



Article Changes in Tumor Necrosis Factor α (TNFα) and Peptidyl Arginine Deiminase 4 (PAD-4) Levels in Serum of General Treated Psoriatic Patients

Joanna Czerwińska *🕑, Marta Kasprowicz-Furmańczyk 🔍, Waldemar Placek and Agnieszka Owczarczyk-Saczonek 🗅

Department of Dermatology, Sexually Transmitted Diseases and Clinical Immunology, School of Medicine, University of Warmia and Mazury in Olsztyn, 10-719 Olsztyn, Poland; martak03@wp.pl (M.K.-F.); w.placek@wp.pl (W.P.); aganek@wp.pl (A.O.-S.)

* Correspondence: joannaj061@gmail.com

Abstract: Psoriasis is an autoimmune disease in which the disturbed dependencies between lymphocytes, dendritic cells, keratinocytes and neutrophils play the most important role. One of them is the overproduction of neutrophil extracellular traps (NETs). The release of NETs can be induced by pathogens, as well as antibodies and immune complexes, cytokines and chemokines, including TNF α . The first step of the NET creation is the activation of peptidyl arginine deiminase 4 (PAD-4). PAD-4 seems to be responsible for citrullination of histories and chromatin decondensation, but the data on PAD-4 in NETs is inconclusive. Thus, the current study aimed to determine PAD-4 and TNF α levels in the serum of psoriatic patients by ELISA and observe the response of these factors to systemic (anti-17a, anti-TNF α and methotrexate) therapies. Increased levels of both PAD-4 and its main stimulus factor TNF α in pre-treatment patients have been reported along with the concentrations of proteins correlated with disease severity (PASI, BSA). Before treatment, the irregularities in the case of anti-nuclear antibodies level (ANA) were also observed. All of the applied therapies led to a decrease in PAD-4 and TNF α levels after 12 weeks. The most significant changes, both in protein concentrations as well as in scale scores, were noted with anti-TNF α therapy (adalimumab and infliximab). This phenomenon may be associated with the inhibition of $TNF\alpha$ production at different stages of psoriasis development, including NET creation. The obtained data suggest the participation of PAD-4 in the activation of neutrophils to produce NETs in psoriasis, which may create opportunities for modern therapies with PAD inhibitors. However, further exploration of gene and protein expression in psoriatic skin is needed.

Keywords: PAD-4; NETs; psoriasis; systemic therapy; adalimumab; secukinumab; methotrexate

1. Introduction

Psoriasis is a chronic inflammatory systemic disease with the burden of comorbidities, i.a., respiratory, cardiovascular and gastrointestinal [1–3]. This disorder is a serious global problem, which concerns at least 4.5 million individuals, according to the Global Burden of Disease (2019). The average age of patients with psoriasis is about 58 years (without differences between sexes). The most incident cases of psoriasis were noted in the most developed countries of Europe and North America [4]. Psoriasis significantly worsens the quality of life of patients and their families, often causing them to withdraw from social life. Despite the fact that the symptoms of psoriasis are typical and the diagnosis is usually not problematic, it is still a serious therapeutic challenge, mainly due to the complicated mechanism of its development. Every year, new elements of the immune system that may affect the pathogenesis of psoriasis are discovered. The most important role is played by the autoinflammatory feedback loop between lymphocytes, dendritic cells and keratinocytes, which activate the control axis (TNF α /IL-17/IL-23) to produce pro-inflammatory interleukins [3,5–7]. Additionally, neutrophils, which have the ability to



Citation: Czerwińska, J.; Kasprowicz-Furmańczyk, M.; Placek, W.; Owczarczyk-Saczonek, A. Changes in Tumor Necrosis Factor α (TNFα) and Peptidyl Arginine Deiminase 4 (PAD-4) Levels in Serum of General Treated Psoriatic Patients. *Int. J. Environ. Res. Public Health* **2022**, *19*, 8723. https://doi.org/10.3390/ ijerph19148723

Academic Editor: Paul B. Tchounwou

Received: 10 June 2022 Accepted: 15 July 2022 Published: 18 July 2022

Publisher's Note: MDPI stays neutral with regard to jurisdictional claims in published maps and institutional affiliations.



Copyright: © 2022 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (https:// creativecommons.org/licenses/by/ 4.0/). create extracellular traps (NETs) abundant in the cytoplasmic granules and DNA, strongly affect this interaction. Three typical stages may be distinguished in the NET formation process: neutrophil activation, changes in cell and extracellular protein, and DNA release. Neutrophils are induced by bacteria, fungi, and parasites [8]. The factors described as stimulating NETs include also antibodies and immune complexes, cytokines, and chemokines [9]. The main representative of the immune group is $TNF\alpha$ —the major stimuli factor of the protein arginine deiminase 4 (PAD-4), i.e., enzyme responsible for the onset of changes in the cell starting from chromatin decondensation (deamination/citrullination) [8]. In activated neutrophils, PAD-4 is translocated to the nucleus, wherein it is stimulated by Ca2+ levels above the physiological concentration [10]. It has been shown that three key molecules are engaged in the phenomena inside the cell: neutrophil elastase (NE) responsible for histone degradation, myeloperoxidase (MPO) changing chromatin structure, and cathelicidin antimicrobial peptide (LL-37) connecting histories with DNA. In the intracellular space, there is also increased ROS generation caused by the activation of nicotinamide adenine dinucleotide phosphate (NADPH) oxidase. The lack of a positive charge of arginine leads to the breaking of nucleosomes and nuclear envelope, and to hampered degranulation. The last stage entails the release of the chain of DNA and protein (e.g., MPO, NE and LL-37) into the extracellular space [5–7] (Figure 1).



Figure 1. The dependence between NET creation and TNF/IL-23/IL-17 axis in psoriatic skin (the box shows the relation of TNF α and PAD-4; \uparrow indicates an increase in the secretion of presented factors; X indicates the sites of inhibition of TNF α production by using of anti-TNF therapy). PAD-4—peptidyl arginine deiminase 4; NE—neutrophil elastase; MPO—myeloperoxidase; cDC—conventional dendritic cell; IFN—interferon; IL—interleukin; LL37—cathelicidin; NETs—neutrophil extracellular traps; pDC—plasmacytoid dendritic cells; Th—T helper lymphocyte; TNF—tumor necrosis factor.

In physiological conditions, NETs dispose of various pathogens, acting as the first line of the body's defenses by direct trapping microorganisms [7,8,11,12]. However, the dysregulated NET activity may lead to aggravated inflammation, impairment of microcirculation, and cause tissue damage and then trigger autoimmune diseases, such as psoriasis [13,14]. The key role is played by LL-37 that creates complexes with DNA/RNA and causes Th1/Th17/Th22 differentiation. Additionally, the pathogenesis of psoriasis follows two more mechanisms: plasmacytoid dendritic cell (pDC) activation via Toll-like receptors (TLRs) and interferon- α (INF- α) release [15–17], which confirms the complex and complicated mechanism of this disease development (Figure 1). Thus, the NETs are found at each stage of psoriasis development, where they are engaged in an immune response manifested by T-cell imbalance, keratinocyte hyperproliferation, and the formation of autoantigens [18,19] (Figure 1).

The immune system is able to produce, for example, anti-neutrophil cytoplasmic antibodies (ANCA), directed against proteins present in neutrophil granules found in systemic small vessel vasculitis (SVV) or anti-nuclear antibodies (ANA, i.a. anti-dsDNA/ssDNA, anti-histone) against nucleus components, which are characteristic for systemic lupus erythematosus (SLE) and they are very often found in psoriasis [20,21]. Hence, autoantibodies accumulate in the tissues in the form of immune complexes (e.g., together with platelets, erythrocytes and plasma proteins), initiating cytotoxic reactions [11] that cause modifications of host proteins, including their citrullination and the production of anti-citrullinated protein antibodies (ACPA). In turn, it leads to the loss of immunological tolerance to these proteins [11–20]. Additionally, it was noted that patients with autoimmune diseases have a limited ability to degrade NETs by decreased activity of DNase, which is related to the increased presence of its inhibitors [21]. Moreover, DNase is able to remove DNA (NET's backbone), but not the proteins attached. This is due to the fact that NET components bind secondarily to proteins present on the host vascular endothelium [22], which contributes to damage to its own tissues and the development of diseases [23]. The excessive NET formation connecting with the externalization of citrullinated autoantigens (vimentin, α -enolase), pro-inflammatory cytokines (TNF α , interleukin IL-6, IL-8), and immunostimulatory molecules impair immune response by leading to inflammation [24,25].

The course of psoriasis is usually characterized by a high accumulation of neutrophils, both in plaques as well as in the blood [26–28]. The cells circulating with blood, low-density granulocytes (LDGs) with ability to synthesize TNF α , are the most essential to NET production [20] in psoriatic patients. Interestingly, the decreased amount of NETotic cells in the blood correlates with the regression of psoriasis [7,29,30]. Moreover, the sera from psoriatic patients, added to healthy neutrophils, are able to induce NET production [7]. Additionally, biological therapies may indirectly affect the function and number of active neutrophils [30]. It is worth noting that both severe lymphocyte infiltration in psoriatic skin lesions, as well as subcorneal Munro's microabscesses filled with neutrophils, are characteristic of the pathological image of the diseases [8].

Interestingly, disruptions in NET production caused by irregularities in the activity of the PAD family (especially PAD-1 and 2) are also indicated. Isoforms are clearly differentiated in terms of occurrence and functions, e.g., PAD-1 occurs in the epidermis, whereas PAD-2 is expressed in various organs, i.a., in the brain, female reproductive tissues, and skeletal muscle, but it is also found in the periphery of granular keratinocytes. Hair follicles and epithelium exhibit PAD-3 expression, and PAD-6 is found in germ cells. Hence, PAD-1 and PAD-3 are engaged in epidermal homeostasis and barrier function, whereas PAD2 is responsible for cytoskeletal degeneration and apoptosis. In turn, PAD-6 regulates mainly female infertility [24,25]. In turn, the highest stability and activity is presented by PAD-4, whose expression is restricted to immune cells, such as neutrophils, eosinophils, granulocytes, and monocytes, where it participates in the regulation of inflammatory processes [24].

Despite the fact that numerous studies have addressed different disorders (mainly rheumatoid arthritis-RA and Bechet's disease-BD), data concerning the necessity of PAD-4 involvement in NET generation still remain contradictory. Both reports showing that the enzyme is essential to NET generation as well as works rejecting this hypothesis may be found in the literature [31–35]. Thus, the role of PAD-4 in the transformation of chromatin structure in neutrophils remains unclear. Moreover, there is still a paucity of research on the involvement of this enzyme in the formation of NETs in human psoriasis. Therefore, the aim of the current study was to determine the levels of PAD-4 and its main activator (TNF α) in the serum of psoriatic patients and observe the response of these factors to systemic

therapy: anti-IL-17A (ixekizumab, secukinumab), anti-TNF α (adalimumab, infliximab), and methotrexate (MTX), mostly deployed in psoriasis treatment.

2. Material and Methods

2.1. Study Group

The study group included 50 patients aged 18–65 without significant burdens (Table 1) and treated in the Clinic and at the Department of Dermatology, Sexually Transmitted Diseases and Clinical Immunology in Olsztyn, for plaque psoriasis. Patients with other inflammatory diseases, neoplastic diseases, previous cardiovascular complications, heart, kidney and liver failure were excluded. The control group consisted of healthy volunteers (n = 16) with no personal and family history of autoimmune and inflammatory diseases. Patients were classified into three research groups. The first group received methotrexate (MTX): orally at a dose of 15 mg/week. The second therapy included the administration of anti-TNF (adalimumab): subcutaneously, initial dose 80 mg, after seven days, 40 mg every two weeks. The third group took an IL-17a inhibitor (secukinumab): subcutaneously, 300 mg at 0–4 weeks, then 300 mg every month. Skin condition assessment (Psoriasis Area Severity Index–PASI, Body Surface Area (BSA), and Dermatology Life Quality Index (DLQI)) were performed on each visit and always by the same qualified person.

Table 1. Clinical characteristics of patients with psoriasis.

Variables	Results	Range
patients	50 (39M, 11F)	
age (years)	52.64 ± 2.1	18–65
illness duration (years)	17.21 ± 2.3	2–36

2.2. Methods

Before the start of treatment (week 0), the level of neutrophils, C Reactive Protein (CRP) and titer of anti-nuclear antibodies (ANA, Euroimmun, Lübeck, Germany) were detected. Hematology and biochemical analyses of blood were performed in the Laboratory of the Municipal Polyclinical Hospital in Olsztyn. All ANA analyses were followed by an Immunoblot test (ANA profile V and ANA Profile 23, Euroimmun, Germany). PAD-4 and TNF α levels in blood serum were assessed by the enzyme-linked immunosorbent assay (ELISA) using commercially available kits (Cayman Chemical, MI, USA; EIAAB, Wuhan, China) with a wide detection range: 156–10,000 pg/mL (PAD-4) and 15.6–1000 pg/mL (TNF α). The procedures were performed according to the manufacturer's protocol. The assay's validity was confirmed based on parallelism between the standard curve and dilutions of randomly chosen serum samples. The intra- and inter-assay coefficient of variation for factors was <4%. The sensitivity of these assays was defined as the least concentration that could be differentiated from zero samples, and it was determined at 78 and 7.8 pg/mL for PAD-4 and TNF α , respectively. Absorbance values were measured at 450 nm using Multiskan FC (Thermo Fisher Scientific, CO, USA).

2.3. Statistical Analysis

The results were processed statistically by the following non-parametric tests: the Mann–Whitney U test (comparison of two independent groups), Friedman's ANOVA (comparison of many dependent groups) and Spearman's correlation test. The results were expressed as mean \pm SEM. All calculations were performed using Statistica software, version 13.1 (Statsoft, Inc., Tulsa, OK, USA). Differences were regarded as statistically significant at *p* < 0.05.

3. Results

In the presented study, the concentrations of NET protein–PAD-4 and its main stimulus factor, TNF α , were analyzed before and during (week 4 and 12) the application of three types of systemic therapy used in patients with psoriasis: MTX, anti-TNF α and anti-IL 17a.

3.1. Neutrophils, C Reactive Protein (CRP) and Anti-Nuclear Antibodies (ANA)

Before starting the treatment, the level of neutrophils, C Reactive Protein (CRP) and the titer of anti-nuclear antibodies (ANA) were detected (Table 2). Interestingly, in all of the patients, the level of neutrophils, as well as CRP, were normal, whereas the titer of ANA was increased. All qualified patients showed a titer level higher than 1:320. The ANA pattern was defined as granular or mixed. An analysis of ANA was followed by an immunoblot test (ANA Profile V and ANA Profile 23). The results were mostly negative, and the type of antibodies was not detected. However, three patients indicated positive mitochondria (AMA), double-stranded DNA (dsDNA) and an Ro-52 antigen band. After 12 weeks of treatment, the levels of described factors did not change significantly. However, the use of anti-TNF α antibodies elevated the titers of ANA by at least one level (1:640–1:250) in most patients. In other groups (MTX and anti-Il 17a), the titer of ANA changed in only 10% of patients.

Table 2. Laboratory analysis of patients' serum before starting the therapy.

Results	Range
57.74 ± 2.5	40–70
4.23 ± 0.3	-
2.58 ± 0.6	<0.5
1:320–1:1280	<1:80
	Results 57.74 ± 2.5 4.23 ± 0.3 2.58 ± 0.6 $1:320-1:1280$

3.2. Psoriasis Area Severity Index (PASI), Body Surface Area (BSA), and Dermatology Life Quality Index (DLQI)

On each measurement point, the Psoriasis Area Severity Index (PASI), Body Surface Area (BSA) and Dermatology Life Quality Index (DLQI) were evaluated. Before therapy, 38 patients indicated a PASI score of more than 10 and a BSA score of more than 16, whereas all of them (n = 50) had a DLQI higher than 10 (Table 3).

Variables	Week 0	Mean
	<10 (<i>n</i> = 12)	8.13 ± 0.6
PASI	>10 (<i>n</i> = 38)	18.31 ± 0.9
DC A	<16 (<i>n</i> = 12)	10.46 ± 0.8
DSA	>16 (<i>n</i> = 38)	30.72 ± 4.0
DLQI	>10 (n = 50)	19.97 ± 1.4

Table 3. Levels of PASI, BSA and DLQI indicators before the study.

During the study, scores on all scales (PASI, BSA, DLQI) decreased (Table 4). The best results were observed for therapy with anti-TNF α (Table 5), where the skin condition assessment indicated changes of more than 65% (p < 0.05).

	Week 0	Week 12	Effect (↓)
PASI	15.74 ± 1.6	5.93 ± 1.0	* 62%
BSA	24.03 ± 4.1	8.15 ± 2.2	* 66%
DLQI	20.09 ± 7.6	8.84 ± 1.2	* 55%

Table 4. Levels of PASI, BSA and DLQI indicators before the study (week 0) and during treatment (week 12). * indicates a statistically significant difference (p < 0.05).

Table 5. The comparison of PASI, BSA and DLQI scores between therapies with: MTX (therapy 1, n = 20), anti-TNF α (therapy 2, n = 20), anti-IL17a (therapy 3, n = 10) in patients with plaque psoriasis at two measurement points (week 0, week 12). * indicates a statistically significant difference (p < 0.05).

		Week 0	Week 12	Effect (↓)
Thomas 1	PASI	12.11 ± 1.1	5.01 ± 0.8	* 58%
Therapy 1	BSA	9.28 ± 3.2	7.84 ± 1.3	15%
	DLQI	22.86 ± 4.3	11.87 ± 1.1	48%
	PASI	19.83 ± 1.4	4.49 ± 0.9	* 77%
Therapy 2	BSA	42.23 ± 5.4	10.41 ± 2.4	* 75%
	DLQI	19.03 ± 1.2	6.16 ± 1.2	* 67%
	PASI	15.3 ± 2.1	8.29 ± 1.3	45%
Therapy 3	BSA	20.60 ± 3.8	6.20 ± 2.9	* 69%
	DLQI	18.40 ± 2.1	8.50 ± 1.1	* 53%

3.3. Peptidyl Arginine Deiminase 4 (PAD-4)

Among a full group of patients (n = 50), there was a correlation between PAD-4 with PASI score (r = 0.3141, p < 0.05). The analyses of protein levels also showed a correlation between PAD-4 and BSA (r = 0.6401, p < 0.05). However, no correlation was found between the duration of the disease (in years) and the concentration of the studied factor before the start of therapy (r = 0.2218). The concentration of PAD-4 was significantly higher in the group of PASI > 10 and BSA > 16 in comparison with PASI ≤ 10 and BSA ≤ 16 . The concentration of PAD-4 in psoriatic patients was also higher than in healthy volunteers (p < 0.05) (Table 6).

Table 6. Pre-treatment levels of PAD-4 (pg/mL) in patients with plaque psoriasis (n = 50) with PASI score ≤ 10 (n = 12), PASI > 10 (n = 38), BSA ≤ 16 (n = 12), BSA > 16 (n = 38) and in the control group (n = 16).

Variable	Group	Mean (pg/mL)
	$PASI \le 10$	391.78 ± 26.3
	PASI > 10	1019.35 ± 62.3
PAD-4	$BSA \le 16$	361.97 + 20.1
	BSA > 16	1022.31 ± 60.7
	control	244.88 ± 28.8

The analysis of the entire group of patients (n = 50) showed a decrease in concentrations (p < 0.05) of the tested factor after four weeks of treatment. The level of PAD-4 remained at a similar level also after 12 weeks (Table 7).

lable 7.	Changes of PAD-4 level	(pg/mL) during	treatment ($n = 50$).	* indicates a statistically
significa	nt difference ($p < 0.05$).			

Variable (pg/mL)	Week 0	Week 4	Effect (↓)	Week 12	Effect (↓)
PAD-4	905.26 ± 29.9	344.91 ± 21.7	* 61%	297.82 ± 21.3	* 67%

The division of the research group in relation to the therapy allowed for the analysis of changes in the concentrations of the tested factor during the course of each of them. It was observed that the level of PAD-4–the enzyme protein initiating the formation of NET –decreased (p < 0.05) as a result of treatment with all three therapies (Figure 2, Table 8). However, in the case of treatment with therapy 2 (anti-TNF α), the greatest decrease in PAD-4 concentration was observed (81%, p < 0.05). The protein concentration was lower than in the group of healthy volunteers (p < 0.05). In the remaining cases (therapy 1 and 3), the concentration of the PAD-4 after 12 weeks of treatment was still higher (p < 0.05) or equal in comparison with the control group. The smallest differences were observed in therapy with anti-IL17a (therapy 3).



Figure 2. PAD-4 level (pg/mL) before (week 0) and during (week 4, week 12) therapy with: MTX (therapy 1, n = 20), anti-TNF α (therapy 2, n = 20) and anti-IL-17A (therapy 3, n = 10) in patients with plaque psoriasis compared with a control consisting of healthy volunteers (n = 16). * indicates a statistically significant difference (p < 0.05) between weeks: 0 and 12.

Table 8. PAD-4 level (pg/mL) between treatments: MTX (therapy 1, n = 20), anti-TNF α (therapy 2, n = 20) and anti-IL-17A (therapy 3, n = 10) in patients with plaque psoriasis at three measurement points (week 0, week 4, week 12). * indicates a statistically significant difference (p < 0.05).

PAD-4	Week 0	Week 4	Effect (↓)	Week 12	Effect (↓)
Therapy 1	686.92 ± 20.8	225.61 ± 19.3	* 67%	206.52 ± 18.2	* 69%
Therapy 2	783.89 ± 11.34	223.26 ± 4.1	* 71%	148.1 ± 2.8	* 81%
Therapy 3	1245.1 ± 57.6	568.0 ± 45.47	* 54%	539.0 ± 45.3	* 56%

3.4. Tumor Necrosis Factor α (TNF α)

Similar to PAD-4, the concentration of TNF α was also correlated with PASI and BSA (r = 0.3668 and r = 0.4142, respectively, *p* < 0.05). Thus, the correlation between TNF α and PAD-4 was noted (0.4061, *p* < 0.05). The level of TNF α was elevated in moderate and severe psoriasis (PASI > 10 and BSA > 16) compared with mild forms of the disease (Table 9). The concentration of this factor in psoriatic patients was also significantly higher than in the control group (*p* < 0.05).

_

Variable	Group	Mean (pg/ml)
	$PASI \le 10$	450.81 ± 325.9
	PASI > 10	598.8 ± 121.61
TNFα	$BSA \le 16$	500.1 ± 272.6
	BSA > 16	605.76 ± 130.2
	control	32.15 ± 1.99

Table 9. Pre-treatment levels of PAD-4 (pg/mL) in patients with plaque psoriasis (n = 50) with PASI score ≤ 10 (n = 12), PASI > 10 (n = 38), BSA ≤ 16 (n = 12), BSA > 16 (n = 38) and in the control group (n = 16).

In the entire population with the disease (n = 50), TNF α level decreased during treatment (p < 0.05). The observed changes were comparable to that of PAD-4 (Table 10).

Table 10. Changes of TNF α level (pg/mL) during treatment (n = 50). * indicates a statistically significant difference (p < 0.05).

Variable (pg/mL)	Week 0	Week 4	Effect (↓)	Week 12	Effect (↓)
TNFα	529.72 ± 164.9	341.37 ± 146.1	35%	256.71 ± 90.7	* 51%

The analyses of protein levels in different research groups (therapy 1–3) indicated a significant decrease in only two applied therapies: MTX and anti-TNF α . However, a larger change was noted with anti-TNF α treatment (78%, *p* < 0.05). Additionally, the TNF α level after 12 weeks of using different therapies (1–3) still remained higher (*p* < 0.05) than in the group of healthy volunteers (Figure 3, Table 11).



Figure 3. TNF α level (pg/mL) before (week 0) and during (week 4, week 12) therapy with: MTX (therapy 1, *n* = 20), anti-TNF α (therapy 2, *n* = 20), anti-IL-17A (therapy 3, *n* = 10) in patients with plaque psoriasis compared with a control consisting of healthy volunteers (*n* = 16). * indicates a statistically significant difference (*p* < 0.05) between weeks: 0 and 12.

Table 11. TNF α level (pg/mL) between treatments with: MTX (therapy 1, *n* = 20), anti-TNF α (therapy 2, *n* = 20) and anti-IL-17A (therapy 3, *n* = 10) in patients with plaque psoriasis at three measurement points (week 0, week 4, week 12). * indicates a statistically significant difference (*p* < 0.05).

TNFα	Week 0	Week 4	Effect (\downarrow)	Week 12	Effect (↓)
Therapy 1	505.26 ± 223.1	329.19 ± 161.8	34%	336.77 ± 103.9	* 33%
Therapy 2	722.34 ± 196.1	352.92 ± 127.5	* 51%	155.07 ± 91.5	* 78%
Therapy 3	361.57 ± 75.6	342.0 ± 149.0	5%	278.2 ± 76.7	23%

4. Discussion

The results of the current study provide novel evidence for the presence of PAD-4 protein in human serum. To the best of the authors' knowledge, this is the first study demonstrating the level of this enzyme in psoriatic patients. The PAD-4 and TNF α responses to three types of systemic treatment were also analyzed. Before treatment, neutrophils and CRF levels, as well as ANA titer, were evaluated. The irregularities were observed only in the case of ANA. Additionally, after 12 weeks of therapy with anti-TNF α , elevated ANA titers were noted in most patients but without reference to clinical symptoms. Similar observations were presented in psoriatic patients by Yasuda et al. (2019) [36].

The study was performed at three different measurement points (before the study and in the 4th and 12th week of therapy). The study confirmed the role of TNF α as an indicator of the disease severity [37], but it also determined, for the first time, that this role may be played by PAD-4. Similar to TNF α , its concentration also correlated with the PASI and BSA score. Moreover, the levels of both TNF α and PAD-4 were significantly increased in psoriatic patients compared with the control group.

The study found that the levels of tested factors varied under the influence of all treatments. TNF α concentration was best influenced by its inhibitors: adalimumab and infliximab. Likewise, the greatest decrease in PAD-4 concentration was observed under general therapy with anti-TNF α . The PASI, BSA and DLQI scores confirmed these observations. The spectacular improvement of the indices was observed in the group treated with biological therapy with anti-TNF α . This phenomenon may be associated with a broad spectrum of action of therapies and inhibition of TNF α production at three stages of psoriatic pathogenesis: from activated LDGs, cDC and Th1 cells (Figure 1). It also confirmed the participation of NET in psoriasis development. Interestingly, in contrast to the current study, a review by Bergen (2020) found treatment with inhibitors of IL-17 to be the most clinically beneficial [32].

In the available literature, there are few studies analyzing PAD-4 protein concentrations in human serum. Thus, the discussion of the current results is difficult. To date, anti-PAD-4 antibodies have been analyzed in the blood of patients with RA. It has been shown that antibodies against PAD-4 precede the clinical onset of RA in a significant proportion of patients and may be important in the initial stage of the disease [38,39], which indirectly confirms the current results. In turn, studies with patients with active BD exhibit higher mRNA PAD4 expression than in inactive BD and healthy volunteer groups. Additionally, neutrophils of healthy subjects exposed to sera from BD patients induced NADPH oxidase protein expression and affected NET formation [40]. PAD-4 is also involved in skin tumorigenesis, in which the inhibition of this enzyme results in apoptosis as well as in the repression of p53 target genes [28]. The main role of the stimuli factor of NETs in autoimmune diseases is assigned to the TNF α /IL-17/IL-23 axis [10–12]. These signals cause the activation of the NADPH oxidase and thus the increase in ROS. Interestingly, there are also reports indicating pathways independent of NADPH [41,42]. It is suggested that only in this case are the participation of PAD-4 and chromatin decondensation needed for the NET formation [38]. The studies performed by Doud et al. (2013) and Neeli et al. (2013) excluded the absolute necessity of the PAD-4 enzyme in the initiation of NADPH-dependent NET. In the described experiments, stimulation of NET with phorbol myristate acetate (PMA) led to the formation of traps without detectable histone deimination [41,42]. Additionally, the use

of PMA, which is also an activator of protein kinase C (PKC)-an inhibitor of PAD-4, showed no relationship between the NET formation and histone citrullination [43]. However, contrary evidence indicating that the NET formation requires PAD-4 activation was presented. Research on PAD-4 was mainly conducted using the mouse animal model. It was reported that histone citrullination correlated with chromatin decondensation during the NET formation. Blocking PAD-4 was found to disrupt network formation and therefore increase the risk of bacterial infection in animals [26]. In addition, the study of Ding et al. (2022) showed remarkably ameliorated psoriasis-like symptoms in PAD-4 knockout mice [44]. Thus, studies using knock-out animals indicated the participation of PAD-4 as a required component for NET-dependent innate antimicrobial immunity [26] as well as a factor firmly associated with psoriasis development [44]. Additionally, PAD-4 has been shown to participate in the regulation of proliferation and hematopoiesis [42]. Studies using the HL-60 cell line clearly indicated that PAD-4 citrullination of histories mediates chromatin decondensation and the inhibition of PAD-4 reduces histone hypercitrullination and the formation of NET structures [25,45,46]. The cellular events that lead to the NET formation were presented by Thiam et al. (2020) for human and mouse primary blood neutrophils, as well as in differentiated HL-60 neutrophil-like cells (dHL-60). Those authors showed that extracellular DNA release required the enzymatic and nuclear localization activities of PAD-4 [46]. Studies using the knock-out mouse model of RA showed reduced severity of symptoms by inducing immune reactions such as helper T-cell development, inflammatory cytokine production and apoptosis [47,48]. In addition to RA, the autoimmune disease in which PAD-4 was studied was also lupus (LE). Again, using a mouse animal model and PAD-4 inhibitors, a reduction in the autoimmune response was observed [49]. In sick mice, the inhibitors of PAD suppressed the formation of NETs, reduced the accumulation of immune complexes, i.a., in the kidneys and improved the condition of the skin [50]. In addition, it has been shown that PAD-4 inhibition reduces NET-related citrulline histones, and minimizes immune cell recruitment [51] and liver damage associated with ischemiareperfusion injury [52]. In contrast, PAD-4 from bone-marrow-derived cells and NETs did not influence chronic atherogenesis, but its involvement in acute thrombotic complications of intimal lesions was noted. Thus, the contribution of NETs and PAD-4 to atherosclerosis and its thrombotic complications seems to be stage- specific [53].

In the newest reports, the hypothesis that abnormality in NET creation is strongly associated with psoriasis development is discussed more and more frequently. The high levels of NETs in the lesions of psoriatic patients are still described [54–56]. However, huge attention is also paid to irregularities in PAD isoform activity. Interestingly, a new direction of PAD analyses was proposed by Bawadeker et al. in 2017. The authors induced the NET formation in mice with TNF α -induced arthritis and noted the participation of PAD-2 in protein citrullination [57]. In turn, Padhi et al. (2022) have recently reported the correlation between II-22+cells and PAD-1 in lesional psoriatic skin, where IL-22 inhibited the expression of PAD-1 in epidermal keratinocytes [58]. Thus, the lack of citrullinated by PAD-1 keratin K1 in skin samples from psoriasis patients may be a contributing factor to disease [54]. The results of both studies confirmed the important role of PAD isoforms as significant elements regulating cytokine secretion in the control axes, which may be crucial in the mechanism of various diseases.

Concurrently, we may find the reports providing information about additional mechanisms engaged in the NET generation. Meng (et al. 2022) noticed that decreased receptorinteracting serine/threonine-protein kinase 1 (RIPK1) expression in psoriasis neutrophils may enhance traps [59], whereas Ding et al. (2022) indicated that Src homology-2 domaincontaining protein tyrosine phosphatase-2 (SHP2) had a high impact on psoriasis severity [44].

The presented data indicated different pathogenetic mechanisms of autoimmune diseases that involve both PAD-dependent and PAD-independent pathways. Simultaneously, it appeared to be an extensive field for NET research because knowledge of the contribution of PAD-4 to the formation of NET networks in psoriasis is still scarce. The current results, showing a clear decrease in PAD-4 concentration in serum, may indicate the enzyme's involvement in the activation of NETs and psoriasis development. However, fully defining PAD-4 as an indicator of psoriasis severity requires additional analyses at both the protein and the gene levels in the skin.

Due to the fact that PAD-4 shows the highest stability and activity in inflammation and it occurs in distinctive cells, this enzyme seems to be particularly dangerous in the pathogenesis of skin barrier defects, including psoriasis. Hence, the PAD-4 may be a primary target for modern therapies with PAD-4-specific inhibitors. These therapies may impede the functioning of the feedback loop between neutrophils (by inhibiting the NET formation without damaging beneficial defensive properties), dendritic cells, lymphocytes and keratinocytes. A reduction in disease symptoms using PAD-inhibitors has been noted in the mouse models of RA and LE. Interestingly, the current results also showed that cytokine-targeted therapies indirectly altered PAD-4 levels. However, further research is still needed regarding PAD activation as a main component of NET in the pathogenesis of psoriasis.

Author Contributions: Conceptualization, J.C.; methodology, J.C. and M.K.-F.; software, J.C.; validation, J.C. and M.K.-F.; formal analysis, J.C.; investigation, M.K.-F.; resources, A.O.-S.; data curation, J.C., A.O.-S. and W.P.; writing—original draft preparation, J.C.; writing—review and editing, A.O.-S. and W.P.; visualization, J.C.; supervision, A.O.-S. and W.P.; project administration, J.C. and M.K.-F.; funding acquisition, J.C. and A.O.-S. All authors have read and agreed to the published version of the manuscript.

Funding: This study was supported by the Ministry of Higher Education [project No. 61.610.006-110].

Institutional Review Board Statement: The study was approved by the Bioethical Committee of the Warmia and Mazury University in Olsztyn on 27 June 2019 (Resolution 60/2019). Informed consent was obtained from each patient enrolled in the study.

Informed Consent Statement: The study was approved by the Bioethical Committee of the Warmia and Mazury University in Olsztyn (Resolution 60/2019). Informed consent was obtained from each patient enrolled in the study.

Data Availability Statement: Not applicable.

Conflicts of Interest: The authors declare no conflict of interest.

References

- Santus, P.; Rizzi, M.; Radovanovic, D.; Airoldi, A.; Cristiano, A.; Conic, R.; Petrou, S.; Pigatto, P.D.; Bragazzi, N.C.; Colombo, D.; et al. Psoriasis and respiratory comorbidities: The added value of fraction of exhaled nitric oxide as a new method to detect, evaluate, and monitor psoriatic systemic involvement and therapeutic efficacy. *BioMed Res. Int.* 2018, 23, 3140682. [CrossRef] [PubMed]
- Conic, R.; Damiani, G.; Schrom, K.P.; Ramser, A.E.; Zheng, C.; Xu, R.; McCormick, T.S.; Cooper, K.D. Psoriasis and psoriatic arthritis cardiovascular disease endotypes identified by red blood cell distribution width and mean platelet volume. *J. Clin. Med.* 2020, *9*, 186. [CrossRef]
- Damiani, G.; Franchi, C.; Pigatto, P.; Altomare, A.; Pacifico, A.; Petrou, S.; Leone, S.; Pace, M.C.; Fiore, M. Outcomes assessment of hepatitis C virus-positive psoriatic patients treated using pegylated interferon in combination with ribavirin compared to new Direct-Acting Antiviral agents. *World J. Hepatol.* 2018, 10, 329–336. [CrossRef]
- Damiani, G.; Bragazzi, N.L.; Aksut, C.K.; Wu, D.; Alicandro, G.; McGonagle, D.; Guo, C.; Dellavalle, R.; Grada, A.; Wong, P.; et al. The global, regional, and national burden of psoriasis: Results and insights from the global burden of disease 2019 study. *Front. Med.* 2021, *8*, 743180. [CrossRef] [PubMed]
- 5. Chiang, C.C.; Cheng, W.J.; Korinek, M.; Lin, C.Y.; Hwang, T.L. Neutrophils in Psoriasis. Front. Immunol. 2019, 10, 2376. [CrossRef]
- Greb, J.E.; Goldminz, A.M.; Elder, J.T.; Lebwohl, M.G.; Gladman, D.D.; Wu, J.J. Psoriasis. Nat. Rev. Dis. Prim. 2016, 2, 16082. [CrossRef]
- 7. Hu, S.C.; Yu, H.S.; Yen, F.L.; Lin, C.L.; Chen, G.S.; Lan, C.C. Neutrophil extracellular trap formation is increased in psoriasis and induces human beta-defensin-2 production in epidermal keratinocytes. *Sci. Rep.* **2016**, *6*, 31119. [CrossRef] [PubMed]
- Wang, Y.; Wysocka, J.; Sayegh, J.; Lee, Y.; Perlin, J.R.; Leonelli, L.; Sonbuchner, L.S.; McDonald, C.C.; Cook, R.G.; Dou, Y.; et al. Human PAD4 regulates histone arginine methylation levels via demethylimination. *Science* 2004, 306, 279–283. [CrossRef] [PubMed]

- 9. Gupta, A.K.; Hasler, P.; Holzgreve, W.; Gebhardt, S.; Hahn, S. Induction of neutrophil extracellular DNA lattices by placental microparticles and IL-8 and their presence in preeclampsia. *Hum. Immunol.* **2005**, *66*, 1146–1154. [CrossRef] [PubMed]
- Vossenaar, E.R.; Radstake, T.R.; van der Heijden, A.; van Mansum, M.A.M.; Dieteren, C.; de Rooij, D.-J.; Barrera, P.; Zendman, A.J.W.; van Venrooij, W.J. Expression and activity of citrullinating peptidylarginine deiminase enzymes in monocytes and macrophages. *Ann. Rheum. Dis.* 2004, 63, 373–381. [CrossRef] [PubMed]
- 11. Kenny, E.F.; Herzig, A.; Krüger, R.; Muth, A.; Mondal, S.; Thompson, P.R.; Brinkmann, V.; von Bernuth, H.; Zychlinsky, A. Diverse stimuli engage different neutrophil extracellular trap pathways. *eLife* 2017, *6*, e24437. [CrossRef]
- Munoz, L.E.; Bilyy, R.; Biermann, M.H.C.; Kienhöfer, D.; Maueröder, C.; Hahn, J.; Brauner, J.M.; Weidner, D.; Chen, J.; Scharin-Mehlmann, M.; et al. Nanoparticles size-dependently initiate self-limiting NETosis-driven inflammation. *Proc. Natl. Acad. Sci. USA* 2016, *113*, E5856–E5865. [CrossRef]
- 13. Ishida-Yamamoto, A.; Senshu, T.; Takahashi, H.; Akiyama, K.; Nomura, K.; Iizuka, H. Decreased deiminated keratin K1 in psoriatic hyperproliferative epidermis. *J. Investig. Dermatol.* **2000**, *114*, 701–705. [CrossRef]
- Chang, X.; Han, J.; Pang, L.; Zhao, Y.; Yang, Y.; Shen, Z. Increased PADI4 expression in blood and tissues of patients with malignant tumors. *BMC Cancer* 2009, 9, 40. [CrossRef] [PubMed]
- Nakashima, K.; Hagiwara, T.; Yamada, M. Nuclear localization of peptidylarginine deiminase V and histone deimination in granulocytes. J. Biol. Chem. 2002, 277, 49562–49568. [CrossRef]
- Arita, K.; Hashimoto, H.; Shimizu, T.; Nakashima, K.; Yamada, M.; Sato, M. Structural basis for Ca²⁺-induced activation of human PAD4. *Nat. Struct. Mol. Biol.* 2004, 11, 777–783. [CrossRef]
- 17. Zabieglo, K.; Majewski, P.; Majchrzak-Gorecka, M.; Wlodarczyk, A.; Grygier, B.; Zegar, A. The inhibitory effect of secretory leukocyte protease inhibitor (SLPI) on formation of neutrophil extracellular traps. *J. Leukoc. Biol.* **2015**, *98*, 99–106. [CrossRef]
- 18. Brinkmann, V.; Reichard, U.; Goosmann, C.; Fauler, B.; Uhlemann, Y.; Weiss, D.S.; Weinrauch, Y.; Zychlinsky, A. Neutrophil extracellular traps kill bacteria. *Science* 2004, *303*, 1532–1535. [CrossRef]
- 19. Ganguly, D.; Chamilos, G.; Lande, R.; Gregorio, J.; Meller, S.; Facchinetti, V. Self-RNAantimicrobial peptide complexes activate human dendritic cells through TLR7 and TLR8. *J. Exp. Med.* **2009**, *206*, 1983–1994. [CrossRef]
- Denny, M.F.; Yalavarthi, S.; Zhao, W.; Thacker, S.G.; Anderson, M.; Sandy, A.R.; McCune, W.J.; Kaplan, M.J. A distinct subset of proinflammatory neutrophils isolated from patients with systemic lupus erythematosus induces vascular damage and synthesizes type I IFNs. J. Immunol. 2010, 184, 3284–3297. [CrossRef] [PubMed]
- Hakkim, A.; Fürnrohr, B.G.; Amann, K.; Laube, B.; Abuabed, U.; Brinkmann, V.; Herrmann, M.; Voll, R.E.; Zychlinsky, A. Impairment of neutrophil extracellular trap degradation is associated with lupus nephritis. *Proc. Natl. Acad. Sci. USA* 2010, 107, 9813–9818. [CrossRef] [PubMed]
- Kolaczkowska, E.; Jenne, C.N.; Surewaard, B.G.J.; Thanabalasuriar, A.; Lee, W.-Y.; Sanz, M.-J.; Mowen, K.; Opdenakker, G.; Kubes, P. Molecular mechanisms of NET formation and degradation revealed by intravital imaging in the liver vasculature. *Nat. Commun.* 2015, *6*, 6673. [CrossRef] [PubMed]
- Hoffmann, J.H.O.; Enk, A.H. Neutrophil extracellular traps in dermatology: Caught in the NET. J. Dermatol. Sci. 2016, 84, 3–10. [CrossRef] [PubMed]
- Koushik, S.; Joshi, N.; Nagaraju, S.; Mahmood, S.; Mudeenahally, K.; Padmavathy, R.; Jegatheesan, S.K.; Mullangi, R.; Rajagopal, S. PAD4: Pathophysiology, current therapeutics and future perspective in rheumatoid arthritis. *Expert Opin. Ther. Targets.* 2017, 21, 433–447. [CrossRef] [PubMed]
- Méchin, M.-C.; Takahara, H.; Simon, M. Deimination and peptidylarginine deiminases in skin physiology and diseases. *Int. J. Mol. Sci.* 2020, 21, 566. [CrossRef] [PubMed]
- Wang, Y.; Li, M.; Stadler, S.; Correll, S.; Li, P.; Wang, D.; Hayama, R.; Leonelli, L.; Han, H.; Grigoryev, S.A.; et al. Histone hypercitrullination mediates chromatin decondensation and neutrophil extracellular trap formation. *J. Cell Biol.* 2009, 184, 205–213. [CrossRef]
- 27. Toichi, E.; Tachibana, T.; Furukawa, F. Rapid improvement of psoriasis vulgaris during drug-induced agranulocytosis. *J. Am. Acad. Dermatol.* **2000**, *43*, 391–395. [CrossRef] [PubMed]
- Naik, H.B.; Natarajan, B.; Stansky, E.; Ahlman, M.A.; Teague, H.; Salahuddin, T.; Ng, Q.; Joshi, A.A.; Krishnamoorthy, P.; Dave, J.; et al. Severity of psoriasis associates with aortic vascular inflammation detected by FDG PET/CT and neutrophil activation in a prospective observational study. *Arterioscler. Thromb. Vasc. Biol.* 2015, *35*, 2667–2676. [CrossRef]
- Li, P.; Li, M.; Lindberg, M.R.; Kennett, M.J.; Xiong, N.; Wang, Y. PAD4 is essential for antibacterial innate immunity mediated by neutrophil extracellular traps. J. Exp. Med. 2010, 207, 1853–1862. [CrossRef] [PubMed]
- Teague, H.L.; Varghese, N.J.; Tsoi, L.C.; Dey, A.K.; Garshick, M.S.; Silverman, J.I.; Baumer, Y.; Harrington, C.L.; Stempinski, E.; Elnabawi, Y.A.; et al. Neutrophil subsets, platelets, and vascular disease in Psoriasis. *JACC Basic Transl. Sci.* 2019, 4, 1–14. [CrossRef] [PubMed]
- Skrzeczynska-Moncznik, J.; Zabieglo, K.; Bossowski, J.P.; Osiecka, O.; Wlodarczyk, A.; Kapinska-Mrowiecka, M.; Kwitniewski, M.; Majewski, P.; Dubin, A.; Cichy, J. Eosinophils Regulate interferon alpha production in plasmacytoid dendritic cells stimulated with components of neutrophil extracellular traps. *J. Interf. Cytokine Res.* 2017, *37*, 119–128. [CrossRef] [PubMed]
- Bergen, L.L.; Petrovic, A.; Aarebrot, A.K.; Appel, S. The TNF/IL-23/IL-17 axis-head-to-head trials comparing different biologics in psoriasis treatment. *Scand. J. Immunol.* 2020, 92, e12946. [CrossRef] [PubMed]

- Gordon, R.A.; Herter, J.A.; Rosetti, F.; Campbell, A.M.; Nishi, H.; Kashgarian, M.; Bastacky, S.I.; Marinov, A.; Nickerson, K.M.; Mayadas, T.N.; et al. Lupus and proliferative nephritis are PAD4 independent in murine models. *JCI Insight* 2017, 2, e92926. [CrossRef] [PubMed]
- 34. Rohrbach, A.S.; Hemmers, S.; Arandjelovic, S.; Corr, M.; Mowen, K.A. PAD4 is not essential for disease in the K/BxN murine autoantibody-mediated model of arthritis. *Arthritis Res. Ther.* **2012**, *14*, R104. [CrossRef] [PubMed]
- 35. Hemmers, S.; Teijaro, J.R.; Arandjelovic, S.; Mowen, K.A. PAD4-mediated neutrophil extracellular trap formation is not required for immunity against influenza infection. *PLoS ONE* **2011**, *6*, e22043. [CrossRef]
- Yasuda, M.; Morimoto, N.; Yonemoto, Y.; Sunaga, N.; Ishikawa, O. Concurrent development of psoriatic arthritis, drug-induced interstitial pneumonia, and increased anti-nuclear antibodies in a patient with psoriasis vulgaris undergoing adalimumab treatment. *Eur. J. Dermatol.* 2019, 1, 81–82. [CrossRef]
- Watanabe, S.; Iwata, Y.; Fukushima, H.; Saito, K.; Tanaka, Y.; Hasegawa, Y.; Akiyama, M.; Sugiura, K. Neutrophil extracellular traps are induced in a psoriasis model of interleukin-36 receptor antagonist-deficient mice. *Sci. Rep.* 2020, 19, 20149. [CrossRef]
- Arican, O.; Aral, M.; Sasmaz, S.; Ciragil, P. Serum levels of TNF-alpha, IFN-gamma, IL-6, IL-8, IL-12, IL-17, and IL-18 in patients with active psoriasis and correlation with disease severity. *Mediat. Inflamm.* 2005, 24, 273–279. [CrossRef]
- Kolfenbach, J.R.; Deane, K.D.; Derber, L.A.; O'Donnell, C.I.; Gilliland, W.R.; Edison, J.D. Autoimmunity to peptidyl arginine deiminase type 4 precedes clinical onset of rheumatoid arthritis. *Arthritis Care Res.* 2010, 62, 2633–2639. [CrossRef]
- Safi, R.; Kallas, R.; Bardawil, T.; Mehanna, C.J.; Abbas, O.; Hamam, R.; Uthman, I.; Kibbi, A.G.; Nassar, D. Neutrophils contribute to vasculitis by increased release of neutrophil extracellular traps in Behcet's disease. J. Dermatol. Sci. 2018, 92, 143–150. [CrossRef]
- Douda, D.N.; Khan, M.A.; Grasemann, H.; Palaniyar, N. SK3 channel and mitochondrial ROS mediate NADPH oxidaseindependent NETosis induced by calcium influx. *Proc. Natl. Acad. Sci. USA* 2015, 112, 2817–2822. [CrossRef] [PubMed]
- 42. Neeli, I.; Radic, M. Opposition between PKC isoforms regulates histone deimination and neutrophil extracellular chromatin release. *Front. Immunol.* **2013**, *4*, 38. [CrossRef] [PubMed]
- 43. Nakashima, K.; Arai, S.; Suzuki, A. PAD4 regulates proliferation of multipotent haematopoietic cells by controlling c-myc expression. *Nat. Commun.* 2013, *4*, 1836. [CrossRef]
- 44. Ding, Y.; Ouyang, Z.; Zhang, C.; Zhu, Y.; Xu, Q.; Sun, H.; Qu, J.; Sun, Y. Tyrosine phosphatase SHP2 exacerbates psoriasis-like skin inflammation in mice via ERK5-dependent NETosis. *MedComm* **2022**, *3*, 120. [CrossRef] [PubMed]
- 45. Tatsiy, O.; McDonald, P.P. Physiological stimuli induce PAD4-dependent, ROS-independent NETosis, with early and late events controlled by discrete signaling pathways. *Front. Immunol.* **2018**, *9*, 2036. [CrossRef] [PubMed]
- Thiam, H.R.; Wong, S.L.; Qiu, R.; Kittisopikul, M.; Vahabikashi, A.; Goldman, A.E.; Goldman, R.D.; Wagner, D.D.; Waterman, C.M. NETosis proceeds by cytoskeleton and endomembrane disassembly and PAD4-mediated chromatin decondensation and nuclear envelope rupture. *Proc. Natl. Acad. Sci. USA* 2020, 117, 7326–7337. [CrossRef] [PubMed]
- 47. Seri, Y.; Shoda, H.; Suzuki, A. Peptidylarginine deiminase type 4 deficiency reduced arthritis severity in a glucose-6-phosphate isomerase-induced arthritis model. *Sci. Rep.* **2015**, *5*, 13041. [CrossRef]
- Izuki, A.; Kochi, Y.; Shoda, H. Decreased severity of experimental autoimmune arthritis in peptidylarginine deiminase type 4 knockout mice. BMC Musculoskelet. Disord. 2016, 17, 205. [CrossRef]
- Knight, J.S.; Zhao, W.; Luo, W.; Subramanian, V.; O'Dell, A.A.; Yalavarthi, S.; Hodgin, J.B.; Eitzman, D.T.; Thompson, P.R.; Kaplan, M.J. Peptidylarginine deiminase inhibition is immunomodulatory and vasculoprotective in murine lupus. *J. Clin. Investig.* 2013, 123, 2981–2993. [CrossRef]
- Knight, J.S.; Subramanian, V.; O'Dell, A.A.; Yalavarthi, S.; Zhao, W.; Smith, C.K.; Hodgin, J.B.; Thompson, P.R.; Kaplan, M.J. Peptidylarginine deiminase inhibition disrupts NET formation and protects against kidney, skin and vascular disease in lupusprone MRL/lpr mice. *Ann. Rheum. Dis.* 2015, 74, 2199–2206. [CrossRef]
- Savchenko, A.S.; Borissoff, J.I.; Martinod, K.; De Meyer, S.F.; Gallant, M.; Erpenbeck, L.; Brill, A.; Wang, Y.; Wagneret, D.D. VWF-mediated leukocyte recruitment with chromatin decondensation by PAD4 increases myocardial ischemia/reperfusion injury in mice. *Blood* 2014, 123, 141–148. [CrossRef] [PubMed]
- Huang, H.; Tohme, S.; Al-Khafaji, A.B.; Tai, S.; Loughran, P.; Chen, L.; Wang, S.; Kim, J.; Billiar, T.; Wang, Y.; et al. Damageassociated molecular pattern-activated neutrophil extracellular trap exacerbates sterile inflammatory liver injury. *Hepatology* 2015, 62, 600–614. [CrossRef] [PubMed]
- Franck, G.; Mawson, T.L.; Folco, E.J.; Molinaro, R.; Ruvkun, V.; Engelbertsen, D.; Liu, X.; Tesmenitsky, Y.; Shvartz, E.; Sukhova, G.K.; et al. Roles of PAD4 and NETosis in experimental atherosclerosis and arterial injury: Implications for superficial erosion. *Circ. Res.* 2018, 123, 33–42. [CrossRef] [PubMed]
- 54. Ciesielski, O.; Biesiekierska, M.; Panthu, B.; Soszyński, M.; Pirola, L.; Balcerczyk, A. Citrullination in the pathology of inflammatory and autoimmune disorders: Recent advances and future perspectives. *Cell. Mol. Life Sci.* 2022, 79, 94. [CrossRef]
- Nakabo, S.; Romo-Tena, J.; Kaplan, M.J. Neutrophils as drivers of immune dysregulation in autoimmune diseases with skin manifestations. *J. Investig. Dermatol.* 2022, 142, 823–833. [CrossRef]
- Huang, S.U.; O'Sullivan, K.M. The expanding role of extracellular traps in inflammation and autoimmunity: The new players in casting dark webs. *Int. J. Mol. Sci.* 2022, 23, 3793. [CrossRef] [PubMed]
- Bawadekar, M.; Shim, D.; Johnson, C.J.; Warner, T.F.; Rebernick, R.; Damgaard, D.; Nielsen, C.H.; Pruijn, G.J.M.; Nett, J.E.; Shelef, M.A. Peptidylarginine deiminase 2 is required for tumor necrosis factor alpha-induced citrullination and arthritis, but not neutrophil extracellular trap formation. *J. Autoimmun.* 2017, *80*, 39–47. [CrossRef] [PubMed]

- Padhi, A.; Srivastava, A.; Ramesh, A.; Ehrström, M.; Simon, M.; Sonkoly, E.; Eidsmo, L.; Bergman, P.; Lysell, J. IL-22 downregulates peptidylarginine deiminase-1 in human keratinocytes: Adding another piece to the IL-22 puzzle in epidermal barrier formation. *J. Investig Dermatol.* 2022, 142, 333–342. [CrossRef] [PubMed]
- 59. Meng, X.; Guo, R.; Fan, C.; Li, X.; Liu, X.; Chen, X.; Lu, L. RIPK1 downregulation enhances neutrophil extracellular traps in psoriasis. *Adv. Dermatol. Allergol.* **2022**, *39*, 72–80. [CrossRef]