

Review

# **Current Trends in Nanomaterial-Based Amperometric Biosensors**

Akhtar Hayat 1,2, Gaëlle Catanante 1 and Jean Louis Marty 1,\*

- BIOMEM, Universitéde Perpignan, 52 Avenue Paul Alduy, 66860 Perpignan Cedex, France; E-Mails: akhtarloona@gmail.com (A.H.); gaelle.catanante@univ-perp.fr (G.C.)
- Interdisciplinary Research Centre in Biomedical Materials (IRCBM), COMSATS Institute of Information Technology (CIIT), Lahore 54000, Pakistan
- \* Author to whom correspondence should be addressed; E-Mail: jlmarty@univ-perp.fr; Tel.: +33-468-661-756.

External Editor: Roberto Pilloton

Received: 11 October 2014; in revised form: 17 November 2014 / Accepted: 1 December 2014 /

Published: 8 December 2014

**Abstract:** The last decade has witnessed an intensive research effort in the field of electrochemical sensors, with a particular focus on the design of amperometric biosensors for diverse analytical applications. In this context, nanomaterial integration in the construction of amperometric biosensors may constitute one of the most exciting approaches. The attractive properties of nanomaterials have paved the way for the design of a wide variety of biosensors based on various electrochemical detection methods to enhance the analytical characteristics. However, most of these nanostructured materials are not explored in the design of amperometric biosensors. This review aims to provide insight into the diverse properties of nanomaterials that can be possibly explored in the construction of amperometric biosensors.

**Keywords:** electrochemical sensing; amperometric biosensors; nanomaterials; sensing design; analytical applications

#### 1. Introduction

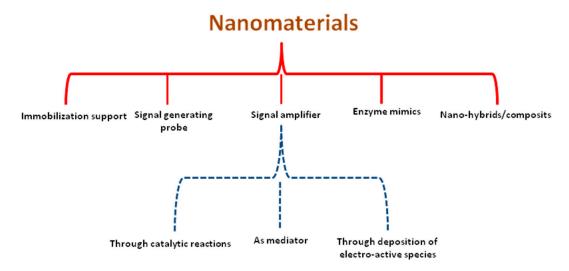
Biosensors have emerged as an alternative tool to classical chromatographic methods for the analysis of analyte samples. A biosensor functions by converting some biological response into a detectable signal. The commonly used biomolecules in biosensor design include DNA/RNA, antibodies, enzyme, peptides, microorganisms and whole cells. The signals obtained are processed and transferred to a display or data storage device [1,2]. The main advantageous characteristic of a biosensor is the direct spatial contact between the bio-recognition element and the transducer surface [3]. The output signal of biosensor can be measured with electrochemical, quartz crystal microbalance, optical absorption fluorescence, surface plasmon resonance, radioactive, magnetic and other transducer platforms. Among all these transduction methods, electrochemistry provides an attractive way of encompassing the advantages of instrumental simplicity, moderate cost, and portability for decentralized analysis. The modern and newly emerged electrochemical methods are sensitive, selective, facile and rapid making them suitable candidate for diverse analytical applications. A number of electrochemical methodologies have been explored in the field of electrochemical biosensors such as differential pulse voltammetry, cyclic voltammetry, linear sweep voltammetry, square wave voltammetry, amperometry and impedimetric analysis [4,5]. Amperometry is the common most electrochemical technique which has been widely employed in the domain of electrochemical biosensors. Amperometry measures the amplitude of a reduction or oxidation current at a given specific potential over a fixed period of time. Despite much progress, increased interest in the development of new materials applicable in amperometric techniques to fulfill the necessity of control specific biomolecular interaction still exists. Important advances have been achieved with the utilization of nanomaterials in the design of electrochemical biosensors to improve their analytical performance [6].

#### 2. Functions of Nanomaterials

The last decade has witnessed an enormous impact of nanomaterials on the field of biosensors [7]. Significant progress has been achieved in synthetic approaches to prepare nanomaterials of desired properties such as controllable size, shape, surface charge and physiochemical characteristics [8,9]. Nanomaterials can be decorated with polymers and biomolecules to achieve better biocompatibility and precise targeting. These features have made it possible to integrate nanomaterials in biosensors for any desired function, and led to an increasing use of nanomaterials in the design of electrochemical amperometric sensors. Thousands of scientific articles describing associations between nanomaterials and biomolecules have been published in the literature [10]. In this context, different techniques have been explored to integrate nanomaterials of diverse nature in the electrochemical transduction platform. The Langmuir-Blodgett method was the pioneering approach in the fabrication of thin films formed by transferring an amphiphilic materials dispersed at air/water interface onto a solid substrate [11]. Another interesting method providing high control of thickness at the nanoscale level was developed by Decher [12]. Despite the specific chemical interactions, the proposed layer by layer methodologies rely on the coulombic electrostatic that provides multilayer growth. This strategy has been proven by various researchers as the simplest tool for film fabrication which can be applied to various types of materials like polyelectrolytes [13], metallophthalocyanines [14], carbon nanotubes [15] and all types of

nanoparticles [16]. Recent trends have also shown the direct conjugation of nanomaterials with biomolecules or target analyte through various kinds of interactions such as physical adsorption, covalent attachment and electrostatic interaction with week hydrogen bonding. However, in some cases nanomaterials are directly employed in the interacting medium on transducer surface to enhance the electroanalytical performance of the system [17]. There is no doubt on the importance of nanostructured materials and their implications in biosensor fabrication. In this review paper, we discuss recent trends in the field of amperometric biosensors exploring the diverse functions of nanomaterials to improve the analytical figures of the biosensors.

**Figure 1.** Possible functions that can be achieved with nanomaterial integration in amperometric biosensors.



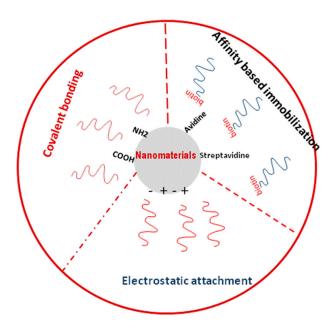
Nanotechnolgy is defined as the creation of functional systems through control of matter at the 1–100 nm scale. Nanomaterials exhibit interesting physical, chemical, and electronic properties such as size and shape dependent surface plasmon resonance, high surface energy and surface to volume ratio and tunable surface properties [18,19]. For example, the physical properties of nanomaterials can be explored for cell imaging, drug delivery systems and to design optical assays and bioassays. Similarly, the chemical properties of the nanomaterials facilitate nano surface modification with a variety of functional groups to attach molecules of diverse nature. The integration of the electronic properties of nanomaterials has dramatically reduced the size of electronic devices. In the same context, these intrinsic and distinguished features make nanomaterials the best fit for designing novel biosensor platforms. A wide variety of nanomaterials including metal nanoparticles, oxide nanoparticles, quantum dots, nanowires, nanoshells, carbon nanotubes and even composite nanoparticles have found attractive applications in many kind of amperometric biosensors for a broad spectrum of analytical applications. Nanomaterials of different nature, size, shape and composition can play various roles in different sensing systems [20]. However, the organization of nanomaterials into controlled surface architectures is essential for the successful realization of these properties. The important possible functions that can be achieved with nanomaterial integration are listed below, presented in Figure 1, and will be discussed in detail in the next sections.

- (1) Immobilization platform for biomolecules
- (2) Amperometric signal amplification
- (3) Nanomaterials as signal generating probe
- (4) Nanomaterials as enzyme mimics
- (5) Hybrid and composite nanomaterials in amperometric biosensors

## 2.1. Immobilization Platform for Biomolecules

Recent interest in nanotechnology has provided a wealth of diverse nanoscaffolds that could be used to support biomolecules/analyte immobilization due to their potential features (Figure 2) [21,22]. Immobilization of biomolecules is an advantageous step towards the commercialization of amperometric sensors due to the possible resulting increase in physiological stability. The premise of employing nanomaterials for immobilization on a transducer surface is to reduce diffusion limits and maximize the surface area to increase biomolecule loading [23]. In the last years several types of nanomaterials have been produced in various nanostructure shapes like nanorods, nanowires, nanotubes and nanorings for immobilization purposes [24,25]. Nanomaterials have a strong tendency to adsorb biomolecules and to help the immobilization of biomolecules in biosensor construction. The adsorption of biomolecules directly onto bulk materials may result in denaturation and loss of bioactivity, while nanosize materials retain the bioactivity of biomolecules. It is of vital importance that several nanoparticles carry charges and hence they can provide electrostatic surface to attach the biomolecules with different charges.

Figure 2. Nanomaterial-based immobilization methodologies.



Among all the nanomaterials, magnetic nanoparticles have been used extensively to immobilize both biomolecules and analytes on the transducer surface [26–28]. Several types of magnetic nanoparticles and magnetic supports such as microspheres of diverse biomaterials encapsulating the magnetic nanoparticles and copolymers with magnetic nanoparticles have been employed in biosensor fabrication with good results [29,30]. The high surface to volume ratio of magnetic nanoparticles facilitates high binding capacity and high catalytic specificity of the conjugated biomolecules. Magnetic nanoparticles

decorated with streptavidin, and having amine and carboxylic functional groups to immobilize specific modified biomolecules/analytes are commercially available [31–33]. Similarly protein G magnetic beads to specifically immobilize antibodies are available [34]. The magnetic beads-based immobilization methodology is easy to operate with simple application of a magnet to provide a magnetic field to capture modified the modified nanoparticles on the transducer surface [35,36]. However, the application of magnetic-based methods is restricted due to lack of reproducibility and renewal of the sensor surface. Future work in this regard may focus on improving the pitfalls of magnetic nanoparticles used by designing new strategies. For example, due to the ease of manipulation and separation using a magnetic field, it is possible to simply 'switch off' the magnetic effects by removing the magnetic induction field, which can be advantageous for bead recovery. Moreover, magnetic beads can be manipulated independent of normal microfluidic or biological processes which results in improved exposure of the functionalized bead surface to the surrounding viscous liquid, due to the increased relative motion of the bead with respect to the fluid. Additionally, the application of a revolving external magnetic during manipulation could also be beneficial to disperse the magnetic beads uniformly on the reactor surface to obtain reproducible sensing surfaces. Besides magnetic nanoparticles, gold nanoparticles serve as an excellent platform for the immobilization of biomolecules since the interaction between amino and cysteine groups of biomolecules can be explored apart from gold and thiol chemistry [37–39]. Several workers have reported a one-step method for immobilization of biomolecues on transducer surfaces through the integration of poly(methyl methacrylate) nanoparticles [40]. Non-metallic polystyrene nanoparticles have also been used in the literature to improve the biomolecule immobilization efficiency [23]. Nanoparticles of silver, silica, zinc oxide, aluminium oxide, zirconium oxide, manganese oxide and nickel oxide were explored to immobilize biomolecules, specifically enzymes, to construct amperometric biosensors [41-44]. Recent research has focused on the polymerization of different molecules on the nanoparticles, and subsequently using them as anchoring supports in the fabrication of biosensors. Martin et al., prepared novel core-shell Fe<sub>3</sub>O<sub>4</sub>@poly(dopamine) magnetic nanoparticles through an *in situ* self-polymerization method. The core-shell nanoparticles were employed as solid supports for the covalent immobilization of horseradish peroxidase (HRP), and the resulting biofunctionalized magnetic nanoparticles were employed to construct an amperometric biosensor for H<sub>2</sub>O<sub>2</sub>. The enzyme biosensor showed a high sensitivity, a low limit of detection, a wide linear range and high stability for 1 month [29]. Lu et al., proposed a novel, high-yield and template-free method for the synthesis of Ag nanoparticle-decorated thionine/infinite coordination polymer (AgNP/THI/ICP) fibers. The thionine was adsorbed to the AgNP/THI/ICP fibers by  $\pi$ -conjugation and acted as the redox probe. The AgNP/THI/ICP fibers not only favored the antibody immobilization but also facilitated the electron transfer [45]. Turkmen et al., fabricated a new biosensor based on immobilization of glucose oxidase (Gox) using an enzyme solution containing o-phenylenediamine(oPD) on platinum nanoparticles (PtNPs) electrodeposited polyvinylferrocenium perchlorate matrix (PVF+ClO<sub>4</sub><sup>-</sup>). Gox was immobilized simultaneously with the electropolymerization of oPD on PtNPs/PVF+ClO<sub>4</sub>-/Pt, and the resulting biosensor showed excellent anti-interference ability to ascorbic acid and uric acid [46]. Despite the much progress in nanomaterials integration in electrochemical biosensors, some of the above described nanomethods are vet to be explored in the construction of amperometric biosensors.

Recent research to explore the hidden properties of nanomaterials has indicated that most nano-materials exhibit intrinsic enzyme-like properties that can be used to construct enzyme-free sensors

to detect target analytes. This reactive nature of nanomaterials can be advantageous in one way, but on the other hand, it may limit the applications of nanomaterials as immobilization supports. One such example is the construction of a glucose biosensor based on the reactivity of ferromagnetic nanoparticles towards hydrogen peroxide [47]. This indicates that use of nanomaterials as immobilization support may increase the non-specific signal and yield irreproducible results due to their reactivity with commonly used optical and electrochemical signal-generating probes. To control the surface reactivity, different strategies, including use of surfactants, encapsulation of the nanomaterial surface and variation in pH have been employed in recent years. Based on these findings, it will be of vital importance and interest to use inert nanomaterials in the construction of biosensors to overcome the problem of non-specific reactivity.

# 2.2. Amperometric Signal Amplification

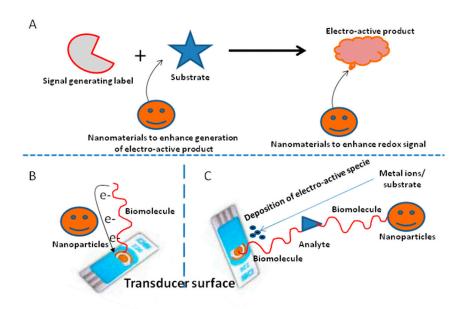
The detection of ultralow analyte concentrations is required not only in the chemical laboratory, but also in variety of areas including clinical diagnostic, food safety and environmental protection. Analytical figures of merits are continuously being pushed down by the gradual improvement of analytical technologies. The sensitivity of any protocol is related to the correlation between the analyte concentration and the strength of the output signal. Many efforts have been devoted to the exploration of novel means to realize ultrasensitive detection. Such strategies include the use of new labels (electroactive molecules, redox complexes, and metal ions), polymerase chain reaction, mass spectrometery and the integration of enzyme-assisted signal amplification processes [17]. Although these methods have increased the sensitivity, they are destructive and mainly suffer from time-consuming derivatization, high cost and the need for professional operation. With the introduction of nanotechnology and nanoscience, nanomaterial-based signal amplification has attained great importance in realizing high sensitivity and selectivity for the detection of analytes due to the rapid analysis procedures and easy miniaturization. A lot of nanomaterials including metal nanoparticles, semiconductor nanoparticles and carbon nanosized structures have emerged as electrochemical signal amplifiers in the design of electrochemical devices [48]. The possible mechanisms for nanomaterial signal amplification are depicted in the Figure 3.

#### 2.2.1. Signal Amplification through Catalytic Reaction

The combination of the catalytic properties of nanomaterials with the unique properties of biosensors can results in the fabrication of highly sensitive sensors. Bio-functionalized nanoparticles can produce synergistic effects between catalytic activity to accelerate electrochemical signals, leading to lower and lower detection limits [49], even reaching zeptomolar concentrations. Furthermore, the linear correlation ranges becomes wider and wider. For example, carboxy-modified graphene oxide has been shown as a catalytic material for the oxidation of hydrogen peroxide [50], and carboxy-modified electrodes may have a synergistic effect towards the oxidation of hydrogen peroxide with subsequent improvement in the sensitivity of the method [51]. Platinum nanoparticles also exhibit catalytic properties and have been used in the fabrication of electrochemical sensors [52,53]. The platinum modified electrodes showed sensitive responses to H<sub>2</sub>O<sub>2</sub> and lowered the oxidation potential. Since H<sub>2</sub>O<sub>2</sub> is the product of many enzymatic reactions, the proposed platinum modified electrodes may find potential application in

electrochemical biosensors for many substances. Similarly Ni nanoparticle-modified electrodes have been shown to possess electrocatalytic activity in the electro-oxidation of sugars, including glucose, sucrose, fructose and lactose [54]. The application of copper nanoparticle-modified transducer surfaces for sensitive detection of amino acids has been demonstrated [55]. Copper oxide nanoparticle-modified electrode surfaceshave shown catalytic properties in the detection of amikacin in another study [56]. Based on their redox properties, nanoceria-modified electrode surfaces have been employed to improve the analytical performance of electrochemical biosensors for a variety of target analytes [49,57]. Electrochemical biosensors based on the catalytic properties of other nanoparticles have also been reported in the literature, for example silver and gold nanoparticles have been used in the construction of electrochemical biosensors for the detection of glucose [58,59].Fe<sub>2</sub>O<sub>3</sub>-, silica-, Pd-, ZnO- and MnO<sub>2</sub>-modified electrodes have been explored for this purpose. Apart from the nanomaterials described above, carbon-based materials such as single walled and multiwalled carbon nanotubes have been also used to generate electrochemical signals [60]. For example, Yang et al., fabricated a sandwich-type amperometric immunosensor for carcinoembryonic antigen by using functionalized carbon nanotubes decorated with concanavalin A along with horseradish peroxidase-labeled carcinoembryonic secondary antibodies. The HRP-Ab2 was assembled onto carbon nanotubes templates and the carcinoembryonic antibody was immobilized on the gold electrode modified by a cysteine monolayer. The developed amperometric immunosensor showed excellent analytical characteristics in terms of sensitivity and linear range [61]. Some of these reported materials are not investigated in the construction of amperometric biosensors, so future work in the domain of amperometric biosensors may focus on this.

**Figure 3.** Nanomaterial-based signal amplification through **(A)** catalytic properties; **(B)** mediator; **(C)** deposition of electrochemically active species.



# 2.2.2. Nanomaterials as Mediator

Electrical contact between redox biomolecules and electrode surface is of vital importance in the construction of electrochemical sensors. However, biomolecules have no direct communication with the

electrode surface because the active sites of biomolecules are surrounded by considerably thick insulating protein shells and the electron transfer between the two moieties is blocked, posing a problem for the electrochemical measurements. In this context, the conductive properties of the metallic nanoparticles to enhance the electron transfer rate between the biomolecules and the transducer surface are very well established. Metallic nanoparticles play the role of electron transfer mediators or electrical wires to replace the commonly used mediator in the construction of electrochemical biosensors. Although, the role of metallic nanoparticles as electron transfer mediators has been extensively explored for amperometric biosensors, however, some non-metallic nanoparticles such as oxide nanoparticles and semiconductor nanoparticles can also improve the electron transfer rate between proteins and electrodes. The improvement in electron flow is related to the conductivity of nanoparticles and arrangement between nanoparticles and biomolecules. The creation of well defined and ordered arrangments of nanoparticles using advanced and novel nano-based methods is a promising approach for the fabrication of amperometric biosensors with enhanced electron transfer properties. Gold nanoparticles have been used to improve the electron transfer between biomolecules and electrode surfaces in the construction of an enzymatic biosensor [62]. A seven-fold faster response was observed in the presence of nanoparticles as compared to the enzyme signal with its natural substrate [63]. Gold nanoparticles have also been integrated in the construction of redox protein-based amperometric biosensors to improve the electron transfer rate. Besides gold nanoparticles, silver nanoparticles have also good electrical conductivity and hence can be used to enhance the electron transfer between biomolecules and the transducer surface. Silver nanoparticles in conjugation with pyrolytic graphite electrodes have been reported to act as electrical bridges that wire the electron transfer between biomolecules and the electrode surface [64]. In the same context, Jimenez et al. investigated the electrocatalytic performance of electrodes modified with Pt nanoparticles (PtNPs) and two dendritic hyperbranched carbosilane polymers for the NADH oxidation. The proposed strategy permitted them to develop efficient biosensors capable of measuring NADH from +0.3 V (vs. SCE) providing a total protection vs. the poisoning of the electrodes. Based on these interesting and useful characteristics, the authors constructed an amperometric alcohol biosensor with alcohol dehydrogenase (ADH) [65].

# 2.2.3. Nanomaterials as Seeds/Mediators to Deposit Electrochemically Active Species and Electrocatalysts

The nanomaterial-modified electrode surfaces can be used as seeds to initiate the deposition of metals through a seed-mediated nucleation or growth mechanism [66,67]. The modifying medium consists of metal salts and a reducing agent, and nanoparticles on the electrode surface serve as nucleation sites to facilitate the metal deposition, which results in higher loading of electroactive species and subsequently improves the nanoparticles-transducer surface communication. This improvement can be attributed to the fact that the deposited nanoparticles are closer to the electrode surface as compared to the nanoparticle labels. Such a feature is of vital importance in the case of competitive and labeled electrochemical immuno- and aptasensors [68], where the distance between the signal label and the electrode surface is too large for an efficient electron transfer due to the large size of both the analyte and the biomolecules. Additionally, the freshly seed-mediated nanoparticles are more electroactive, since they are free from any stabilizer, making them a highly attractive and useful tool in the design of

affinity-based amperometric biosensors. This method has been employed to deposit electrochemically active metallic species including silver, copper and gold from their salts using a reducing agent for signal amplification [69–71]. So far, an enormous number of electrochemical biosensors based on this methods with developer solutions has been reported in the literature [48].

In another methodology, the seed approach is employed to deposit electrocatalysts. For the detection of an analyte, the transducer surface is incubated in a solution containing a high concentration of substrate which can be electrochemically oxidized or reduced by the electrocatalyst. The concentration of the deposited electrocatalyst can be indirectly measured on the basis of the catalytic current resulting from electrocatalysis of a substrate [72,73]. Since a clean surface can provide efficient binding of substrates, the catalytic properties of these particle-modified electrode surfaces is expected to be very high. In this concept, Pt has been shown to bean excellent catalyst for electrocatalysis of a range of substrates such as hydrazine, protons and H<sub>2</sub>O<sub>2</sub>. Therefore Pt is an ideal for signal amplification in the design of electrochemical amperometric biosensors for the above described compounds based on this strategy, if a suitable developer solution is available [74,75]. However, the advantages of seed-mediated deposition of Pt remain relatively unexplored, due to the difficulty in finding a suitable and an appropriate developer solution. In this context, some efforts have been made to explored different developer solutions [76], but an ideal solution for signal amplification employing this strategy is still not available.

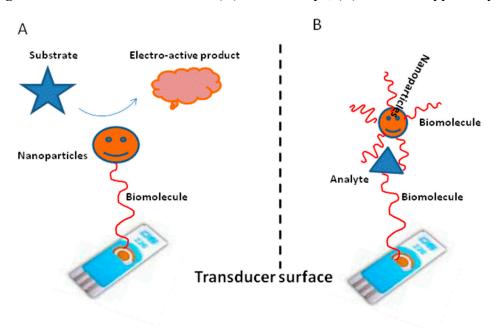
Nanomaterials have a tendency to strongly adsorb on different supports. Nanoparticles may carry negative or positive zeta potential values, making their surfaces negative or positive. The presence of such a charge further enhances the non-specific adsorption of nanomaterials due to electrostatic interactions. The non-specific adsorption of nanomaterials surface may impact the generated signals in three ways: (a) by blocking the surface and generating electron transfer resistance to the flow of current which may result in a shift of redox potentials to higher values, increasing the probability of interfering signals due to elevated redox potentials; (b) the charged surface of nanomaterials may increase or decrease the rate of flow of electrons depending on the charge; (c)the reactivity associated with the non-specifically adsorbed nanoparticles may also impact the obtained signal. Compared to nanomaterials, it is very easy and simply to control the non-specific adsorption of biomolecules through the use of different proteins. However, use of these blocking proteins is not so fruitful in the case of nanomaterials. Future research in the field may focus on overcoming the problem of non-specific adsorption of nanomaterials on the transducer surface. Similarly, the other big challenge with the re-usability of nanomaterials may hinder their application in amperometric biosensors, and consequently future research may focus on this issue to solve the problem. However, based on the intrinsic physical and chemical properties of nanomaterials, it is expected that they will provide building blocks for the construction of amperometric biosensors to improve the analytical performance of the system.

# 2.3. Nanomaterials as Signal Generating Probes

Different approaches have been employed to obtain the electrochemical output signal in the construction of biosensors (Figure 4). Most conventional approaches involve the use of labels such as enzymes, electroactive molecules, redox complexes and metal ions. The emergence of nanotechnology has opened new horizons for the use of nanomaterial labels to obtain the electrochemical signal in the

biosensors. The power and scope of such nanomaterials can be greatly enhanced by modifying the target analyte or biomolecules with these particles. Biomolecules labeled with nanomaterials retain their bioactivity and indentify their counterparts. The enormous signal enhancement associated with the use of nanomaterial labels provides the basis of ultra-sensitive affinity-based electrochemical biosensors. The strategies which utilize nanoparticles as labels can be sub-classified into two main groups: (a) nanomaterials that increase the loading of electroactive species; (b) nanoparticles that act as ultramicroelectrode arrays for the electrolysis of large amounts of substrates. Among the many kind of nanomaterials, gold nanoparticles are the most frequently used labels in electrochemical biosensors. Semiconductor and oxide nanoparticles are also available as labels to replace the commonly used enzyme labels in bioaffinity biosensors. Immuno- and aptasensors employing gold nanoparticles as labels were extensively reported for diverse analytical applications in the literature in recent years [77–79]. Besides gold nanoparticles, silver nanoparticles are also widely used as labels in the construction of both immuno- and aptasensors [80,81]. Similarly, DNA probes are modified with different types of semiconductor nanoparticles such as ZnS, CdS and PbS in the fabrication of electrochemical DNA-based aptasensors [82–84]. Some of the above described nanomaterials have been only demonstrated with voltammetry detection methods, but they may find potential applications in the amperometric detection in the future.

Figure 4. Nanomaterials as labels: (A) Direct assays; (B) Sandwich type assays.



Nanoparticles are composed of thousands of electrochemically active atoms. When biomolecules/analytes are labeled with nanoparticles, the loading of electroactive species on the electrode surface increases, and subsequently the sensitivity of the system is improved. Nanoparticles of gold, ZnS, CdS, PbS and silver have been explored in this regard [85–89]. However, many nanoparticles are electrochemically inactive within the potential range of aqueous electrolyte media or have redox potentials close to the solvent potential limit where the background signal may interfere. This drawback is associated with the intrinsic stability of nanoparticles in addition to the presence of stabilizer agents on nanosurfaces. Consequently, this drawback makes it difficult to detect nanoparticle labels directly

using electrochemical methods. To overcome this problem, nanoparticles are dissolved in solution using a strong oxidizing agent and subsequently electrochemical measurements are performed [90]. Since the concentration of metallic ions is proportional to the amount of nanoparticles, hence it can be related to the concentration of biomolecules in the sample solution. Alternatively, in order to avoid the need for an additional step to dissolve nanoparticle labels for their detection, another strategy is to explore the catalytic property of nanoparticles and monitor the amount of nanoparticles on the electrode surface indirectly by the process they catalyze. In this approach, after performing the labeling process, biosensors are incubated with the solution of nanoparticle electrochemically active substrates. Under these conditions, the biosensor acts like an ultramicroelectrode array [91]. This strategy works well only in case of nanoparticle labels that have very high catalytic activity, such as Pd, Au, Pt, Ag-Au alloy and Pt alloy [92-96]. Since most of the nanomaterials are highly resistant to electrooxidation, a direct detection strategy by employing catalytic properties is an attractive alternative. For example, Wang et al., used Au/Ag/Au core/double shell nanoparticles (Au/Ag/Au NPs) as novel enzyme-mimetic labels for the development of a sandwich-type electrochemical immunosensor for SCCA. Au NPs decorated mercapto-functionalized graphene sheets (Au@SH-GS) were used as platform for the immobilization of primary antibody (Ab1), while Au/Ag/Au NPs were employed as the secondary antibody (Ab2) label. Due to the excellent electrocatalytic activity of Au/Ag/Au NPs towards the reduction of hydrogen peroxide, the designed immunosensor displayed an excellent analytical performance with good reproducibility, high selectivity and stability [97]. Similarly, Lu et al., reported carbon nanotube/manganese dioxide (CNT-MnO2) composites as electrochemical tags for non-enzymatic signal amplification in immunosensing. The synthesized CNT-MnO<sub>2</sub> composites showed good electrochemical activity, electrical conductivity and stability in a hydrogen peroxide environment. The electrochemical immunosensor demonstrated a linear response on a log-scale for AFP [98]. Li et al., developed a novel sandwich-type nonenzymatic electrochemical immunosensor for the cancer biomarker carbohydrate antigen 15-3. Hierarchical nanoporous PtFe alloys were used as carriers of the signaling antibody anti-CA15-3 in order to achieve signal amplification. The electrochemical signal of the immunosensor was based on the high electrocatalytic activity of PtFe alloy for the electro-oxidation of H<sub>2</sub>O<sub>2</sub>. Theimmunosensor showed good accuracy, stability and fabrication reproducibility [99]. Despite the many advancements, many nanopaticles having catalytic properties are yet to be explored.

The analytical performance of labeled affinity sensors based on antibodies and aptamers is strongly dependent on the pH of the buffer, nature of the buffer, temperature and salt concentration in the buffer. Each affinity-based sensor has a specific optimized experimental condition range, and any change in the experimental conditions and binding medium may affect the characteristics of biosensors. On the other hand, nanomaterials are also very sensitive to the dispersion medium and pH. Nanomaterials may aggregate or agglomerate at any given pH, and the presence of salts is also known to aggregate the nanoparticles. For example, silver and gold nanoparticles aggregate in the presence of salts of sodium, potassium and manganese. Phosphate buffer is the commonly used medium to perform the affinity based electrochemical biosensors. Researchers have also shown the adsorption of phosphate ions on the surface of nanomaterials. It is always very hard to find optimal experimental conditions and medium where both affinity interaction and nanomaterials retain their functionalities. Future research in this context may focus on the synthesis of nanomaterials that can retain the intrinsic properties under any experimental conditions, and their functions will be independent of the pH of the medium.

#### 2.4. Nanomaterials as Enzyme Mimics

Recently, nanomaterials have attracted tremendous interest in enzyme mimetic research because of their several distinct properties, such as high surface area to volume ratio, abundance of reactive groups on their surface, unique optical properties and fascinating catalytic activity. To date, catalase-, oxidase-, peroxidase- or oxidase-like activities have been demonstrated for various types of nanomaterials [100]. The most commonly investigated nanomaterials are gold nanoparticles, Fe<sub>3</sub>O<sub>4</sub> nanoparticles, sheet like-FeS nanostructures, single walled carbon nanotubes, graphene oxide, AgM nanoparticles, nanoceria and metallic nanocomposites [101]. These nanomaterial-based enzyme mimetics offer the advantages of low cost, high stability and sustained catalytic activities. As a consequence, biosensors based on these nanomaterials have been extensively employed for many applications in the bioassay, biotechnology and biomedical field. For example, many electrochemical biosensors based on the enzyme-like activity of silver, gold, graphene oxide and Fe<sub>3</sub>O<sub>4</sub> for electrochemical detection of hydrogen peroxide, glucose and many other analytes have been reported in the literature [59,102–104]. However, nanomaterials do not have not real enzyme-like properties, as it is very difficult to regenerate the nanomaterial surface for subsequent measurement, limiting their applications in amperometric biosensors. Moreover, controlling the reactivity of nanomaterials against certain interfering molecules is a very difficult task. The reactivity of nanomaterials mainly relies on the functional groups of the analytes, and closely related interfering molecules share a very similar structure to the focus analyte. This reactivity may result in generation of signals even in the absence of analyte and produce false positive results. To replace enzymes for biosensing applications, it is highly desirable to design selective and specific nanomaterials to overcome the matrix interferences.

#### 2.5. Composite Nanomaterials in Amperometric Biosensors

The nanomaterial hybrids made by combining nanostructures (nanoparticles, nanorods) exhibit properties of both materials, *i.e.*, the electronic structure and characteristics of the individual nanomaterials [105]. With appropriate design, these conjugates can be employed in electrochemical biosensors. Recently, such hybrid and composite nanomaterials have attracted great attraction in the field of electrochemical biosensors, along with other domains. These hybrid structures can serve as immobilization supports, signal amplifiers and electrochemical signal generating probe. For examples, Iron-silica composites have been used for biomolecule immobilization due to their better biocompatibility [106]. Other commonly used nanocomposites for immobilization purposes include magnetic nanoparticles-silica, Fe<sub>3</sub>O<sub>4</sub>-chitosan, PAni/PVA-AgNP and Pt/carbon nanotubes [107–110]. Similarly, composites of silver and gold nanoparticles have been explored to amplify the electrochemical signal as well as electrochemical signal generating probes in affinity-based sensors [111]. Pt and ZnOhave the potential to be used in combination with various types of nanomaterials to enhance the performance of amperometric biosensors [112]. It is anticipated that the combination of diverse properties of nanomaterials into a single composite structure will open new gateways for research in many fields

**Table 1.** Some of the examples of nanomaterials functions in electrochemical biosensors.

Sr No	Functions	Nanomaterial Use	Type of Biosensor	Ref.
1	Immobilization support	iron oxide-chitosan nanocomposite	Electrochemical enzymatic biosensor	[113]
2	Immobilization support	Gold nanoparticles and graphene	Electrochemical DNA biosensor	[114]
3	Immobilization support	Gold nanoparticles/polyaniline nanofibers	Electrochemical immunosensor	[115]
4	Immobilization support	Graphene nanoplatelet–titanate nanotube composite	Electrochemical enzymatic biosensor	[116]
5	Immobilization support	Graphene oxide and silver nanoparticles	Electrochemical enzymatic biosensor	[117]
6	Signal amplification	Silver nanoparticles	Electrochemical DNA biosensor	[118]
7	Signal amplification	Platinum nanotubes modified with polyamidoamine	Electrochemical DNA biosensor	[119]
8	Signal amplification	Gold nanoparticles	Electrochemical immunosensor	[120]
9	Signal amplification	Ferroferric oxide nanoparticle	Electrochemical immunosensor	[121]
10	Signal amplification	Graphene, platinum nanoparticles	Electrochemical immunosensor	[122]
11	Signal generating probe	Cu@Ag (Cu@Ag-CD) core–shell nanoparticles	Electrochemical immunosensor	[123]
12	Signal generating probe	Magnetic beads	Electrochemical immunosensor	[124]
13	Signal generating probe/mediator	PtCo Alloy and graphene	Electrochemical immunosensor	[125]
14	Improve electrical properties	Carboxygraphene	Enzymatic biosensors	[126]
15	Improve electrical properties	Nano gold/graphene	Electrochemical myoglobin biosensor	[127]
16	Enzyme mimics	Graphene oxide (ERGO)–silver nanoparticle	Hydrogen per oxide sensor	[128]
17	Enzyme mimics	$MnO_2$	Hydrogen peroxide sensor	[129]

#### 3. Conclusions

Nanomaterial-based amperometric biosensors present a novel and attractive paradigm in terms of new and augmented functionality that encompasses a wide variety of application in the analytical domain. This review provided a brief survey of the enormous variety of functions of nanomaterials, which are attributed to the unique physiochemical properties of the constituent nanomaterials, that could find potential applications in amperometric biosensors. Table 1 presents some of the examples of nanomaterial functionalities in electrochemical biosensors. Amperometric biosensors based on nanomaterials may offer various advantages for enhancing and superseding the capabilities of current analytical methodology by permitting rapid and highly accurate analysis. Despite much progress, this field is still new, and there are many points in the development of nanomaterial-based amperometric biosensors to be addressed such as: (a) many complex biological systems require specific physiological environments and a certain degree of biocompatibility, and the biosensor-integrated nanomaterial must

fulfill this requirement; (b) it is highly desirable to find nanomaterials with sufficient binding sites for biomolecules; (c) the possibility to controllably tune the nanomaterials' properties for simultaneous analysis of various anlytes is yet to be explored; (d) although there are reports on the joint use of nanomaterials, most of nanomaterial combinations still need to be explored; and (e) the problem of nanomaterial aggregation in complex media should be resolved. Future research may focus on improving the above described parameters to further improve the analytical characteristic of nanomaterial-based amperometric biosensors.

# Acknowledgments

Akhtar HAYAT acknowledge collaborative research group of IAMGES to support his short research stay in France.

#### **Author Contributions**

Jean Louis Marty provided the main theme of presented work, while Akhtar Hayat described the main part of work under the direction of Jean Louis Marty. Gaëlle Catanante helped to design schematics, tables and as well supported in the literature survey

#### **Conflicts of Interest**

The authors declare no conflict of interest.

## References

- 1. Grieshaber, D.; MacKenzie, R.; Vörös, J.; Reimhult, E. Electrochemical biosensors—Sensor principles and architectures. *Sensors* **2008**, *8*, 1400–1458.
- 2. Hayat, A.; Marty, J.L. Aptamer based electrochemical sensors for emerging environmental pollutants. *Front.Chem.* **2014**, *2*, doi:10.3389/fchem.2014.00041.
- 3. Luz, R.S.; Iost, R.; Crespilho, F. Nanomaterials for biosensors and implantable biodevices. In *Nanobioelectrochemistry*; Crespilho, F.N., Ed.; Springer: New York, NY, USA, 2013; pp. 27–48.
- 4. Meng, Y.; Aldous, L.; Belding, S.R.; Compton, R.G. The formal potentials and electrode kinetics of the proton/hydrogen couple in various room temperature ionic liquids. *Chem. Commun.* **2012**, *48*, 5572–5574.
- 5. Hayat, A.; Marty, J.L. Disposable screen printed electrochemical sensors: Tools for environmental monitoring. *Sensors* **2014**, *14*, 10432–10453.
- 6. Hayat, A.; Yang, C.; Rhouati, A.; Marty, J.L. Recent advances and achievements in nanomaterial-based, and structure switchable aptasensing platforms for ochratoxin a detection. *Sensors* **2013**, *13*, 15187–15208.
- 7. Dequaire, M.; Degrand, C.; Limoges, B. An electrochemical metalloimmunoassay based on a colloidal gold label. *Anal. Chem.* **2000**, *72*, 5521–5528.
- 8. Chen, A.; Chatterjee, S. Nanomaterials based electrochemical sensors for biomedical applications. *Chem. Soc. Rev.* **2013**, *42*, 5425–5438.

9. Li, H.; Liu, S.; Dai, Z.; Bao, J.; Yang, X. Applications of nanomaterials in electrochemical enzyme biosensors. *Sensors* **2009**, *9*, 8547–8561.

- 10. Song, S.; Qin, Y.; He, Y.; Huang, Q.; Fan, C.; Chen, H.-Y. Functional nanoprobes for ultrasensitive detection of biomolecules. *Chem. Soc. Rev.* **2010**, *39*, 4234–4243.
- 11. Iost, R.M.; da Silva, W.C.; Madurro, J.M.; Madurro, A.G.; Ferreira, L.F.; Crespilho, F.N. Recent advances in nano-based electrochemical biosensors: Application in diagnosis and monitoring of diseases. *Front. Biosci. (Elite Ed.)* **2011**, *3*, 663–689.
- 12. Decher, G. Fuzzy nanoassemblies towards layer polymeric multicomposites. *Science* **1997**, *277*, 1232–1237.
- 13. Bricaud, Q.; Fabre, R.M.; Brookins, R.N.; Schanze, K.S.; Reynolds, J.R. Energy transfer between conjugated polyelectrolytes in layer-by-layer assembled films. *Langmuir* **2011**, *27*, 5021–5028.
- 14. Alencar, W.S.; Crespilho, F.N.; Santos, M.R.M.C.; Zucolotto, V.; Oliveira, O.N.; Silva, W.C. Influence of film architecture on the charge-transfer reactions of metallophthalocyanine layer-by-layer films. *J. Phys. Chem. C* **2007**, *111*, 12817–12821.
- 15. De Sousa Luz, R.A.; Martins, M.V.A.; Magalhães, J.L.; Siqueira, J.R., Jr.; Zucolotto, V.; Oliveira, O.N., Jr.; Crespilho, F.N.; da Silva, W.C. Supramolecular architectures in layer-by-layer films of single-walled carbon nanotubes, chitosan and cobalt (ii) phthalocyanine. *Mater. Chem. Phys.* **2011**, *130*, 1072–1077.
- Alencar, W.S.; Crespilho, F.N.; Martins, M.V.A.; Zucolotto, V.; Oliveira, J.O.N.; Silva, W.C. Synergistic interaction between gold nanoparticles and nickel phthalocyanine in layer-by-layer (lbl) films: Evidence of constitutional dynamic chemistry (cdc). *Phys. Chem. Chem. Phys.* 2009, 11, 5086–5091.
- 17. Lei, J.; Ju, H. Signal amplification using functional nanomaterials for biosensing. *Chem. Soc. Rev.* **2012**, *41*, 2122–2134.
- 18. Pandiaraj, M.; Madasamy, T.; Gollavilli, P.N.; Balamurugan, M.; Kotamraju, S.; Rao, V.K.; Bhargava, K.; Karunakaran, C. Nanomaterial-based electrochemical biosensors for cytochrome c using cytochrome c reductase. *Bioelectrochemistry* **2013**, *91*, 1–7.
- 19. Wang, J. Nanomaterial-based electrochemical biosensors. *Analyst* **2005**, *130*, 421–426.
- 20. Kerman, K.; Saito, M.; Tamiya, E.; Yamamura, S.; Takamura, Y. Nanomaterial-based electrochemical biosensors for medical applications. *TrAC Trends Anal. Chem.* **2008**, *27*, 585–592.
- 21. Putzbach, W.; Ronkainen, N.J. Immobilization techniques in the fabrication of nanomaterial-based electrochemical biosensors: A review. *Sensors* **2013**, *13*, 4811–4840.
- 22. Ansari, S.A.; Husain, Q. Potential applications of enzymes immobilized on/in nano materials: A review. *Biotechnol. Adv.* **2012**, *30*, 512–523.
- 23. Jia, H.; Zhu, G.; Wang, P. Catalytic behaviors of enzymes attached to nanoparticles: The effect of particle mobility. *Biotechnol. Bioeng.* **2003**, *84*, 406–414.
- 24. Ali, M.; Winterer, M. Zno nanocrystals: Surprisingly 'alive'. Chem. Mater. 2009, 22, 85–91.
- 25. Ni,Y.;Cao, X.;Wu, G.; Hu,G.; Yang,Z.; Wei, X. Preparation, characterization and property study of zinc oxide nanoparticles via a simple solution-combusting method. *Nanotechnology* **2007**, *18*, 155–161.

26. Hayat, A.; Barthelmebs, L.; Marty, J.-L. Enzyme-linked immunosensor based on super paramagnetic nanobeads for easy and rapid detection of okadaic acid. *Anal. Chim. Acta* **2011**, *690*, 248–252.

- 27. Sassolas, A.; Hayat, A.; Marty, J.-L. Immobilization of enzymes on magnetic beads through affinity interactions. In *Immobilization of Enzymes and Cells*; Humana Press: New York, NY, USA, 2013; pp. 139–148.
- 28. Pinacho, D.; Sánchez-Baeza, F.; Pividori, M.-I.; Marco, M.-P. Electrochemical detection of fluoroquinolone antibiotics in milk using a magneto immunosensor. *Sensors* **2014**, *14*, 15965–15980.
- 29. Martin, M.; Salazar, P.; Villalonga, R.; Campuzano, S.; Pingarron, J.M.; Gonzalez-Mora, J.L. Preparation of core-shell Fe<sub>3</sub>O<sub>4</sub>@poly(dopamine) magnetic nanoparticles for biosensor construction. *J. Mater. Chem. B* **2014**, *2*, 739–746.
- 30. Yang, C.-H.; Kuo, L.-S.; Chen, P.-H.; Yang, C.-R.; Tsai, Z.-M. Development of a multilayered polymeric DNA biosensor using radio frequency technology with gold and magnetic nanoparticles. *Biosens. Bioelectron.* **2012**, *31*, 349–356.
- 31. Rhouati, A.; Hayat, A.; Hernandez, D.B.; Meraihi, Z.; Munoz, R.; Marty, J.-L. Development of an automated flow-based electrochemical aptasensor for on-line detection of ochratoxin a. *Sens. Actuators B Chem.* **2013**, *176*, 1160–1166.
- 32. Dominguez, R.B.; Hayat, A.; Sassolas, A.; Alonso, G.A.; Munoz, R.; Marty, J.-L. Automated flow-through amperometric immunosensor for highly sensitive and on-line detection of okadaic acid in mussel sample. *Talanta* **2012**, *99*, 232–237.
- 33. Barthelmebs, L.; Hayat, A.; Limiadi, A.W.; Marty, J.-L.; Noguer, T. Electrochemical DNA aptamer-based biosensor for OTA detection, using superparamagnetic nanoparticles. *Sens. Actuators B Chem.* **2011**, *156*, 932–937.
- 34. Hayat, A.; Barthelmebs, L.; Sassolas, A.; Marty, J.-L. Development of a novel label-free amperometric immunosensor for the detection of okadaic acid. *Anal. Chim. Acta* **2012**, *724*, 92–97.
- 35. Zhuo, Y.; Yuan, P.-X.; Yuan, R.; Chai, Y.-Q.; Hong, C.-L. Bienzyme functionalized three-layer composite magnetic nanoparticles for electrochemical immunosensors. *Biomaterials* **2009**, *30*, 2284–2290.
- 36. Liu, Z.; Liu, Y.; Yang, H.; Yang, Y.; Shen, G.; Yu, R. A phenol biosensor based on immobilizing tyrosinase to modified core-shell magnetic nanoparticles supported at a carbon paste electrode. *Anal. Chim. Acta* **2005**, *533*, 3–9.
- 37. Zheng, B.; Xie, S.; Qian, L.; Yuan, H.; Xiao, D.; Choi, M.M.F. Gold nanoparticles-coated eggshell membrane with immobilized glucose oxidase for fabrication of glucose biosensor. *Sens. Actuators B Chem.* **2011**, *152*, 49–55.
- 38. Wang, L.; Wen, W.; Xiong, H.; Zhang, X.; Gu, H.; Wang, S. A novel amperometric biosensor for superoxide anion based on superoxide dismutase immobilized on gold nanoparticle-chitosan-ionic liquid biocomposite film. *Anal. Chim. Acta* **2013**, *758*, 66–71.
- 39. Florea, A.; Taleat, Z.; Cristea, C.; Mazloum-Ardakani, M.; Săndulescu, R. Label free mucl aptasensors based on electrodeposition of gold nanoparticles on screen printed electrodes. *Electrochem. Commun.* **2013**, *33*, 127–130.

40. Ho, K.M.; Mao, X.; Gu, L.; Li, P. Facile route to enzyme immobilization: Core-shell nanoenzyme particles consisting of well-defined poly(methyl methacrylate) cores and cellulase shells. *Langmuir* **2008**, *24*, 11036–11042.

- 41. Rawal, R.; Chawla, S.; Pundir, C.S. Polyphenol biosensor based on laccase immobilized onto silver nanoparticles/multiwalled carbon nanotube/polyaniline gold electrode. *Anal. Biochem.* **2011**, *419*, 196–204.
- 42. Li, H.; He, J.; Zhao, Y.; Wu, D.; Cai, Y.; Wei, Q.; Yang, M. Immobilization of glucose oxidase and platinum on mesoporous silica nanoparticles for the fabrication of glucose biosensor. *Electrochim. Acta* **2011**, *56*, 2960–2965.
- 43. Devi, R.; Thakur, M.; Pundir, C.S. Construction and application of an amperometric xanthine biosensor based on zinc oxide nanoparticles—polypyrrole composite film. *Biosens. Bioelectron.* **2011**, *26*, 3420–3426.
- 44. MansouriMajd, S.; Teymourian, H.; Salimi, A.; Hallaj, R. Fabrication of electrochemical theophylline sensor based on manganese oxide nanoparticles/ionic liquid/chitosan nanocomposite modified glassy carbon electrode. *Electrochim. Acta* **2013**, *108*, 707–716.
- 45. Lu, W.; Cao, X.; Tao, L.; Ge, J.; Dong, J.; Qian, W. A novel label-free amperometric immunosensor for carcinoembryonic antigen based on ag nanoparticle decorated infinite coordination polymer fibres. *Biosens. Bioelectron.* **2014**, *57*, 219–225.
- 46. Turkmen, E.; Bas, S.Z.; Gulce, H.; Yildiz, S. Glucose biosensor based on immobilization of glucose oxidase in electropolymerized poly(o-phenylenediamine) film on platinum nanoparticles-polyvinylferrocenium modified electrode. *Electrochim. Acta* **2014**, *123*, 93–102.
- 47. Gao, L.; Zhuang, J.; Nie, L.; Zhang, J.; Zhang, Y.; Gu, N.; Wang, T.; Feng, J.; Yang, D.; Perrett, S.; *et al.* Intrinsic peroxidase-like activity of ferromagnetic nanoparticles. *Nat. Nanotechnol.* **2007**, *2*, 577–583.
- 48. Ding, L.; Bond, A.M.; Zhai, J.; Zhang, J. Utilization of nanoparticle labels for signal amplification in ultrasensitive electrochemical affinity biosensors: A review. *Anal. Chim. Acta* **2013**, *797*, 1–12.
- 49. Hayat, A.; Andreescu, S. Nanoceria particles as catalytic amplifiers for alkaline phosphatase assays. *Anal. Chem.* **2013**, *85*, 10028–10032.
- 50. Song, Y.; Qu, K.; Zhao, C.; Ren, J.; Qu, X. Graphene oxide: Intrinsic peroxidase catalytic activity and its application to glucose detection. *Adv. Mater.* **2010**, *22*, 2206–2210.
- 51. Liu, Y.; Yu, D.; Zeng, C.; Miao, Z.; Dai, L. Biocompatible graphene oxide-based glucose biosensors. *Langmuir* **2010**, *26*, 6158–6160.
- 52. Polsky, R.; Gill, R.; Kaganovsky, L.; Willner, I. Nucleic acid-functionalized Pt nanoparticles: Catalytic labels for the amplified electrochemical detection of biomolecules. *Anal. Chem.* **2006**, 78, 2268–2271.
- 53. Kajita, M.; Hikosaka, K.; Iitsuka, M.; Kanayama, A.; Toshima, N.; Miyamoto, Y. Platinum nanoparticle is a useful scavenger of superoxide anion and hydrogen peroxide. *Free Radic. Res.* **2007**, *41*, 615–626.
- 54. You, T.; Niwa, O.; Chen, Z.; Hayashi, K.; Tomita, M.; Hirono, S. An amperometric detector formed of highly dispersed ni nanoparticles embedded in a graphite-like carbon film electrode for sugar determination. *Anal. Chem.* **2003**, *75*, 5191–5196.

55. Heli, H.; Hajjizadeh, M.; Jabbari, A.; Moosavi-Movahedi, A.A. Fine steps of electrocatalytic oxidation and sensitive detection of some amino acids on copper nanoparticles. *Anal. Biochem.* **2009**, *388*, 81–90.

- 56. Xu, J.-Z.; Zhu, J.-J.; Wang, H.; Chen, H.-Y. Nano-sized copper oxide modified carbon paste electrodes as an amperometric sensor for amikacin. *Anal. Lett.* **2003**, *36*, 2723–2733.
- 57. Özel, R.E.; Ispas, C.; Ganesana, M.; Leiter, J.C.; Andreescu, S. Glutamate oxidase biosensor based on mixed ceria and titania nanoparticles for the detection of glutamate in hypoxic environments. *Biosens. Bioelectron.* **2014**, *52*, 397–402.
- 58. Lin, J.; He, C.; Zhao, Y.; Zhang, S. One-step synthesis of silver nanoparticles/carbon nanotubes/chitosan film and its application in glucose biosensor. *Sens. Actuators B Chem.* **2009**, 137, 768–773.
- 59. Jv, Y.; Li, B.; Cao, R. Positively-charged gold nanoparticles as peroxidiase mimic and their application in hydrogen peroxide and glucose detection. *Chem. Commun.* **2010**, *46*, 8017–8019.
- 60. Wang, J. Nanomaterial-based amplified transduction of biomolecular interactions. *Small* **2005**, *I*, 1036–1043.
- 61. Yang, P.; Li, X.; Wang, L.; Wu, Q.; Chen, Z.; Lin, X. Sandwich-type amperometric immunosensor for cancer biomarker based on signal amplification strategy of multiple enzyme-linked antibodies as probes modified with carbon nanotubes and concanavalin a. *J. Electroanal. Chem.* **2014**, *732*, 38–45.
- 62. Liu, S.; Ju, H. Reagentless glucose biosensor based on direct electron transfer of glucose oxidase immobilized on colloidal gold modified carbon paste electrode. *Biosens. Bioelectron.* **2003**, *19*, 177–183.
- 63. Xiao, Y.; Patolsky, F.; Katz, E.; Hainfeld, J.F.; Willner, I. "Plugging into enzymes": Nanowiring of redox enzymes by a gold nanoparticle. *Science* **2003**, *299*, 1877–1881.
- 64. Ghalkhani, M.; Shahrokhian, S.; Ghorbani-Bidkorbeh, F. Voltammetric studies of sumatriptan on the surface of pyrolytic graphite electrode modified with multi-walled carbon nanotubes decorated with silver nanoparticles. *Talanta* **2009**, *80*, 31–38.
- 65. Jiménez, A.; Armada, M.P.G.; Losada, J.; Villena, C.; Alonso, B.; Casado, C.M. Amperometric biosensors for nadh based on hyperbranched dendritic ferrocene polymers and Pt nanoparticles. *Sens. Actuators B Chem.* **2014**, *190*, 111–119.
- 66. Wang, J.; Xu, D.; Kawde, A.-N.; Polsky, R. Metal nanoparticle-based electrochemical stripping potentiometric detection of DNA hybridization. *Anal. Chem.* **2001**, *73*, 5576–5581.
- 67. Guo, S.; Wang, E. Synthesis and electrochemical applications of gold nanoparticles. *Anal. Chim. Acta* **2007**, *598*, 181–192.
- 68. Kienberger, F.; Kada, G.; Mueller, H.; Hinterdorfer, P. Single molecule studies of antibody–antigen interaction strength versus intra-molecular antigen stability. *J. Mol. Biol.* **2005**, *347*, 597–606.
- 69. Lai, G.; Yan, F.; Wu, J.; Leng, C.; Ju, H. Ultrasensitive multiplexed immunoassay with electrochemical stripping analysis of silver nanoparticles catalytically deposited by gold nanoparticles and enzymatic reaction. *Anal. Chem.* **2011**, *83*, 2726–2732.

70. Dungchai, W.; Siangproh, W.; Chaicumpa, W.; Tongtawe, P.; Chailapakul, O. Salmonella typhi determination using voltammetric amplification of nanoparticles: A highly sensitive strategy for metalloimmunoassay based on a copper-enhanced gold label. *Talanta* **2008**, *77*, 727–732.

- 71. Tan, E.; Yin, P.; Lang, X.; Wang, X.; You, T.; Guo, L. Functionalized gold nanoparticles as nanosensor for sensitive and selective detection of silver ions and silver nanoparticles by surface-enhanced raman scattering. *Analyst* **2012**, *137*, 3925–3928.
- 72. Pei, X.; Zhang, B.; Tang, J.; Liu, B.; Lai, W.; Tang, D. Sandwich-type immunosensors and immunoassays exploiting nanostructure labels: A review. *Anal. Chim. Acta* **2013**, *758*, 1–18.
- 73. Omidfar, K.; Khorsand, F.; Darziani Azizi, M. New analytical applications of gold nanoparticles as label in antibody based sensors. *Biosens. Bioelectron.* **2013**, *43*, 336–347.
- 74. Das, R.S.; Singh, B.; Banerjee, R.; Mukhopadhyay, S. Pvp stabilized Pt nano particles catalyzed de-oxygenation of phenoxazine group by hydrazine in physiological buffer media: Surfactant competes with reactants for the same surface sites. *Dalton Trans.* **2013**, *42*, 4068–4080.
- 75. Xu, Z.; Zhang, H.; Zhong, H.; Lu, Q.; Wang, Y.; Su, D. Effect of particle size on the activity and durability of the Pt/C electrocatalyst for proton exchange membrane fuel cells. *Appl. Catal. B Environ.* **2012**, *111–112*, 264–270.
- 76. Zhang, J.; Ting, B.P.; Khan, M.; Pearce, M.C.; Yang, Y.; Gao, Z.; Ying, J.Y. Pt nanoparticle label-mediated deposition of Pt catalyst for ultrasensitive electrochemical immunosensors. *Biosens. Bioelectron.* **2010**, *26*, 418–423.
- 77. Tang, H.; Chen, J.; Nie, L.; Kuang, Y.; Yao, S. A label-free electrochemical immunoassay for carcinoembryonic antigen (cea) based on gold nanoparticles (aunps) and nonconductive polymer film. *Biosens. Bioelectron.* **2007**, *22*, 1061–1067.
- 78. Ozsoz, M.; Erdem, A.; Kerman, K.; Ozkan, D.; Tugrul, B.; Topcuoglu, N.; Ekren, H.; Taylan, M. Electrochemical genosensor based on colloidal gold nanoparticles for the detection of factor v leiden mutation using disposable pencil graphite electrodes. *Anal. Chem.* **2003**, *75*, 2181–2187.
- 79. Das, J.; Aziz, M.A.; Yang, H. A nanocatalyst-based assay for proteins: DNA-free ultrasensitive electrochemical detection using catalytic reduction of p-nitrophenol by gold-nanoparticle labels. *J. Am. Chem. Soc.* **2006**, *128*, 16022–16023.
- 80. Cai, H.; Wang, Y.; He, P.; Fang, Y. Electrochemical detection of DNA hybridization based on silver-enhanced gold nanoparticle label. *Anal. Chim. Acta* **2002**, *469*, 165–172.
- 81. Ting, B.P.; Zhang, J.; Gao, Z.; Ying, J.Y. A DNA biosensor based on the detection of doxorubicin-conjugated Ag nanoparticle labels using solid-state voltammetry. *Biosens. Bioelectron.* **2009**, *25*, 282–287.
- 82. Feng, L.-N.; Bian, Z.-P.; Peng, J.; Jiang, F.; Yang, G.-H.; Zhu, Y.-D.; Yang, D.; Jiang, L.-P.; Zhu, J.-J. Ultrasensitive multianalyte electrochemical immunoassay based on metal ion functionalized titanium phosphate nanospheres. *Anal. Chem.* **2012**, *84*, 7810–7815.
- 83. Feng, L.-N.; Peng, J.; Zhu, Y.-D.; Jiang, L.-P.; Zhu, J.-J. Synthesis of Cd<sup>2+</sup>-functionalized titanium phosphate nanoparticles and application as labels for electrochemical immunoassays. *Chem. Commun.* **2012**, *48*, 4474–4476.
- 84. Gao, H.; Zhong, J.; Qin, P.; Lin, C.; Sun, W.; Jiao, K. Electrochemical DNA hybridization assay for the fmv 35s gene sequence using pbs nanoparticles as a label. *Microchim. Acta* **2009**, *165*, 173–178.

85. Zhugesi Formation. In *Geological Formation Names of China (1866–2000)*; Zhang, S., Ed.; Springer: New York, NY, USA, 2009; pp. 1414–1414.

- 86. Omidfar, K.; Zarei, H.; Gholizadeh, F.; Larijani, B. A high-sensitivity electrochemical immunosensor based on mobile crystalline material-41–polyvinyl alcohol nanocomposite and colloidal gold nanoparticles. *Anal. Biochem.* **2012**, *421*, 649–656.
- 87. Wang, J.; Liu, G.; Merkoçi, A. Electrochemical coding technology for simultaneous detection of multiple DNA targets. *J. Am. Chem. Soc.* **2003**, *125*, 3214–3215.
- 88. Numnuam, A.; Chumbimuni-Torres, K.Y.; Xiang, Y.; Bash, R.; Thavarungkul, P.; Kanatharana, P.; Pretsch, E.; Wang, J.; Bakker, E. Potentiometric detection of DNA hybridization. *J. Am. Chem. Soc.* **2007**, *130*, 410–411.
- 89. Zhu, N.; Zhang, A.; Wang, Q.; He, P.; Fang, Y. Lead sulfide nanoparticle as oligonucleotides labels for electrochemical stripping detection of DNA hybridization. *Electroanalysis* **2004**, *16*, 577–582.
- 90. Zhou, N.; Li, J.; Chen, H.; Liao, C.; Chen, L. A functional graphene oxide-ionic liquid composites-gold nanoparticle sensing platform for ultrasensitive electrochemical detection of Hg<sup>2+</sup>. *Analyst* **2013**, *138*, 1091–1097.
- 91. Chazalviel, J.-N.; Allongue, P. On the origin of the efficient nanoparticle mediated electron transfer across a self-assembled monolayer. *J. Am. Chem. Soc.* **2010**, *133*, 762–764.
- 92. Leng, C.; Wu, J.; Xu, Q.; Lai, G.; Ju, H.; Yan, F. A highly sensitive disposable immunosensor through direct electro-reduction of oxygen catalyzed by palladium nanoparticle decorated carbon nanotube label. *Biosens.Bioelectron.***2011**, *27*, 71–76.
- 93. Lu, J.; Liu, Y.; Wang, X.; Wang, S.; Di, X. Development and validation of a rapid high-performance liquid chromatography-tandem mass spectrometry method for the determination of wj-38, a novel aldose reductase inhibitor, in rat plasma and its application to a pharmacokinetic study. *J. Chromatogr. B* **2012**, *893–894*, 29–33.
- 94. Dong, X.-Y.; Mi, X.-N.; Zhang, L.; Liang, T.-M.; Xu, J.-J.; Chen, H.-Y. Dnazyme-functionalized Pt nanoparticles/carbon nanotubes for amplified sandwich electrochemical DNA analysis. *Biosens. Bioelectron.* **2012**, *38*, 337–341.
- 95. Tang, J.; Tang, D.; Li, Q.; Su, B.; Qiu, B.; Chen, G. Sensitive electrochemical immunoassay of carcinoembryonic antigen with signal dual-amplification using glucose oxidase and an artificial catalase. *Anal. Chim. Acta* **2011**, *697*, 16–22.
- 96. Li, Y.; Yuan, R.; Chai, Y.; Song, Z. Electrodeposition of gold–platinum alloy nanoparticles on carbon nanotubes as electrochemical sensing interface for sensitive detection of tumor marker. *Electrochim. Acta* **2011**, *56*, 6715–6721.
- 97. Wang, Y.; Zhang, Y.; Su, Y.; Li, F.; Ma, H.; Li, H.; Du, B.; Wei, Q. Ultrasensitive non-mediator electrochemical immunosensors using Au/Ag/Au core/double shell nanoparticles as enzyme-mimetic labels. *Talanta* **2014**, *124*, 60–66.
- 98. Tu, M.-C.; Chen, H.-Y.; Wang, Y.; Moochhala, S.M.; Alagappan, P.; Liedberg, B. Immunosensor based on carbon nanotube/manganese dioxide electrochemical tags. *Anal. Chim. Acta.* **2015**, *853*, 228–233.
- 99. Li, Y.; Xu, C.; Li, H.; Wang, H.; Wu, D.; Ma, H.; Cai, Y.; Du, B.; Wei, Q. Nonenzymatic immunosensor for detection of carbohydrate antigen 15–3 based on hierarchical nanoporous PtFe alloy. *Biosens. Bioelectron.* **2014**, *56*, 295–299.

100. Xiao, F.; Zhao, F.; Mei, D.; Mo, Z.; Zeng, B. Nonenzymatic glucose sensor based on ultrasonic-electrodeposition of bimetallic ptm (m = Ru, Pd and Au) nanoparticles on carbon nanotubes—ionic liquid composite film. *Biosens. Bioelectron.* **2009**, *24*, 3481–3486.

- 101. Xie, J.; Zhang, X.; Wang, H.; Zheng, H.; Huang, Y. Analytical and environmental applications of nanoparticles as enzyme mimetics. *TrAC Trends Anal. Chem.* **2012**, *39*, 114–129.
- 102. He, W.; Wu, X.; Liu, J.; Hu, X.; Zhang, K.; Hou, S.; Zhou, W.; Xie, S. Design of agm bimetallic alloy nanostructures (m = Au, Pd, Pt) with tunable morphology and peroxidase-like activity. *Chem. Mater.* **2010**, *22*, 2988–2994.
- 103. Zhang, Z.; Zhu, H.; Wang, X.; Yang, X. Sensitive electrochemical sensor for hydrogen peroxide using Fe<sub>3</sub>O<sub>4</sub> magnetic nanoparticles as a mimic for peroxidase. *Microchim. Acta* **2011**, *174*, 183–189.
- 104. Zhang, S.; Tang, S.; Lei, J.; Dong, H.; Ju, H. Functionalization of graphene nanoribbons with porphyrin for electrocatalysis and amperometric biosensing. *J. Electroanal. Chem.* **2011**, *656*, 285–288.
- 105. Zhang, Y.; Zhang, K.; Ma, H. Electrochemical DNA biosensor based on silver nanoparticles/poly(3-(3-pyridyl) acrylic acid)/carbon nanotubes modified electrode. *Anal. Biochem.* **2009**, *387*, 13–19.
- 106. Kim, H.; Kwon, H.-S.; Ahn, J.; Lee, C.-H.; Ahn, I.-S. Evaluation of a silica-coated magnetic nanoparticle for the immobilization of a his-tagged lipase. *Biocatal. Biotransform.* **2009**, *27*, 246–253.
- 107. Besanger, T.R.; Chen, Y.; Deisingh, A.K.; Hodgson, R.; Jin, W.; Mayer, S.; Brook, M.A.; Brennan, J.D. Screening of inhibitors using enzymes entrapped in sol-gel-derived materials. *Anal. Chem.* **2003**, *75*, 2382–2391.
- 108. Pan, C.; Hu, B.; Li, W.; Sun, Y.; Ye, H.; Zeng, X. Novel and efficient method for immobilization and stabilization of β-d-galactosidase by covalent attachment onto magnetic Fe<sub>3</sub>O<sub>4</sub>–chitosan nanoparticles. *J. Mol. Catal. B Enzym.* **2009**, *61*, 208–215.
- 109. Crespilho, F.N.; Emilia Ghica, M.; Florescu, M.; Nart, F.C.; Oliveira, O.N., Jr; Brett, C.M.A. A strategy for enzyme immobilization on layer-by-layer dendrimer—gold nanoparticle electrocatalytic membrane incorporating redox mediator. *Electrochem. Commun.* **2006**, *8*, 1665–1670.
- 110. Yang, M.; Yang, Y.; Liu, Y.; Shen, G.; Yu, R. Platinum nanoparticles-doped sol-gel/carbon nanotubes composite electrochemical sensors and biosensors. *Biosens. Bioelectron.* **2006**, *21*, 1125–1131.
- 111. Wang, J.; Polsky, R.; Xu, D. Silver-enhanced colloidal gold electrochemical stripping detection of DNA hybridization. *Langmuir* **2001**, *17*, 5739–5741.
- 112. Ahmad, M.; Pan, C.; Gan, L.; Nawaz, Z.; Zhu, J. Highly sensitive amperometric cholesterol biosensor based on Pt-incorporated fullerene-like ZnO nanospheres. *J. Phys. Chem. C* **2009**, *114*, 243–250.
- 113. Jeyapragasam, T.; Saraswathi, R. Electrochemical biosensing of carbofuran based on acetylcholinesterase immobilized onto iron oxide–chitosan nanocomposite. *Sens. Actuators B Chem.* **2014**, *191*, 681–687.

114. Shi, A.; Wang, J.; Han, X.; Fang, X.; Zhang, Y. A sensitive electrochemical DNA biosensor based on gold nanomaterial and graphene amplified signal. *Sens. Actuators B Chem.* **2014**, *200*, 206–212.

- 115. Zhang, S.; Zhang, L.; Zhang, X.; Yang, P.; Cai, J. An efficient nanomaterial-based electrochemical biosensor for sensitive recognition of drug-resistant leukemia cells. *Analyst* **2014**, *139*, 3629–3635.
- 116. Liu, X.; Zhang, J.; Yan, R.; Zhang, Q.; Liu, X. Preparation of graphene nanoplatelet–titanate nanotube composite and its advantages over the two single components as biosensor immobilization materials. *Biosens. Bioelectron.* **2014**, *51*, 76–81.
- 117. Palanisamy, S.; Karuppiah, C.; Chen, S.M. Direct electrochemistry and electrocatalysis of glucose oxidase immobilized on reduced graphene oxide and silver nanoparticles nanocomposite modified electrode. *Colloids Surf. B Biointerfaces* **2014**, *114*, 164–169.
- 118. Song, W.; Li, H.; Liang, H.; Qiang, W.; Xu, D. Disposable electrochemical aptasensor array by using in situ DNA hybridization inducing silver nanoparticles aggregate for signal amplification. *Anal. Chem.* **2014**, *86*, 2775–2783.
- 119. Xu, W.; Wu, Y.; Yi, H.; Bai, L.; Chai, Y.; Yuan, R. Porous platinum nanotubes modified with dendrimers as nanocarriers and electrocatalysts for sensitive electrochemical aptasensors based on enzymatic signal amplification. *Chem. Commun.* **2014**, *50*, 1451–1453.
- 120. Xiong, P.; Gan, N.; Cui, H.; Zhou, J.; Cao, Y.; Hu, F.; Li, T. Incubation-free electrochemical immunoassay for diethylstilbestrol in milk using gold nanoparticle-antibody conjugates for signal amplification. *Microchim. Acta* **2014**, *181*, 453–462.
- 121. Wang, H.; Li, X.; Mao, K.; Li, Y.; Du, B.; Zhang, Y.; Wei, Q. Electrochemical immunosensor for α-fetoprotein detection using ferroferric oxide and horseradish peroxidase as signal amplification labels. *Anal. Biochem.* **2014**, *465*, 121–126.
- 122. Jia, X.; Chen, X.; Han, J.; Ma, J.; Ma, Z. Triple signal amplification using gold nanoparticles, bienzyme and platinum nanoparticles functionalized graphene as enhancers for simultaneous multiple electrochemical immunoassay. *Biosens. Bioelectron.* **2014**, *53*, 65–70.
- 123. Gao, J.; Guo, Z.; Su, F.; Gao, L.; Pang, X.; Cao, W.; Du, B.; Wei, Q. Ultrasensitive electrochemical immunoassay for cea through host–guest interaction of β-cyclodextrin functionalized graphene and Cu@Ag core–shell nanoparticles with adamantine-modified antibody. *Biosens. Bioelectron.* **2015**, *63*, 465–471.
- 124. Akter, R.; Kyun Rhee, C.; Aminur Rahman, M. Sensitivity enhancement of an electrochemical immunosensor through the electrocatalysis of magnetic bead-supported non-enzymatic labels. *Biosens. Bioelectron.* **2014**, *54*, 351–357.
- 125. Feng, R.; Zhang, Y.; Ma, H.; Wu, D.; Fan, H.; Wang, H.; Li, H.; Du, B.; Wei, Q. Ultrasensitive non-enzymatic and non-mediator electrochemical biosensor using nitrogen-doped graphene sheets for signal amplification and nanoporous alloy as carrier. *Electrochim. Acta* **2013**, *97*, 105–111.
- 126. Liang, B.; Fang, L.; Yang, G.; Hu, Y.; Guo, X.; Ye, X. Direct electron transfer glucose biosensor based on glucose oxidase self-assembled on electrochemically reduced carboxyl graphene. *Biosens. Bioelectron.* **2013**, *43*, 131–136.

127. Li, G.; Li, T.; Deng, Y.; Cheng, Y.; Shi, F.; Sun, W.; Sun, Z. Electrodeposited nanogold decorated graphene modified carbon ionic liquid electrode for the electrochemical myoglobin biosensor. *J. Solid State Electrochem.* **2013**, *17*, 2333–2340.

- 128. Zhao, B.; Liu, Z.; Fu, W.; Yang, H. Construction of 3d electrochemically reduced graphene oxide–silver nanocomposite film and application as nonenzymatic hydrogen peroxide sensor. *Electrochem. Commun.* **2013**, *27*, 1–4.
- 129. Babu, K.J.; Zahoor, A.; Nahm, K.; Ramachandran, R.; Rajan, M.A.J.; Gnana kumar, G. The influences of shape and structure of MnO<sub>2</sub> nanomaterials over the non-enzymatic sensing ability of hydrogen peroxide. *J. Nanoparticle Res.* **2014**, *16*, 1–10.
- © 2014 by the authors; licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution license (http://creativecommons.org/licenses/by/4.0/).