GENOME SEQUENCES



AMERICAN SOCIETY FOR MICROBIOLOGY

Whole-Genome Sequence of *Avian coronavirus* from a 15-Year-Old Sample Confirms Evidence of GA08-like Strain Circulation 4 Years Prior to Its First Reported Outbreak

💿 Iryna V. Goraichuk,ª,b James F. Davis,ʿ Arun B. Kulkarni,ʿ 💿 Claudio L. Afonso,ª 💿 David L. Suarezª

Exotic and Emerging Avian Viral Disease Research Unit, Southeast Poultry Research Laboratory, U.S. National Poultry Research Center, ARS, USDA, Athens, Georgia, USA
National Scientific Center Institute of Experimental and Clinical Veterinary Medicine, Kharkiv, Ukraine
Georgia Poultry Laboratory Network, Gainesville, Georgia, USA

ABSTRACT Here, we report the complete genome sequence of an *Avian coronavirus* strain GA08-like isolate from a fecal sample from a broiler chicken collected in Georgia in 2004. The viral genome in this 15-year-old sample provides evidence for the circulation of the GA08-like strain at least 4 years before its first report in 2008.

A vian infectious bronchitis virus (IBV) is a positive-sense, single-stranded RNA coronavirus (genus *Gammacoronavirus*, family *Coronaviridae*) that causes a highly contagious and economically significant disease in chickens worldwide (1–3). Currently, 7 genotypes exist that together comprise 35 distinct viral lineages and numerous strains of IBV, and new variants emerge due to frequent point mutations and recombination events in the viral genome (4–7). In 2008, a novel IBV strain was detected in Georgia broiler chickens with acute chronic respiratory disease and colibacillosis (8, 9). The GA08 virus became a predominant IBV strain at that time in the southeastern United States (10, 11). Over the years, its incidence and severity have increased dramatically, resulting in significant losses. In this study, we report the complete sequence of an IBV GA08-like strain from a 15-year-old sample.

A fecal sample from a 22-day-old broiler chicken was collected at a commercial farm in Georgia in 2004. Feces were first diluted 3:7 in sterile phosphate-buffered saline and then centrifuged for 10 min at 3,200 rpm. The supernatant was further passed sequentially through 1.2-µm- and 0.45-µm-pore-size filters (Merck Millipore, USA) to remove bacteria and large-cell particles. The filtrate was then stored at -70° C for 15 years. In 2019, preserved filtered lysate was DNase treated using the Turbo DNA-free kit (Ambion, USA) to remove host DNA, followed by total nucleic acid extraction using the DNeasy blood and tissue kit (Qiagen, Germany) according to the manufacturer's recommendations. Sequence-independent single-primer amplification (12) was used to produce random amplicons that were processed using the Nextera XT DNA library preparation kit (Illumina, USA). Paired-end (2×150 bp) sequencing was performed on an Illumina MiSeq instrument using the 300-cycle MiSeq reagent kit v2 (Illumina, USA). A total of 2,088,901 raw paired-end reads were generated. A customized workflow on the Galaxy platform (13) was used to perform preprocessing and assembly of the raw sequencing reads, as described previously (14, 15). Briefly, the raw read quality was assessed using FastQC v0.63 (16), and residual adapter sequences were trimmed using Cutadapt v1.6 (17). De novo assembly was performed utilizing MIRA3 v0.0.1 (18) with default settings. The contigs of interest were subjected to a BLASTn search and mapped using BWA-MEM (19) to the full-length reference genome GA9977/2019 (GenBank accession number MK878536) to obtain a draft genome scaffold. The consensus genome sequence of the strain, designated GA/1472/2004, was then recalled based on BWA-MEM mapping of 207,352 trimmed IBV reads to the genome scaffold. The median read depth of the IBV assembly was 558 reads. The final genome consensus was 27,639 nucleotides long, excluding the poly(A) tail (100% genome coverage as estimated based on the size of the IBV GA08

Citation Goraichuk IV, Davis JF, Kulkarni AB, Afonso CL, Suarez DL. 2021. Whole-genome sequence of *Avian coronavirus* from a 15-yearold sample confirms evidence of GA08-like strain circulation 4 years prior to its first reported outbreak. Microbiol Resour Announc 10:e01460-20. https://doi.org/10.1128/MRA .01460-20.

Editor Jelle Matthijnssens, KU Leuven

This is a work of the U.S. Government and is not subject to copyright protection in the United States. Foreign copyrights may apply. Address correspondence to David L. Suarez, david.suarez@usda.gov.

Received 18 December 2020 Accepted 15 March 2021 Published 15 April 2021

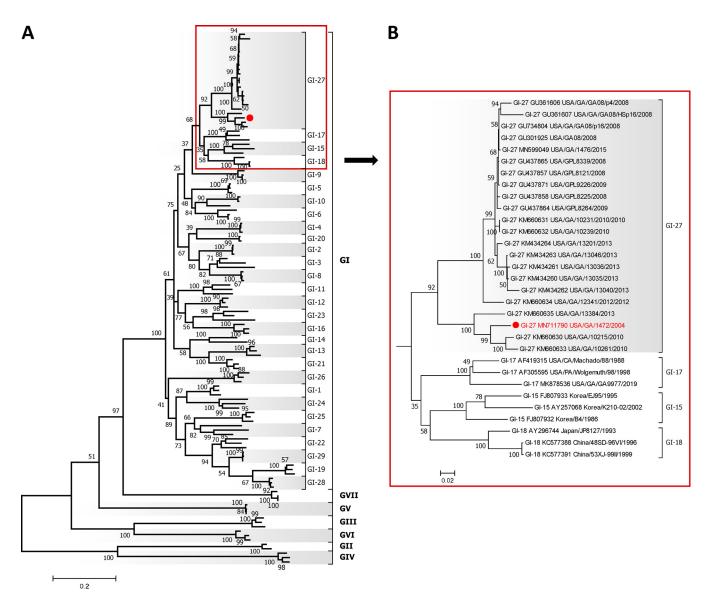


FIG 1 Phylogenetic analysis of IBV isolates of all 35 lineages (121 nucleotide sequences) (A) and GI-27 isolates (B) based on the complete S1 gene sequences constructed with the maximum likelihood method based on the general time-reversible model in MEGA v7.0.26. The tree with the highest log likelihood (-40755.95) is shown. The percentage of replicate trees in which the associated taxa clustered together in the bootstrap test (1,000 replicates) is shown next to each branch. The tree is drawn to scale, with the branch lengths measured in the number of substitutions per site, and the tree is midpoint rooted. All positions containing gaps and missing data were eliminated. There were a total of 1,347 positions in the final data set. The isolate used in this study is shown in red.

reference genome GA/1476/2015; GenBank accession number MN599049.1), and had 38.2% GC content (20). The open reading frames (ORF) were identified using Geneious v11.1.5 and confirmed by alignment with published IBV genome sequences. The genome has the typical genetic structure of all IBV strains and contains 13 ORF (5'-1a/1b-S-3a-3b-E-M-4b-4c-5a-5b-N-6b-3'). A preliminary BLAST comparison of the GA/1472/2004 genome sequence to the currently available full-length IBV genome sequences showed the highest (94.08%) nucleotide identity to the DMV/1639 strain GA9977/2019 belonging to GI-17 (21). However, a new classification of IBV genotypes based on the phylogenetic analysis of the complete spike gly-coprotein S1 coding region has been proposed (4). Therefore, classification using complete S1 nucleotide sequences of all IBV lineages revealed that the GA/1472/2004 isolate clustered with GA08 strains belonging to the GI-27 lineage (Fig. 1), which form two distinct clades of vaccine-type isolates and isolates circulating in backyard birds (9, 20, 22). The IBV isolate characterized in this study clustered together with isolates GA/13384/2013, GA/10215/2010, and GA/10261/2010 (GenBank accession numbers KM660635, KM660630, and KM660633,

respectively) (23) from backyard birds with nucleotide identities ranging from 92.31% to 94.75%.

Therefore, the phylogenetic analysis of the GA/1472/2004 isolate revealed that the IBV GA08-like strain was already circulating in Georgia at least 4 years prior to the first report in 2008. This complete genome sequence information of the 15-year-old IBV GA08 strain would be useful for an in-depth understanding of IBV evolution as well as planning vaccination strategies.

Data availability. The complete genome sequence of the *Avian coronavirus* GA/ 1472/2004 isolate of strain GA08 has been deposited in GenBank under the accession number MN711790. The raw data were deposited in the SRA under accession number SRR10500281, BioSample accession number SAMN13337568, and BioProject accession number PRJNA556282.

ACKNOWLEDGMENTS

We thank Mark W. Jackwood for helpful advice and constructive discussions on IBVs. The mention of trade names or commercial products in this publication is solely for the purpose of providing specific information and does not imply recommendation or endorsement by the U.S. Department of Agriculture.

This study was supported by USDA CRIS project 6040-32000-072 and USDA APHIS interagency agreement 60-6040-5-009.

REFERENCES

- Jackwood MW, de Wit S. 2013. Infectious bronchitis, p 139–159. In Swayne DE, Glisson JR, McDougald LR, Nolan LK, Suarez DL, Nair VL (ed), Diseases of poultry, 13th ed. Wiley-Blackwell Publishing, Ames, IA.
- Cavanagh D. 2007. Coronavirus avian infectious bronchitis virus. Vet Res 38:281–297. https://doi.org/10.1051/vetres:2006055.
- King AMQ, Lefkowitz EJ, Mushegian AR, Adams MJ, Dutilh BE, Gorbalenya AE, Harrach B, Harrison RL, Junglen S, Knowles NJ, Kropinski AM, Krupovic M, Kuhn JH, Nibert ML, Rubino L, Sabanadzovic S, Sanfacon H, Siddell SG, Simmonds P, Varsani A, Zerbini FM, Davison AJ. 2018. Changes to taxonomy and the International Code of Virus Classification and Nomenclature ratified by the International Committee on Taxonomy of Viruses (2018). Arch Virol 163:2601–2631. https://doi.org/10.1007/s00705-018-3847-1.
- Valastro V, Holmes EC, Britton P, Fusaro A, Jackwood MW, Cattoli G, Monne I. 2016. S1 gene-based phylogeny of infectious bronchitis virus: an attempt to harmonize virus classification. Infect Genet Evol 39:349–364. https://doi.org/10.1016/j.meegid.2016.02.015.
- Chen Y, Jiang L, Zhao W, Liu L, Zhao Y, Shao Y, Li H, Han Z, Liu S. 2017. Identification and molecular characterization of a novel serotype infectious bronchitis virus (GI-28) in China. Vet Microbiol 198:108–115. https:// doi.org/10.1016/j.vetmic.2016.12.017.
- Jiang L, Zhao W, Han Z, Chen Y, Zhao Y, Sun J, Li H, Shao Y, Liu L, Liu S. 2017. Genome characterization, antigenicity and pathogenicity of a novel infectious bronchitis virus type isolated from south China. Infect Genet Evol 54:437–446. https://doi.org/10.1016/j.meegid.2017.08.006.
- Ma T, Xu L, Ren M, Shen J, Han Z, Sun J, Zhao Y, Liu S. 2019. Novel genotype of infectious bronchitis virus isolated in China. Vet Microbiol 230:178–186. https://doi.org/10.1016/j.vetmic.2019.01.020.
- de Wit JJ, Cook JKA, van der Heijden HMJF. 2011. Infectious bronchitis virus variants: a review of the history, current situation and control measures. Avian Pathol 40:223–235. https://doi.org/10.1080/03079457.2011.566260.
- Jackwood MW, Hilt DA, Sellers HS, Williams SM, Lasher HN. 2010. Rapid heat-treatment attenuation of infectious bronchitis virus. Avian Pathol 39:227–233. https://doi.org/10.1080/03079451003801516.
- Jackwood M. 2009. Infectious bronchitis in the USA. MSD Animal Health, Kenilworth, NJ. http://www.infectious-bronchitis.com/ibv-usa-2006.aspx.
- Jackwood MW. 2012. Review of infectious bronchitis virus around the world. Avian Dis 56:634–641. https://doi.org/10.1637/10227-043012-Review.1.
- Chrzastek K, Lee D-H, Smith D, Sharma P, Suarez DL, Pantin-Jackwood M, Kapczynski DR. 2017. Use of sequence-independent, single-primer-amplification (SISPA) for rapid detection, identification, and characterization of avian RNA viruses. Virology 509:159–166. https://doi.org/10.1016/j.virol.2017.06.019.

- Afgan E, Baker D, van den Beek M, Blankenberg D, Bouvier D, Čech M, Chilton J, Clements D, Coraor N, Eberhard C, Grüning B, Guerler A, Hillman-Jackson J, Von Kuster G, Rasche E, Soranzo N, Turaga N, Taylor J, Nekrutenko A, Goecks J. 2016. The Galaxy platform for accessible, reproducible and collaborative biomedical analyses: 2016 update. Nucleic Acids Res 44:W3–W10. https://doi.org/10.1093/nar/gkw343.
- Dimitrov KM, Sharma P, Volkening JD, Goraichuk IV, Wajid A, Rehmani SF, Basharat A, Shittu I, Joannis TM, Miller PJ, Afonso CL. 2017. A robust and cost-effective approach to sequence and analyze complete genomes of small RNA viruses. Virol J 14:72. https://doi.org/10.1186/s12985-017-0741-5.
- Goraichuk IV, Davis JF, Kulkarni AB, Afonso CL, Suarez DL. 2020. A 25year-old sample contributes the complete genome sequence of *Avian coronavirus* vaccine strain ArkDPI, reisolated from commercial broilers in the United States. Microbiol Resour Announc 9:e00067-20. https://doi .org/10.1128/MRA.00067-20.
- Andrews S. 2010. FastQC: a quality control tool for high throughput sequence data. http://www.bioinformatics.babraham.ac.uk/projects/fastqc/.
- Martin M. 2011. Cutadapt removes adapter sequences from high-throughput sequencing reads. EMBnet J 17:10–12. https://doi.org/10.14806/ej.17.1.200.
- Chevreux B, Wetter T, Suhai S. 1999. Genome sequence assembly using trace signals and additional sequence information, p 45–56. *In* Computer science and biology: proceedings of the German Conference on Bioinformatics (GCB '99). GCB, Hanover, Germany.
- Li H, Durbin R. 2009. Fast and accurate short read alignment with Burrows-Wheeler transform. Bioinformatics 25:1754–1760. https://doi.org/10 .1093/bioinformatics/btp324.
- Goraichuk IV, Davis JF, Kulkarni AB, Afonso CL, Suarez DL. 2020. Complete genome sequence of *Avian coronavirus* strain GA08 (GI-27 lineage). Microbiol Resour Announc 9:e00068-20. https://doi.org/10.1128/MRA.00068-20.
- Goraichuk IV, Kulkarni AB, Williams-Coplin D, Suarez DL, Afonso CL. 2019. First complete genome sequence of currently circulating infectious bronchitis virus strain DMV/1639 of the Gl-17 lineage. Microbiol Resour Announc 8:e00840-19. https://doi.org/10.1128/MRA.00840-19.
- Kulkarni AB, Resurreccion RS. 2010. Genotyping of newly isolated infectious bronchitis virus isolates from northeastern Georgia. Avian Dis 54:1144–1151. https://doi.org/10.1637/9358-040510-Reg.1.
- Kulkarni A. 2016. Characterization of recent infectious bronchitis virus isolates from broilers and backyard flocks of Georgia, p 139–141. *In* Frame D (ed), Proceedings of the Sixty-Fifth Western Poultry Disease Conference. American College of Poultry Veterinarians, Jacksonville, FL. https://aaap .memberclicks.net/assets/WPDC/wpdc_2016_final.pdf.