

1 **B cell immune repertoire sequencing in tobacco cigarette smoking, vaping,**
2 **and chronic obstructive pulmonary disease in the COPDGene cohort**

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73 **Abstract**

74 Rationale: Cigarette smoking (CS) impairs B cell function and antibody production, increasing
75 infection risk. The impact of e-cigarette use ('vaping') and combined CS and vaping ('dual-use')
76 on B cell activity is unclear.

77 Objective: To examine B cell receptor sequencing (BCR-seq) profiles associated with CS,
78 vaping, dual-use, COPD-related outcomes, and demographic factors.

79 Methods: BCR-seq was performed on blood RNA samples from 234 participants in the
80 COPDGene study. We assessed multivariable associations of B cell function measures
81 (immunoglobulin heavy chain (IGH) subclass expression and usage, class-switching, V-segment
82 usage, and clonal expansion) with CS, vaping, dual-use, COPD severity, age, sex, and race. We
83 adjusted for multiple comparisons using the Benjamini-Hochberg method, identifying significant
84 associations at 5% FDR and suggestive associations at 10% FDR.

85 Results: Among 234 non-Hispanic white (NHW) and African American (AA) participants, CS
86 and dual-use were significantly positively associated with increased secretory IgA production,
87 with dual-use showing the strongest associations. Dual-use was positively associated with class
88 switching and B cell clonal expansion, indicating increased B cell activation, with similar trends
89 in those only smoking or only vaping. We observed significant associations between race and

90 IgG antibody usage. AA participants had higher IgG subclass proportions and lower IgM usage
91 compared to NHW participants.

92 Conclusions: CS and vaping additively enhance B cell activation, most notably in dual-users.

93 Self-reported race was strongly associated with IgG isotype usage. These findings highlight

94 associations between B cell activation and antibody transcription, suggesting potential

95 differences in immune and vaccine responses linked to CS, vaping, and race.

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112 Introduction

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114 Use of electronic cigarettes, i.e. vaping, has increased substantially since their introduction to
115 the U.S. market in 2007¹. Numerous studies have demonstrated that vaping induces
116 inflammatory responses and has adverse health effects²⁻⁶. More precise characterization of the
117 inflammatory effects of vaping may better define its effects on health, both for vaping alone and
118 vaping in conjunction with combustible cigarettes, i.e. dual-use.

119 B cells participate in adaptive immunity largely by producing antibodies that protect mucosal
120 surfaces and provide antigen-specific responses to infection. Combustible tobacco cigarette
121 smoking (CS) has adverse effects on B cells resulting in increased susceptibility to infections⁷.
122 Proper B cell function depends on B cell activation, a process in which naïve B cells are
123 activated by exposure to an antigen. This process triggers clonal expansion of B-cell populations
124 with specifically rearranged B-cell receptor (BCR) genes, which encode the specific
125 immunoglobulin (Ig) produced by each clone. In conjunction with T-cell help, these activated
126 clones undergo somatic hypermutation, becoming optimized to bind specific antigens. BCR
127 sequencing (BCR-seq) allows for the identification and sequence-specific characterization of B
128 cells and expanded B cell clones, providing rich characterization of the B cell response⁸.

129 We hypothesized that vaping and dual-use alter B cell function and the ability of B cells to
130 respond appropriately to antigens through activation and antibody production. To address this
131 question, and to determine whether any of these B cell changes are associated with measures of

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133 chronic obstructive pulmonary disease (COPD), we performed BCR-seq in 234 participants from
134 the Genetic Epidemiology of COPD (COPDGene)⁹ Study, a large study of individuals who
135 currently or previously smoked that is enriched for participants with COPD.

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137 **Methods**

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139 **Study population**

140 Written informed consent was obtained from all study participants, and institutional review
141 board approval was obtained at all study centers. The Genetic Epidemiology of COPD
142 (COPDGene)⁹ study enrolled 10,198 non-Hispanic white (NHW) and African American (AA)
143 individuals who smoked 10 or more pack-years of cigarettes during their lifetime and who were
144 aged 45-80 at study enrollment. COPDGene is an ongoing longitudinal study with completed
145 enrollment, 5-year, and 10-year visits. At each visit, anthropometric measurements, spirometry,
146 chest computed tomography (CT) imaging, and blood samples were collected. At the 5-year
147 follow up visit, we collected questionnaire data on use of cigarette and e-cigarette products. All
148 data in this paper comes from the 5-year study visit where both blood RNA-seq and vaping data
149 are available.

150

151 *Participant selection*

152 Selection of participants for BCR-seq⁸ was performed using a stratified random sampling
153 approach as follows. First, all participants with available blood RNA and complete vaping and
154 CS data from COPDGene Phase 2 at the time of participant selection were considered (n=3,601).
155 They were stratified into five groups based on vaping and CS status – never smokers, former

156 combustible cigarette smokers, current cigarette smokers (without current vaping), current vapers
157 (without current cigarette use), and current dual-users (vaping and cigarette use). All participants
158 in the current vaper (n=41) and dual-user group (n=57) with available samples were selected for
159 BCR-seq, and the remaining 136 participants were randomly sampled from the set of available
160 participants in the other CS groups. Participants taking oral corticosteroids were excluded from
161 the analysis as these medications are known to modify B cell function.

162

163 *Definition of key study variables*

164 CS and vaping behavior were ascertained by self-report. Vapers were participants who
165 reported using at least one e-cigarette within the prior week and had a history of smoking
166 tobacco cigarettes, but not within the last 30 days. Current cigarette users reported current
167 smoking with an average of at least one cigarette per day without any e-cigarette use. Dual-users
168 were vapers who also reported current CS, and former cigarette users were defined as those who
169 reported a history of smoking but did not meet criteria for current CS or vaping. In most of the
170 reported analyses, former cigarette users are used as the reference group.

171 Information on the age, sex, and race of participants was elicited through self-report. For sex,
172 participants were asked if they were male or female. For race, participants were asked if they
173 were NHW, Black or African American, Asian, Pacific Islander, American Indian or Alaska
174 Native, or Other with the option to select multiple categories. By design, inclusion in
175 COPDGene was limited to participants self-identifying as NHW or AA. A separate ethnicity
176 question asked participants if they were Hispanic or Latino. For this study, participants were
177 coded as NHW if they indicated “White” and AA if they indicated “Black or African American.”

178 In the U.S., COPD primarily develops in the setting of cigarette smoking exposure, and B cell
179 lymphoid follicles are associated with COPD severity¹⁰⁻¹². Therefore, we examined the
180 association of BCR-seq measures with COPD and COPD-related traits. COPD status was
181 determined by GOLD spirometry grades based on post-bronchodilator spirometry testing where
182 participants were grouped into normal spirometry (FEV1/FVC \geq 0.7 and FEV1 % predicted \geq
183 80%) or GOLD spirometry grade 1, GOLD grade 2-4, or preserved ratio with impaired
184 spirometry (PRISm)¹³. For all analyses, the reference group included formerly smoking
185 individuals with normal spirometry. Global Lung Initiative (GLI) race-neutral equations were
186 used to calculate % predicted spirometry values. Computed tomography (CT) imaging measures
187 of emphysema and airway wall thickness were generated by Thirona (<https://thirona.eu/>) and the
188 following measures were analyzed: % low attenuation area less than -950 Hounsfield units for
189 emphysema and airway wall thickness as % of overall airway volume (wall area percent¹⁴).

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191 *B-cell receptor sequencing library preparation*

192 Details regarding generation of RNAseq data in COPDGene were previously published¹⁵.
193 Whole blood was collected and stored in PAXgene Blood RNA tubes, and total RNA was
194 extracted using Qiagen PreAnalytiX PAXgene Blood miRNA Kit (Qiagen, Valencia, CA).
195 Sequencing libraries were prepared using 200 ng of total RNA as input following a protocol
196 modified from⁸. Additional details regarding library preparation and data processing can be
197 found in the Supplementary Methods.

198 We generated adaptive immune receptor repertoire sequencing data for B cell receptors
199 (hereafter, 'BCR-seq') data using a set of isotype-specific immunoglobulin heavy chain (IGH)
200 constant region primers. Reads were aligned to International Immunogenetics Information

201 System (IMGT) reference germline sequences, and clonal relationships between BCR sequences
202 were inferred using the spectralClones function from the scoper R package contained within the
203 Immcantation suite of software packages
204 (<https://immcantation.readthedocs.io/en/stable/about.html>). Mutated sequences were defined as
205 sequences that were aligned but differed from the IMGT reference by one or more bases.

206 Uniquely identified BCR sequences were quantified to represent antibody isotype expression
207 (\log_2 counts of the number of unique BCR sequences present within each isotype class) and
208 usage (number of unique BCR sequences present within each isotype class divided by the total
209 number of BCR sequences), B cell activation measured through class switching (number of
210 unique BCR sequences in the IgA, IgG, and IgE isotypes divided by the total number of BCR
211 sequences), length of the CDR3 region in nucleotides, and the clonal diversity of the B cell
212 population in each individual as measured by Hill numbers¹⁶. V-segment usage was defined as
213 the number of unique and mutated BCR sequences containing a specific V-segment (as defined
214 by IGHV genes from the IMGT reference) divided by total number of unique BCR sequences.
215 For each BCR-seq measure, we analyzed only those measures where the isotype or V-segment
216 class in question was present at >1% of the total unique sequences for 25% of the participants or
217 more. The one exception was measurement of the IgE isotype which was analyzed despite being
218 below this threshold due to its established clinical importance.

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220 *B-cell Receptor Sequencing Measures*

221 BCR sequencing involves sequencing transcripts of the B cell receptor using a set of primers
222 targeting the Fc-region of the immunoglobulin heavy chain (IGH). This provides comprehensive
223 assessment of the BCR repertoire including antibody isotype (IgM, IgD, IgA, IgG, and IgE), V-

224 segments corresponding to the variable region that determines antibody specificity, and clonal
225 expansion and somatic hypermutation of specific B cell populations. These sequence counts are
226 summarized into quantitative measures of 1) isotype usage (proportion of antibody transcripts for
227 each isotype within each individual), 2) isotype expression (log₂ transformed counts that
228 represent the number of unique B cells per isotype within each individual), 3) class switching
229 (proportion of class-switched B cells per individual), 4) V-segment usage (proportion of
230 antibody transcripts for each V-segment within each individual), 5) CDR3 length by isotype, and
231 6) B cell clonal diversity measured by Hill numbers. For duplicated sequences, we only counted
232 the sequence once, which means our measures represent numbers of B cells rather than number
233 of transcripts.

234

235 *Statistical Analysis*

236 We performed analyses in R version >4.0 (www.r-project.org). We assessed normality of
237 continuous variables by visual inspection of histograms. Results are shown as mean ± standard
238 deviation or median [interquartile range], as appropriate. Differences in continuous variables
239 were assessed with Student t-tests or Wilcoxon tests. Categorical variables were compared by
240 ANOVA or Kruskal-Wallis tests, as appropriate. We considered false discovery rate (FDR)-
241 adjusted Benjamini-Hochberg¹⁷ p-values less below 0.05 to be significant and between 0.05 and
242 0.1 to be suggestive.

243 For each of the BCR-seq measures, we used univariable analysis and multivariable regression
244 to examine the association between each of these measures with CS, vaping, and dual-use.
245 Further, we examined associations to age, sex, and race as well as COPD affection status (GOLD

246 2-4), CT emphysema measures (% low attenuation area (LAA) < -950 Hounsfield units (HU)¹⁴),
247 and CT airway wall thickness (wall area %¹⁴).

248 For analyses of smoking/vaping, demographic variables, and GOLD spirometry grade,
249 multivariable models included the following covariates: age, sex, self-identified race,
250 vaping/smoking behavior, GOLD grade, pack-years of smoking, and inhaled corticosteroid use.
251 For CT imaging measures, models were additionally adjusted for CT scanner model. To
252 visualize the results, we constructed violin plots and heatmaps.

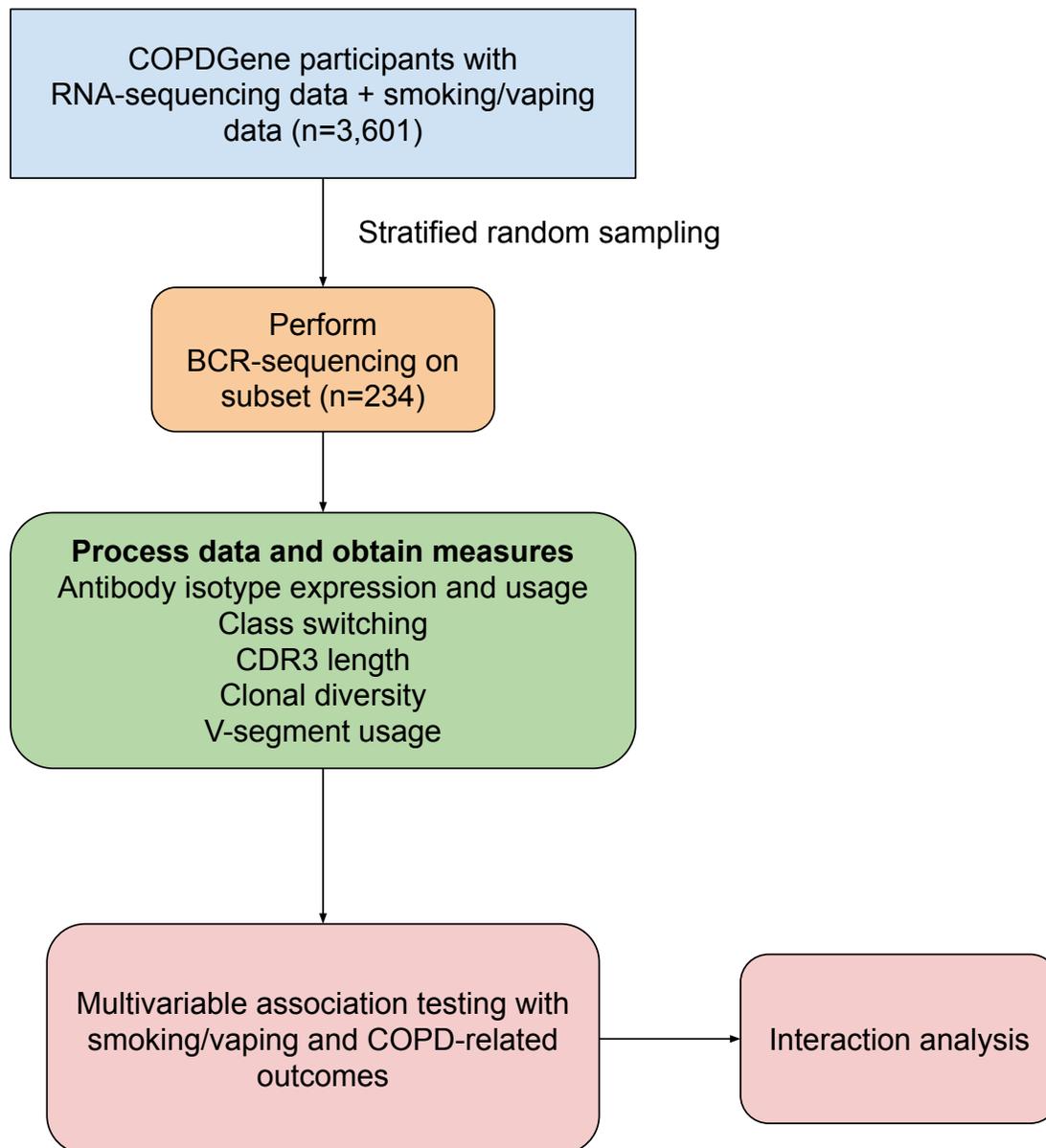
253 Sensitivity analyses were performed for the significant associations observed with
254 smoking/vaping status and self-reported race adjusting for self-reported income level and social
255 deprivation index, a measure of area-level deprivation¹⁸, and principal components of genetic
256 ancestry. We additionally performed interaction analyses between self-reported race and
257 smoking/vaping variables by including the main effects and cross-product interaction terms in a
258 regression model.

259

260 **Results**

261 *Characteristics of study participants*

262 A schematic of our study design is shown in Figure 1. We included 234 COPDGene NHW
263 and AA participants with smoking/vaping and BCR-seq data, and a table of their characteristics
264 is shown in Table 1. Compared to other groups, dual-users were more likely to be younger,
265 NHW, have more pack-years of cigarette smoking (CS), lower FEV₁ % predicted, and thicker
266 airway walls. Compared to dual-users, individuals who only vaped were slightly older, were less
267 likely to be male, had similar pack-years of smoking, but had higher FEV₁ % predicted, and more
268 quantitative emphysema (% LAA < -950 HU).



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270 **Figure 1: Schematic of study design.** COPDGene = Genetic Epidemiology of COPD study.

271 BCR = B cell receptor. COPD = chronic obstructive pulmonary disease.

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Table 1. Characteristics of Study Participants

	former smoker (N=44)	never (N=41)	vaping (N=41)	cigarette smoking (N=51)	dual (N=57)	overall (N=234)	p-value
age	69.6 (7.42)	65.3 (9.96)	64.4 (6.56)	62.0 (7.21)	61.3 (6.47)	64.2 (8.04)	<0.001
sex							
female	23 (52.3%)	29 (70.7%)	26 (63.4%)	31 (60.8%)	29 (50.9%)	138 (59.0%)	0.403
male	21 (47.7%)	12 (29.3%)	15 (36.6%)	20 (39.2%)	28 (49.1%)	96 (41.0%)	
race							
AA	6 (13.6%)	2 (4.9%)	4 (9.8%)	28 (54.9%)	15 (26.3%)	55 (23.5%)	<0.001
NHW	38 (86.4%)	39 (95.1%)	37 (90.2%)	23 (45.1%)	42 (73.7%)	179 (76.5%)	
cigarette pack-years	43.0 (23.3)	0 (0)	52.2 (24.6)	48.5 (27.8)	52.9 (25.3)	40.7 (29.8)	<0.001
GOLD spirometry grade							
Normal spirometry	15 (34.1%)	39 (95.1%)	19 (46.3%)	22 (43.1%)	19 (33.3%)	114 (48.7%)	<0.001
GOLD 1	6 (13.6%)	0 (0%)	5 (12.2%)	3 (5.9%)	6 (10.5%)	20 (8.5%)	
GOLD 2,3,4	21 (47.7%)	1 (2.4%)	13 (31.7%)	20 (39.2%)	29 (50.9%)	84 (35.9%)	
PRISm	2 (4.5%)	1 (2.4%)	4 (9.8%)	6 (11.8%)	3 (5.3%)	16 (6.8%)	
FEV1, % of predicted	75.3 (27.7)	74.1 (26.8)	107 (13.2)	72.8 (21.1)	82.0 (24.9)	81.3 (26.5)	<0.001
FEV1/FVC	0.633 (0.149)	0.795 (0.0479)	0.672 (0.142)	0.688 (0.141)	0.642 (0.158)	0.682 (0.146)	<0.001
CT emphysema	6.30 (7.55)	1.24 (1.44)	3.43 (4.90)	3.43 (7.13)	3.10 (5.76)	3.45 (5.99)	0.002
CT airway wall thickness	51.1 (6.34)	42.7 (4.81)	48.4 (8.79)	52.6 (8.97)	52.6 (9.14)	49.8 (8.67)	<0.001

Values are mean (standard deviation) for continuous variables and N (%) for categorical variables. CT emphysema measurement is % low attenuation area <-950 Hounsfield units. CT airway wall thickness is defined as (area of segmental airway walls / overall airway area). FEV1 = forced expiratory volume in 1 second. FEV1/FVC = FEV1/forced vital capacity. PRISm = preserved ratio with impaired spirometry. AA= African American. NHW = non-Hispanic white. CT = computed tomography. Global Lung Initiative (GLI) race-neutral equations were used to calculate % predicted values. P-values test differences across all groups (analysis of variance (ANOVA)).

275 *Associations between BCR-seq measures and cigarette smoking, vaping, and dual-use*

276 Significant (q-value < 0.05) and suggestive (q-value <0.1) associations for vaping, CS, and
277 dual-use are shown in Table 2 (Tables E1-E5 contain complete model results). Overall, we
278 observed that the most pronounced changes in antibody production were associated with dual-
279 use. Specifically, dual-use resulted in a shift in isotype usage towards IgA and away from IgM
280 (Figure 2A). It was also associated with increased class switching suggestive of B cell activation
281 (Figure 2B) and increased usage of specific V-segments. Clonality analysis also demonstrated
282 reduced antibody diversity for participants engaged in dual-use (Figure E1), suggesting that there
283 is a greater amount of B cell clonal expansion in this group. Since CS status is often associated
284 with socioeconomic variables, we tested these associations after adjusting for income level and
285 social deprivation index, a composite measure of area-level deprivation, which had minimal
286 effect on the significance of these associations (Table E6).

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Table 2. Significant BCR associations to vaping, cigarette smoking, and dual-use.

BCR measure	Dual-use			Cigarette smoking			Vaping		
	β (se)	P-value	q-value	β (se)	P-value	q-value	β (se)	P-value	q-value
IgA2 count	0.879 (0.294)	3.1E-03	0.05	0.766 (0.306)	0.01	0.11	0.432 (0.301)	0.15	0.48
IgA1 usage	0.095 (0.023)	7.5E-05	2.7E-03	0.061 (0.025)	0.01	0.11	0.051 (0.024)	0.04	0.19
IgA2 usage	0.059 (0.011)	2.8E-07	7.0E-05	0.035 (0.012)	0.00	0.05	0.027 (0.012)	0.02	0.12
IgM usage	-0.173 (0.038)	1.1E-05	8.9E-04	-0.113 (0.04)	0.01	0.07	-0.087 (0.04)	0.03	0.17
class switching proportion	0.111 (0.027)	5.4E-05	2.6E-03	0.064 (0.028)	0.02	0.14	0.05 (0.028)	0.07	0.31
IGHV1.18.01 usage	0.005 (0.001)	4.9E-04	0.01	0.004 (0.001)	0.01	0.11	0.004 (0.001)	0.02	0.12
IGHV3.7.01 usage	0.006 (0.002)	3.2E-04	9.6E-03	0.003 (0.002)	0.05	0.24	0.002 (0.002)	0.28	0.62
IGHV5.51.01 usage	0.009 (0.002)	6.2E-05	2.6E-03	0.005 (0.002)	0.02	0.12	0.006 (0.002)	0.01	0.11
IgE CDR3 length	-0.384 (2.294)	0.87	0.96	0.41 (2.392)	0.86	0.96	6.755 (2.427)	0.01	0.08

Count values are log2 of unique BCR RNA sequence count. Usage is the proportion of all BCR RNA sequences falling into either a specific isotype or v-segment category (calculated separately for isotypes and v-segments). Class switching proportion is the proportion of all BCR RNA sequences that belong to IgA, IgG, or IgE isotypes and have evidence of somatic hypermutation (>1 mutation relative to the IMGT reference database). CDR3 length is the length of the CDR3 sequence in nucleotides. Q-value is calculated using the Benjamini-Hochberg method.

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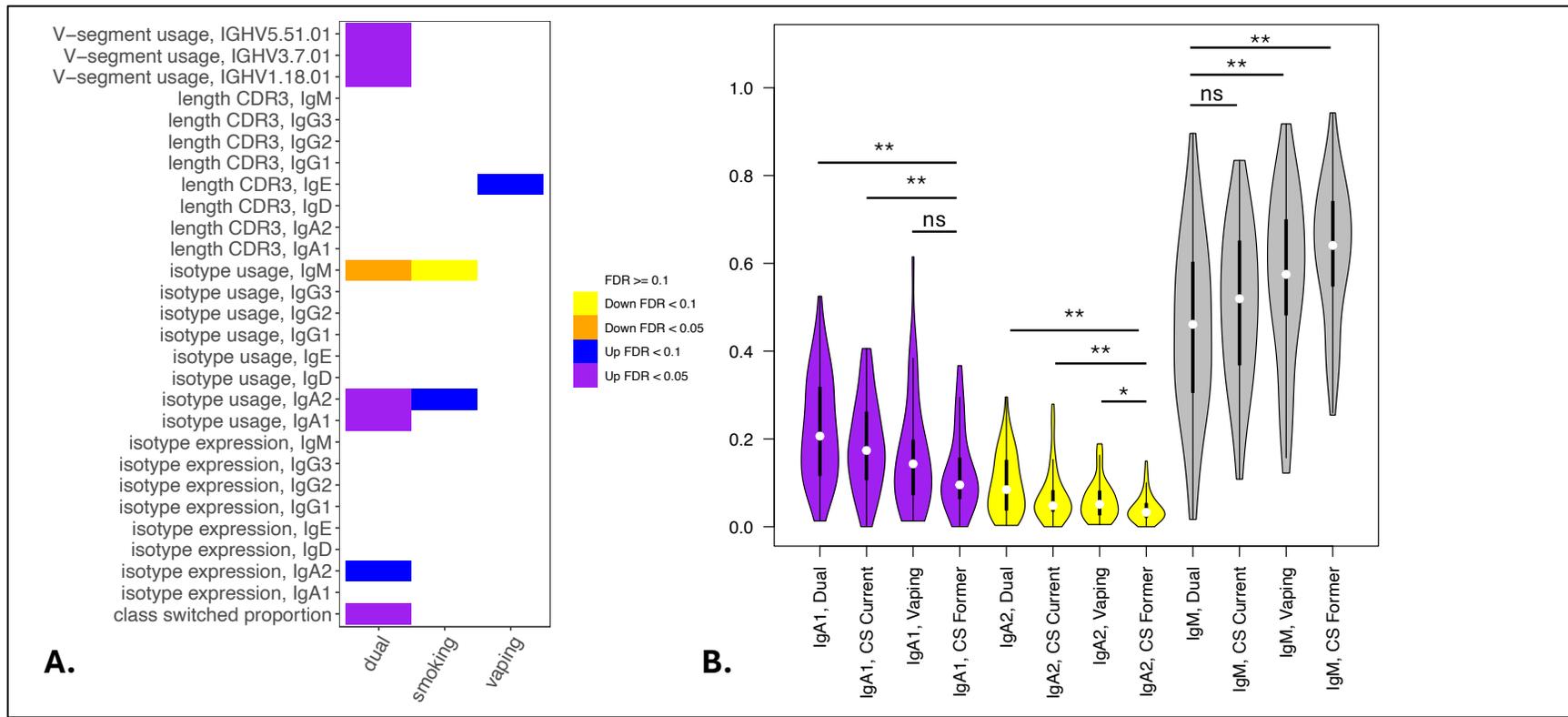
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Figure 2. Associations of BCR Measures with Vaping and Cigarette Smoking. Significant associations between BCR measures

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and vaping, cigarette smoking, and dual-use from multivariable models analyzing class switching, isotype expression and usage, V-

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segment usage, and CDR3 length (in nucleotides) are shown in Panel A. Panel B shows IgA and IgM isotype usage among

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participants engaged in current smoking, vaping, or dual-use with former smokers included for comparison. Significance is assessed

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by t-tests. * p<=0.05, ** p<=0.005, ns p>0.05.

304 Current CS also showed suggestive association to increased secretory IgA usage and
305 decreased IgM, with several other borderline but non-significant associations. Vaping showed a
306 suggestive association to decreased CDR3 length in IgE antibody transcripts as well as several
307 borderline but non-significant associations (Figure E2). Overall, vaping and CS showed a similar
308 trend in effect sizes compared to dual-use, suggesting that the effects were similar but less
309 pronounced in current smokers and vapers relative to dual-users. When comparing dual-use to
310 CS, we found no significant associations with BCR-measures. When comparing dual-use to
311 vaping, we found dual-use was associated with lower IgE CDR3 length ($\beta = -7.14$ (SE: 2.28,
312 adjusted p-value = 0.034)) and higher IgA2 usage ($\beta = 0.032$ (SE: 0.011, adj. p-value = 0.052)).

313 To examine the appropriateness of using former rather than never smokers as the reference
314 group, we performed multivariable linear regressions comparing isotype usage in former versus
315 never smoking individuals (Table E7), which demonstrated no significant differences between
316 these groups after adjusting for multiple comparisons, suggesting that smoking effects on class
317 switching may resolve after cessation.

318

319 *Associations between Sex and Self-Reported Race on B cell antibody production*

320 In multivariable models, some of the strongest observed associations for BCR-seq measures
321 were with self-reported race (Table 3). Comparing self-reported NHW versus AA participants,
322 NHW-identifying participants had decreased usage of IgG1, IgG2, and IgG3 isotypes and
323 increased usage of IgM (Figure 3). To investigate the extent to which these associations may be
324 driven by variables related to income or socioeconomic status, we repeated the analysis adjusting
325 for self-reported income level and area deprivation index, and 4 of the 6 significant associations
326 remained significant, and all 6 associations had a consistent effect direction (Table E8). After

327 adjusting for principal components of genetic ancestry, the isotype usage associations with race
328 were attenuated, though notably, the principal component variables were also not associated with
329 isotype usage. We observed no interaction between self-identified race and CS, vaping, or dual-
330 use on isotype usage (all $p > 0.05$).

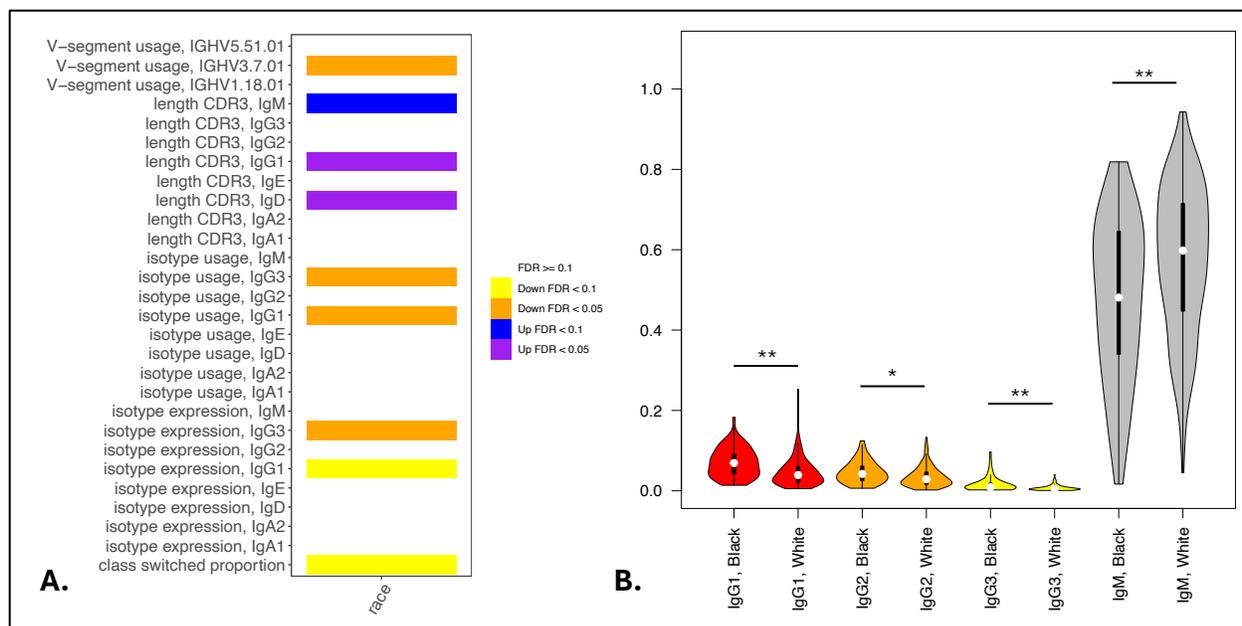
331 In the sex analysis, we observed one significant association in which male compared to
332 female sex was significantly associated with decreased CDR3 length in IgM isotype sequences
333 (β -0.63 (SE: 0.18, $p=0.0008$)).

BCR Measure	Self-reported AA vs NHW Participants		
	β (se)	P-value	q-value
IgG3 usage	-0.009 (0.002)	6.0E-06	7.6E-04
IgG1 usage	-0.026 (0.006)	2.6E-05	1.6E-03
IgG1 CDR3 length	1.038 (0.285)	3.4E-04	9.6E-03
IgG3 count	-0.75 (0.211)	4.6E-04	0.01
IgD CDR3 length	0.96 (0.271)	4.9E-04	0.01
IGHV3.7.01 usage	-0.004 (0.001)	1.8E-03	0.03
IgM CDR3 length	0.699 (0.238)	3.7E-03	0.06
IgG1 count	-0.549 (0.191)	4.5E-03	0.06
class switching	-0.059 (0.022)	7.0E-03	0.08

Count values are log2 of unique BCR RNA sequence count. Usage is the proportion of all BCR RNA sequences falling into either a specific isotype or v-segment category (calculated separately for isotypes and v-segments). Class switching proportion is the proportion of all BCR RNA sequences that belong to IgA, IgG, or IgE isotypes and have evidence of somatic hypermutation (>1 mutation relative to the IMGT reference database). CDR3 length is the length of the CDR3 sequence in nucleotides. NHW is the reference group (i.e. negative β means lower in NHW).

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344 *Associations to COPD and Related Phenotypes*

345 We also examined associations between BCR-seq measures and age, COPD affection status,
346 and CT-related measures of emphysema and airway wall thickness. We observed a significant
347 univariate association between COPD and increased usage of the IGHV5.51.01 V-segment and
348 suggestive associations with increased class switching and a shift from IgM to IgA (Table E9).
349 Similar but non-significant trends were associated with the CT-quantified airway wall thickness
350 (Table E10). However, these associations were not significant in models adjusting for CS/vaping
351 behavior, age, sex, and race. No significant associations were observed with CT emphysema
352 measures.

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354 **Discussion**

355 In this study of 234 individuals with B cell receptor sequencing (BCR-seq) and cigarette
356 smoking (CS), vaping, and COPD-related outcome data, we observed significant effects of CS
357 and vaping leading to increased IgA expression and usage, increased class switching, and lower
358 antibody diversity indicating greater clonal activation of specific B cell populations. Taken
359 together, these results demonstrate the potential dangers of dual-use compared to single product
360 use, an under-recognized but important public health concern.

361 Our study demonstrates associations between dual-use and increased production of IgA,
362 increased class switching, increased usage of specific V-segments and clonal expansion of B
363 cells. These results point to increased B cell activation and increased production of secretory IgA
364 (i.e. IgA2), consistent with mucosal exposure to compounds from vaping devices and
365 combustible cigarettes. We note that CS was suggestively associated with increased IgA usage,
366 and vaping also showed a borderline association with a consistent effect direction (adjusted

367 p=0.12). It is possible that in a larger study these associations would reach statistical
368 significance. IgA is secreted by the airway mucosa and is important in lung immune defense
369 against pathogens¹⁹. Our data suggest that CS and vaping trigger similar host immune responses
370 with a greater effect observed in participants engaged in dual-use. Indeed, many patients who use
371 e-cigarettes for smoking cessation will smoke tobacco cigarettes and vape, and our results
372 underscore the importance of understanding the health effects of dual-use specifically, as well as
373 vaping and CS alone.

374 Our findings are consistent with previous research demonstrating that CS increases IgA
375 production in blood and lung^{20,21}. Higher levels of class-switched memory B cells have been
376 observed in individuals who smoke compared to former and never smokers, irrespective of
377 COPD status²². Vaping is associated with increased circulating club cell protein and decreased
378 transcutaneous oxygen tension³, increased IL-10 and TNF- α ²³, and methylation changes that
379 may cause long-term alterations in cytokine levels^{24,25}. Vaping has also been associated with
380 increased expression of 191 inflammatory proteins from bronchoalveolar lavage fluid, including
381 MUC5AC, which is important in mucin production²⁶. Clinically, vaping is associated with acute
382 lung injury^{2,27}, decreased FEV₁/FVC and peak expiratory flow in asthmatics²³, and chronic
383 bronchitis²⁶. However, the implications of dual-use on adaptive immunity are an important
384 contribution of our study.

385 The role of adaptive immunity and B cells in COPD and emphysema pathogenesis is well-
386 recognized^{10,19,28,29}. Although we observed several univariable associations of blood B cell
387 transcriptomics to COPD and related phenotypes, none of these associations remained significant
388 in multivariable models after adjustment for multiple comparisons. While seemingly in contrast
389 to the well-known increase in lung lymphoid follicles and B cell infiltration in severe COPD¹⁰⁻

390 ^{12,28,29}, this lack of significant associations is not surprising, as by definition the presence of
391 germinal centers in lung lymphoid follicles indicates local B cell division and maturation.
392 Indeed, antigen exposure within the lung leads to local recruitment of memory (and perhaps
393 naïve) B cells, local expansion of B cells and plasma cells within the airway tissue, and
394 subsequent production of antigen-specific antibodies that tend to stay in the lung before getting
395 into the bloodstream^{10,30}. Since our analysis is limited to the transcriptome (cross-sectionally) of
396 circulating B-cells, our results do not rule out spillover of antibody produced by lung-resident B-
397 cells into the blood, and other COPD-specific inflammatory changes in blood such as
398 neutrophilia and the increase in several RNA and protein biomarkers are well-documented^{31–33}.
399 Our results suggest that the circulating B cell population in participants with COPD does not
400 show large COPD-specific changes. Since COPD is strongly associated with CS and older age,
401 participants with COPD would not be expected to have “normal” B cell function reflective of
402 good health, but rather they would have B cell alterations that are characteristic of individuals
403 with similar age and smoking history.

404 An intriguing aspect of BCR-seq is the ability to characterize the B cell response at the level
405 of specific B cell clones. It is of interest that multiple associations between dual-use and race
406 were observed with the usage of specific V-segments. For example, dual-use was strongly
407 associated with increased usage of IGHV5.51.01 which has been associated with immune
408 responses to parainfluenza viruses³⁴, a common cause of upper respiratory illness that can cause
409 severe respiratory illness in older or immunocompromised individuals.

410 Our finding of higher levels of IgG1 and IgG3 in AA compared to NHW participants agrees
411 with a previous report³⁵, which we extend by demonstrating this association in the context of
412 smoking/vaping behaviors. Further, we observed no interaction between race and

413 smoking/vaping variables on isotype usage, though it is important to note that our study is not
414 well-powered to detect interactions of modest effect. Despite the lack of a large literature on
415 racial differences in B cell function, this topic is of substantial interest due to the increased risk
416 of multiple myeloma in AA individuals³⁶. IgG1 and IgG3 both have excellent complement
417 activation and opsonization capabilities, and IgG3 is a potent immune effector, suggesting
418 differences in response to pathogens and toxins. As certain continuous traits, such as height and
419 skin color, can vary with a person's ancestral geographic origins, the observed associations with
420 self-identified race could represent complex gene-by-environment interactions, or like all
421 associations, could just be due to unmeasured confounding. In sensitivity analyses adjusting for
422 income levels and area deprivation index, the associations with self-reported race and
423 smoking/vaping status remained significant. We further adjusted for principal components of
424 genetic ancestry, which attenuated race associations with isotype usage; however, there was no
425 association of the principal components of genetic ancestry with isotype usage, suggesting that
426 genetic principal components alone do not account for the observed differences. These findings
427 need to be confirmed by future studies that focus on identifying the potential biological and
428 socio-economic factors driving these differences as well as exploring forms of genetic analysis
429 that account for admixture. Such studies would likely yield useful data for assessing disease risk
430 and understanding vaccination response.

431 To our knowledge, this is the largest study to date of BCR-seq in humans or model systems.
432 The strengths of this study include the novel use of BCR-seq in a large, deeply phenotyped
433 cohort of participants engaged in current CS, vaping, or both. One limitation is that we did not
434 have a suitable replication cohort, but our current findings highlight the need to obtain BCR-seq
435 data longitudinally and in additional cohorts. We were not able to compare B cell activation in

436 lung versus blood, which is important for understanding the role of adaptive immunity in CS,
437 vaping, and COPD pathogenesis. Single-cell and spatial transcriptomic or proteomic data would
438 provide greater resolution of the adaptive immune responses to CS and vaping. T cell receptor
439 sequencing that coincides with BCR-seq would provide a more comprehensive view of adaptive
440 immune responses in this context as well. A larger sample size would be desirable to examine
441 COPD-related outcomes, particularly longitudinal outcomes such as FEV₁ decline, mortality, and
442 exacerbations.

443 In conclusion, we observed that CS and vaping each enhance B cell activation, and that dual-
444 users show a trend towards greater effects than either alone. Self-identified race was strongly
445 associated with IgG isotype usage. These findings highlight associations between B cell
446 activation and antibody transcription, suggesting potential differences in immune and vaccine
447 responses linked to CS, vaping, and self-identified race.

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459 **References**

460

- 461 1. Erhabor, J. *et al.* E-Cigarette Use Among US Adults in the 2021 Behavioral Risk Factor
462 Surveillance System Survey. *JAMA Netw Open* **6**, e2340859 (2023).
- 463 2. Park, J.-A., Crotty Alexander, L. E. & Christiani, D. C. Vaping and Lung Inflammation and
464 Injury. *Annu Rev Physiol* **84**, 611–629 (2022).
- 465 3. Chaumont, M. *et al.* Fourth generation e-cigarette vaping induces transient lung
466 inflammation and gas exchange disturbances: results from two randomized clinical trials. *Am*
467 *J Physiol Lung Cell Mol Physiol* **316**, L705–L719 (2019).
- 468 4. Moshensky, A. *et al.* Effects of mango and mint pod-based e-cigarette aerosol inhalation on
469 inflammatory states of the brain, lung, heart, and colon in mice. *Elife* **11**, e67621 (2022).
- 470 5. Sayed, I. M. *et al.* Inflammatory phenotype modulation in the respiratory tract and systemic
471 circulation of e-cigarette users: a pilot study. *Am J Physiol Lung Cell Mol Physiol* **321**,
472 L1134–L1146 (2021).
- 473 6. Sharma, A. *et al.* E-cigarettes compromise the gut barrier and trigger inflammation. *iScience*
474 **24**, 102035 (2021).
- 475 7. Qiu, F. *et al.* Impacts of cigarette smoking on immune responsiveness: Up and down or
476 upside down? *Oncotarget* **8**, 268–284 (2016).
- 477 8. Vollmers, C., Sit, R. V., Weinstein, J. A., Dekker, C. L. & Quake, S. R. Genetic measurement
478 of memory B-cell recall using antibody repertoire sequencing. *Proc Natl Acad Sci U S A* **110**,
479 13463–13468 (2013).
- 480 9. Regan, E. A. *et al.* Genetic epidemiology of COPD (COPDGene) study design. *COPD* **7**, 32–
481 43 (2010).

- 482 10. Rojas-Quintero, J. *et al.* Spatial Transcriptomics Resolve an Emphysema-Specific Lymphoid
483 Follicle B Cell Signature in Chronic Obstructive Pulmonary Disease. *Am J Respir Crit Care*
484 *Med* **209**, 48–58 (2024).
- 485 11. Hogg, J. C. *et al.* The nature of small airway obstruction in chronic obstructive pulmonary
486 disease. *N Engl J Med* **350**, 2645–2653 (2004).
- 487 12. Faner, R. *et al.* Network Analysis of Lung Transcriptomics Reveals a Distinct B-Cell
488 Signature in Emphysema. *American journal of respiratory and critical care medicine* **193**,
489 1242–53 (2016).
- 490 13. Wan, E. S. *et al.* Epidemiology, genetics, and subtyping of preserved ratio impaired
491 spirometry (PRISm) in COPDGene. *Respiratory research* **15**, 89 (2014).
- 492 14. Han, M. K. *et al.* Chronic Obstructive Pulmonary Disease Exacerbations in the COPDGene
493 Study: Associated Radiologic Phenotypes. *Radiology* **261**, 274–282 (2011).
- 494 15. Parker, M. M. *et al.* Correction to: RNA sequencing identifies novel non-coding RNA and
495 exon-specific effects associated with cigarette smoking. *BMC medical genomics* **12**, 166
496 (2019).
- 497 16. Alberdi, A. & Gilbert, M. T. P. A guide to the application of Hill numbers to DNA-based
498 diversity analyses. *Mol Ecol Resour* **19**, 804–817 (2019).
- 499 17. Benjamini, Y. & Hochberg, Y. Controlling the False Discovery Rate: A Practical and
500 Powerful Approach to Multiple Testing. *Journal of the Royal Statistical Society. Series B*
501 *(Methodological)* **57**, 289–300 (1995).
- 502 18. Butler, D. C., Petterson, S., Phillips, R. L. & Bazemore, A. W. Measures of social deprivation
503 that predict health care access and need within a rational area of primary care service
504 delivery. *Health Serv Res* **48**, 539–559 (2013).

- 505 19. Tesfaigzi, Y. *et al.* Does Chronic Obstructive Pulmonary Disease Originate from Different
506 Cell Types? *Am J Respir Cell Mol Biol* **69**, 500–507 (2023).
- 507 20. Tarbiah, N., Todd, I., Tighe, P. J. & Fairclough, L. C. Cigarette smoking differentially affects
508 immunoglobulin class levels in serum and saliva: An investigation and review. *Basic Clin*
509 *Pharmacol Toxicol* **125**, 474–483 (2019).
- 510 21. Brusselle, G. G., Joos, G. F. & Bracke, K. R. New insights into the immunology of chronic
511 obstructive pulmonary disease. *Lancet* **378**, 1015–1026 (2011).
- 512 22. Brandsma, C.-A. *et al.* Increased levels of (class switched) memory B cells in peripheral
513 blood of current smokers. *Respir Res* **10**, 108 (2009).
- 514 23. Kotoulas, S.-C. *et al.* Acute effects of e-cigarette vaping on pulmonary function and airway
515 inflammation in healthy individuals and in patients with asthma. *Respirology* **25**, 1037–1045
516 (2020).
- 517 24. Luo, Y. & Stent, S. Smoking’s lasting effect on the immune system. *Nature* **626**, 724–725
518 (2024).
- 519 25. Peng, G. *et al.* Nicotine dose-dependent epigenomic-wide DNA methylation changes in the
520 mice with long-term electronic cigarette exposure. *Am J Cancer Res* **12**, 3679–3692 (2022).
- 521 26. McAlinden, K. D., Eapen, M. S., Lu, W., Sharma, P. & Sohal, S. S. The rise of electronic
522 nicotine delivery systems and the emergence of electronic-cigarette-driven disease. *Am J*
523 *Physiol Lung Cell Mol Physiol* **319**, L585–L595 (2020).
- 524 27. Crotty Alexander, L. E. *et al.* E-Cigarette or Vaping Product Use-associated Lung Injury:
525 Developing a Research Agenda. An NIH Workshop Report. *Am J Respir Crit Care Med* **202**,
526 795–802 (2020).

- 527 28. Polverino, F., Seys, L. J. M., Bracke, K. R. & Owen, C. A. B cells in chronic obstructive
528 pulmonary disease: moving to center stage. *Am J Physiol Lung Cell Mol Physiol* **311**, L687–
529 L695 (2016).
- 530 29. Sullivan, J.-L. *et al.* B Cell-Adaptive Immune Profile in Emphysema-Predominant Chronic
531 Obstructive Pulmonary Disease. *Am J Respir Crit Care Med* **200**, 1434–1439 (2019).
- 532 30. Syeda, M. Z., Hong, T., Huang, C., Huang, W. & Mu, Q. B cell memory: from generation to
533 reactivation: a multipronged defense wall against pathogens. *Cell Death Discov* **10**, 117
534 (2024).
- 535 31. Halper-Stromberg, E. *et al.* Systemic Markers of Adaptive and Innate Immunity Are
536 Associated with Chronic Obstructive Pulmonary Disease Severity and Spirometric Disease
537 Progression. *Am J Respir Cell Mol Biol* **58**, 500–509 (2018).
- 538 32. Agustí, A. *et al.* Persistent systemic inflammation is associated with poor clinical outcomes
539 in COPD: a novel phenotype. *PLoS One* **7**, e37483 (2012).
- 540 33. Regan, E. A. *et al.* Omics and the Search for Blood Biomarkers in Chronic Obstructive
541 Pulmonary Disease. Insights from COPDGene. *Am J Respir Cell Mol Biol* **61**, 143–149
542 (2019).
- 543 34. Abu-Shmais, A. A. *et al.* Potent HPIV3-neutralizing IGHV5-51 Antibodies Identified from
544 Multiple Individuals Show L Chain and CDRH3 Promiscuity. *J Immunol* **212**, 1450–1456
545 (2024).
- 546 35. Bunce, C. M. & Drayson, M. T. Dissecting racial disparities in multiple myeloma-clues from
547 differential immunoglobulin levels. *Blood Cancer J* **10**, 44 (2020).
- 548 36. Bhutani, M. *et al.* Addressing the disparities: the approach to the African American patient
549 with multiple myeloma. *Blood Cancer J* **13**, 189 (2023).

550 **Figure legends**

551

552 **Figure 1: Schematic of study design.** COPDGene = Genetic Epidemiology of COPD study.

553 BCR = B cell receptor. COPD = chronic obstructive pulmonary disease.

554

555 **Figure 2. Associations of BCR Measures with Vaping and Cigarette Smoking.** Significant
556 associations between BCR measures and vaping, cigarette smoking, and dual-use from
557 multivariable models analyzing class switching, isotype expression and usage, V-segment usage,
558 and CDR3 length (in nucleotides) are shown in Panel A. Panel B shows IgA and IgM isotype
559 usage among participants engaged in current cigarette smoking, vaping, or dual-use with former
560 smokers included for comparison. Significance is assessed by t-tests. * $p \leq 0.05$, ** $p \leq 0.005$, ns
561 $p > 0.05$.

562

563 **Figure 3. Associations of BCR Measures with Self-Reported Race.** Significant associations
564 between BCR measures and self-reported AA or NHW participants from multivariable models
565 analyzing class switching, isotype expression and usage, V-segment usage, and CDR3 length (in
566 nucleotides) are shown in Panel A. Panel B shows IgG and IgM isotype usage by self-reported
567 race. Significance is assessed by t-tests. * $p \leq 0.05$, ** $p \leq 0.005$, ns $p > 0.05$.

568

COPDGene participants with
RNA-sequencing data + smoking/vaping
data (n=3,601)

Stratified random sampling

Perform
AIRR-sequencing on
subset (n=234)

Process data and obtain measures
Antibody isotype expression and usage
Class switching
CDR3 length
Clonal diversity
V-segment usage

Multivariable association testing with
smoking/vaping and COPD-related
outcomes

Interaction analysis

