0.36,

respectively. The posttest probabilities of BM after a positive CSF PCT or blood PCT test were 0.77 and 0.94, respectively. The posttest probability values associated with negative CSF PCT or blood PCT tests were 0.12 and 0.03, respectively.

**Subgroup Analysis and Meta-Regression**

Sources of significant heterogeneity between studies investigating blood PCT were determined using subgroup analysis and meta-regression. The type of data collection (prospective or retrospective) was the source of heterogeneity in sensitivity (P < 0.01; Figure 4). In the joint model, none of the study characteristics (data collection, age, test assay, and subject sources) represented sources of heterogeneity.

**Publication Bias**

The funnel plots for publication bias were asymmetrical (Figure 5), suggesting significant publication bias. The statistical significance of this publication bias for both CSF PCT and blood PCT was confirmed using Deeks test (P < 0.05 for both).

**TABLE 2.** Quality Assessment of Eligible Studies

Study	Risk of Bias				Applicability Concerns		
	Patient Selection	Index Test	Reference Standard	Flow and Timing	Patient Selection	Index Test	Reference Standard
Gendrel et al <sup>13</sup>	Low	Low	Low	Low	Low	Low	Low
Viallon et al <sup>14</sup>	Low	High	Low	Unknown	Low	Low	Low
Schwarz et al <sup>15</sup>	Low	Low	Low	Low	Low	Low	Low
Jereb et al <sup>16</sup>	Unknown	Unknown	Low	Unknown	Low	Low	Low
Dubos et al <sup>17</sup>	High	Low	Low	High	Low	Low	Low
Ray et al <sup>18</sup>	High	High	Low	Unknown	Low	Low	Low
Knudsen et al <sup>19</sup>	Unknown	Low	Low	Unknown	Low	Low	Low
Dubos et al <sup>20</sup>	High	Low	Low	High	Low	Low	Low
Onal et al <sup>21</sup>	Unknown	Low	Low	Unknown	Low	Low	Low
Steinberg et al <sup>22</sup>	Unknown	Low	Low	Unknown	Low	Low	Low
Makoo et al <sup>23</sup>	Low	Low	Low	Unknown	Low	Low	Low
Alkholi et al <sup>24</sup>	High	Unknown	Low	Unknown	High	Low	Low
Viallon et al <sup>25</sup>	High	Low	Low	Unknown	Low	Low	Low
Ibrahim et al <sup>26</sup>	Low	Unknown	Low	Low	Low	Low	Low
Choi and Choi <sup>27</sup>	Unknown	Unknown	Low	High	Low	Low	Low
Prasad et al <sup>28</sup>	High	Unknown	Low	Low	High	Low	Low
Casado et al <sup>29</sup>	Unknown	High	Low	High	Low	Low	Low
Konstantinidis et al <sup>30</sup>	Unknown	Low	Low	Low	Low	Low	Low
Abdelkader et al <sup>31</sup>	High	Unknown	Low	Low	Low	Low	Low
Umran and Radhi <sup>32</sup>	Low	Unknown	High	Low	Low	Low	High
Li et al <sup>33</sup>	Low	Low	Low	Low	Low	Low	Low
Shen et al <sup>34</sup>	Low	Low	Low	Low	Low	Low	Low

**TABLE 3.** Meta-Analysis: Key Findings of Eligible Studies

Author	BM/NBM	AUC	Sensitivity	Specificity	Assay	Thresholds, ng/mL	TP	FN	FP	TN
CSF										
Li et al <sup>33</sup>	50/128	0.746	0.68	0.73	VADAS	0.08	34	16	35	93
Konstantinidis et al <sup>30</sup>	19/11	–	1.00	0.73	Lumitest	0.50	19	0	3	8
Jereb et al <sup>16</sup>	20/25	–	0.55	1.00	Lumitest	0.50	11	9	0	25
Makoo et al <sup>23</sup>	19/31	–	0.84	0.94	Lumitest	0.50	16	3	2	29
Shen et al <sup>34</sup>	45/75	0.90	0.82	0.81	Lumitest	0.50	37	8	14	61
Serum										
Choi and Choi <sup>27</sup>	14/30	0.65	0.50	0.80	VADAS	0.15	7	7	6	24
Jereb et al <sup>16</sup>	20/25	–	0.90	1.00	Lumitest	0.50	18	2	0	25
Alkholi et al <sup>24</sup>	20/30	–	1.00	0.66	Lumitest	2.00	20	0	10	20
Ray et al <sup>18</sup>	18/133	0.98	0.87	1.00	Kryptor	2.13	16	2	0	133
Schwarz et al <sup>15</sup>	16/14	–	0.69	1.00	Lumitest	0.50	11	5	0	14
Knudsen et al <sup>19</sup>	10/42	0.75	0.90	0.57	Kryptor	0.25	9	1	18	24
Casado et al <sup>29</sup>	38/47	0.99	0.97	1.00	Elecsys	0.53	37	1	0	47
Viallon et al <sup>25</sup>	35/218	0.99	0.97	1.00	Kryptor	0.28	34	1	0	218
Viallon et al <sup>14</sup>	23/57	1.00	1.00	1.00	Lumitest	0.20	23	0	0	57
Dubos et al <sup>20</sup>	90/100	0.98	0.99	0.83	Lumitest	0.50	89	1	17	83
Dubos et al <sup>17</sup>	18/134	0.95	0.89	0.89	Lumitest	0.50	16	2	15	119
Gendrel et al <sup>13</sup>	18/41	–	0.94	1.00	Lumitest	5.00	17	1	0	41
Ibrahim et al <sup>26</sup>	18/20	–	0.95	0.94	Lumitest	0.50	17	1	1	19
Abdelkader et al <sup>31</sup>	16/24	–	0.69	0.83	Raybiotech	1.20	11	5	4	20
Prasad et al <sup>28</sup>	40/15	0.90	0.92	0.67	Raybiotech	15.00	38	2	5	10
Makoo et al <sup>23</sup>	19/31	–	1.00	0.88	Lumi test	0.50	19	0	4	27
Onal et al <sup>21</sup>	16/14	–	0.93	1.00	Lumi test	0.50	15	1	0	14
Steinberg et al <sup>22</sup>	106/126	–	1.00	1.00	Lumi test	0.50	106	0	0	126
Umran and Radhi <sup>32</sup>	29/16	0.77	0.79	0.81	ELIZA M6	0.05	23	6	3	13
Shen et al <sup>34</sup>	45/75	0.96	0.98	0.65	Lumi test	0.50	43	2	26	49

AUC = area under receiver operating characteristic curves, BM = bacterial meningitis, CSF = cerebrospinal fluid, FN = false negative, FP = false positive, NBM = nonbacterial meningitis, TN = true negative, TP = true positive.

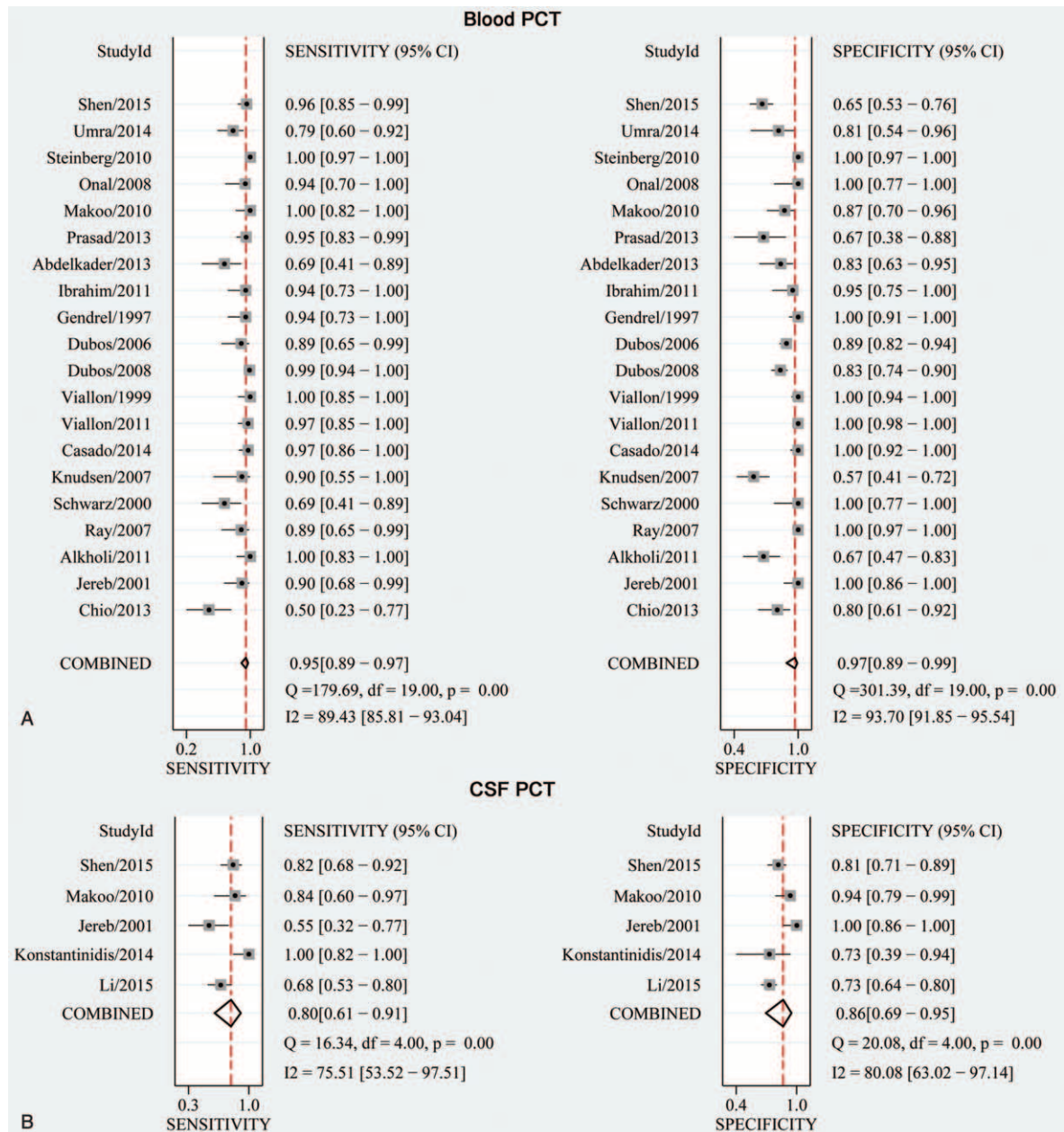


FIGURE 1. Forest plot of the sensitivity and specificity of PCT for BM diagnosis. BM = bacterial meningitis, PCT = procalcitonin.

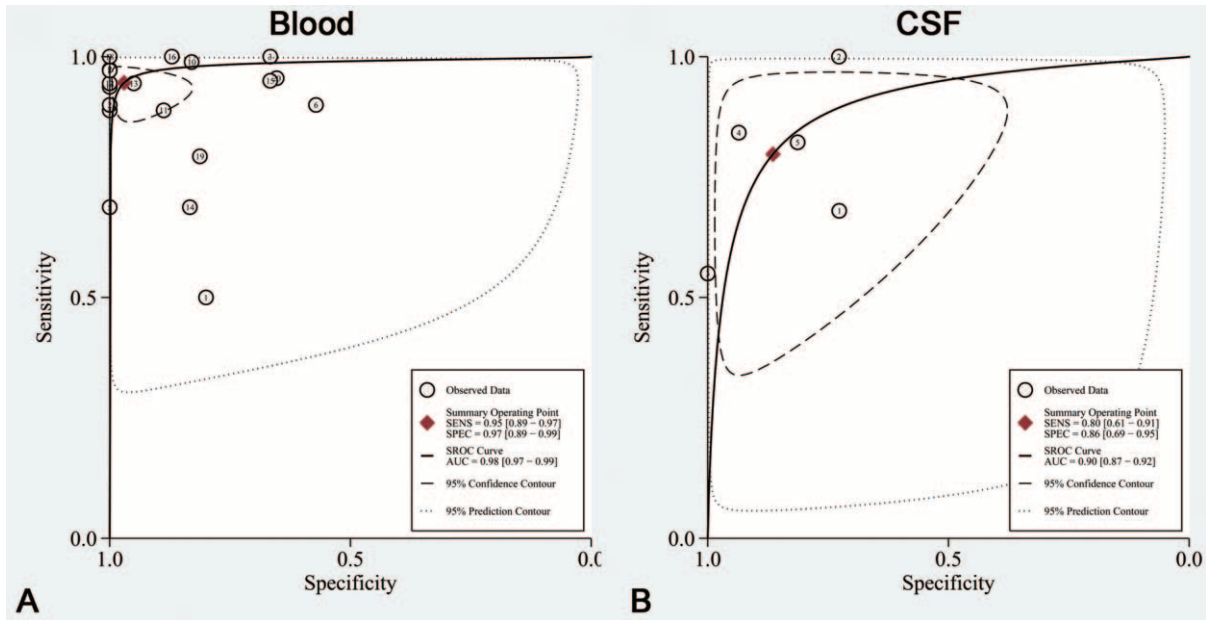
**DISCUSSION**

The results of this meta-analysis indicate that CSF PCT and blood PCT were both effective biomarkers for BM diagnosis. The diagnostic accuracy of elevated blood PCT appeared to be superior to CSF PCT. Additional bias associated with patient selection and partial verification was the major flaw in the design of the eligible studies. Publication bias existed across all eligible studies.

Two meta-analyses investigated the diagnostic value of PCT for BM.<sup>35,36</sup> Compared with the 2 studies, the strengths of our work are as follows. First, we used a bivariate model instead of a random-effects model to pool the sensitivity and specificity in studies. Therefore, the results of our work are more reliable.

Second, previous studies only investigated the diagnostic value of serum PCT for BM, while our study investigated the diagnostic value of both serum and CSF PCT for BM, and therefore, our work is more informative.

We found that blood PCT was associated with a higher pooled sensitivity and specificity when compared with CSF PCT. This finding suggests that blood PCT has superior diagnostic potential when compared with CSF PCT. Furthermore, the superior diagnostic potential of blood PCT was confirmed by SROC analysis, which indicated that the AUCs for CSF PCT were lower than that of blood PCT. Although no single statistical method compared the AUCs of the SROCs, we found no overlap between the 95% CI of the AUCs for CSF PCT and



**FIGURE 2.** The SROC AUC of PCT in BM diagnosis. The overall diagnostic efficiency of PCT in BM is summarized by the regression curve. AUC = area under curve, BM = bacterial meningitis, PCT = procalcitonin, SROC = summary receiver operating characteristic.

blood PCT, demonstrating that the overall diagnostic accuracy of blood PCT was superior to CSF PCT.

The DOR is an independent indicator of test accuracy that compares the odds of TP patients with the odds of FPs.<sup>37</sup> The results of the test range from 0 to infinity, and higher values indicate a better discriminatory test performance.<sup>38</sup> The present meta-analysis yielded DOR values of 568 and 25 for blood PCT and CSF PCT, respectively, indicating that both CSF PCT and blood PCT were effective markers for BM diagnosis. Furthermore, the results indicate that the diagnostic accuracy of blood PCT was superior to CSF PCT.

The pooled PLRs and NLRs are more clinically useful than the sensitivity, specificity, DOR, or AUC. Positive likelihood ratios greater than 10 or negative likelihood ratios below 0.1 generate large and often conclusive shifts from pre- to posttest probability (indicating high accuracy). We found that the PLR for blood PCT was 31.7 indicating that patients with BM have an approximately 32-fold higher chance of being PCT positive

when compared with BM negative patients. Conversely, we found that the NLR for blood PCT was 0.06, suggesting that a negative blood PCT result was associated with a mere 6% probability that the patient had BM.

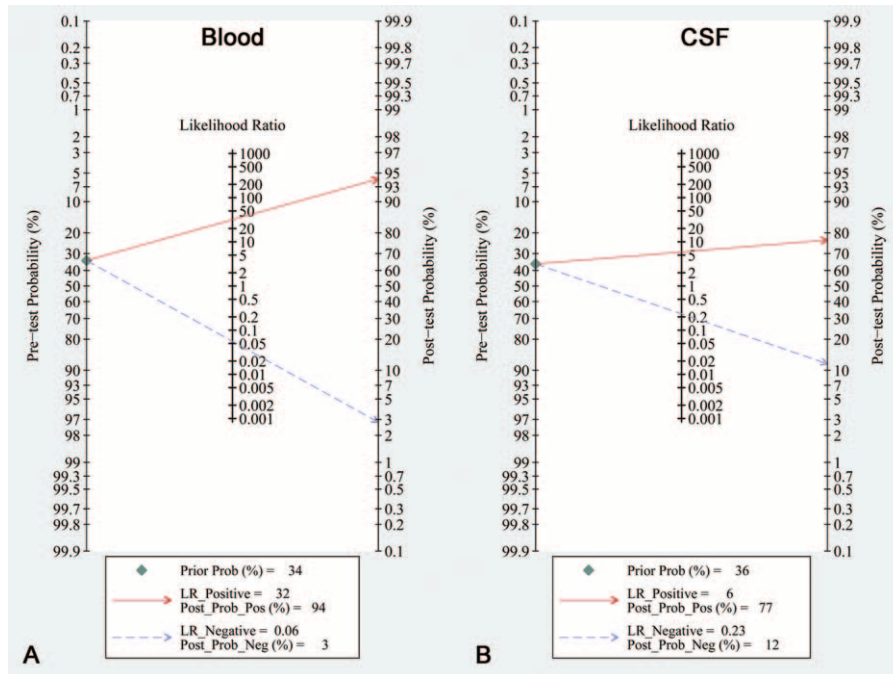
The Fagan nomogram also confirmed the extremely high diagnostic accuracy of blood PCT for BM. The BM pretest was approximately 0.36; however, the posttest probabilities associated with positive and negative PCT were 0.94 and 0.03, respectively. The results indicate that the probability of BM was as high as 94% for patients who tested positive for PCT, but only 3% for patients negative for PCT. These results indicate that positive blood PCT can be used to confirm a diagnosis of BM, while negative blood PCT alone is sufficient to rule out BM. The PLR and NLR for CSF were 5.9 and 0.23, respectively, indicating that CSF PCT alone is insufficient to confirm or rule out BM.

The turnaround time for blood PCT or CSF PCT analysis is shorter than that of traditional bacterial culture. Compared with

**TABLE 4.** Overall Diagnostic Characteristics Associated with Blood PCT and CSF PCT

	Blood PCT	CSF PCT
Number of studies	20	5
Bacterial/nonbacterial	609/1192	153/270
Area under the SROC curve (95% CI)	0.98 (0.97–0.99)	0.90 (0.87–0.92)
Sensitivity (95% CI)	0.95 (0.89–0.97)	0.80 (0.61–0.91)
Specificity (95% CI)	0.97 (0.89–0.99)	0.86 (0.70–0.95)
Positive likelihood ratio (95% CI)	31.7 (8.0–124.8)	5.9 (2.4–14.0)
Negative likelihood ratio (95% CI)	0.06 (0.03–0.11)	0.23 (0.12–0.47)
Diagnostic odds ratio (95% CI)	568 (103–3141)	25 (8–78)
Inconsistency ( $I^2$ ) (95% CI)	0.96 (0.92–0.99)	0.67 (0.26–1.00)

CI = confidence interval, CSF = cerebrospinal fluid, PCT = procalcitonin, SROC = summary receiver operating characteristic.



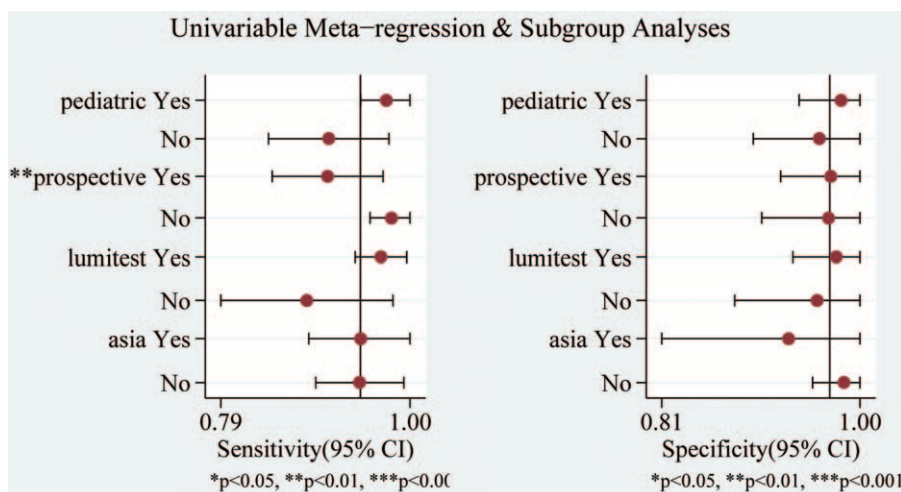
**FIGURE 3.** Fagan nomogram of the blood PCT and CSF PCT tests for BM diagnosis. BM = bacterial meningitis, CSF = cerebrospinal fluid, PCT = procalcitonin.

CSF Gram staining, PCT is an objective test that can be reliability implemented independent of laboratory technical expertise. The present meta-analysis revealed a high level of heterogeneity among all the eligible studies, and only a small portion of the heterogeneity was explained by the threshold effect. Meta-regression analysis revealed that the type of data collection (prospective or retrospective), the age of subjects (pediatric or nonpediatric), test assay (Lumi test), and sources of subjects (Asian or other) were not the source of heterogeneity. Further studies are needed to explore the sources of heterogeneity.

Our analysis revealed the following design flaws in the eligible studies:

Some of the eligible studies failed to incorporate inclusion and exclusion criteria.<sup>39</sup> Additionally, they also failed to report whether or not the subjects were consecutively enrolled. Because these design flaws resulted in subject populations that may not reflect clinical reality, they introduced a large amount of bias.<sup>8,40</sup>

Partial verification bias was not completely ruled out in some of the eligible studies as they usually confirmed the diagnosis of BM using microbiological examination (reference), but failed to report whether other types of meningitis were excluded. Therefore, additional well-designed studies are needed to rigorously assess the diagnostic accuracy of PCT for BM.



**FIGURE 4.** Subgroup analysis of PCT sensitivity and specificity in BM diagnosis. BM = bacterial meningitis, PCT = procalcitonin.

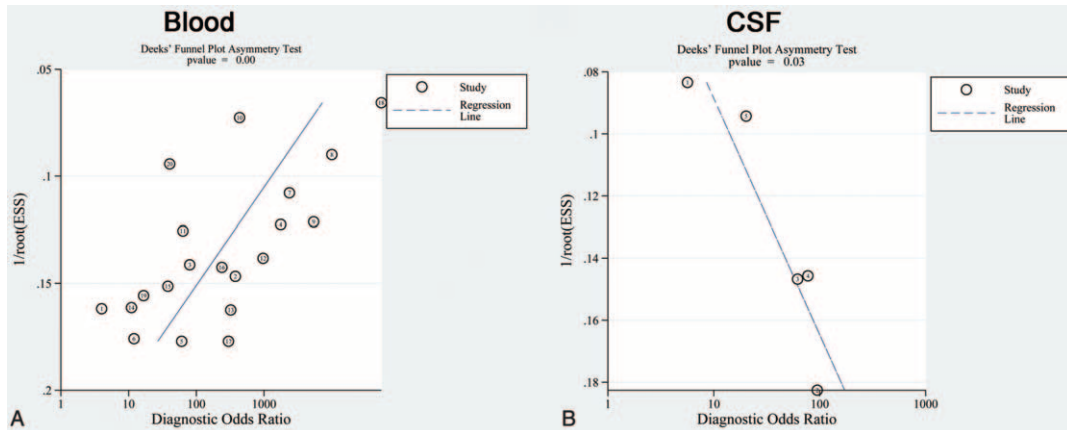


FIGURE 5. Funnel plot of potential publication bias. Each solid rectangle in the funnel plot represents an eligible study.

Some of the limitations of this meta-analysis are related to the small sample sizes, especially among studies investigating the diagnostic value of CSF PCT. Furthermore, the thresholds in eligible studies were not consistent, which may be due to various PCT assays used in eligible studies. Finally, our analysis indicated the presence of publication bias, indicating that this report may overestimate the diagnostic accuracy of PCT for BM.

In summary, this meta-analysis reveals that both CSF PCT and blood PCT are effective diagnostic markers for BM. However, blood PCT appears to exhibit superior diagnostic accuracy when compared with CSF PCT, and blood PCT alone is sufficient to confirm or exclude BM diagnosis. Additional well-designed studies are needed to corroborate the results of this meta-analysis.

#### ACKNOWLEDGMENTS

The authors thank Medjaden Bioscience Limited, Hong Kong, China, for assisting with the preparation of this manuscript. The authors also thank the study program supported by grants from the National Natural Science Foundation of China (81302541, 81471608).

#### REFERENCES

- Spanos A, Harrell FE Jr, Durack DT. Differential diagnosis of acute meningitis. An analysis of the predictive value of initial observations. *JAMA*. 1989;262:2700–2707.
- van de Beek D, de Gans J, Spanjaard L, et al. Clinical features and prognostic factors in adults with bacterial meningitis. *N Engl J Med*. 2004;351:1849–1859.
- Mekitarian Filho E, Horita SM, Gilio AE, et al. Cerebrospinal fluid lactate level as a diagnostic biomarker for bacterial meningitis in children. *Int J Emerg Med*. 2014;7:14.
- White K, Ostrowski K, Maloney S, et al. The utility of cerebrospinal fluid parameters in the early microbiological assessment of meningitis. *Diagn Microbiol Infect Dis*. 2012;73:27–30.
- Meisner M. Update on procalcitonin measurements. *Ann Lab Med*. 2014;34:263–273.
- Wacker C, Prkno A, Brunkhorst FM, et al. Procalcitonin as a diagnostic marker for sepsis: a systematic review and meta-analysis. *Lancet Infect Dis*. 2013;13:426–435.
- McCann FJ, Chapman SJ, Yu WC, et al. Ability of procalcitonin to discriminate infection from non-infective inflammation using two pleural disease settings. *PLoS One*. 2012;7:e49894.
- Whiting PF, Rutjes AW, Westwood ME, et al. QUADAS-2: a revised tool for the quality assessment of diagnostic accuracy studies. *Ann Intern Med*. 2011;155:529–536.
- Moher D, Liberati A, Tetzlaff J, et al. Preferred reporting items for systematic reviews and meta-analyses: the PRISMA statement. *Ann Intern Med*. 2009;151:264–269W264.
- Reitsma JB, Glas AS, Rutjes AW, et al. Bivariate analysis of sensitivity and specificity produces informative summary measures in diagnostic reviews. *J Clin Epidemiol*. 2005;58:982–990.
- Deeks JJ, Macaskill P, Irwig L. The performance of tests of publication bias and other sample size effects in systematic reviews of diagnostic test accuracy was assessed. *J Clin Epidemiol*. 2005;58:882–893.
- Hu ZD, Wei TT, Yang M, et al. Diagnostic value of osteopontin in ovarian cancer: A meta-analysis and systematic review. *PLoS One*. 2015;10:e0126444.
- Gendrel D, Raymond J, Assicot M, et al. Measurement of procalcitonin levels in children with bacterial or viral meningitis. *Clin Infect Dis*. 1997;24:1240–1242.
- Viallon A, Zeni F, Lambert C, et al. High sensitivity and specificity of serum procalcitonin levels in adults with bacterial meningitis. *Clin Infect Dis*. 1999;28:1313–1316.
- Schwarz S, Bertram M, Schwab S, et al. Serum procalcitonin levels in bacterial and abacterial meningitis. *Crit Care Med*. 2000;28:1828–1832.
- Jereb M, Muzlovic I, Hojker S, et al. Predictive value of serum and cerebrospinal fluid procalcitonin levels for the diagnosis of bacterial meningitis. *Infection*. 2001;29:209–212.
- Dubos F, Moulin F, Gajdos V, et al. Serum procalcitonin and other biologic markers to distinguish between bacterial and aseptic meningitis. *J Pediatr*. 2006;149:72–76.
- Ray P, Badarou-Acossi G, Viallon A, et al. Accuracy of the cerebrospinal fluid results to differentiate bacterial from non bacterial meningitis, in case of negative gram-stained smear. *Am J Emerg Med*. 2007;25:179–184.
- Knudsen TB, Larsen K, Kristiansen TB, et al. Diagnostic value of soluble CD163 serum levels in patients suspected of meningitis: comparison with CRP and procalcitonin. *Scand J Infect Dis*. 2007;39:542–553.
- Dubos F, Korczowski B, Aygun DA, et al. Serum procalcitonin level and other biological markers to distinguish between bacterial and



- aseptic meningitis in children: a European multicenter case cohort study. *Arch Pediatr Adolesc Med.* 2008;162:1157–1163.
21. Onal H, Onal Z, Ozdil M, et al. A new parameter in the differential diagnosis of bacterial and viral meningitis. *Neurosciences.* 2008;13:91–92.
  22. Steinberg AV, Korzhenevich VI, Mihailova EV. Optimization of a diagnostic procedure for children with preliminary diagnosis of “meningitis”. *J Pediatr Infect Dis.* 2010;5:57–63.
  23. Makoo ZB, Soltani HR, Hasani A, et al. Diagnostic value of serum and cerebrospinal fluid procalcitonin in differentiation bacterial from Aseptic meningitis. *Am J Infect Dis.* 2010;6:93–97.
  24. Alkhali UM, Abd Al-Monem N, Abd El-Azim AA, et al. Serum procalcitonin in viral and bacterial meningitis. *J Glob Infect Dis.* 2011;3:14–18.
  25. Viallon A, Desseigne N, Marjollet O, et al. Meningitis in adult patients with a negative direct cerebrospinal fluid examination: value of cytochemical markers for differential diagnosis. *Crit Care.* 2011;15:R136.
  26. Ibrahim KA, Abdel-Wahab AA, Ibrahim AS. Diagnostic value of serum procalcitonin levels in children with meningitis: a comparison with blood leukocyte count and C-reactive protein. *J Pak Med Assoc.* 2011;61:346–351.
  27. Choi SH, Choi SH. Predictive performance of serum procalcitonin for the diagnosis of bacterial meningitis after neurosurgery. *Infect Chemother.* 2013;45:308–314.
  28. Prasad R, Kapoor R, Mishra OP, et al. Serum procalcitonin in septic meningitis. *Indian J Pediatr.* 2013;80:365–370.
  29. Casado MI, Alonso FM, Pinedo BL, et al. Acute meningitis in the pediatric emergency department: diagnostic yield of procalcitonin and C-reactive protein. *Pediatr Emerg Care.* 2014;30:849–850.
  30. Konstantinidis T, Cassimos D, Gioka T, et al. Can procalcitonin in cerebrospinal fluid be a diagnostic tool for meningitis? *J Clin Lab Anal.* 2014;29:169–174.
  31. Abdelkader NA, Mahmoud WA, Saber SM. Serum procalcitonin in Egyptian patients with acute meningitis and a negative direct cerebrospinal fluid examination. *J Infect Public Health.* 2014;7:106–113.
  32. Umran RM, Radhi NH. Diagnostic value of serum procalcitonin level in differentiating bacterial from nonbacterial meningitis in children. *Iran J Pediatr.* 2014;24:739–744.
  33. Li Y, Zhang G, Ma R, et al. The diagnostic value of cerebrospinal fluids procalcitonin and lactate for the differential diagnosis of post-neurosurgical bacterial meningitis and aseptic meningitis. *Clin Biochem.* 2015;48:50–54.
  34. Shen HY, Gao W, Cheng JJ, et al. Direct comparison of the diagnostic accuracy between blood and cerebrospinal fluid procalcitonin levels in patients with meningitis. *Clin Biochem.* 2015;48:1079–1082.
  35. Vikse J, Henry BM, Roy J, et al. The role of serum procalcitonin in the diagnosis of bacterial meningitis in adults: a systematic review and meta-analysis. *Int J Infect Dis.* 2015;38:68–76.
  36. Henry BM, Roy J, Ramakrishnan PK, et al. Procalcitonin as a serum biomarker for differentiation of bacterial meningitis from viral meningitis in children: evidence from a meta-analysis. *Clin Pediatr (Phila).* 2015. epub ahead of print. doi: 10.1177/0009922815606414.
  37. Cleophas TJ, Zwinderman AH. Meta-analyses of diagnostic studies. *Clin Chem Lab Med.* 2009;47:1351–1354.
  38. Hu ZD, Liu XF, Liu XC, et al. Diagnostic accuracy of osteopontin for malignant pleural mesothelioma: a systematic review and meta-analysis. *Clin Chim Acta.* 2014;433:44–48.
  39. Rutjes AW, Reitsma JB, Vandenbroucke JP, et al. Case-control and two-gate designs in diagnostic accuracy studies. *Clin Chem.* 2005;51:1335–1341.
  40. Schmidt RL, Factor RE. Understanding sources of bias in diagnostic accuracy studies. *Arch Pathol Lab Med.* 2013;137:558–565.