Rare but specific:

5-bp Composite Motifs Define SMAD Binding in BMP Signaling

Jerome Jatzlau^{1,2}, Sophie-Nhi Do¹, Rebeca A. Mees², Paul-Lennard Mendez¹, Rameez Jabeer Khan², Lukas Maas¹, Lidia Ruiz², Pau Martin-Malpartida², Maria J. Macias^{2,3,*}, Petra Knaus^{1,*}

Supplementary Figures

- Fig. S1. SMAD-MH1 purity control and experimental set-up
- Fig. S2. BRE₂-Luc and CAGA₁₂-Luc define experimental conditions for selective comparison of BMP6 and TGFβ1
- Fig. S3. BMP6 dose-response curves of SMAD composite motif reporters
- Fig. S4. Composite motif spacers below 5 bp do not inhibit SMAD1-MH1 double binding
- Fig. S5. BMP-responsive SMAD homocomposite motif reporters are unresponsive to TGFβ1
- Fig. S6. BMP-responsive SMAD heterocomposite motif reporters are unresponsive to $\mathsf{TGF}\beta1$
- Fig. S7. BMP-specificity of SMAD composite motif reporters is maintained in U2OS cells

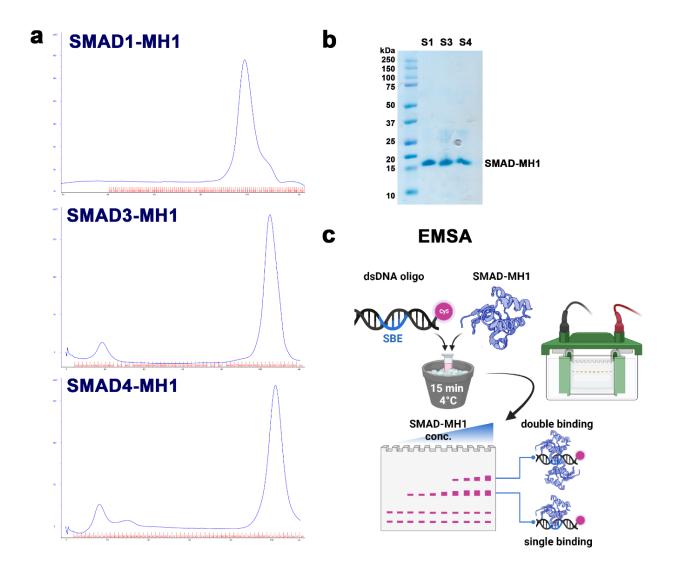


Fig. S1. SMAD-MH1 purity control and experimental set-up: *a, SEC traces of purified SMAD-MH1 domains. b, Coomassie staining of purified SMAD-MH1 domains. c, Increasing concentration of SMAD1/3/4-MH1 domains were incubated with Cy5-dsDNA oligos containing known SMAD-binding element (SBE)s and analyzed for binding using Electro Mobility Shift Assay. Single or double binding of SMAD-MH1 is detected by weight shift of Cy5-dsDNA signal.*

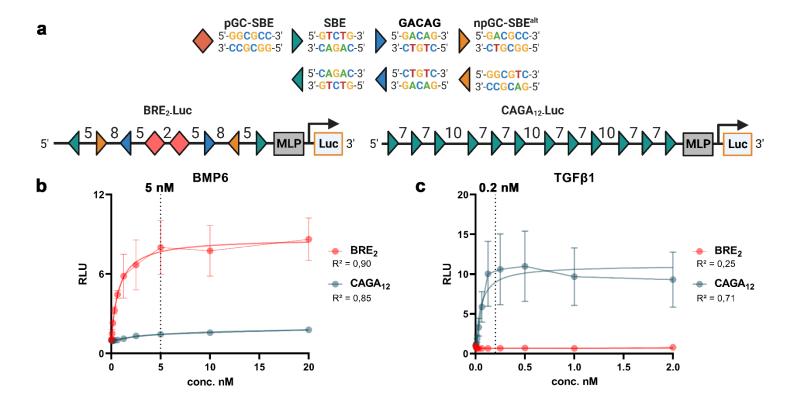


Fig. S2. BRE₂-Luc and CAGA₁₂-Luc define experimental conditions for selective comparison of BMP6 and TGF β 1: a, SMAD motifs in the two luciferase reporters are indicated. b, BRE₂-Luc or CAGA₁₂-Luc was transfected together with TK-renilla luciferase into HEK293t cells, cells were then starved and stimulated with BMP6 (0.03 - 20 nM) or TGF β 1 (0.003 - 2 nM) for 24 h before analysis using a microplate reader. b-c, Data are shown as mean fold induction in relative luciferase units (RLU) \pm SD (n=3 independent experiments). 5 nM BMP6 and 0.2 nM TGF β 1 were selected for all consecutive experiments as they gave a high response close to saturation of the signal.

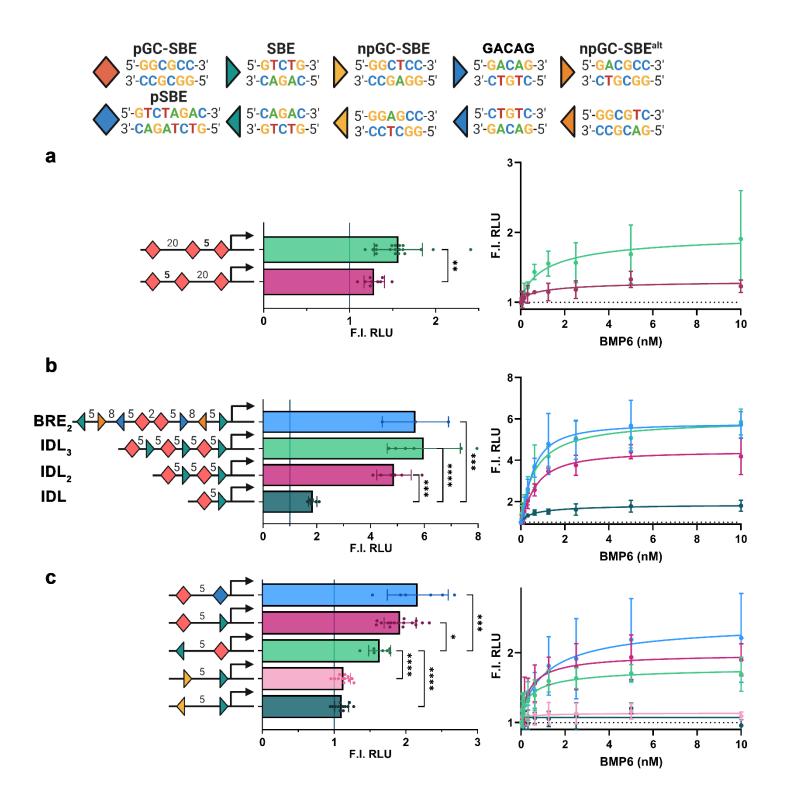


Fig. S3. BMP6 dose-response curves of SMAD composite motif reporters: *a-c,* HEK293t cells were co-transfected with SBE-firefly-Luc constructs and TK-renilla luciferase, starved and stimulated with BMP6 for 24 hrs before analysis using a microplate reader. Bar charts represent response to 5 nM BMP6 (left) and dose curves represent the response to 0.04 nM to 10 nM BMP6 (right). **a,** Dual luciferase reporter assay displaying decreased BMP6-responsiveness, if 5bp-spaced pGC-SBE

homocomposite motif is 20+6 bp further away from the MLP. **b**, Dual luciferase reporter assay displaying elevated BMP6-responsiveness towards multiplied (2-3x) pGC-SBE/SBE heterocomposite motifs. **c**, Dose response comparison of heterocomposite motifs from Fig.3. **a-c**, Data are shown as mean fold induction to unstimulated cells (grey line) in relative luciferase units (RLU) \pm SD (5nM BMP6 n=3–19; dose curves n=3 independent experiments). Statistical significance was calculated between samples using one-way ANOVA and Tukey's post-hoc test.

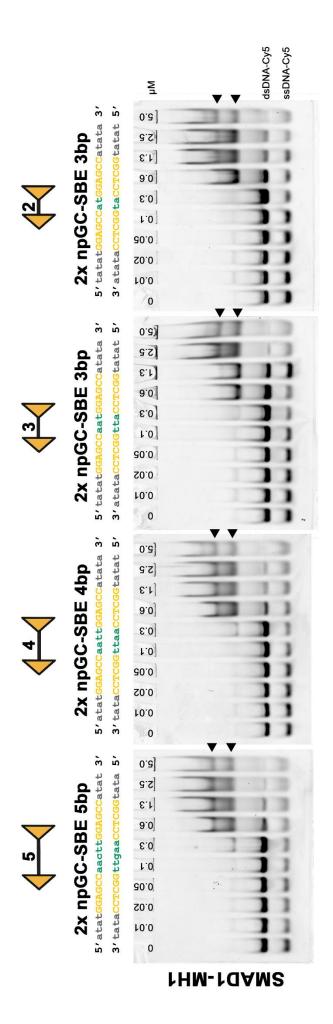


Fig. S4. Composite motif spacers below 5 bp do not inhibit SMAD1-MH1 double binding: EMSA experiments were performed testing the binding of human SMAD1-MH1 domains to differently spaced (2-5 bp) npGC-SBE homocomposite motifs. Protein concentrations (µM) are shown on top of the EMSA. Abbreviations for the DNA oligonucleotides and dsDNA sequence are shown above. Single and double SMAD-MH1 binding to dsDNA is indicated with black triangles.

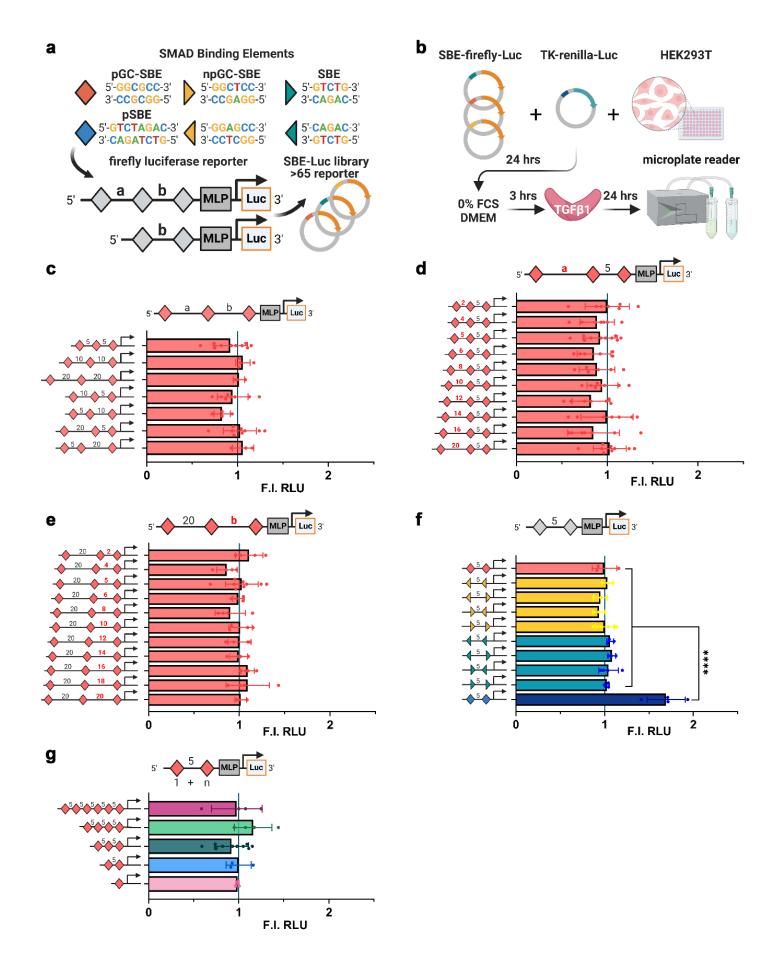


Fig. S5. BMP-responsive SMAD homocomposite motif reporters are unresponsive to TGFβ1: a, A library of synthetic firefly luciferase constructs was cloned with 1 to 6 SMAD motifs positioned 10 bp before a minimal promotor (MLP) with varying spacer length for pGC-SBEs and varying orientation for npGC-SBE and SBE motifs. b, HEK293t cells were co-transfected with SBE-firefly-Luc constructs and TK-renilla luciferase, starved and stimulated with TGFβ1 (0.2 nM) for 24 hrs before analysis using a microplate reader. c-g, Dual luciferase reporter assay displaying no TGFβ1-responsiveness towards constructs with differently spaced SMAD homocomposite motifs, except to two 5-bp spaced pSBE motifs. c-g, Data are shown as mean fold induction to unstimulated cells (grey line) in relative luciferase units (RLU) \pm SD (n=3–10 independent experiments). Statistical significance was calculated between samples using one-way ANOVA and Tukey's post-hoc test.

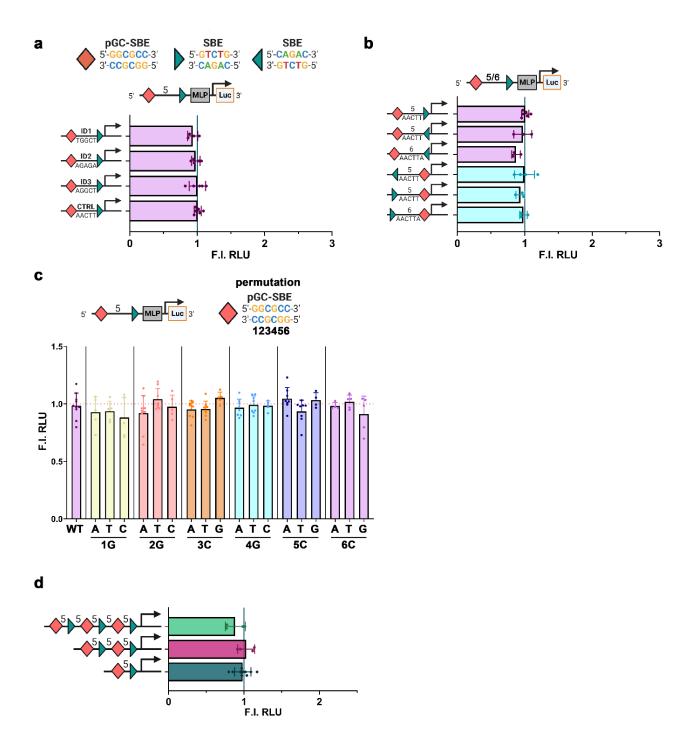


Fig. S6. BMP-responsive SMAD heterocomposite motif reporters are unresponsive to TGFβ1: $\it a-c$, Dual luciferase reporter assay displaying no TGFβ1-responsiveness (0.2 nM TGFβ1, 24 hrs) towards constructs with differently spaced ($\it a$), oriented ($\it b$), or permutated ($\it c$) or multiplied ($\it d$) SMAD heterocomposite motifs. Data are shown as mean fold induction to unstimulated cells (line) in relative luciferase units (RLU) ±SD ($\it n=3-10$ independent experiments). Statistical significance was calculated between samples using one-way ANOVA and Tukey's post-hoc test ($\it a-b,d$) and between samples ($\it c$) or relative to WT ctrl ($\it c$) using one-way ANOVA and Šídák's or Dunnett's multiple comparisons test.

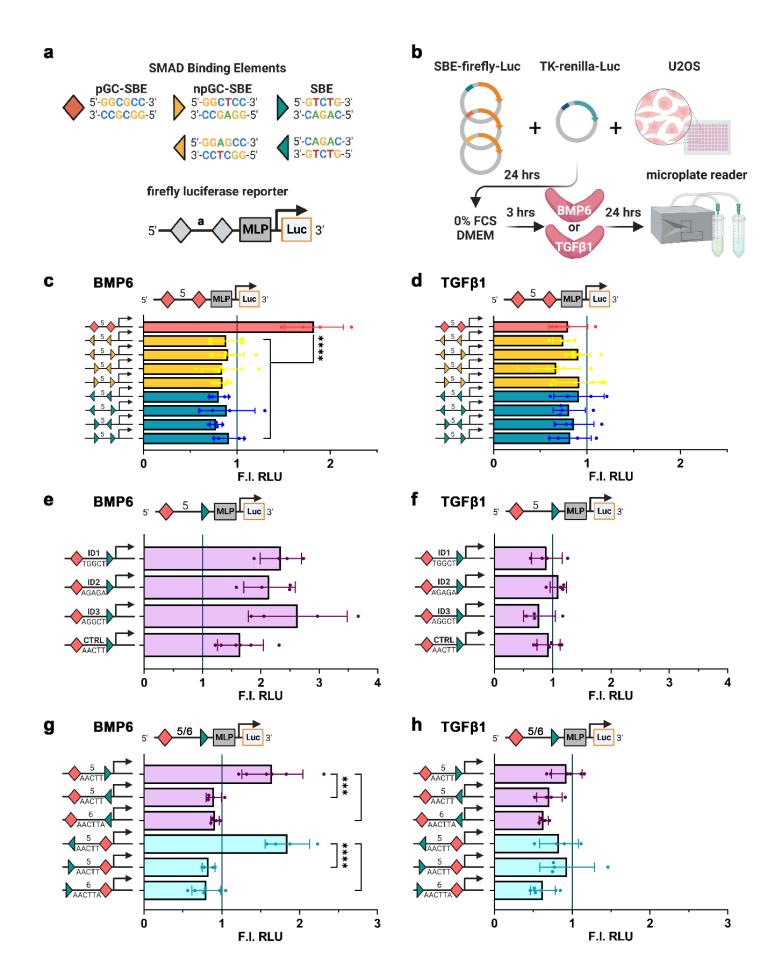


Fig. S7. BMP-specificity of SMAD composite motif reporters is maintained in U2OS cells: a, A library of synthetic firefly luciferase construct was cloned with 2 SMAD motifs positioned before a minimal promotor (MLP) with varying spacer length and orientation of SMAD motifs. b, U2OS cells were co-transfected with SBE-firefly-Luc constructs and TK-renilla luciferase, starved, and stimulated with BMP6 (5 nM) or TGF61 (0.2 nM) for 24 hrs before analysis using a microplate reader. c-h, Dual luciferase reporter assay displaying BMP6-responsiveness towards 5-bp spaced SMAD homo- and heterocomposite motif constructs. Data are shown as mean fold induction to unstimulated cells in relative luciferase units (RLU) \pm SD (n=4–6 independent experiments). Statistical significance was calculated between samples using one-way ANOVA and Tukey's post-hoc test.